

Cancer vaccines

from ex vivo production to in vivo vaccination



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I have no financial relationships to disclose

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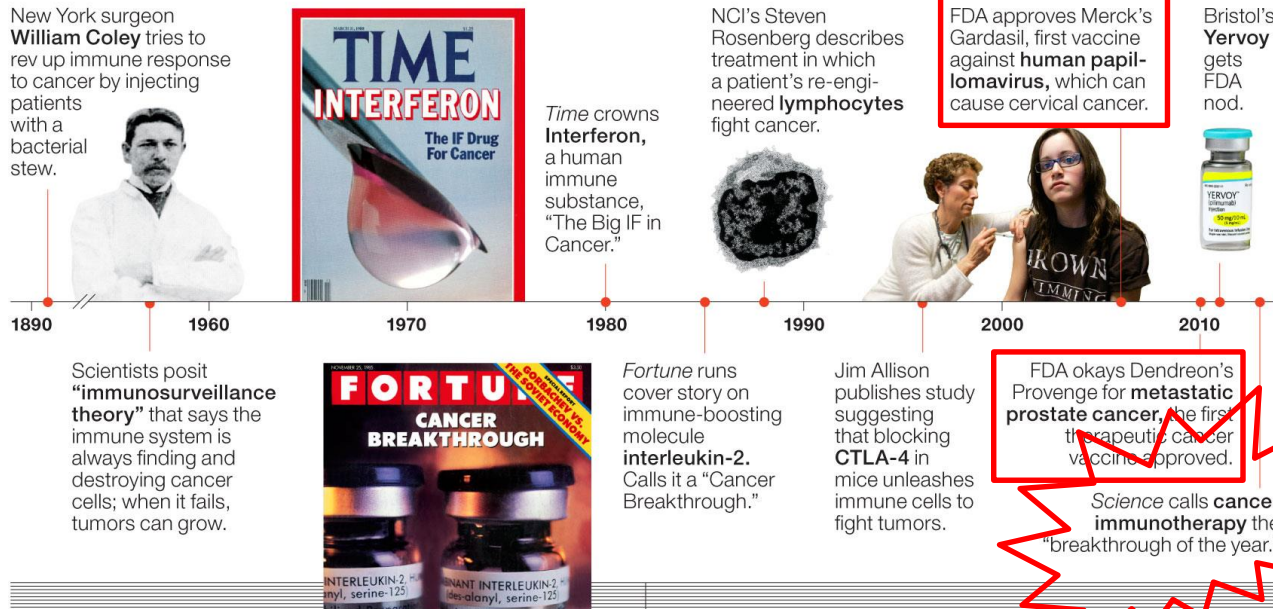
Learning goals

1. **Understanding the stumbling blocks and requirements for the design of successful therapeutic vaccines: choice of antigens and adjuvants**
2. **Becoming familiar with some of the current approaches to vaccination against cancer: broader definition includes *in vivo* vaccination**
3. **Acquiring an understanding of the positioning of cancer vaccines in the developing field of cancer immunotherapy**

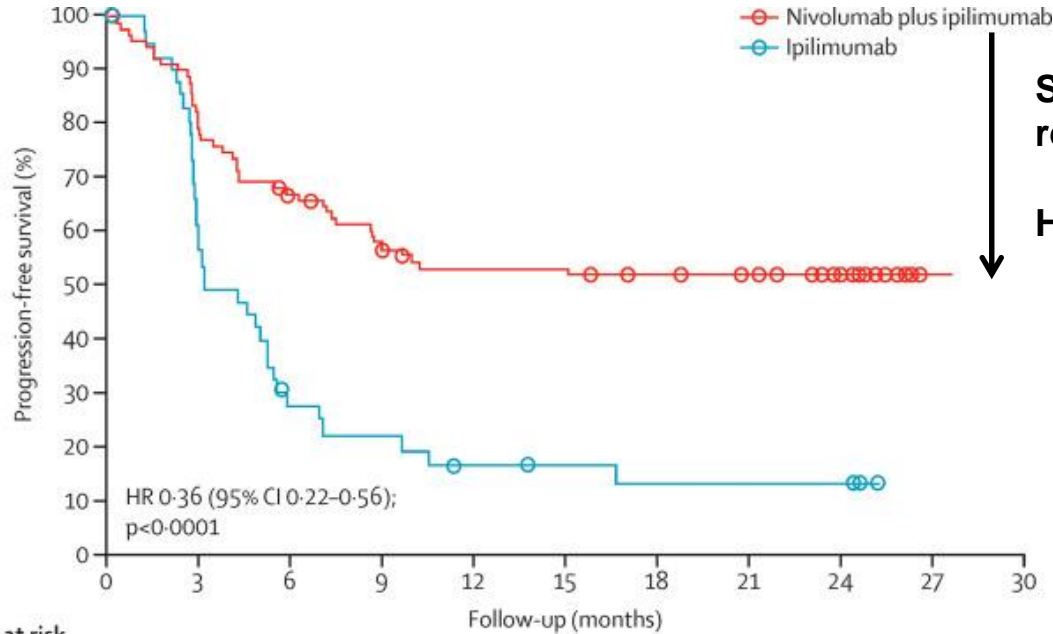
Cancer Immunotherapy: a bit of history

WAKING THE BODY'S DEFENDERS

For more than a century, researchers have tried to harness the human immune system to fight cancer. But high hopes, too often, have been followed by disappointment. Here, some milestones.

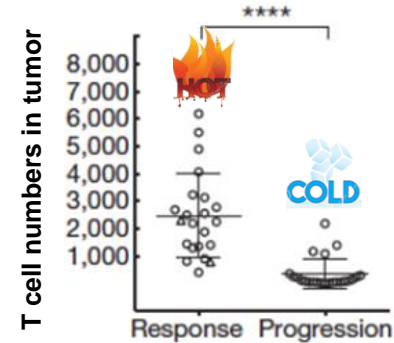


The promise of cancer immunotherapy: durable responses



Still...not everyone responds.

How come?



Number at risk (censored)	0	3	6	9	12	15	18	21	24	27	30
Nivolumab plus ipilimumab	95 (0)	69 (5)	58 (7)	47 (9)	43 (10)	43 (10)	40 (12)	38 (14)	24 (28)	2 (50)	0 (52)
Ipilimumab	47 (0)	22 (6)	10 (7)	8 (7)	5 (8)	4 (9)	3 (9)	3 (9)	3 (9)	0 (12)	0 (12)

T cell infiltration: role for vaccination!

Cancer vaccination: the basics (definitely not passive!)

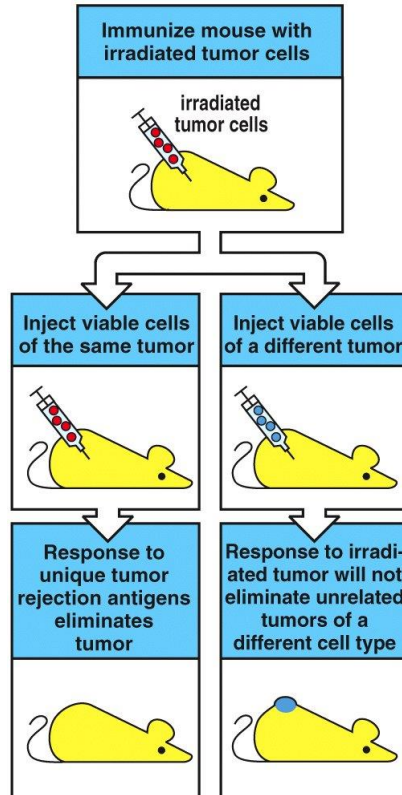


Figure 14-10 Immunobiology, 6/e. (© Garland Science 2005)

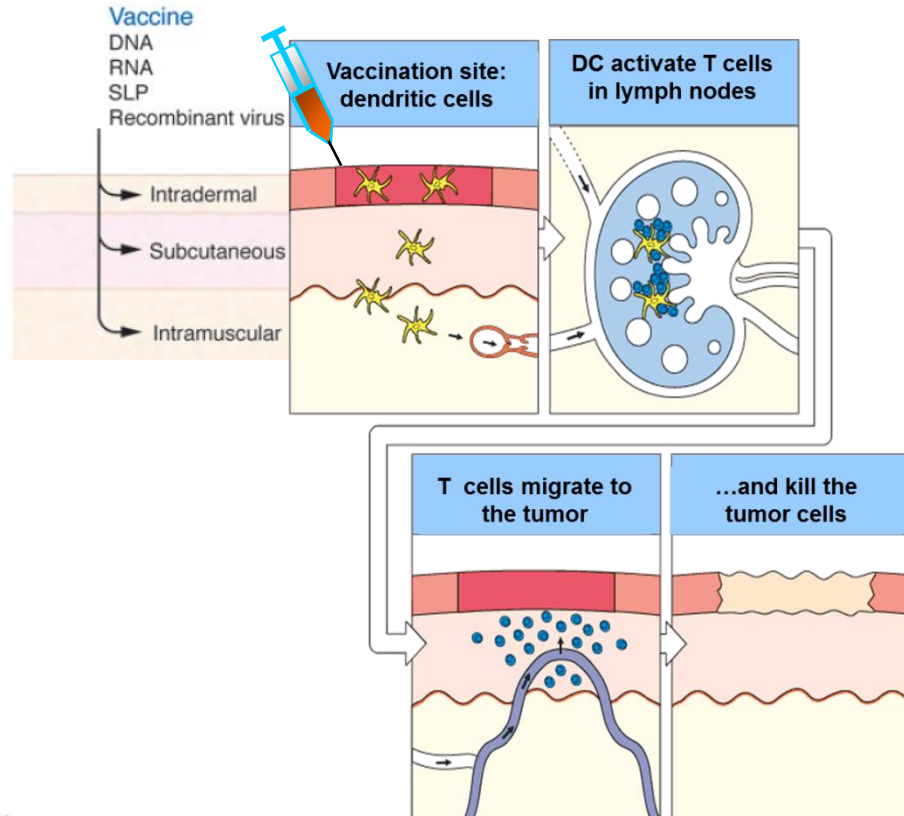
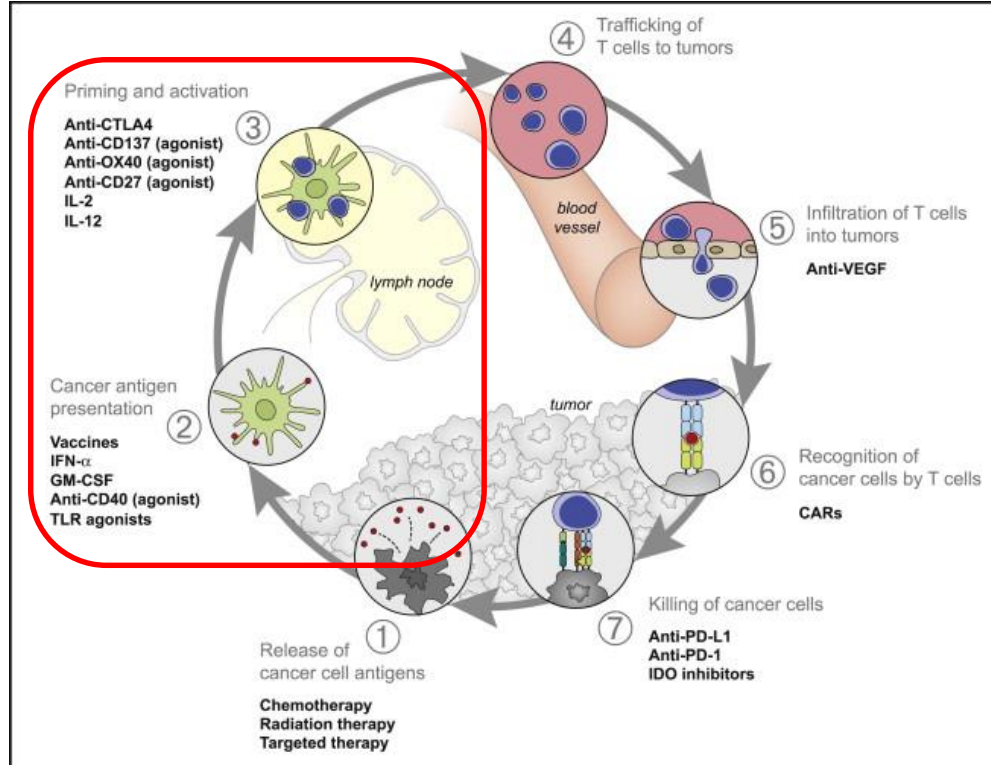


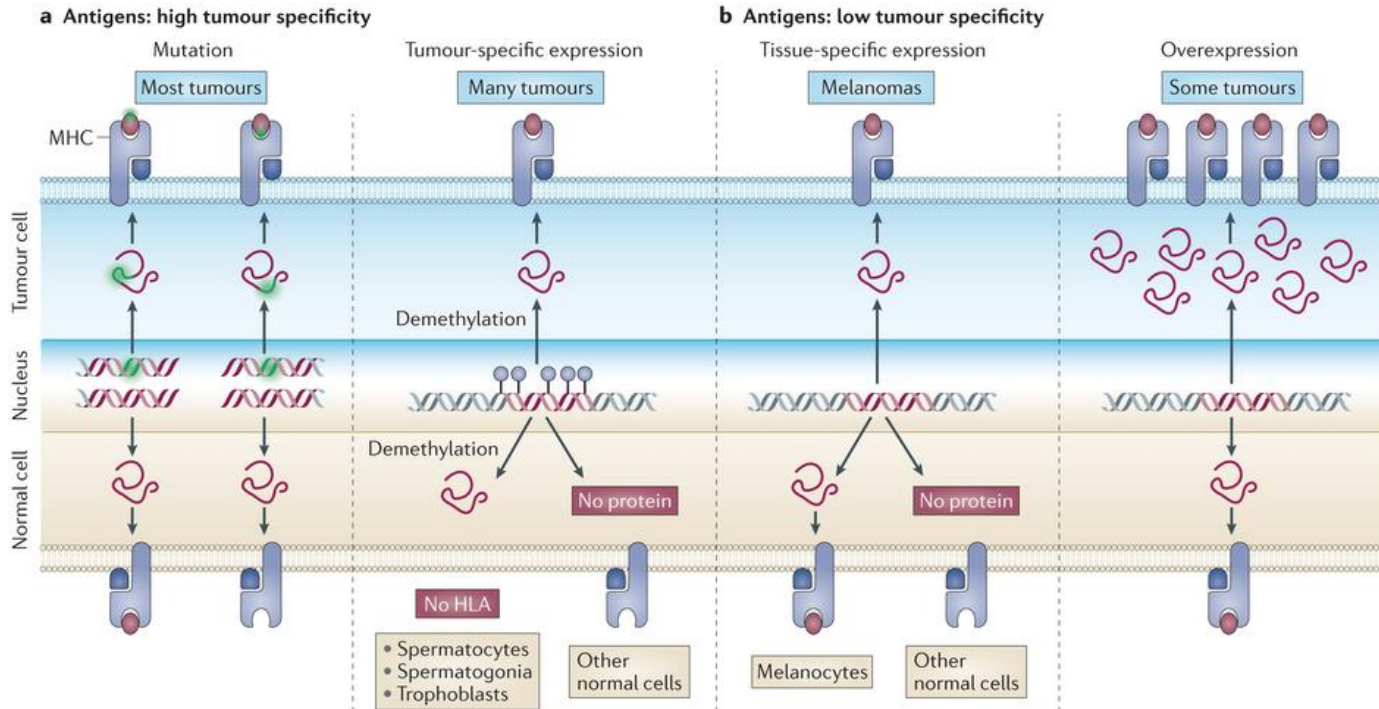
Fig 13.26 © 2001 Garland Science

Cancer Immunotherapy: therapeutic windows

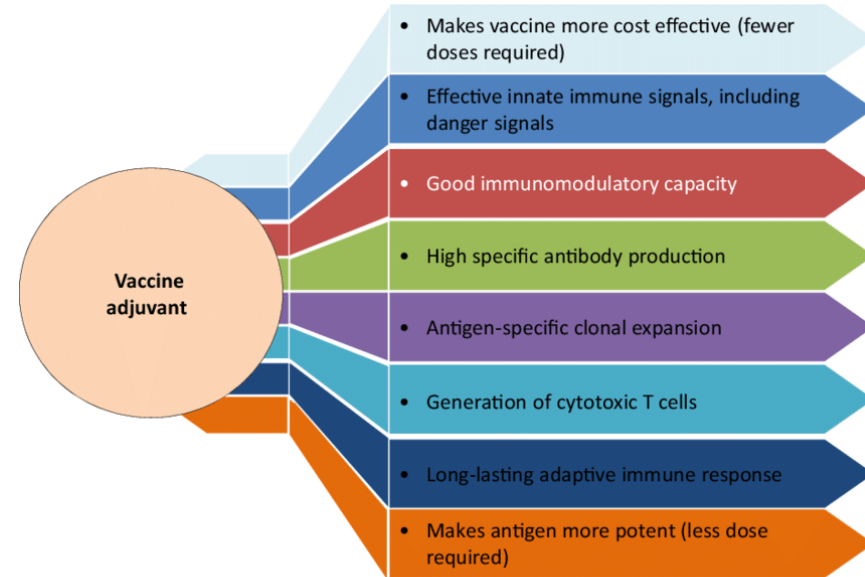
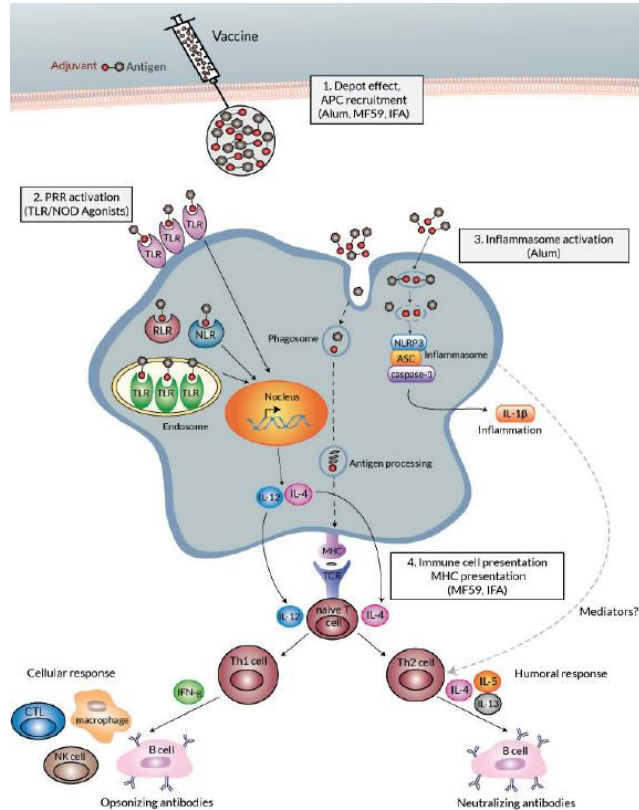
vaccination



Vaccine=Antigen+Adjuvant



Vaccine=Antigen+Adjuvant



Trends in Pharmacological Sciences

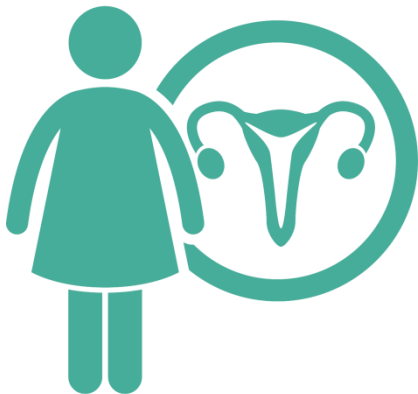


Successful (cancer) vaccines: prophylactic and antibody based



Doug Lowy & John Schiller

**Prophylactic
and active:**





BBC NEWS UK EDITION

Cervical cancer jab 'in a year'

A vaccine shown to be 100% effective against two virus strains that cause most cervical cancer could be available within a year, say manufacturers.

Gardasil worked against the sexually transmitted human papillomavirus (HPV). Some 12,167 women aged 16 to 23 from 13 countries, including the UK, took part in the drug company study.

Researchers believe a vaccine could work best if given before adolescence, but critics fear this could encourage under-age sex.

Merck's vaccine is in head-to-head competition with a rival from UK-based GlaxoSmithKline called Cervarix.

The two-year Future II trial found Gardasil was **100% effective at preventing early stage cancers and pre-cancerous abnormalities** caused by the two key strains of HPV - the 16 and 18 strains - which cause 70% of cervical cancers.




Friday, 7 October 2005, 04:31 GMT 05:31 UK



More challenging cancer vaccines: therapeutic and T cell based

**Therapeutic
and active:**

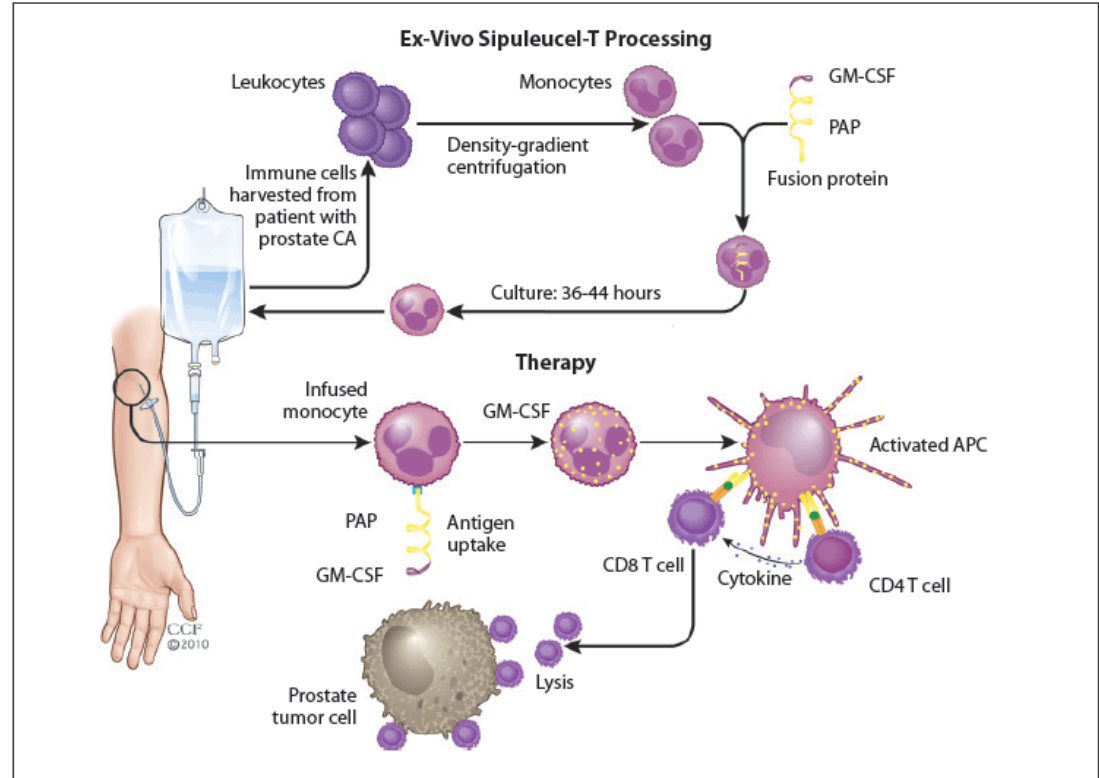
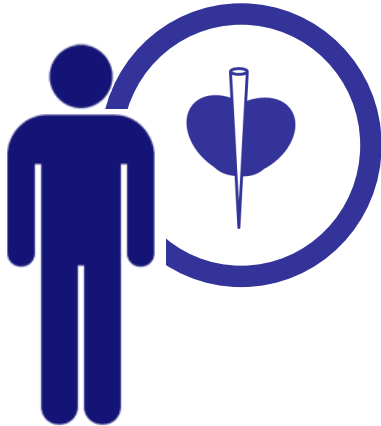
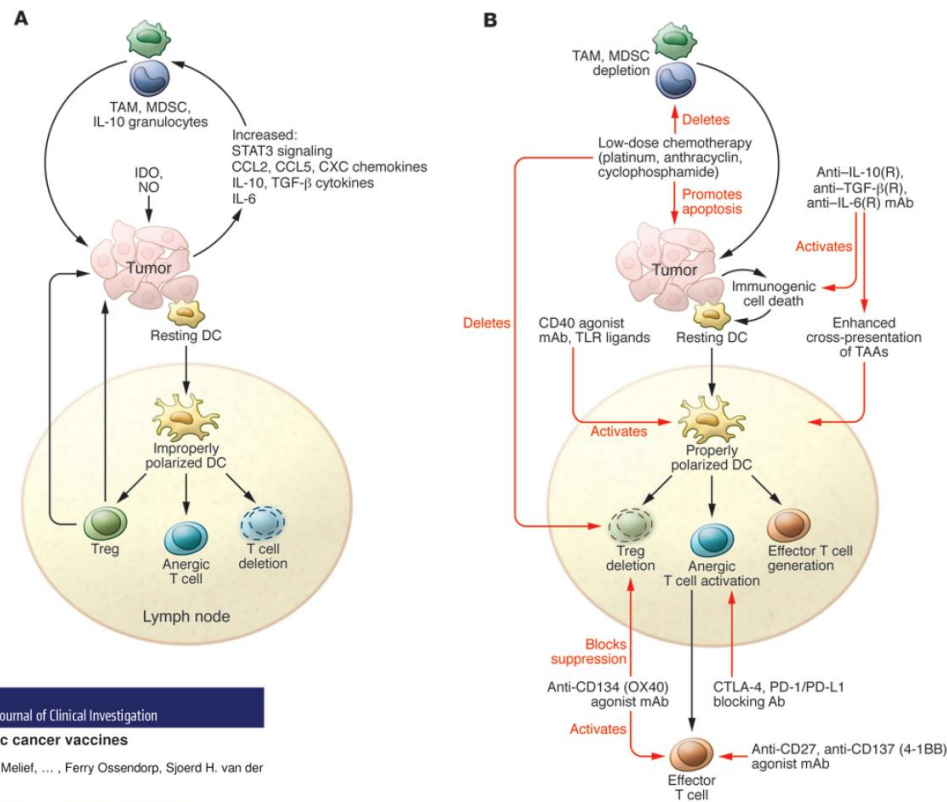


Table 5 Results of clinical vaccine studies in patients with metastatic cancers

Vaccine type	Reference	Cancer type	Vaccine	Total patients	Patients responding
Peptide	43	Melanoma	Tyrosinase + GMCSF	16	0
	44	Melanoma	Peptides in IFA or on DC	26	3
	45	Melanoma	MART-1 + IL-12	28	2
	46	Prostate	Peptides	10	0
	47	Melanoma	Peptides on PBMC + IL-12	20	2
	48	Breast and prostate	Telomerase	7	0
	49	Cervix	HPV16 E7	17	0
	50	Colorectal	Peptides in IFA	10	0
	51	Multiple	NY-ESO-1	12	0
	52	Multiple	Ras in DETOX adjuvant	15	0
Virus	53	Multiple	Peptides in IFA	14	0
	29	Prostate	Vaccinia-PSA	33	0
	54	Prostate	Vaccinia-PSA	42	0
	55	Colorectal	Vaccinia-CEA	20	0
	56	Colorectal	Vaccinia-CEA and B7-1	18	0
	57	Multiple	Avipox-CEA (IGMCSF)	60	0
	58	Multiple	Avipox-CEA	15	0
	59	Multiple	Vaccinia + avipox-CEA	18	0
Tumor cells	60	Melanoma	Transduced with GM-CSF	26	1
	61	Melanoma	Membranes on silicone beads	17	1
	62	Lung	Transduced with GMCSF	26	1
	63	Lung	Transduced with GMCSF	43	3
	64	Breast	Transduced with B7-1	30	0
Dendritic cells	65	Melanoma	Pulsed with peptides	17	0
	66	Melanoma	Pulsed with peptides or lysates	33	3
	67	Melanoma	Pulsed with peptides or lysates	16	5
	68	Melanoma	Pulsed with peptides	24	1
	22	Melanoma	Pulsed with MAGE-3A1 peptide	11	0
	69	Childhood cancers	Pulsed with lysates	15	1
	70	Kidney	Transfected with RNA	15	0
	71	Colorectal	Pulsed with CEA peptides	12	1
	72	Kidney	Pulsed with tumor lysates	35	3
	23	Multiple	Pulsed with tumor lysates	20	0
Heat shock protein	73	Melanoma	Hsp-96	28	2
	74	Multiple	Hsp-96	16	0
		Total		765	29

Objective response rate = 3.8%

The trouble with therapeutic cancer vaccines... Rosenberg *et al* Nat Med 2004





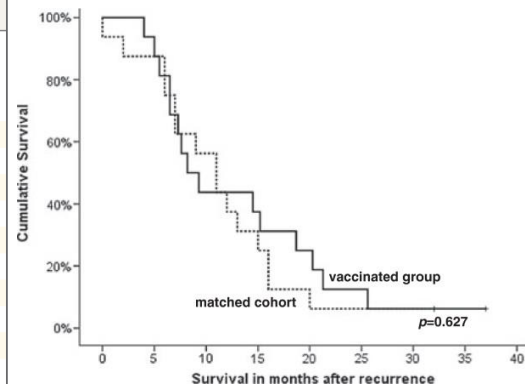
One solution: go early

HPV16 SLP vaccine in VIN: 16/20 PR/CR

...but in cancer: none!

Table 3. Clinical Results at 3, 12, and 24 Months after the Last Vaccination.*

Patient No.	No. of Vaccinations	At 3 Months				At 12 Mo		At 24 Mo
		Symptoms	Lesion Response	Histologic Findings	Type of HPV Infection	Symptoms	Lesion Response	Lesion Response
1	4	Mild to moderate	Partial	VIN 2	16	Mild to moderate	Partial	Partial†
2	4	Severe	None	VIN 3	16		Carcinoma	
3	4	Severe	None	VIN 3	16	None	Partial	Partial‡
6	4	None	Complete	Normal	16	None	Complete	Complete
7	4	None	Complete	Normal	None	None	Complete	Complete
8	4	Mild to moderate	Complete	Normal	6b	None	Complete§	Complete
9	3	None	Complete	Normal	None	None	Complete	Complete
10	4	None	Partial	VIN 3	16	Lost to follow-up¶		
11	4	None	None	VIN 3	16	None	Complete	Complete
12	4	Mild to moderate	None	VIN 3	16	Mild to moderate	Partial	None
13	4	Mild to moderate	Partial	VIN 3	16	Mild to moderate	Partial	Partial
16	4	Mild to moderate	Partial	VIN 1	16	Mild to moderate	Complete	Complete
18	4	Severe	None	VIN 3	16	Severe	None	None
22	4	Mild to moderate	None	VIN 3	16	Severe	Partial	Partial
23	4	Mild to moderate	Partial	VIN 2	16	None	Partial	Microinvasive carcinoma**
26	4	None	None	VIN 3	16	None	None	None
27	3	None	Partial	VIN 3	16	None	Complete	Complete
28	4	None	None	VIN 3	16	None	None	None
29	4	None	Complete	Normal	None	None	Complete	Complete
30	4	Mild to moderate	Partial	VIN 2	16	None	Complete	Complete



van Poelgeest et al. *Journal of Translational Medicine* 2013, 11:88
<http://www.translational-medicine.com/content/11/1/88>



RESEARCH

Open Access

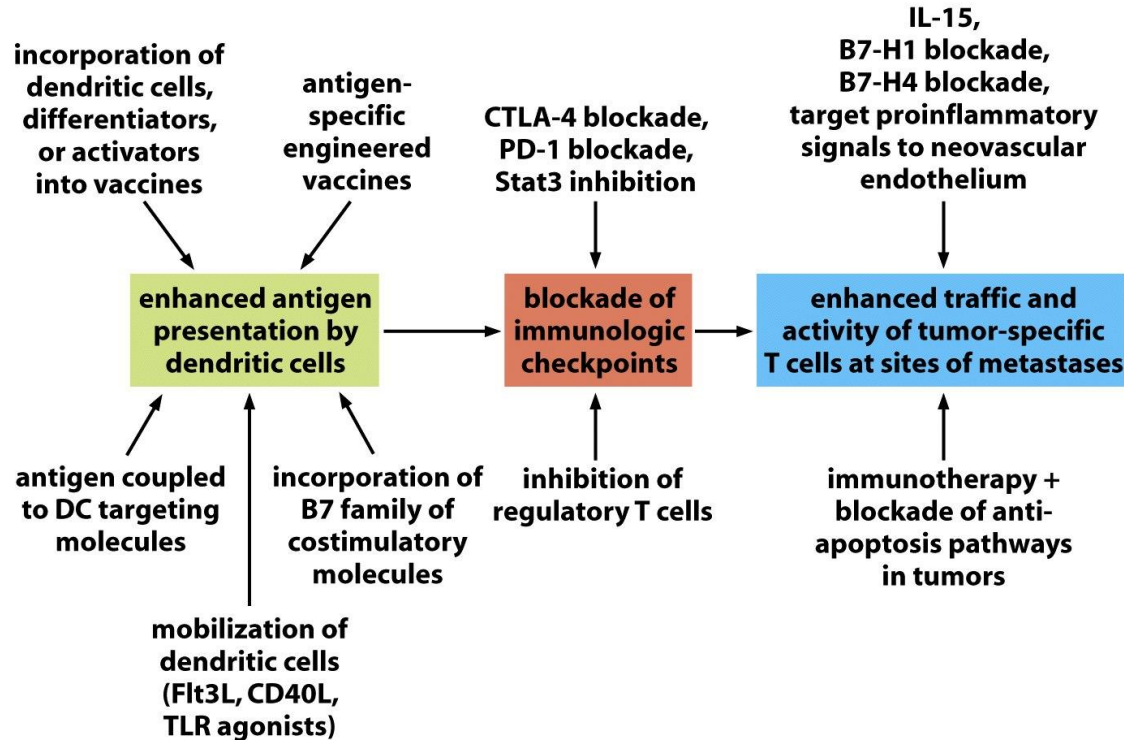
HPV16 synthetic long peptide (HPV16-SLP) vaccination therapy of patients with advanced or recurrent HPV16-induced gynecological carcinoma, a phase II trial

Mariette I E van Poelgeest^{1†}, Marij J P Welters^{2†}, Edith M G van Esch³, Linda F M Stynenbosch⁴, Gjo Kerpeshoek⁵, Els L van Persijn van Meerten⁶, Muriel van den Hende⁷, Margriet J G Löwik⁸, Dorien M A Berends-van der Meer⁹, Lorraine M Fathers¹⁰, A Rob P M Valentijn¹¹, Jaap Oostendorp¹², Gert Jan Fleuren¹³, Cornelis J M Melief¹⁴, Gemma G Kenter¹⁵ and Sjoerd H van der Burg¹⁶

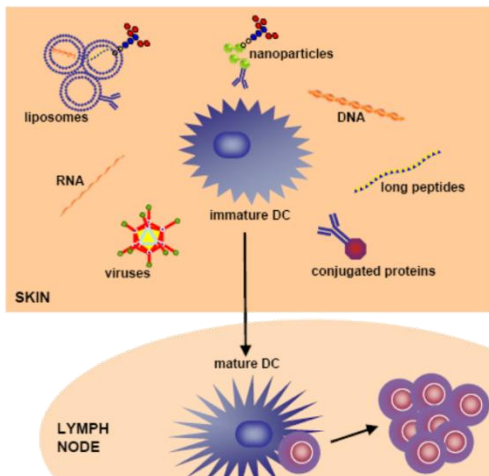
Vaccination against HPV-16 Oncoproteins for Vulvar Intraepithelial Neoplasia

Gemma G. Kenter, M.D., Ph.D.,¹ Marij J. P. Welters, Ph.D.,²
 A. Rob P. M. Valentijn, Ph.D.,³ Margriet J. G. Löwik,⁴
 Dorien M. A. Berends-van der Meer, Annelies P. G. Vloot, Farah Essahsah,
 Lorraine M. Fathers, Rienk Offringa, Ph.D.,⁵ Jan Wouter Drijfhout, Ph.D.,⁶
 Amon R. Wafelman, Ph.D.,⁷ Jaap Oostendorp, Ph.D.,⁸ Gert Jan Fleuren, M.D., Ph.D.,⁹
 Sjoerd H. van der Burg, Ph.D., and Cornelis J. M. Melief, M.D., Ph.D.¹⁰

Another solution: optimizing vaccines and combination therapies



Cancer vaccine formulations and choice of adjuvant



Vaccine format	Advantages	Challenges
Synthetic peptides (45)	Cell-free manufacturing Automated synthesis established Proven clinical activity of long peptides Compatible with a wide range of formulations to improve delivery	Lack of clinical-grade manufacturability of a substantial portion of sequences High variability in the physicochemical properties of individual peptides, complicating manufacturing Irrelevant immune responses against artificial epitopes created by peptide degradation in the extracellular space
Messenger RNA (46)	Cell-free manufacturing Inherent adjuvant function via TLR7, TLR8, and TLR3 signaling Proven clinical activity Highly efficient systemic delivery into DCs established Transient activity and complete degradation All types of epitopes can be encoded	Fast extracellular degradation of mRNA if not protected by appropriate formulation Interpatient variability of TLR7-driven adjuvant activity
DNA plasmids (47)	Cell-free manufacturing Inherent adjuvant activity driven by TLR9 Cost-effective and straightforward manufacturing All types of epitopes can be encoded	Potential safety risks by insertional mutagenesis Successful transfection requires entry into nucleus, thereby limiting effective delivery of vaccines into DCs
Viral vectors (48) (adenoviral and vaccinia)	Strong immunostimulatory activity Extensive clinical experience with vector formats in the infectious disease field All types of epitopes can be encoded	Complex manufacturing Immune responses against components of the viral vector backbone, limiting successful in vivo vaccine delivery and efficacy
Engineered attenuated bacterial vectors (49) (<i>Salmonella</i> , <i>Listeria</i>)	Strong immunostimulatory activity Could be combined with plasmid DNA All types of epitopes can be encoded	Complex manufacturing and "sterility" testing Immune responses against bacterial components, limiting vaccine delivery and vaccine immunogenicity Potential safety risks due to delivery of live, replication-competent bacteria
Ex vivo antigen-loaded DCs (50)	Strong immunostimulatory activity Proven clinical efficacy of DC vaccines Can be loaded with various antigen formats	Higher costs and resources required for adoptive cell therapy approaches

Oosterhoff et al. in: T.J. Curiel (ed.), *Cancer Immunotherapy*, 2013

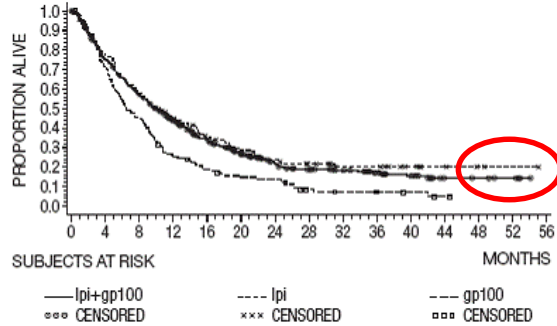
Science

Personalized vaccines for cancer immunotherapy

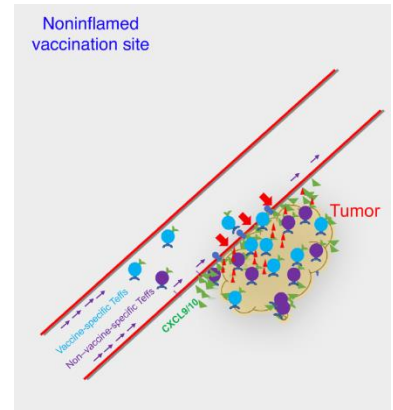
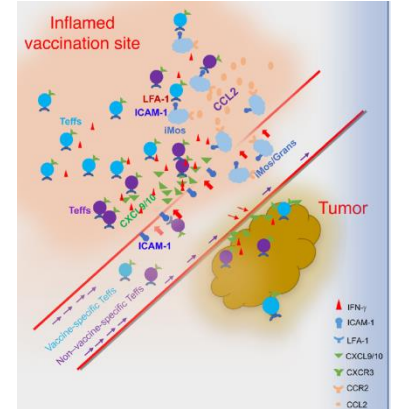
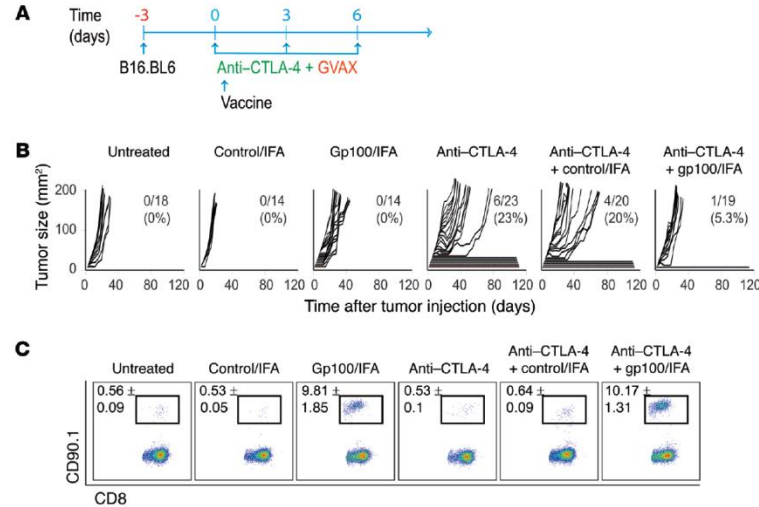
Ugur Sahin and Özlem Türeci

Science 359 (6382), 1355-1360.

Cancer vaccine formulations: avoid depot formation



3 weeks for four treatments. In the vaccine groups, patients received two modified HLA-A*0201-restricted peptides, injected subcutaneously as an emulsion with incomplete Freund's adjuvant (Montanide ISA-51): a gp100:209-217(210M) peptide, 1 mg injected in the right anterior thigh, and a gp100:280-288(288V) peptide, 1 mg injected in the left anterior thigh.



Improved Survival with Ipilimumab in Patients with Metastatic Melanoma

F. Stephen Hodi, M.D., Steven J. O'Day, M.D., David F. McDermott, M.D., Robert W. Weber, M.D., Jeffrey A. Sosman, M.D., John B. Haanen, M.D., Rene Gonzalez, M.D., Caroline Robert, M.D., Ph.D., Dirk Schadendorf, M.D., Jevica C. Hamel, M.D., Wallace Haskins, M.D., Allison M. van den Broek, M.D., Ph.D., Jemel Lubala, M.D., Paul Langer, M.D., Julia M. Veeke, M.D., Gerald P. Linette, M.D., Ph.D., David Figg, M.D., Christian H. Ottaviano, M.D., Ph.D., Collette Lobb, M.D., Christian Reusch, M.D., Ian Qian, M.D., Joseph J. Clark, M.D., Jedd D. Wolchok, M.D., Ph.D., Jeffrey S. Weber, M.D., Ph.D., Jason Tien, Ph.D., Michael J. Yellin, M.D., Geoffrey M. Nichol, M.B., Ch.B., Axel Haus, M.D., Ph.D., and Walter J. Ullrich, M.D., Ph.D.

JCI The Journal of Clinical Investigation

Cancer vaccine formulation dictates synergy with CTLA-4 and PD-L1 checkpoint blockade therapy

Yared Hailemichael, ... , Victor H. Engelhard, Willem W. Overwijk

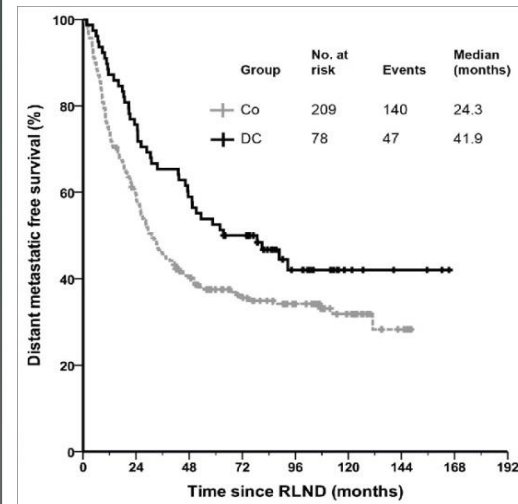
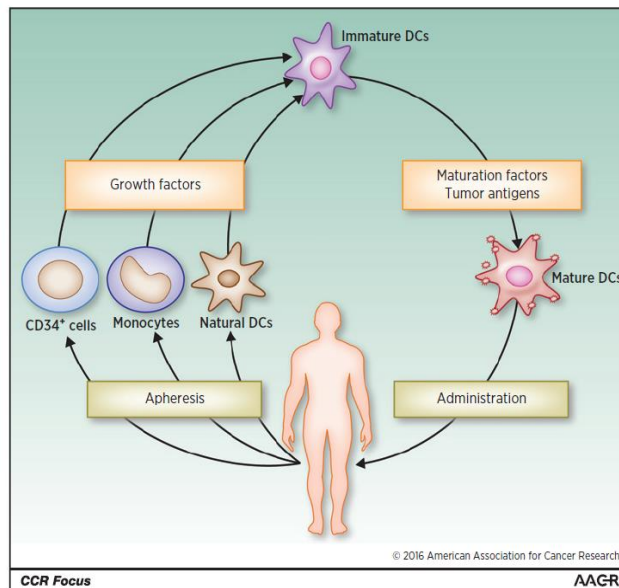
J Clin Invest. 2018;128(4):1338-1354. <https://doi.org/10.1172/JCI93303>.



Dendritic cell vaccines: classic approach



http://www.scharphoto.com/fine_art_prints/archives/000605.php



CCR FOCUS

Dendritic Cell-Based Immunotherapy: State of the Art and Beyond

Kalijn F. Bol^{1,2}, Gerty Schreibeit¹, Winald R. Gerritsen³, I. Jolanda M. de Vries^{1,2}, and Carl G. Figdor¹

Clin Cancer Res; 22(8) April 15, 2016

Journal of Clinical Oncology 33, #107673, January 2015; © 2015 Taylor & Francis Group, LLC

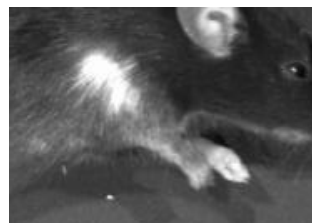
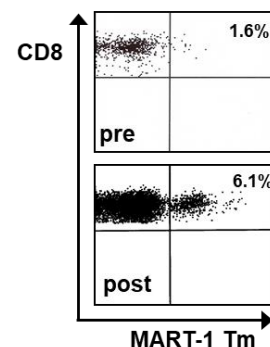
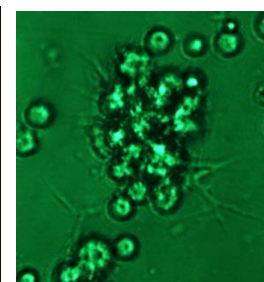
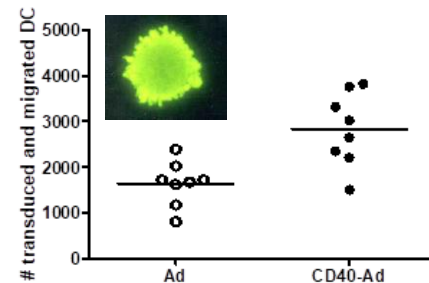
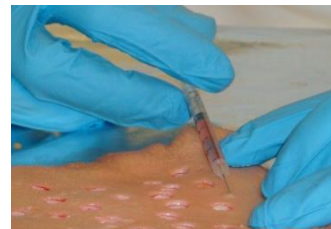
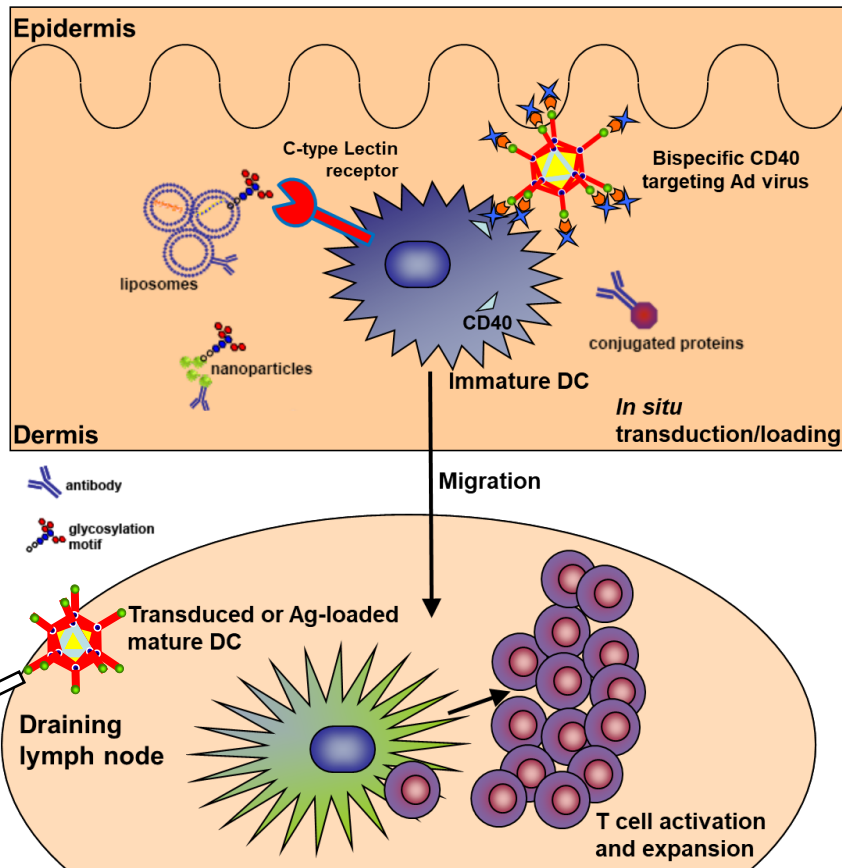
Favorable overall survival in stage III melanoma patients after adjuvant dendritic cell vaccination

Kalijn F. Bol^{1,2}, Erik H. J. G. Aarnouts^{1,2,3}, Florentien E. M. in 't Hout^{1,4}, Gerty Schreibeit¹, Jeroen H. A. Cnossen¹, W. Joost Lesterhuis^{1,2,3}, Winald R. Gerritsen³, Dirk J. Grunhagen⁵, Cornelis Verhoef⁶, Cornelis J. A. Punt⁷, Johannes J. Borenkamp⁸, Johannes H. W. de Wit⁹, Carl G. Figdor¹, and I. Jolanda M. de Vries^{1,2,4}

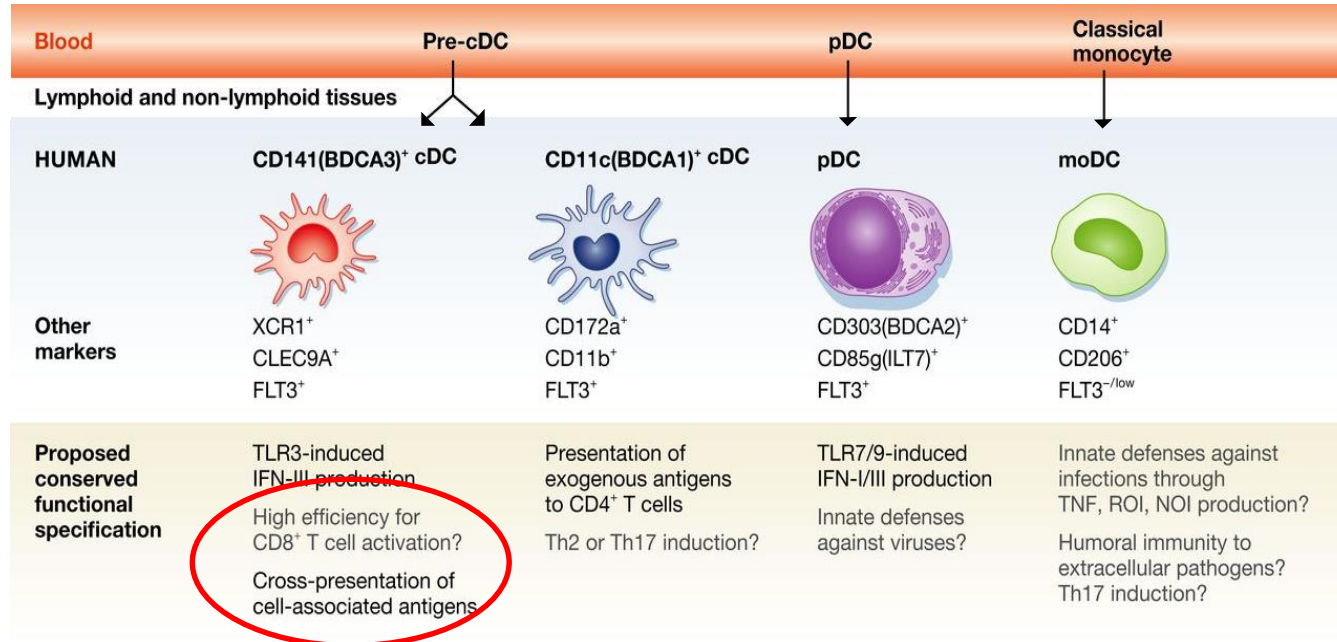
¹Department of Tumor Immunology, Radboud Institute for Molecular Life Sciences, Radboud University Medical Center, Nijmegen, The Netherlands; ²Department of Medical Oncology, Radboud University Medical Center, Nijmegen, The Netherlands; ³Department of Radiology and Nuclear Medicine, Radboud University Medical Center, Nijmegen, The Netherlands; ⁴Department of Surgical Oncology, Radboud University Medical Center, Nijmegen, The Netherlands; ⁵Department of Medicine and Pharmacology, University of Western Australia, Crawley, Australia; ⁶Department of Surgical Oncology, Erasmus MC Cancer Institute, Rotterdam, The Netherlands; ⁷Department of Medical Oncology, Academic Medical Center, Amsterdam, The Netherlands



Alternative DC vaccines: *in vivo* targeting

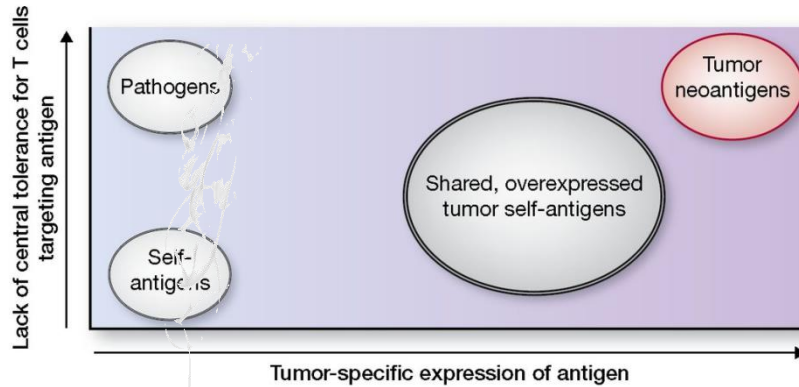


DC vaccines: what subset to target?



Choice of antigens: what do T cells react to in tumors?

Hacohen et al.
CIR 2013



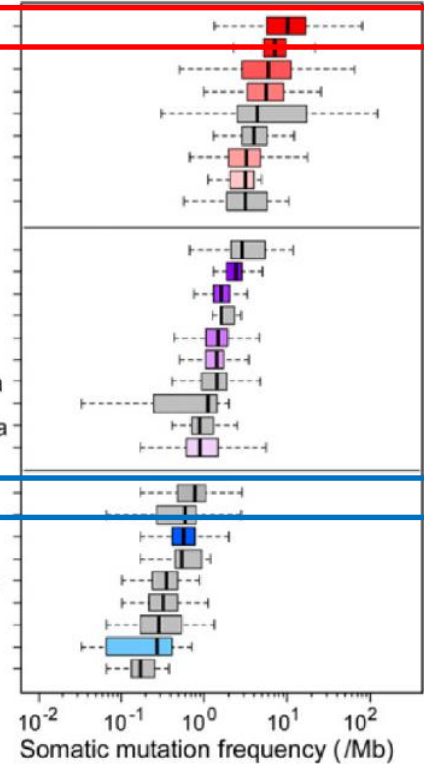
Melanoma

LUSC
LUAD
Bladder
Stomach
ESO AD
Head and neck
Colon
DLBCL

Cervical
Rectum
GBM
Papillary RCC
Ovarian
Clear cell RCC
Multiple myeloma
Pancreas
Low-grade-glioma
Breast

Prostate

Neuroblastoma
CLL
Carcinoid
Medulloblastoma
Thyroid
Ewing Sarcoma
AML
Rhabdoid tumor

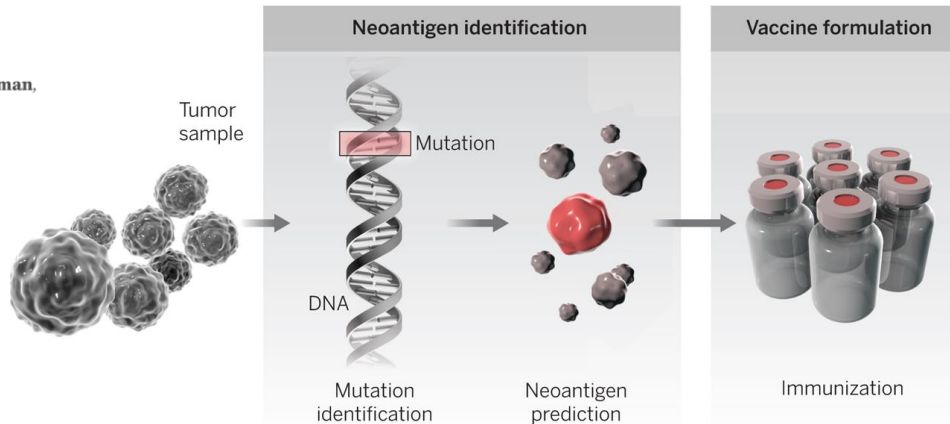


Classic vaccines with novel antigens: personalized!

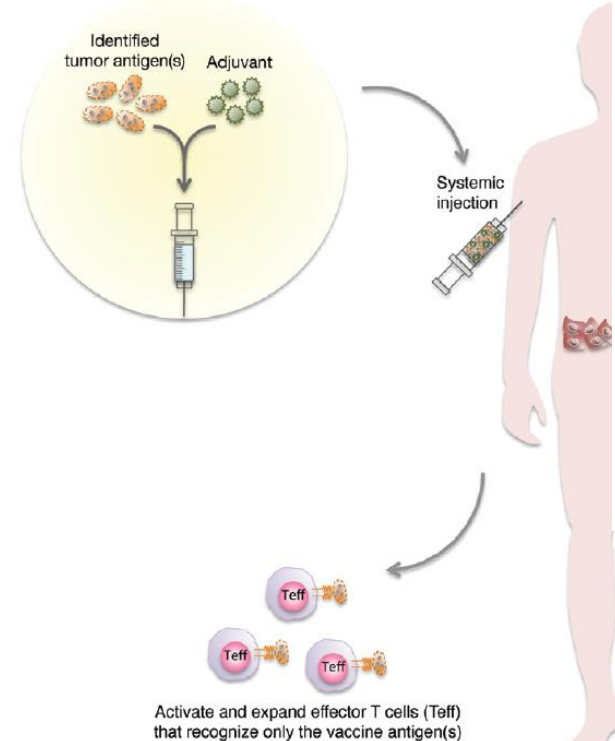
Neo approaches to cancer vaccines

A neoantigen-based vaccine elicits T cell responses in cancer patients

By Lélia Delamarre, Ira Mellman, Mahesh Yadav

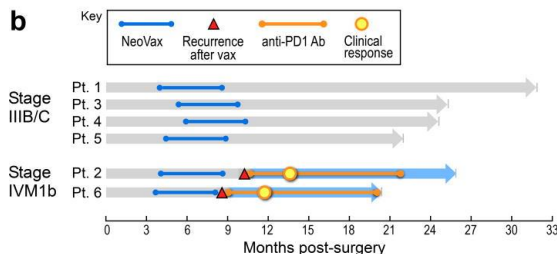
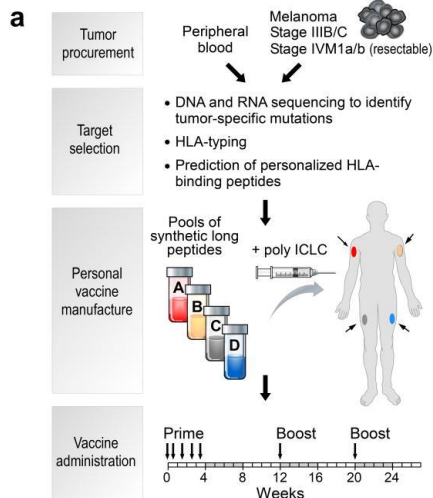


(a) Conventional vaccination





Neoantigen vaccines



LETTER

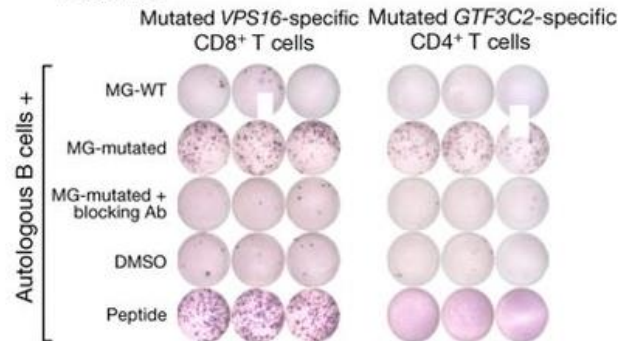
doi:10.1038/nature22991

An immunogenic personal neoantigen vaccine for patients with melanoma

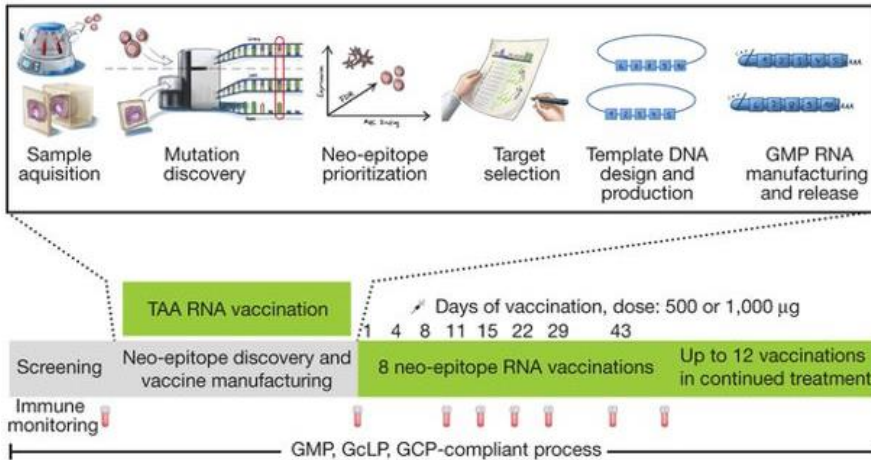
Patrick A. Ott^{1,2,3*}, Zhuting Hu^{1*}, Derin B. Keskin^{1,3,4}, Sachet A. Shukla^{1,4}, Jing Sun¹, David J. Bozym¹, Wandi Zhang¹, Adrienne Luoma⁵, Anita Giobbie-Hurder⁶, Lauren Peter^{7,8}, Christina Chen¹, Oriol Olive¹, Todd A. Carter⁴, Shuang Li⁴, David J. Lieb⁴, Thomas Eisenhaure⁴, Evisa Gjini⁹, Jonathan Stevens¹⁰, William J. Lane¹⁰, Indu Javeri¹¹, Kaliappanadar Nellaippan¹¹, Andres M. Salazar¹², Heather Daley¹, Michael Seaman⁷, Elizabeth I. Buchbinder^{1,2,3}, Charles H. Yoon^{1,13}, Maegan Harden⁴, Niall Lennon⁴, Stacey Gabriel⁴, Scott J. Rodig^{9,10}, Dan H. Barouch^{3,7,8}, Jon C. Aster^{3,10}, Gad Getz^{3,4,14}, Kai Wücherpfennig^{3,5}, Donna Neuberg⁶, Jerome Ritz^{1,2,3}, Eric S. Lander^{1,4}, Edward F. Fritsch^{1,4}, Nir Hacohen^{3,4,15} & Catherine J. Wu^{1,2,3,4}

13 JULY 2017 | VOL 547 | NATURE | 217

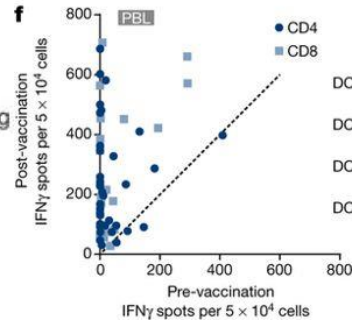
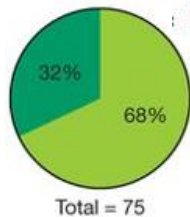
Patient 3



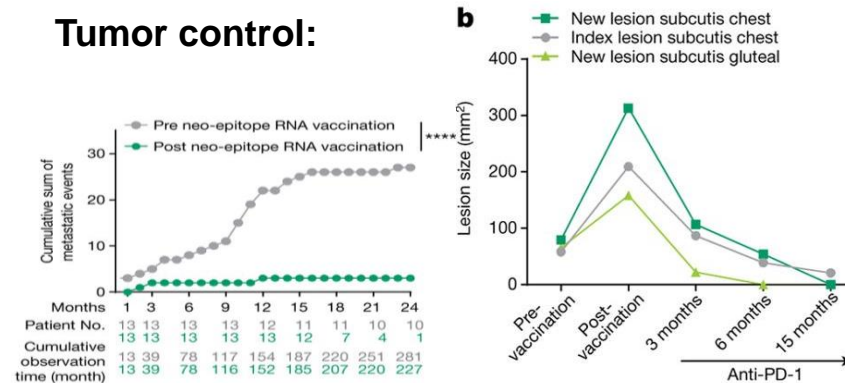
Neoantigen vaccines



Immune response:

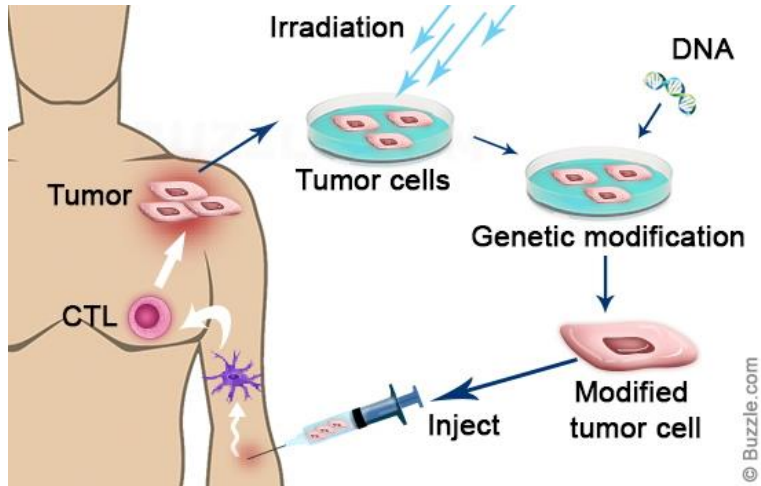


Tumor control:



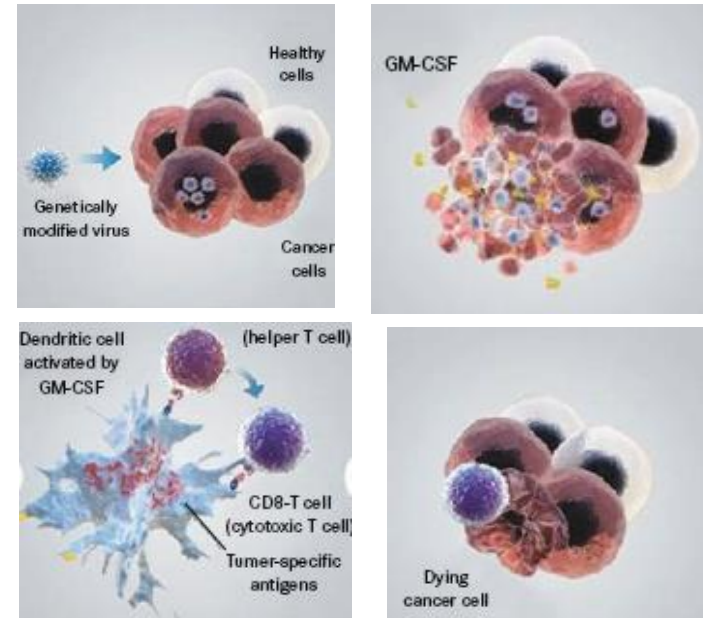
Antigen agnostic cancer vaccination approaches

1) Whole cancer cell vaccines



<https://healthhearty.com/strategies-for-cancer-vaccine-development>

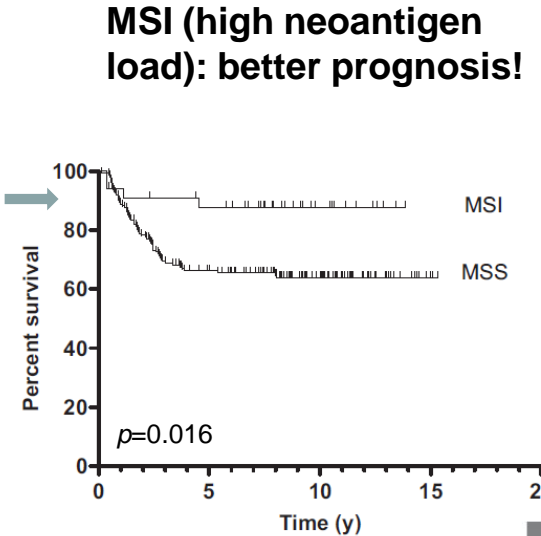
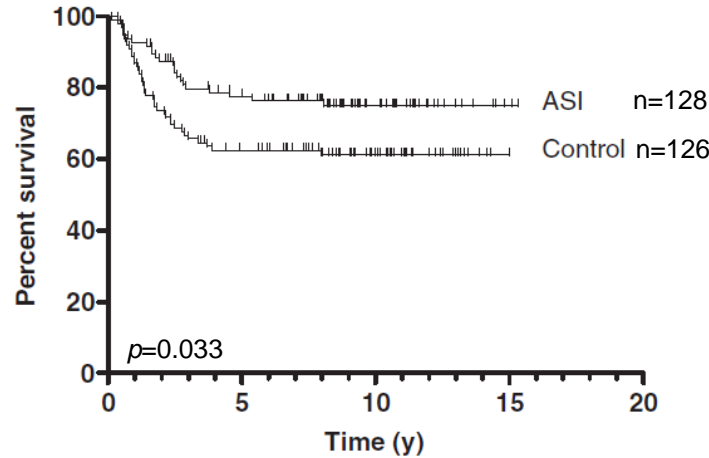
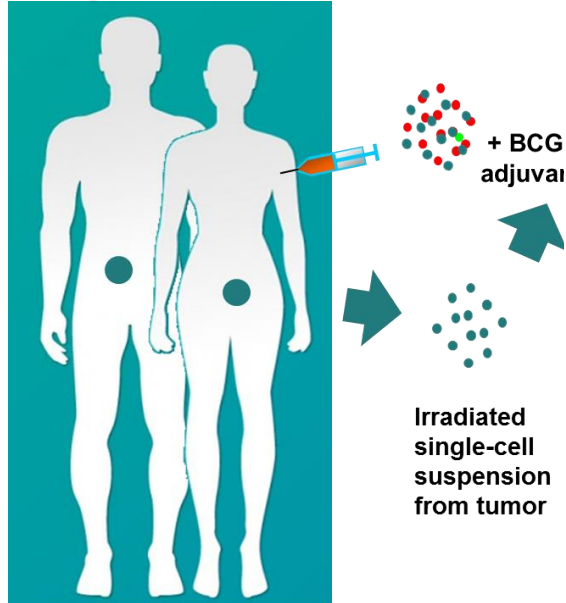
2) Oncolytic virotherapy



<https://www.gotoper.com/publications/ajho/2016/2016apr/an-update-on-talimogene-laherparepvec>



Whole Cancer cell vaccines: CRC vaccination –who benefits?



THE LANCET • Vol 353 • January 30, 1999

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Predictive Biomarkers and Personalized Medicine

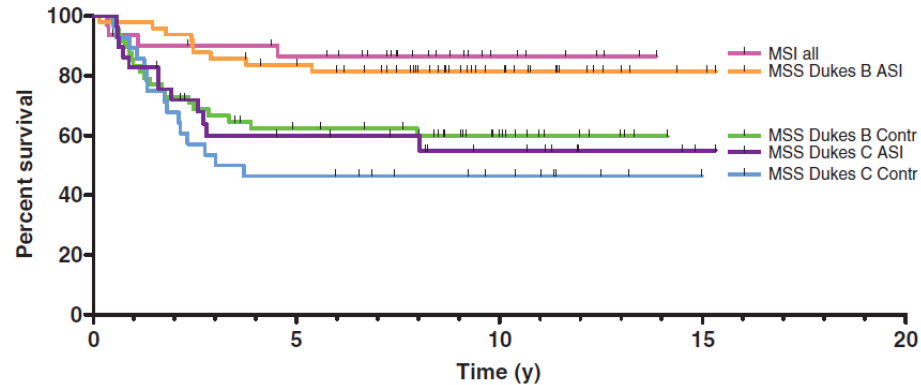
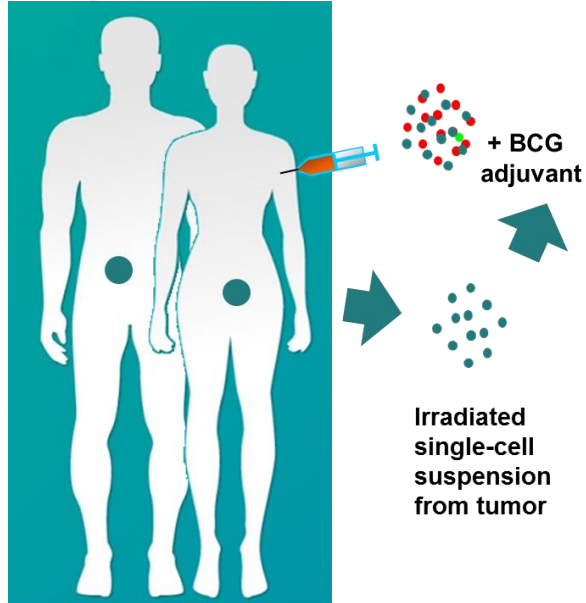
Clinical Effects of Adjuvant Active Specific Immunotherapy Differ between Patients with Microsatellite-Stable and Microsatellite-Unstable Colon Cancer

Vincent A. de Weger¹, Annelies W. Turkema², Quirinus J.M. Vooshuis¹, Zaida Ester¹, Herman Brij¹, Alfons J. van den Endevegh¹, Elisabeth Bloemena¹, Herbert M. Pinedo¹, Jan B. Vermorken¹, Harm van Tinteren¹, Gerrit A. Meijer¹, and Erik Hoogberg¹

Clin Cancer Res; 18(3) February 1, 2012



Whole Cancer cell vaccines: CRC vaccination –who benefits?



Only patients with MSS stage II (low neoantigen load!) benefit from vaccination

**Actionable tumor antigens!
Neoantigens?**

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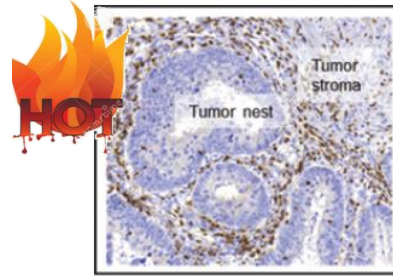
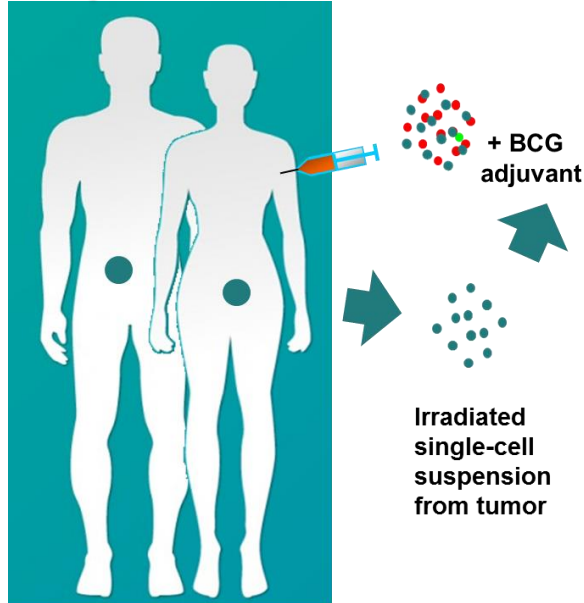
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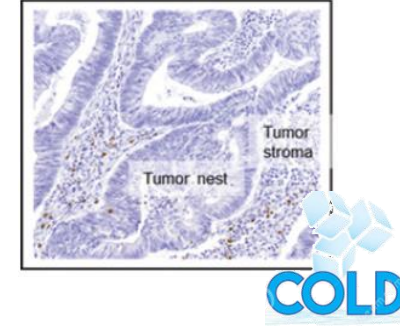
Clinical
Cancer
Research



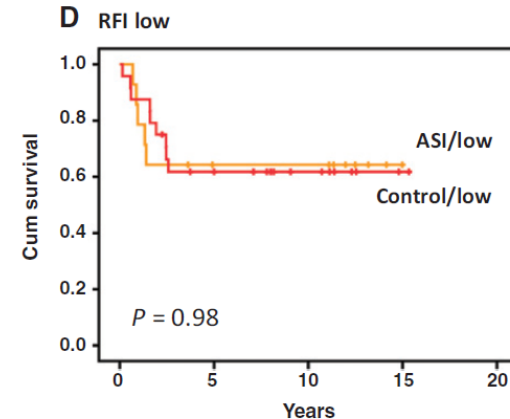
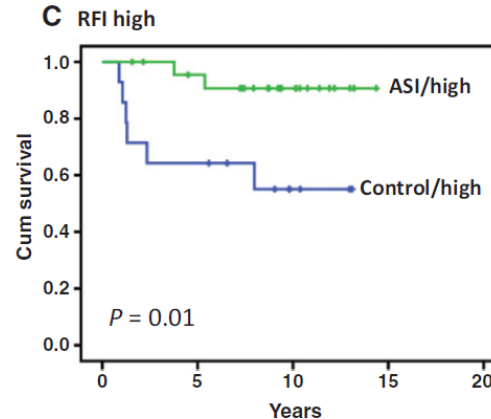
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T cell infiltration



Only patients with MSS and T cell infiltrates benefit from vaccination



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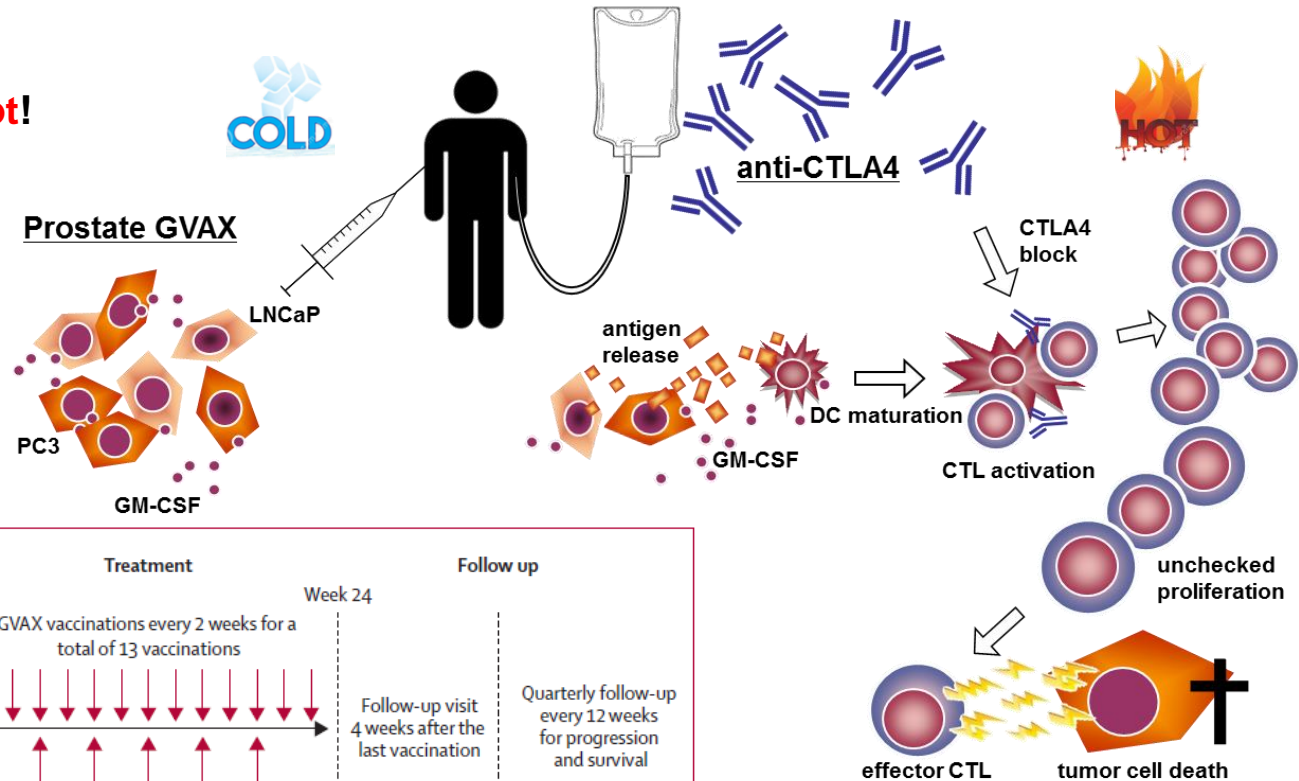
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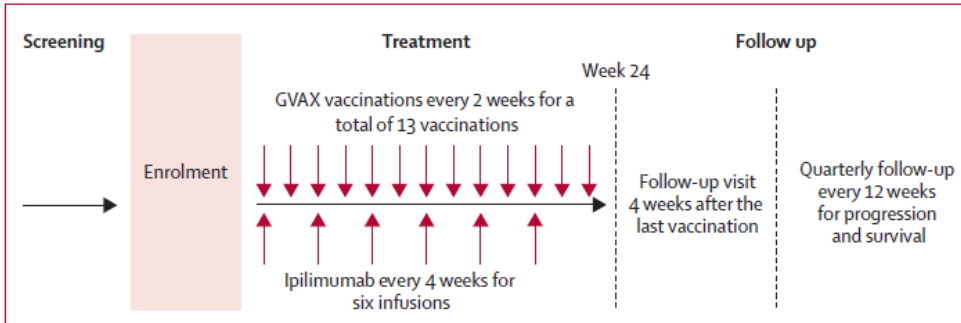


Turning a **cold** tumor **hot**!

Whole Cancer cell vaccines: GVAX with checkpoint blockade -turning up the heat



Lancet Oncol 2012; 13: 509-17



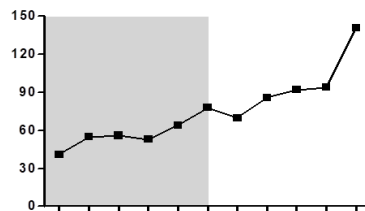
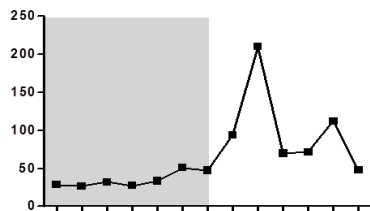
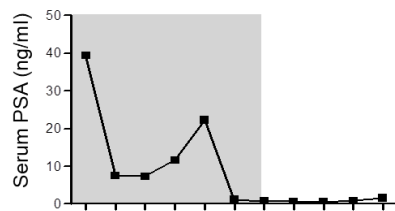


Whole Cancer cell vaccines: GVAX with checkpoint blockade -turning up the heat

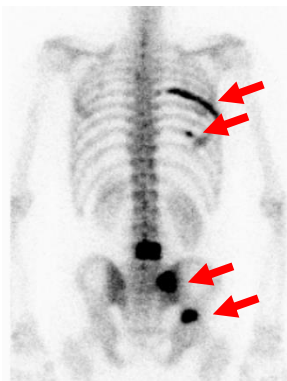
Partial Response (PR);
>50% PSA decline

Stable Disease (SD);
No PR or PD

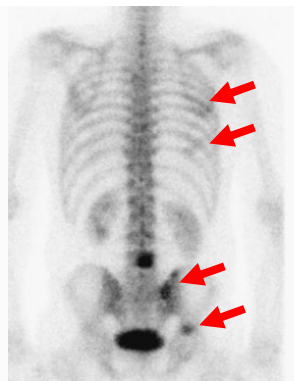
Progressive Disease (PD);
>25% PSA increase



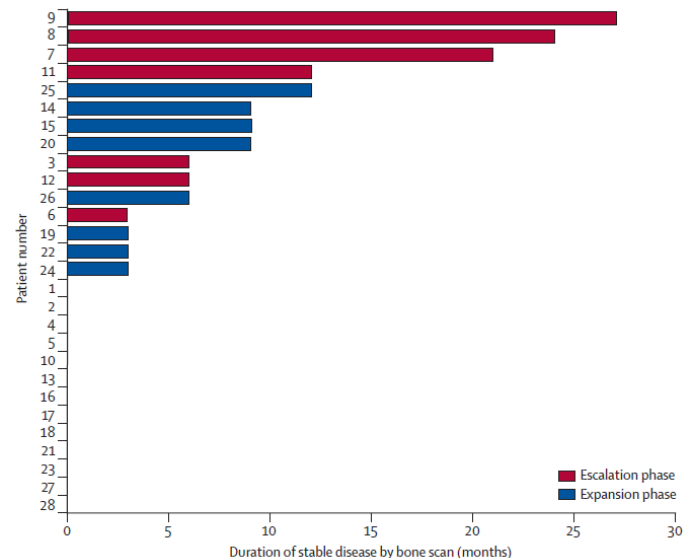
Category:	Number of patients
PSA Partial Response (PR)	5 / 28
PSA Stable Disease (SD)	12 / 28
PSA Progressive Disease (PD)	11 / 28



15-9-2005

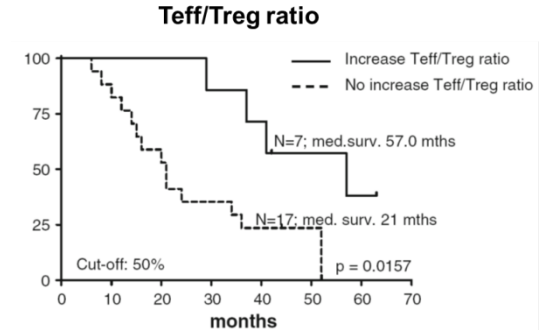
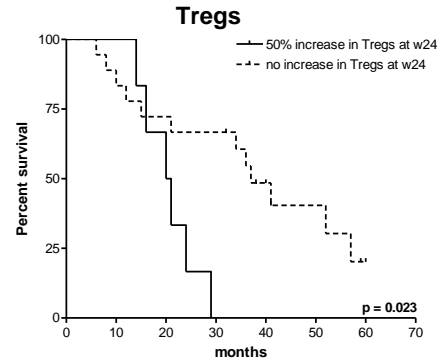
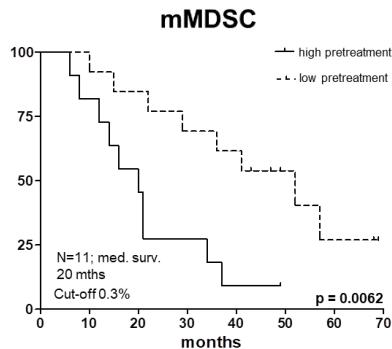
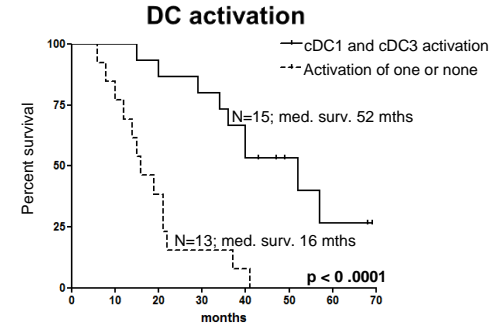
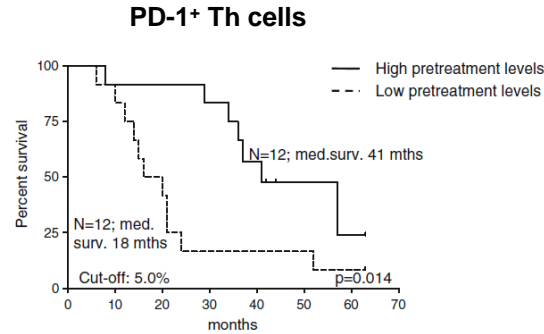
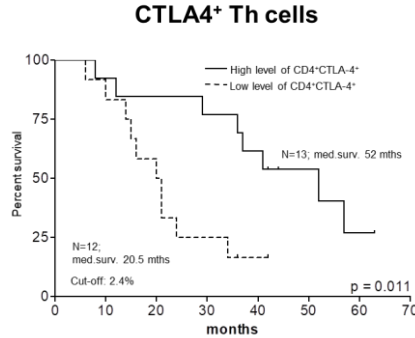


29-3-2006

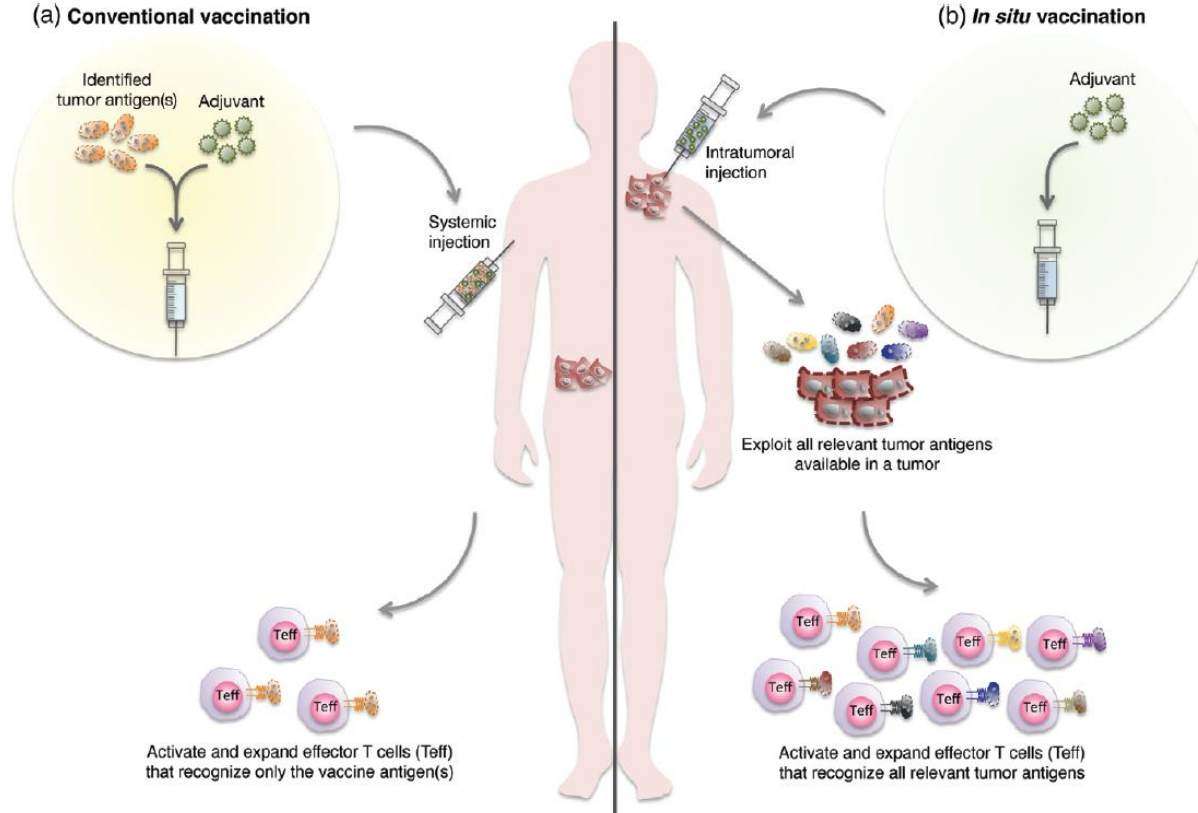




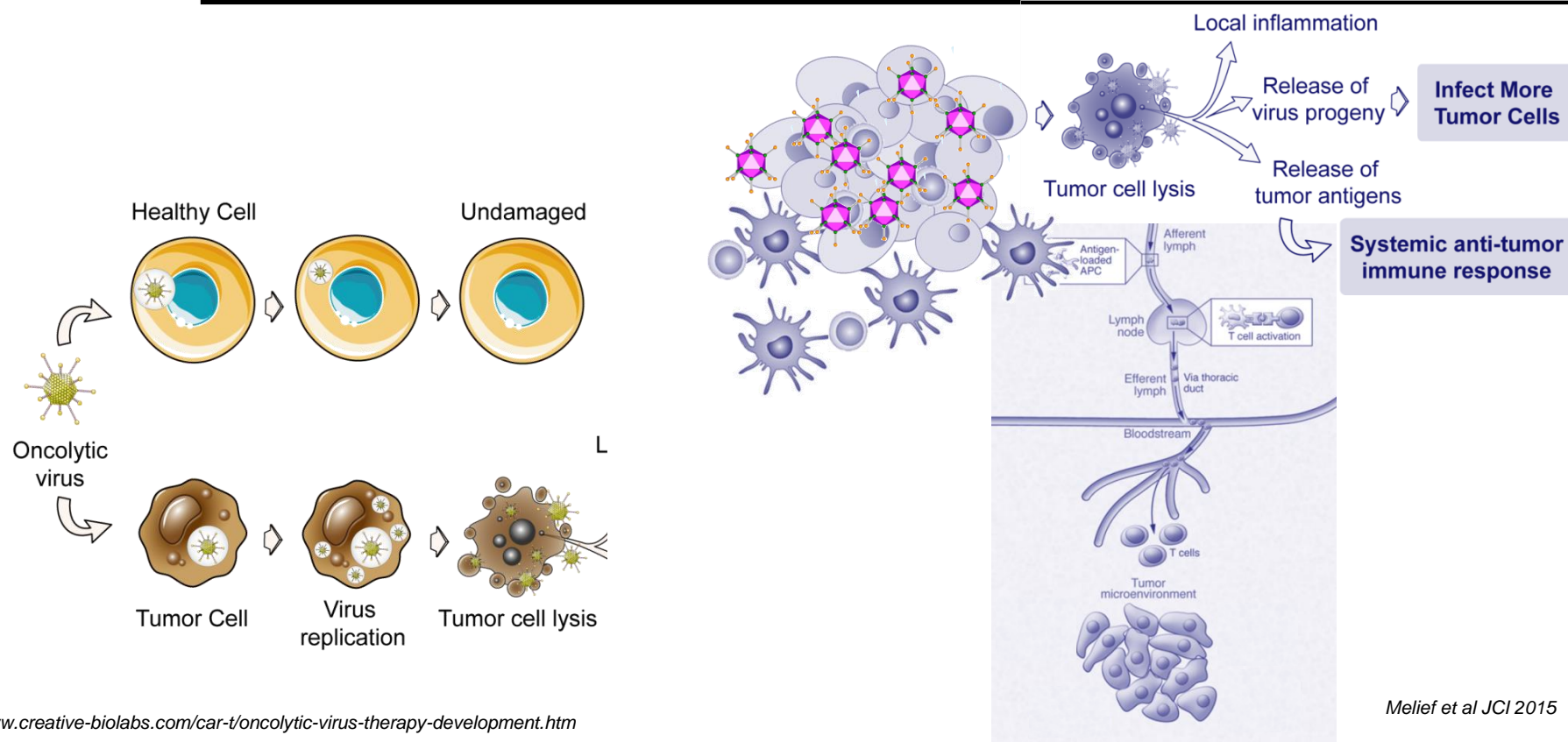
Whole Cancer cell vaccines: GVAX with checkpoint blockade -turning up the heat



Conventional versus *in vivo* vaccination



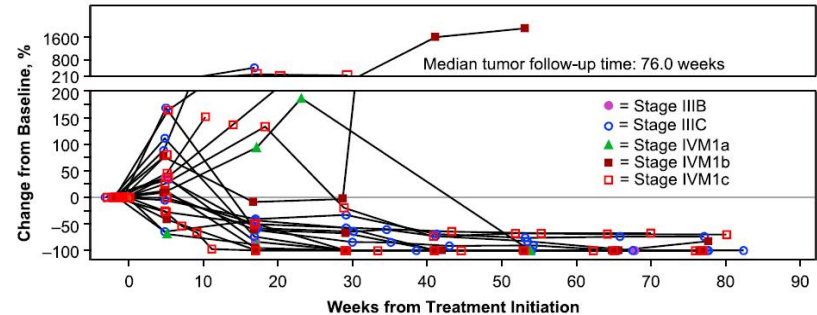
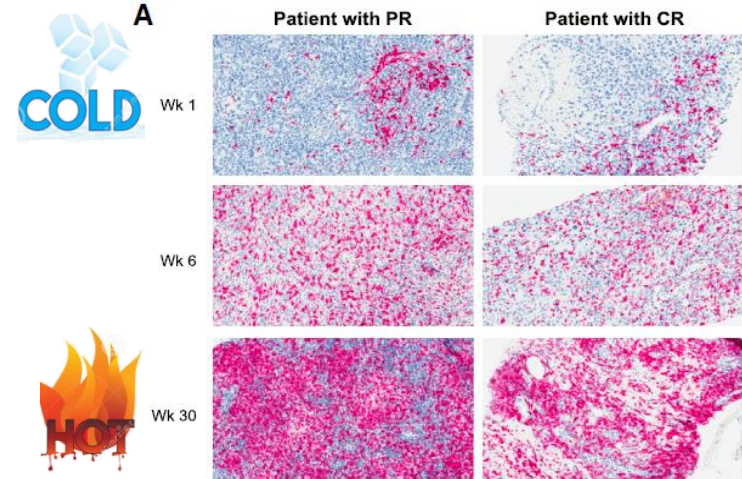
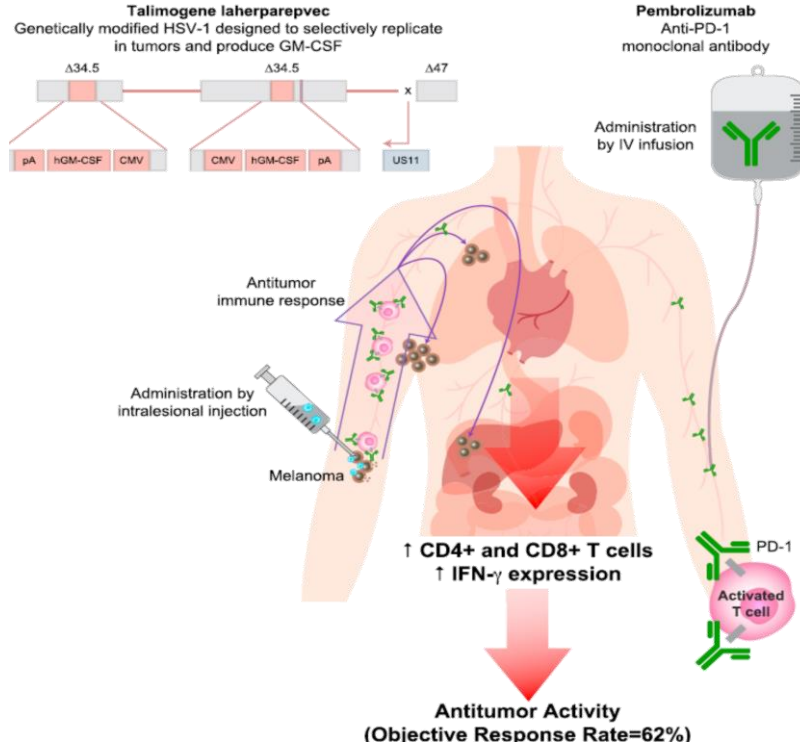
Oncolytic virotherapy: *in vivo* vaccination





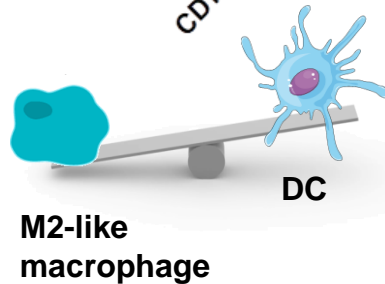
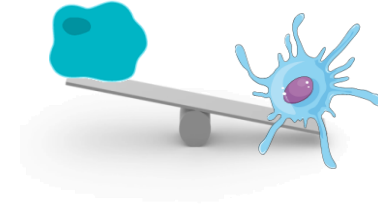
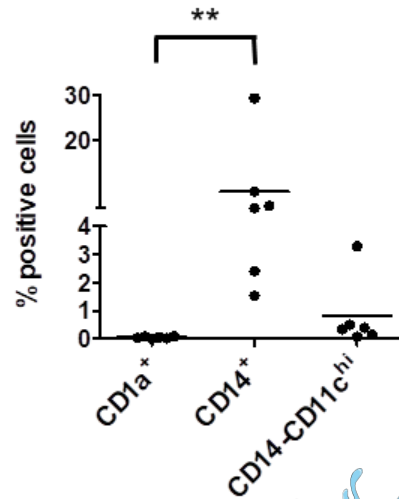
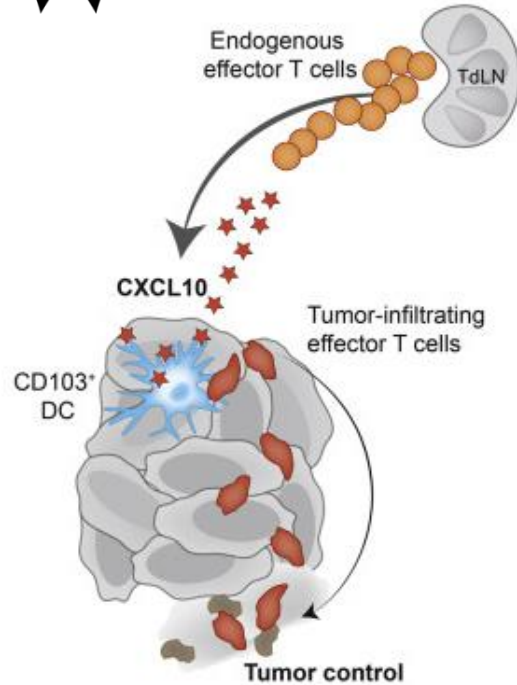
Oncolytic viruses (T-vec) and immune checkpoint blockade

Ribas et al. Cell 2017





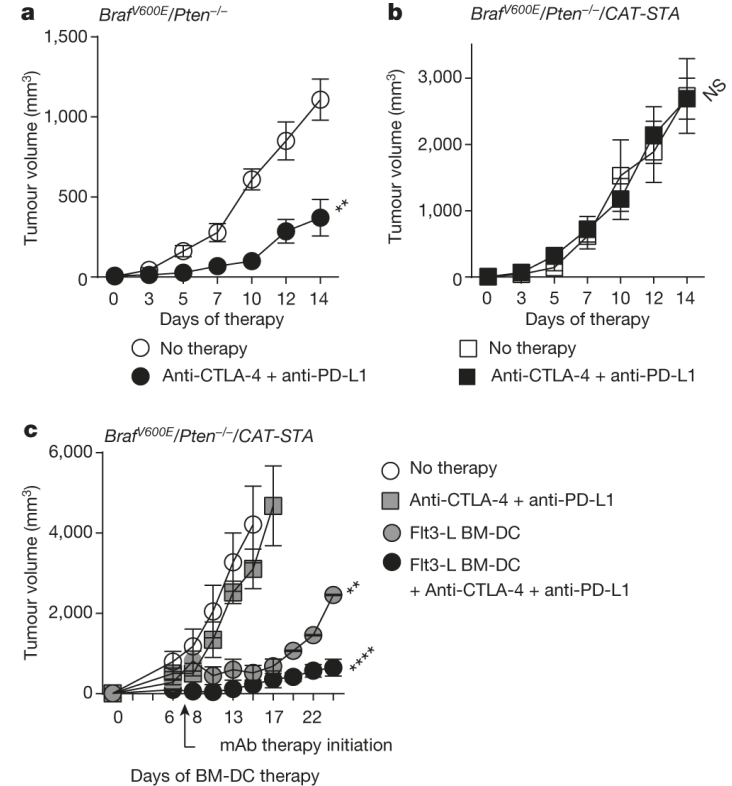
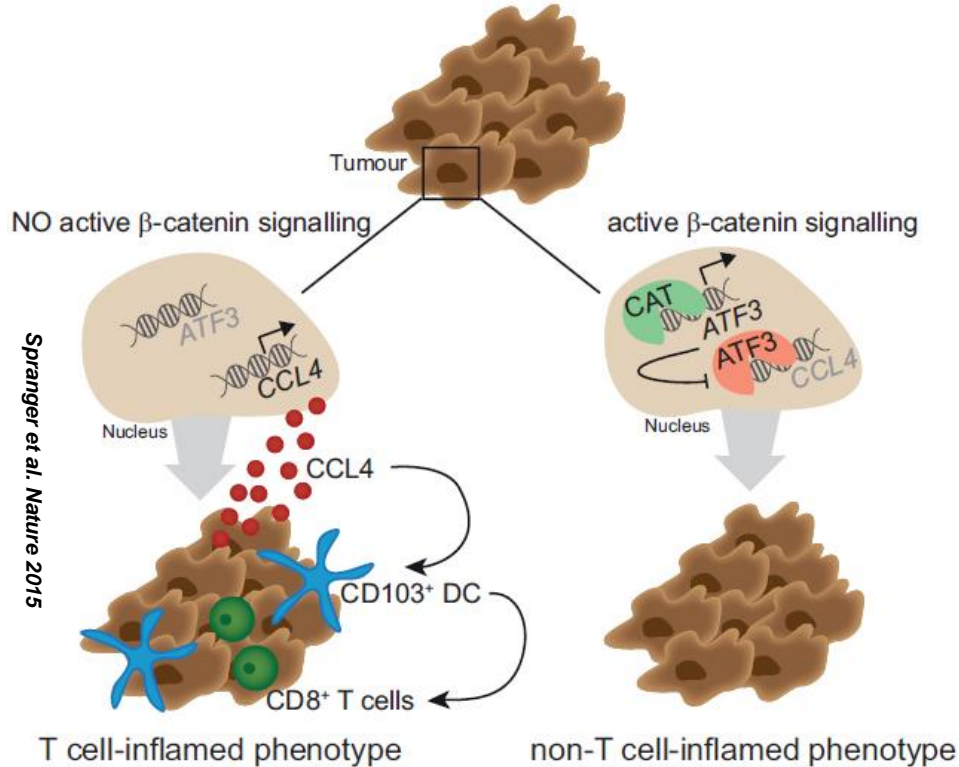
Melanoma T cell infiltration depends on DC



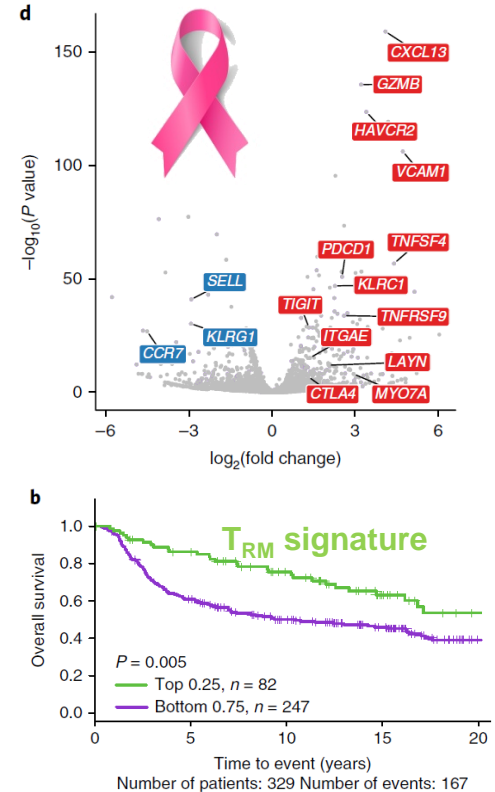
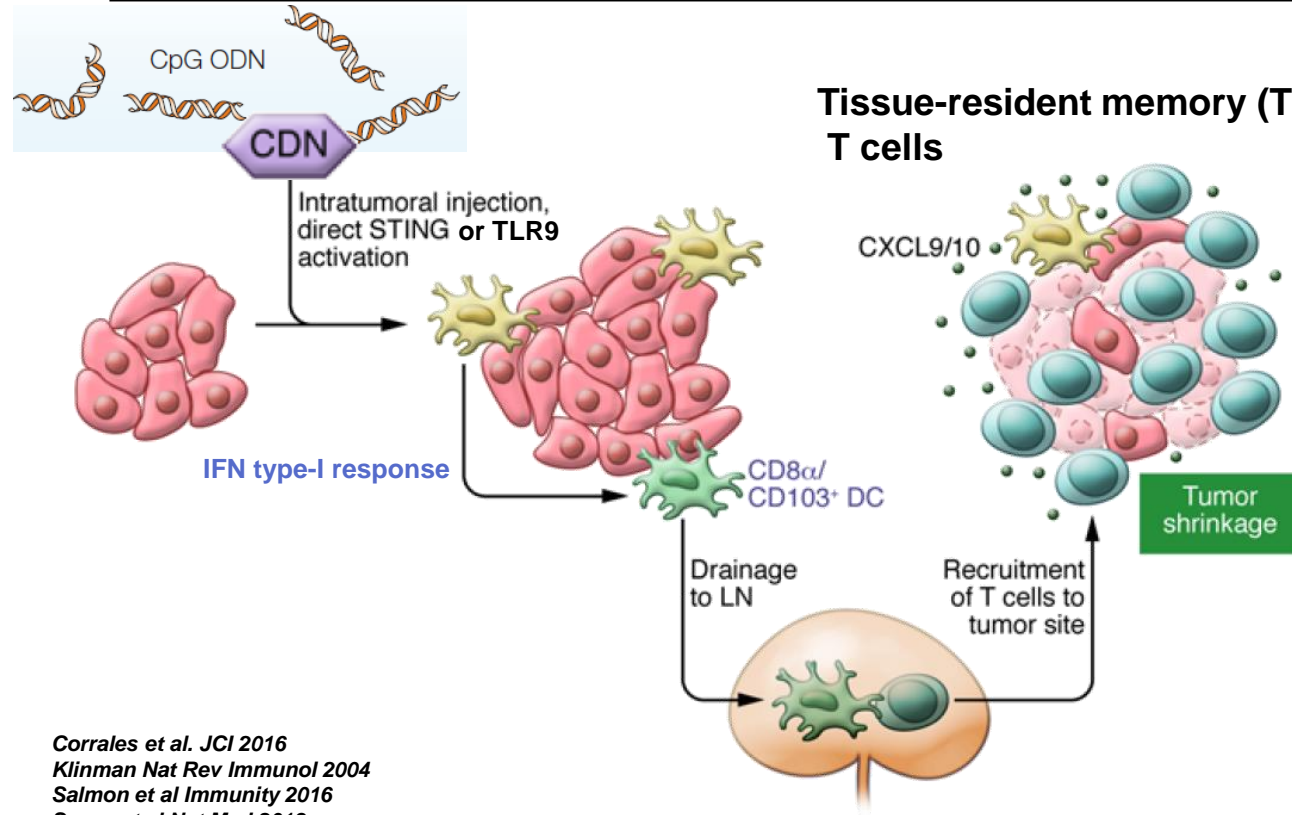
Aim: to safeguard proper DC differentiation in melanoma and enable *in vivo* vaccination



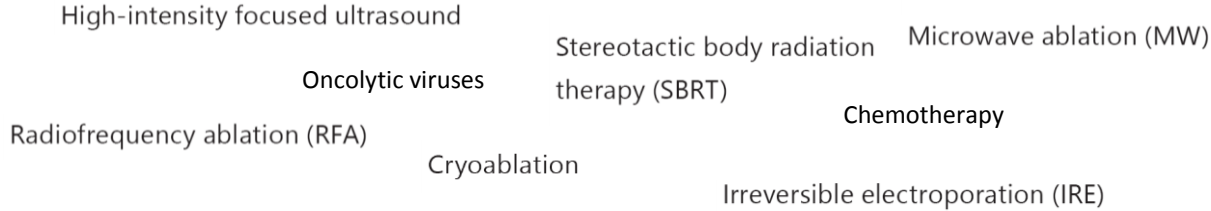
Role for Wnt in melanoma: DC and T cell infiltration



Leveraging DC *in vivo*: priming and attracting effective T cells



Options and advantages of *in vivo* vaccination



Immunogenic cell death: Ag and DAMP release
Type-I IFN response
Ensure CD103+ DC and T cell recruitment

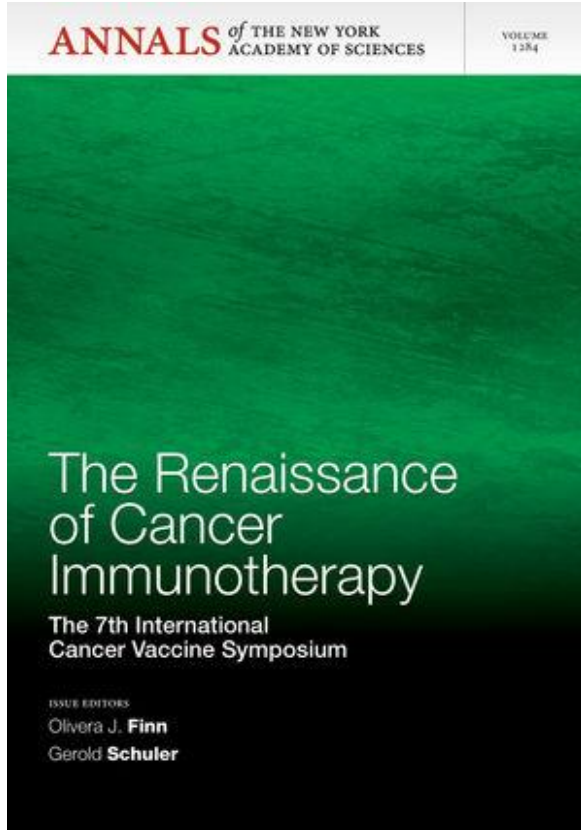


- ISV exploits all relevant antigens in the tumor, avoiding the need to identify tumor antigens or consider the HLA type
- ISV is simple and cost effective because it utilizes standard reagents and does not require patient-specific vaccines
- ISV takes advantage of the entire antigenic repertoire of a tumor to minimize immune escape
- ISV utilizes feasible local delivery with minimal systemic side effects
- ISV has the potential for synergy when combined with other therapies
- ISV can be effectively performed prior to surgery as neoadjuvant therapy

Learning goals

1. **Understanding the stumbling blocks and requirements for the design of successful therapeutic vaccines: choice of antigens and adjuvants**
2. **Becoming familiar with some of the current approaches to vaccination against cancer: broader definition includes *in vivo* vaccination**
3. **Acquiring an understanding of the positioning of cancer vaccines in the developing field of cancer immunotherapy**

Cancer vaccines: a renaissance in the golden age of immunotherapy?



<https://owlcation.com/humanities/>