# Immune monitoring of cellular and humoral responses in immunotherapy

Sacha Gnjatic, PhD

Associate Professor, Department of Medicine Division of Hematology and Medical Oncology Tisch Cancer Institute and Immunology Institute Icahn School of Medicine at Mount Sinai



The Tisch Cancer Institute

SITC PRIMER – NATIONAL HARBOR, MD – NOV 2015

## **Disclosure information: Sacha Gnjatic, PhD**

The following relationships exist related to this presentation:

*Immune Design:* Research Support *Janssen R&D:* Research Support *Third Rock Ventures:* Consulting Fees (e.g., advisory boards)

# Basic immunology is transforming cancer clinical care

- Accelerating FDA approval of a series of immune agents for the treatment of cancer, including first combinations
  - Anti-CTLA-4 for advanced melanoma (BMS 03/2011)
  - Anti-PD-1 for advanced melanoma (Merck 09/2014)
  - Anti-PD-1 for advanced melanoma (BMS 12/2014)
  - Bi-specific antibody for ALL (Amgen 12/2014)
  - Anti-PD-1 for NSCLC (BMS 03/2015)
  - Anti-PD-1 for PD-L1<sup>+</sup> NSCLC (Merck 10/2015) with companion test
  - Anti-PD-1 + Anti-CTLA-4 for BRAF<sup>WT</sup> melanoma (BMS 10/2015)
  - Anti-CTLA-4 for adjuvant melanoma (BMS 10/2015)
  - T-VEC for melanoma (Amgen 10/2015)
- Novartis invests \$20 million in CAR therapy and JUNO therapeutics raises \$265 million



• Mode of action is immunological, prompting intense search for correlative markers to understand and improve efficacy

## Immune monitoring goals

- To provide a comprehensive assessment of the immune status in patients
- To discover immune profiles of disease, leading to new biomarkers of diagnosis, prognosis, and response to therapy
- To quantitate immune responses to help optimize dose, delivery, schedule, and combinations and to identify immunomodulatory effects of novel drugs

## Human immunomonitoring



#### Single cell level

#### **BLOOD AND TISSUES**

#### Comprehensive

BIOTHERAPEUTICS

Biomarker of disease Target identification CELL COMPOSITION FLOW CYTOMETRY / MASS CYTOMETRY

SPECIFICITY & ENUMERATION ANTIGEN-SPECIFIC T CELL ASSAYS SERUM PROFILING OF ANTIBODIES

FUNCTION & QUALITY ELISPOT, INTRACELLULAR CYTOKINES, TETRAMERS, SORTING, AVIDITY, POLYFUNCTIONALITY, ISOTYPE

TRANSCRIPTOMICS OF PURIFIED POPULATIONS TCR SEQUENCING

PROTEOMICS

MULTIPLEX (antibody or aptamer-based) SEROMICS and ELISA for autoantibodies PHOSPHO-CYTOMETRY

TISSUE ANALYSIS & IMAGING IMMUNOSCORE, CONTEXTURE, MULTIPLEX IMMUNOHISTOCHEMISTRY

DATA ANALYSIS BIOINFORMATICS, DATA MANAGEMENT, HARMONIZATION, QUALITY CONTROL

### Case study: neoadjuvant therapy trial design

Hypothetical study of the immunomodulatory effect of a new drug administered prior to surgery followed by checkpoint blockade

#### At the tumor site

• Fresh biopsies, surgical material

➔ Mass cytometry profiling of tumor composition, cell sorting for expansion or functional characterization

Frozen material

→ Genomic analyses of microenvironment, immunofluorescence

## Paraffin-embedded blocks → Multiplex immunohistochemistry for immunoscore-type analyses, TCR sequencing

#### In the periphery

- Serially collected serum or plasma
  - → Antibody profiling
- Serially collected peripheral blood mononuclear cells (PBMC)
   Mass cytometry profiling of phenotypic and functional changes, antigen-specific T cell characterization, TCR sequencing

## Scope

In situ immune monitoring

#### Immunotherapy can make tumors "hot"

#### Measuring T cell infiltration by immunohistochemistry after neoadjuvant ipilimumab treatment in melanoma patients



### **CD8 tumor infiltration as a biomarker**

Predictive value of immunocytic infiltration in the context of immunotherapy

Immunohistochemical analysis of CD8<sup>+</sup> T cells in samples obtained before and during pembrolizumab treatment in advanced melanoma.



PC Tumeh et al. Nature 515, 568-571 (2014)

## **Immunomics.** Establishing an immune landscape using genomic or transcriptomic immune signatures of tumors



Adapted from Bindea et al., Immunity, 2013;39:782-95

Validation by immunohistochemistry of CRC tumors

### Identifying T cell repertoire and functionality in tumors

#### From fresh/frozen tissues:

- Use of CD154, 4-1BB, IFN-γ capture, etc. to sort cells following stimulation with antigen or tumor
- RNASeq of sorted populations

#### From paraffin-embedded tissue:

- Multiplex IHC with functional markers, needs development:
  - better tools to identify functional status of cells, including exhaustion markers and metabolic stress
  - better tools to assess master regulators of T cell differentiation and lineage: STAT1, T-bet, GATA-3, RORγt, etc.
- TCR sequencing to look for clonality status at baseline and diversification of repertoire after treatment



### Identifying antigens recognized at the tumor site

 Whole exome sequencing and RNASeq to identify tumor-specific mutations that may give rise to neoepitopes, followed by highthroughput tetramer screening or cytokine production of T cells (Schumacher et al)
 → Mutational load linked to clinical

response rate in checkpoint blockade

 Immunohistochemistry and RT-PCR to confirm presence of known tumor antigens



 Serology to quickly screen for immunogenic target antigens, as a surrogate for T cells, possible from fresh tissue after expansion



#### Screening supernatants from tumor-infiltrating B cells in NSCLC



 $\rightarrow$  Production of tumor-specific antibodies by intratumoral plasma cells

2014;189:832-44

## Scope

Peripheral immune monitoring

#### Mass cytometry to explore phenotypic changes during treatment



Nature Biotechnology

## Multi-dimension reduction algorithms and software allow to visualize complex data in 2D plots in an unbiased manner



Mass cytometry allows the analysis, at the single cell level, up to 40 markers simultaneously with minimal signal overlap.

### Seromics. Methodology for antibody profiling with protein microarrays



Protein microarrays contain >9000 proteins mostly full-length baculovirus-produced GST-fusion proteins randomly selected, both known and predicted sequences



## Seromics detects antigen-specific changes in autoantibody profiles during treatment (H. Wada, Osaka; H. Shiku, Mie)



Only 1 significant change in antibody reactivity out of 9000 possible proteins



40 other non-vaccine related proteins react, including cancer-testis antigens

## Correlation of NY-ESO-1 antibody with clinical course following anti-CTLA-4 treatment with ipilimumab

In collaboration with Jedd Wolchok and Jim Allison MSKCC/Ludwig Center and with Ruth Halaban and Mario Sznol, Yale University - Melanoma sera

#### Sera from melanoma patients taken at baseline, before CTLA-4 treatment

Status at wk 24	# patients (%)	NY-ESO-1 SERONEGATIVE # (%)	NY-ESO-1 SEROPOSITIVE # (%)
CR	4 (2.9%)	3	1
PR	14 (10.0%)	10	4
SD	30 (21.4%)	23	7
Clinical Benefit	<b>48</b> (34.3%)	<b>36</b> (30.5%)	<b>12</b> (54.6%)
No Clinical Benefit	<b>92</b> (65.7%)	<b>82</b> (69.5%)	<b>10</b> (45.4%)
Total	140 (100%)	118	22

According to Immune-related response criteria:

#### Clinical Benefit

- CR: Complete Response
- **PR: Partial Response**
- SD: Stable Disease

#### **No Clinical Benefit**

- POD: Progression of Disease (includes MR: mixed response)
- DOD: Dead of Disease

Fisher's exact test (two-tailed): P value 0.0481 RR=1.8(1.1-2.9)

#### MANIFOLD VARIABLES CLINICAL TRIAL STRATEGY CENTERED ON NY-ESO-1

#### CVC Trials Network

45 clinical trials of different vaccine combinations and strategies completed or ongoing, involving 950 patients worldwide

C C C C C C C C C C C C C C C C C C C	Antigen NY-ESO-1	Cancer PopulationsBladderLungBreastMelanomEsophagealOvarian/GastricTubeHead and NeckProstateSarcoma	ia Peritoneal/Fallopian	
Antigen Forms Peptides – Class I and II • Short Peptides • Long Peptides • Overlapping Peptides Protein DNA	Adjuvants Bacillus Calmette-Guerin CpG 7909 GM-CSF Imiquimod Mixed Bacterial Vaccine Montanide ISA-51 Poly ICLC Resiquimod Streptococcal OK432 AS02B	Delivery Systems Viral Vectors • Vaccinia-NY-ESO-1 • Fowlpox-NY-ESO-1 • Canarypox – ALVAC-NY- ESO-1/TRICOM Antigen Presenting Cells • DCs pulsed with NY- ESO-1 peptide Other	Inoculation Strategy Method • Gene Gun • Intradermal • Intramuscular • Subcutaneous • Topical Timing • Serial • Intensive course • Prime-boost	
Modulators of Immunosuppression Anti-CTLA-4 (ipilimumab) Cyclophosphamide		<ul> <li>Cholesteryl-bearing Hydrophobized Pullulan</li> <li>ISCOMATRIX</li> </ul>		

## Comparative summary of immune responses elicited following various NY-ESO-1-based vaccine trials

Trial, Publication, Comments	Ab	CD8	CD4	Integrated Ab, CD4 and CD8 responses
Vaccinia/Fowlpox PMID: 16984998, Jäger et al. 2006 3 baseline seropositive, various ca.	11/23 (48%)	19/23 (83%)	13/23 (57%)	7/23 (30%)
<b>Protein + CpG</b> PMID: 21163871, Karbach et al. 2011 2 baseline seropositive, prostate ca.	13/13 (100%)	6/13 (46%)	9/13 (69%)	6/13 (46%)
<b>Protein + Montanide + CpG</b> PMID: 17517626, Valmori et al. 2007 No baseline seropositive, various ca.	18/18 (100%)	9/18 (50%)	17/18 (94%)	9/18 (50%)
<b>Protein + CHP</b> PMID: 17441676, Uenaka et al. 2007 2 baseline seropositive, esophageal ca.	9/9 (100%)	7/9 (78%)	7/9 (78%)	5/9 (56%)
<b>OLP+ Montanide + Poly-ICLC</b> PMID: 23032745, Sabbatini et al. 2012 1 baseline seropositive, ovarian ca.	10/11 (91%)	10/11 (91%)	11/11 (100%)	10/11 (91%)

CHP: Cholesteryl Pullulan delivery adjuvant

OLP: Overlapping Long Peptides (30-32mers)

Ca.: Cancer

## Induction of immunity by vaccine strategies with different antigen formulations

Vaccine antigen formulation	Ab	CD8	CD4	Recognition of naturally processed antigen
Short HLA Class I peptides	-	+++	-	Rarely
Short HLA Class II peptides	-	+/	+++	Rarely
rV-NY-ESO-1 / rF-NY-ESO-1	+/_	++/_	++/_	Yes
DNA	-	+/	++	Yes
Protein	+++	++/_	+++	Yes
Overlapping long peptides	++	+++	+++	Yes

### Take home message

Comprehensive immune monitoring strategies to guide and inform future immunotherapy designs

Multiplexing and sample-sparing techniques are becoming critical to address the complexity of immune responses and suppression

Plan ahead: need to consider sampling of tissues and samples carefully

Defining antigen specificity and quality of immune responses is important to validate the mechanism of action of drugs

Era of biomarker discovery for companion diagnostics and patient pre-selection

## **Future directions**

Microbiome

**Single-cell genomics** 

Integration with systems biology and bioinformatics

Plasticity, ontogeny of immune cells – Variability over time

In situ specificity (tetramers for IHC, microdissection and functional analyses)

## Acknowledgments

Icahn School of Medicine at Mount Sinai, New York NY Romain Remark Adeeb Rahman Seunghee Kim-Schulze Miriam Merad Nina Bhardwaj Hearn J. Cho

Ludwig Institute for Cancer Research, New York NY

Achim Jungbluth Erika Ritter Nikoletta Lendvai Ralph Venhaus Linda Pan Lloyd J. Old

Institut des Cordeliers, INSERM, Paris FR Marie-Caroline Dieu-Nosjean Claire Germain Jérôme Galon Diane Damotte Catherine Sautès-Fridman Wolf-Hervé Fridman

Cancer Research Institute, New York, NY Jill O'Donnell-Tormey MSKCC, New York, NY Jianda Yuan Taha Merghoub Jedd Wolchok

MD Anderson, Houston, NY Jim Allison

Yale University, New Haven, CT Mario Sznol Ruth Halaban

Osaka and Mie University, Japan Hisashi Wada Hiroyoshi Nishikawa Hiroshi Shiku

Roswell Park Cancer Institute, Buffalo NY Takemasa Tsuji

Support from: Cancer Research Institute Cancer Vaccine Collaborative ARRA RC2 NCI Ludwig Institute for Cancer Research