The Good, the Bad and the Ugly:

Clinical trials which assess vaccine characteristics

ISBT Meeting, San Francisco, CA November 4-8, 2004

Ideal cancer vaccine trial

- 1. An informative immune assay
- 2. Ability to derive data on immune response
- 3. Toxicity and clinical response/survival data

- 4. Correlate ability of the assay in #1 to be a surrogate for #3.
- 5. Problem: #3 not the idea scenario for #2!

What is needed in a vaccine trial?

- Sufficient <u>number</u> of highly <u>avid</u> T cells that are antigen specific
- Ability of the T cells in question to <u>traffic</u> to lymph nodes and sites
- T cells generated must be in a proper state of activation and able to overcome both passive (antigen and MHC down-regulation) and active (Tregs, IL-10, TGF-beta) defences

What is the evidence that immune monitoring has clinical relevance?

- We need to determine if any immune assay correlates with relapse-free or overall survival
- Is there a surrogate endpoint for survival and/or clinical benefit?
- If simple enumeration is not useful, why not?
- Immune monitoring, if it correlates with clinical benefit can help us decide what qualities are important for a therapeutic T cell

Do we have the right assay, in the right type of trial?

- Different immune assays need to be prioritized
- Is there a place for pure immune surrogate trials, in patients without evidence of disease?
- Should we concentrate on patients with measurable disease, or are NED patients OK?
- What clinical endpoints are proper for vaccine trials; survival vs. response vs. stability?

Are we measuring the correct thing, and in the right place?

- Measurement of circulating T cells in PBMC is important, but what about draining nodes and tumor infiltrating T cells?
- Should we be measuring circulating or tumor Treg cells as well as effector cells?
- How important are circulating cytokines, both proinflammatory and suppressive?
- Are NK, NKT or DC relevant as a measure of immunity?
- Should we be measuring cytokine gene polymorphisms and cytokine gene epigenetic modifications and changes after vaccination as a surrogate marker for the ability to immunize?

Case studies in immune monitoring of clinical vaccine trials

- CanVaxin: cell based vaccine with BCG
- Peptide vaccines: melanoma differentiation antigens
- Dendritic cells: pulsed with peptides, lysates and fusion products
- This is not a comprehensive assessment, more a set of instructive examples to assess whether immune assays correlate with clinical benefit

Canvaxin: cellular vaccine

Chung et al JCO 2003 21: 313

- Three melanoma cell lines administered with BCG for two injections, then alone for 6 months total
- Induces antibody responses against a 90 kD tumor associated glycoprotein TA-90
- 54 patients: (-)SNB, all had >4 mm melanoma
- 43 got vaccine, 11 were observed
- DFS and OS correlated with maximal TA-90 IgM response (p=0.006 and 0.06) in the vaccine group, but not the observation group
- Non-randomized, but encouraging result

Canvaxin: cellular vaccine Morton et al. Ann Surg 2002 236: 438

- 2602 patients had complete lymphadenectomy in the period 1984-1998; 935 received Canvaxin, and 1667 did not
- Comparison group had no therapy or IFN 1971-1998
- They were matched for 7 co-variates
- Median OS was 49% vs. 37% favoring vacche
- The authors claim OS was the same in the observed group pre-1985 and post-1985, which disagrees with SWOG data
- Canvaxin correlated with OS p=0.001; RR death = 0.64
- Justifies a randomized phase III trial, just concluded in over 1100 patients of Canvaxin/BCG vs. BCG alone

CanVaxin: Phase III trials

- Two randomized trials, one ongoing, one just finished in resected stages III and IV melanoma
- A lower than expected rate of events will slow down the final interpretation of the trial; BCG effect?
- Evidence that Canvaxin/BCG may be beneficial:
 - Vaccinated patients have increased DTH to the vaccine, which correlates with survival
 - Vaccinated patients have a reduction in TA-90 IgM levels, which correlates with survival
 - Anti-ganglioside antibodies are induced by CanVaxin
 - T cell responses can be detected to known antigens

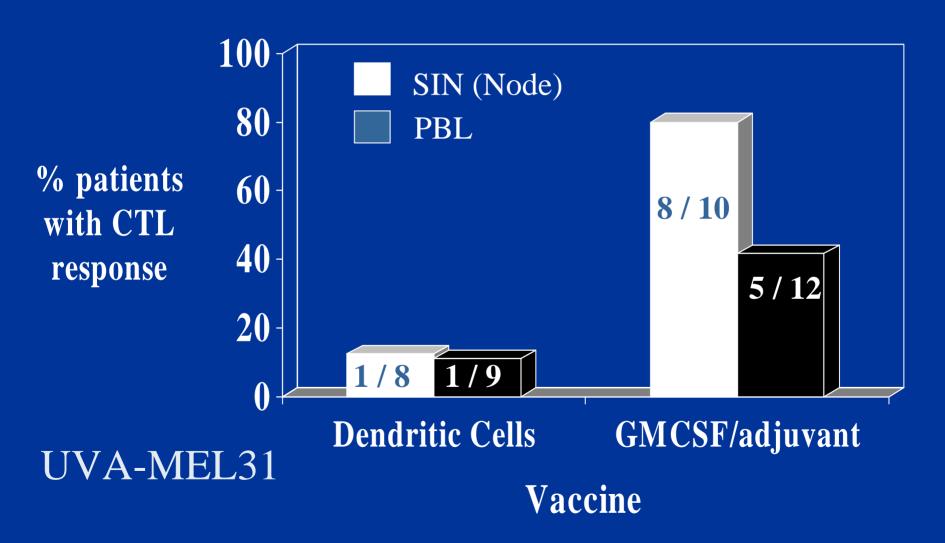
Peptide vaccine for resected melanoma: Walker et al Clin Can Res 2004

- 35 patients received gp100 209-217 (210M) with Montanide ISA 51
- Tetramer staining shows median of 0.36% post-vaccine (0.05 to 8.9%)
- Cells were CCR7(-) CD45RA (+) or (-) > suggesting effector or effector-memory type
- Virtually all cells expressed gamma interferon after *ex vivo* expansion

Peptide vs. DC vaccine for stage IV melanoma: Slingluff et al *J Clin Oncol* 2004

- 26 patients, stage IV melanoma, 13 each randomly allocated to receive peptides with Montanide/GM-CSF or pulsed onto DC
- Higher overall immune response with restimulated ELISPOT in peptide arm p<.02
- Vitiligo seen in 2 peptide but no DC patient
- 4 SD + PR in the peptide arm, versus 2 SD + PR in the DC arm
- Immune response appeared to correlate with PR/SD

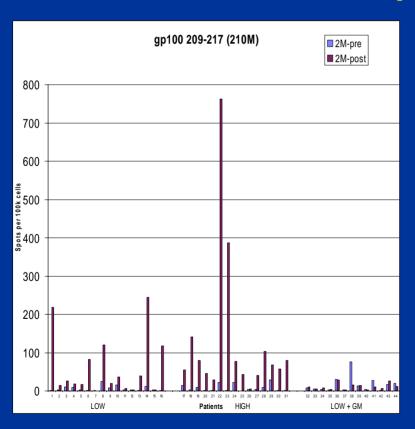
Evaluation of CTL Responses to Vaccination with GMCSF-in-Adjuvant or DC+peptide in Patients with Substantial Tumor Burden (Stage IV)

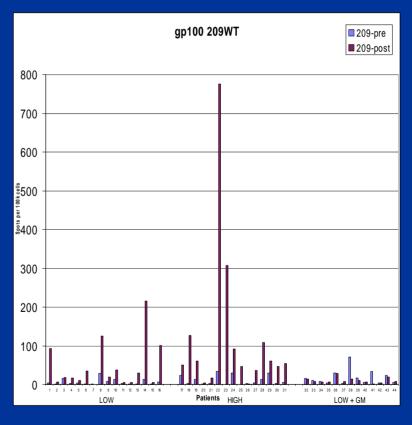


Peptide vaccines for melanoma: Clinical data

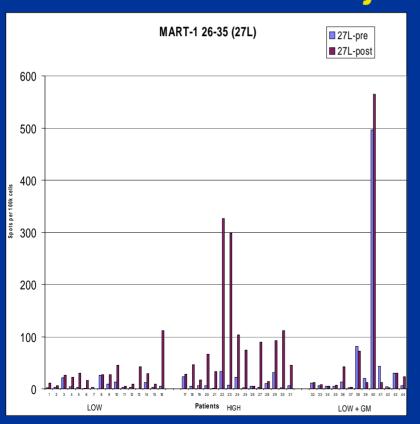
- gp100/tyrosinase/IFA+IL-12 trial for resected stage III/IV patients: 26 with stage III, 22 with stage IV disease; median relapse-free survival 20 months, median survival greater than 57 months, 85% had augmented immunity to gp100 by tetramer staining, with increase from 0.03 to 0.08% IL-12 vs. no IL-12 Lee et al J Clin Oncol 2001
- In an ongoing trial, <u>three</u> peptides with IFA were used to vaccinate stage III/IV resected patients with low dose IL-12/alum, low dose IL-12+GM-CSF or high dose IL-12/alum.

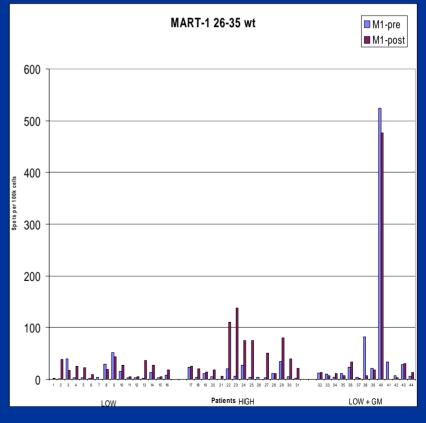
Reactivity to melanoma antigen gp100: are higher doses of IL-12 with alum a superior adjuvant?





Reactivity to melanoma antigen MART-1: are higher doses of IL-12 with alum a superior adjuvant?





Conclusions: Peptide vaccines with Montanide, alum and IL-12

- ELISPOT responses greater for both gp100 and MART-1/Melan-A heteroclitic and wild type in high dose IL-12 than either low dose group, p values ranging from 0.04 to 0.005
- WT immune responses equal to heterclitic
- More deaths (3 versus 1) and more relapses (10 vs. 4) in low dose groups than high dose group; correlation seen with immune response and time to relapse

Fowlpox gp100 vaccine: no correlation of immunity with response

- Three consecutive trials were done with 7, 14 and 16 pts who received a fowlpox-native gp100, fowlpox modified gp100, and folwpox –gp100 minigene (ER targeted)
- Rosenberg et al Clin Can Res 2003
- Responses to gp100 seen in 0/7, 10/14 and 12/16 patients respectively
- Restimulation assays done for cytokine release
- No correlation of assays with response and benefit
- The group immunized with the fowlpox gp100 minigene later received IL-2 with a 50% response rate

Class II peptide-pulsed DC Schuler-Thurner et al J Exp Med 2002

- Five biweekly SC vaccinations with peptide pulsed mature DC; only 16 received all DC
- Good responses seen to MAGE-3 243-258 by fresh *ex vivo* ELISPOT, and to KLH
- No clear correlation of immune response with clinical response; 1 CR with very low immunity seen, also 7 stable disease patients with no clear pattern of immunity

hTERT peptide-pulsed DC induce functional T cell responses

- Four of seven patients immunized with hTERT peptide/KLH pulsed DC demonstrated an immune response
- The only objective response in a breast cancer patient was associated with a potent CD8 T cell response Vonderheide et al *Clin Can Res* 2004
- The same hTERT I540 peptide with Montanide did not induce immune responses with CD8 T cells that recognized native cell lines; 0 responses were observed Parkhurst et al *Clin Can Res* 2004

Peptide-pulsed CD34+ derived DC

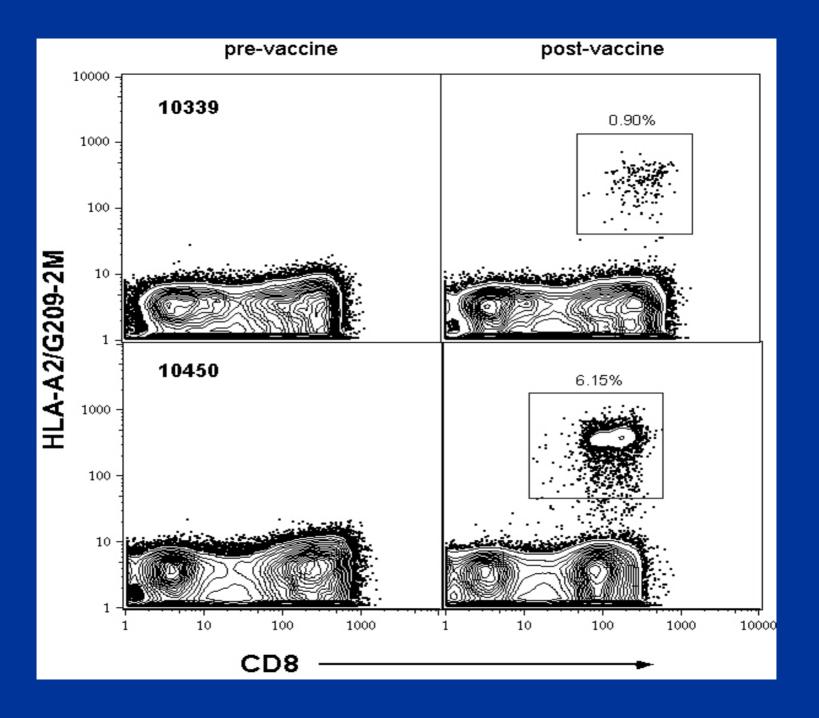
- 18 patients were treated with multiple melanoma peptidepulsed DC generated from CD34+ progenitor cells
- 16/18 responded by ELISPOT to *ex vivo* or restimulated cells
- 6/7 pts with response to 2 peptides or less progressed, versus only 1 of 9 with an immune response with p=0.02; the authors felt that response correlated with benefit
- Follow-up suggests that survival does correlate with immune response to more than 2 antigens
- Palucka et al *Cancer Res* 2001

CEA peptide-pulsed flt3L derived DC: immune response correlation

- Patients were treated with heteroclitic CEA peptide-pulsed DC after flt3L treatment
- 2 clinical responses of 12 seen
- Correlation of clinical response with CD8 tetramer-specific immune response to CEA
- Fong et al *P.N.A.S.* 2001

Immune Assays for tumor specific T cells: strengths and weaknesses

- Choice of surrogate assay in important to guide future development
- ELISPOT methodology is limited in its reliability, flexibility and reproducibility, but is today's choice
- Flow assays can be standardized and easily controlled, but are not functional assays
- New tetramer assay generates functional CD8 T cell data; it is based on staining with CD107a, a lysosomal membrane protein, to denote lytic T cells
- Tetramer array in development yields quantitative data on T cell phenotype and function



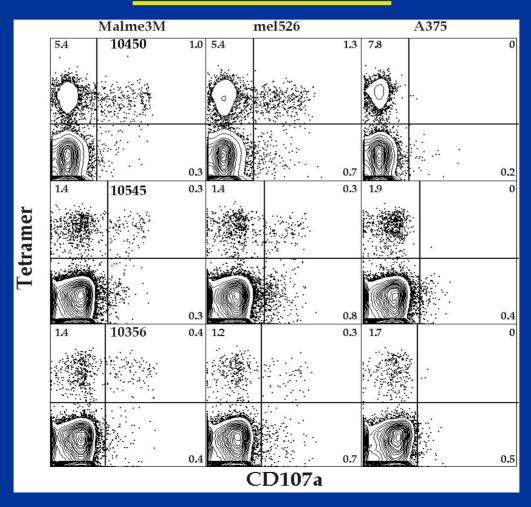
High avidity T cell clones are CD107a positive Lee et al Nat Med 2003

- CD8 T cell clones were raised from gp100 peptide-vaccinated melanoma patients
- Most were low avidity and did not recognize tumor cells or APC pulsed with low peptide concentration; some were high avidity but all bound gp100 tetramer
- The high avidity clones were lytic, recognized tumor cells and expressed CD107a

Tetramer+ CD8 high avidity T cell clones are CD107a positive and recognize tumor cells

| Average % | cytotoxicity | Functional avidity (M) | | | |
|-----------|--------------|------------------------|--|--|--|
| CD107+ | CD107- | CD107+ | | | |
| 45 | -2 | 10-12 | | | |
| 13 | -3 | 10-11 | | | |
| 42 | -3 | 10-11 | | | |
| 35 | -2 | 10-11 | | | |
| 46 | -5 | 10-12 | | | |
| 31 | -5 | 10-11 | | | |
| 35.3 | -3.3 | | | | |

CD107a/tetramer flow assay: high avidity T cells recognize tumor cells

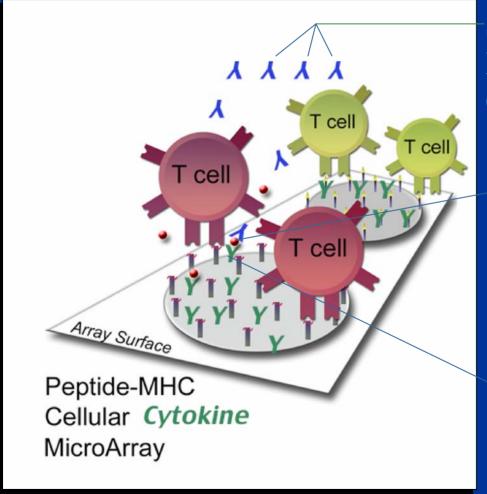


Functional status of TAA specific immune response: endogenous* vs. vaccine induced T cells:

| | | | % functional response | | | | | |
|------------|-------------|----------------|-----------------------|------|------|--------|--------------|-------------------------|
| Patient ID | <u>TAA</u> | T2- peptide | | | | mel526 | Malm e-3M | <u>A-</u> <u>375</u> |
| 422 | G209- 2M | 98.2 | | 27.2 | 23.8 | 0.5 | | |
| 476 | G209- 2M | 99.6 | | 99.6 | | 32.5 | 27.8 | 2.6 |
| 132* | G209- 2M | 99.2 | | 87 | 82.2 | 2.7 | | |
| 517 | M26 | 86 | | 86 | | 29.3 | 28.3 | 6.4 |
| 520 | M26 | 93 | | 93 | | 28.5 | 25.1 | 2.8 |
| 461* | M26 | 95.3 | | 54.9 | 36.8 | 3 | | |

| | <i>,</i> | | • | | | | |
|--------------|----------------|-----------------------|--------------------------|-----------------|---|--|--|
| | | | % functional response | | | | |
| Patie n t ID | TAA | % tetr am er | <u>mel</u> <u>526</u> | Malm e 3M | <u>A-</u> <u>3</u> <u>7</u> <u>5</u> | | |
| 713 | M26 | 0.49 | 18.2 | 13 | 6.1 | | |
| 721 | M26 | 0.19 | 49.6 | 45.2 | 4.8 | | |
| 721 | G209- 2 | 0.23 | 20.8 | 24.9 | 5.2 | | |
| 735 | M26 | 0.21 | 30.6 | 42.1 | 1.8 | | |
| 735 | G209- | 2.7 | 32.6 | 35 | 3 | | |
| 722 | $G209_{M}^{2}$ | 0.19 | 37.4 | 21.5 | 6.1 | | |

MHC-Cytokine Arrays Cytokine Sandwich Assays



Secondary cytokine
Detection antibody
Conjugated to a flurophore

Co-spotted Cytokine Capture antibody

Cytokine secreted by T cell after recognition of Peptide/MHC

T Cell Functional Profile

Capture Probes: αCD8, gp100 209/A2, MART1 25/A2, CMVpp65/A2, αCD3/αCD28

Cytokine Detector Probes:

| IL4 | IFNγ | IL12 |
|------|------------|--------|
| IL5 | TNFα | IL15 |
| IL10 | GranzymeB | VEGF |
| IL13 | GM-CSF | VEGF-D |
| TGFβ | IL1b | |
| IL2 | IL6 | |
| IL7 | No Co-Spot | |
| | | |

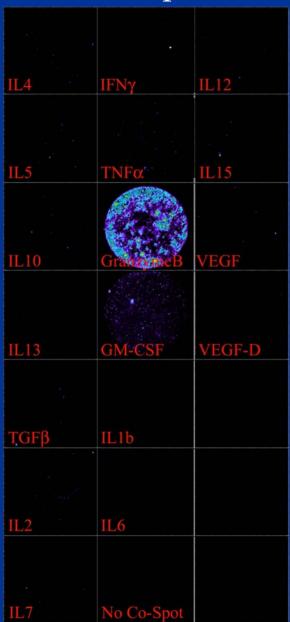
IL4 IL5 IL10 IL13 $TGF\beta$ IL2 IL7 $IFN\gamma$ $TNF\alpha$ GranzymeB

GM-CSF IL1b IL6 IL12 IL15 VEGF VEGF-D Regulatory cytokine Regulatory cytokine Immunosuppressive Regulatory cytokine Growth factor (f)Stimulatory cytokine Cytokine growth *f* Stimulatory pleiotropic *f* Stimulatory pleiotropic *f* Mediator of CTL killing, apoptotic *f* Hematologic growth *f* Inflammatory cytokine Stimulatory cytokine Stimulatory cytokine Stimulatory cytokine Angiogenic *f* Angiogenic/lymphogenic

αCD8 Co-Spots

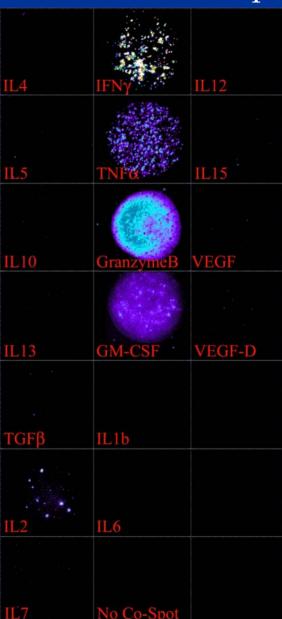


αCD8 brightfield



MART1/A2 Co-Spots





Functional T Cell Responses to Peptide Vaccines

| Pt | ID | Vax | Adj | Stage | Outcome | IFNγ | TNFα | GranzB | IL-2 | TGFb | IL1b | IL6 | GMCSF |
|----|-------|-------|-----------|-------|------------------------|------|------|--------|------|------|------|-----|-------|
| 1 | 68w | 3 рер | High IL12 | IV | Alive, 13m | | | | | | | | |
| 2 | 76m | 3 рер | High IL12 | IV | Alive, 13m | | | | | | | | |
| 3 | 72m | 3 рер | High IL12 | III | Recur m8/deceased | | | | | | | | |
| 4 | 65w | 3 рер | Low IL12 | III | Relapsed m11, resected | | | | | | | | |
| 5 | 52w | 3 рер | Low IL12 | III | Alive, 16m | | | | | | | | |
| 6 | 74m | 3 рер | Low IL12 | III | Alive, 16m | | | | | | | | |
| | | | | | | | | | | | | | |
| 7 | 37m | 3 рер | Montanide | III | Alive, 16m | | | | | | | | |
| | | | | | | | | | | | | | |
| 8 | 42m | 2 pep | GMCSF | III | Alive, 5yrs | | | | | | | | |
| 9 | 51m | 2 pep | GMCSF | III | Alive, 5yrs | | | | | | | | |
| | | | | | | | | | | | | | |
| G | clone | | no | | - | | | | | | | | |
| М | clone | | no | | - | | | | | | | | |

of Blocks
Denotes gp100
Specific Activity

Overview

- Analysis of T cell specificity and function
- Single cell resolution
- High throughput

- Few peptide-specific T cells are responsive
- Different vaccination strategies result in different functional profiles
- Interferon-γ and TNF-α discordance correlates with poor outcomes
- IL-1b and IL-6 secretion is associated with good outcomes
- Representation of complex cellular interplay

Conclusions and Lessons Learned

- Immune monitoring is more rigorously and carefully done and ex vivo tetramer and ELISPOT assays are more widespread than when we last met in 2001
- More evidence on the correlation between immune response and clinical benefit seen, but most trials have failed to show any correlation
- State of the art <u>functional</u> *ex vivo* assays are necessary, and new assays and arrays are likely to be useful
- Immune response assays provide feedback on optimal vaccine development and mechanistic understanding
- High avidity, long lasting T cells capable of recognizing antigen on tumor cells are needed
- We need to think outside the box on the development of new surrogate assays of immunity in cancer