

Adoptive T-Cell Transfer for Metastatic Melanoma:

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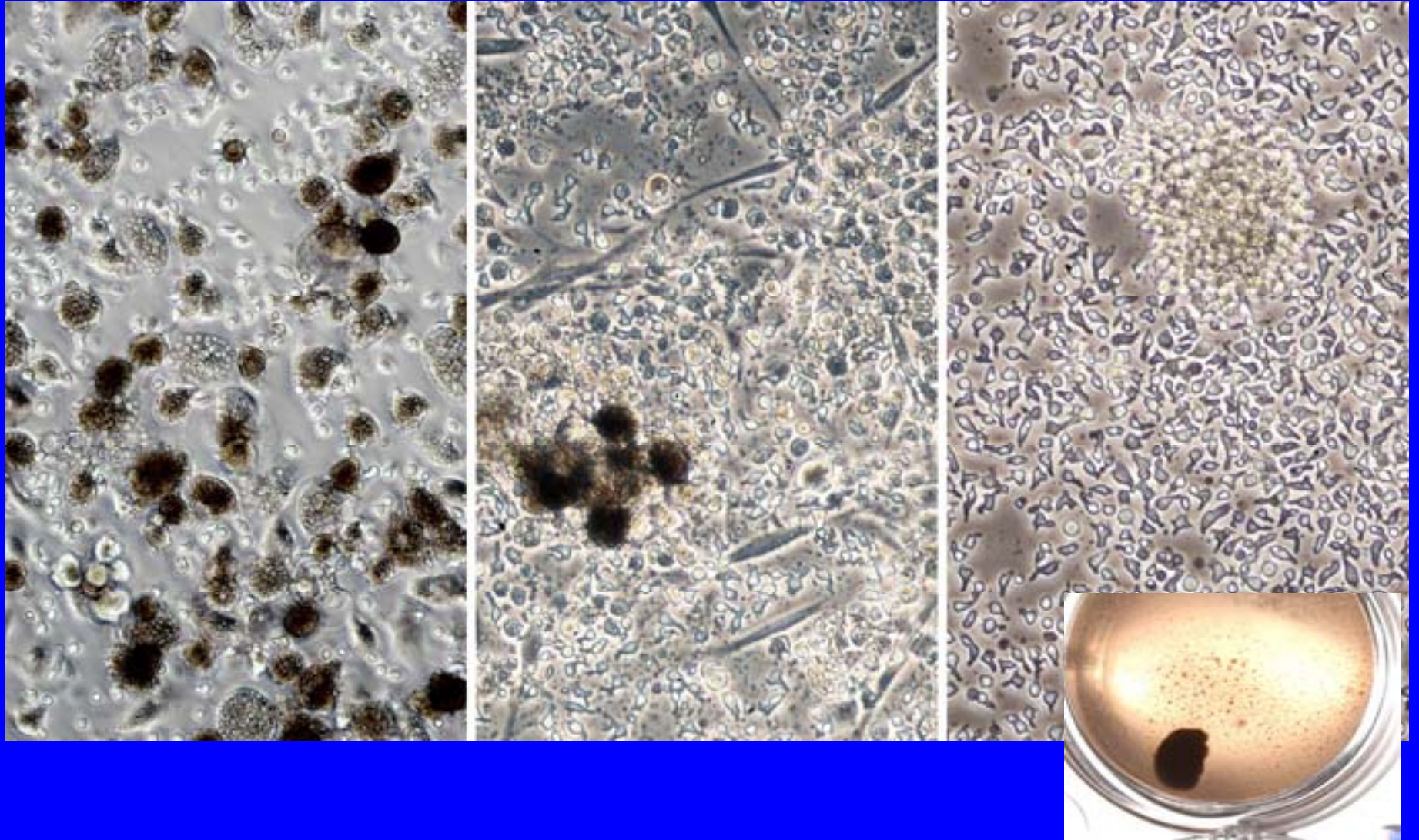
T-Cell-Mediated Tumor Rejection

- Dozens of human tumor-associated proteins which provoke T-cell responses have been discovered, many by studying the T-cells that infiltrate melanoma (TIL; tumor infiltrating lymphocytes)
- T-cell tolerance has been a major impediment to tumor rejection by these cells
- The tumor microenvironment and other host factors play active roles in regulating T-cell activity

Hypotheses

- Ex-vivo stimulation and expansion of tumor-reactive T-cells might overcome some of these impediments to T-cell function
- Manipulation of the recipient prior to the transfer of ex-vivo expanded T-cells could augment the efficacy of adoptive cellular therapy

Melanoma TIL (Tumor Infiltrating Lymphocytes)



Giving TIL as Therapy

- From 1987-1994, eighty-six patients with metastatic melanoma were given their autologous, unselected, bulk-cultured TIL along with supportive systemic IL-2
- Some received one dose of cyclophosphamide (25 mg/kg) prior to cell transfer
- Twenty-eight had prior high-dose IL-2 therapy

Objective Responses

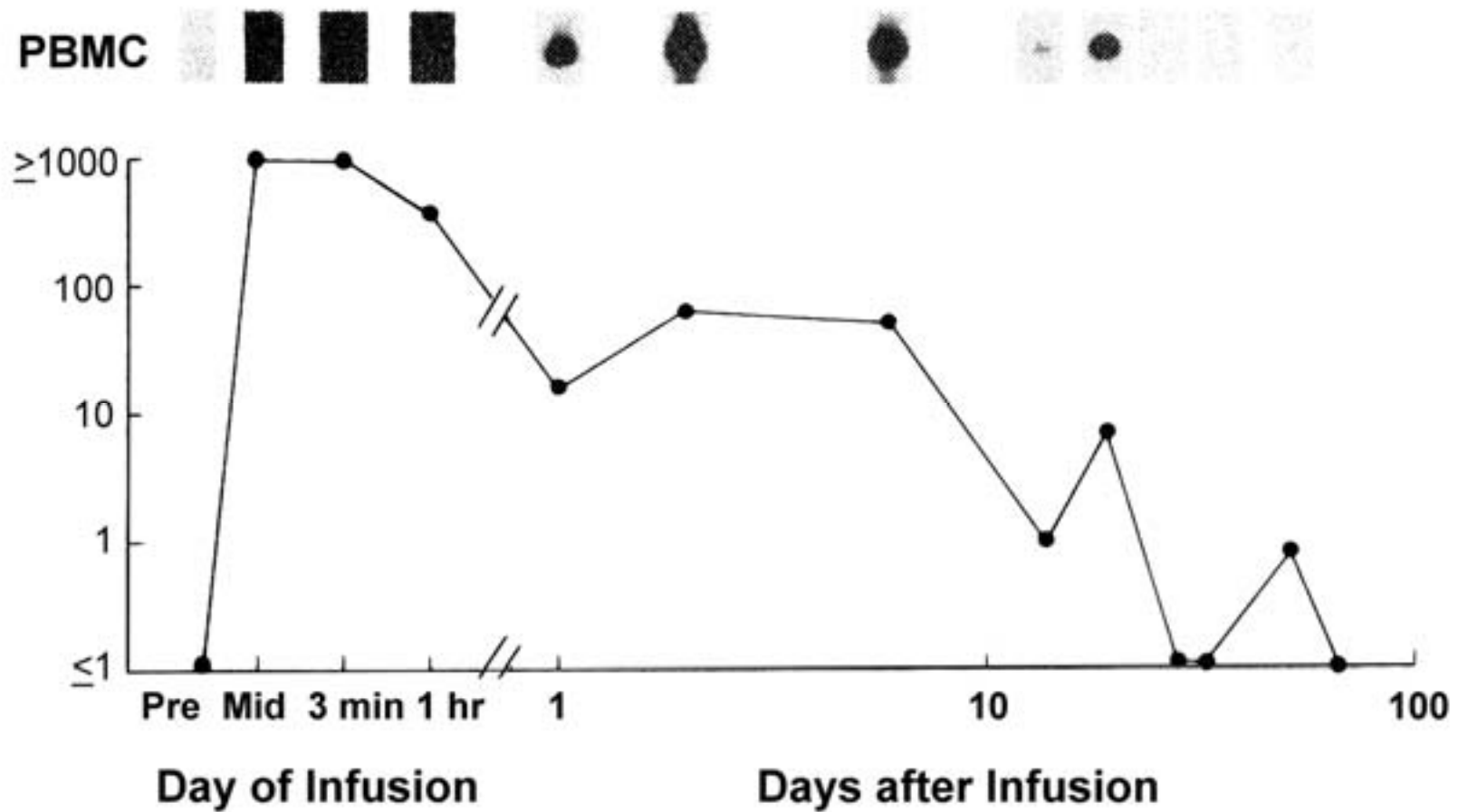
- Objective RR= 32% prior IL-2
 34% no prior IL-2
-
- Response Durations (months):
 - PRs: 53+, 9, 8, 7, 7, 7, 7, 6,
5, 5, 4, 4, 4, 4, 2, 1, 1
 - CRs: 46+, 38, 21+, 23, 20

(5/6 durable or complete responses
were in IL-2 naïve pts.)

Survival/Persistence of TIL in Vivo

- 2×10^{11} was the median number of TIL given
- When genetic marking was done with the neophosphotransferase gene and in vivo survival tracked after infusion, TIL could only be briefly detected by PCR

Gene-Transduced Cells per 10^5 Cells



TIL Recognition of Tumor (IFN-gamma Release Assay)

T-Cells

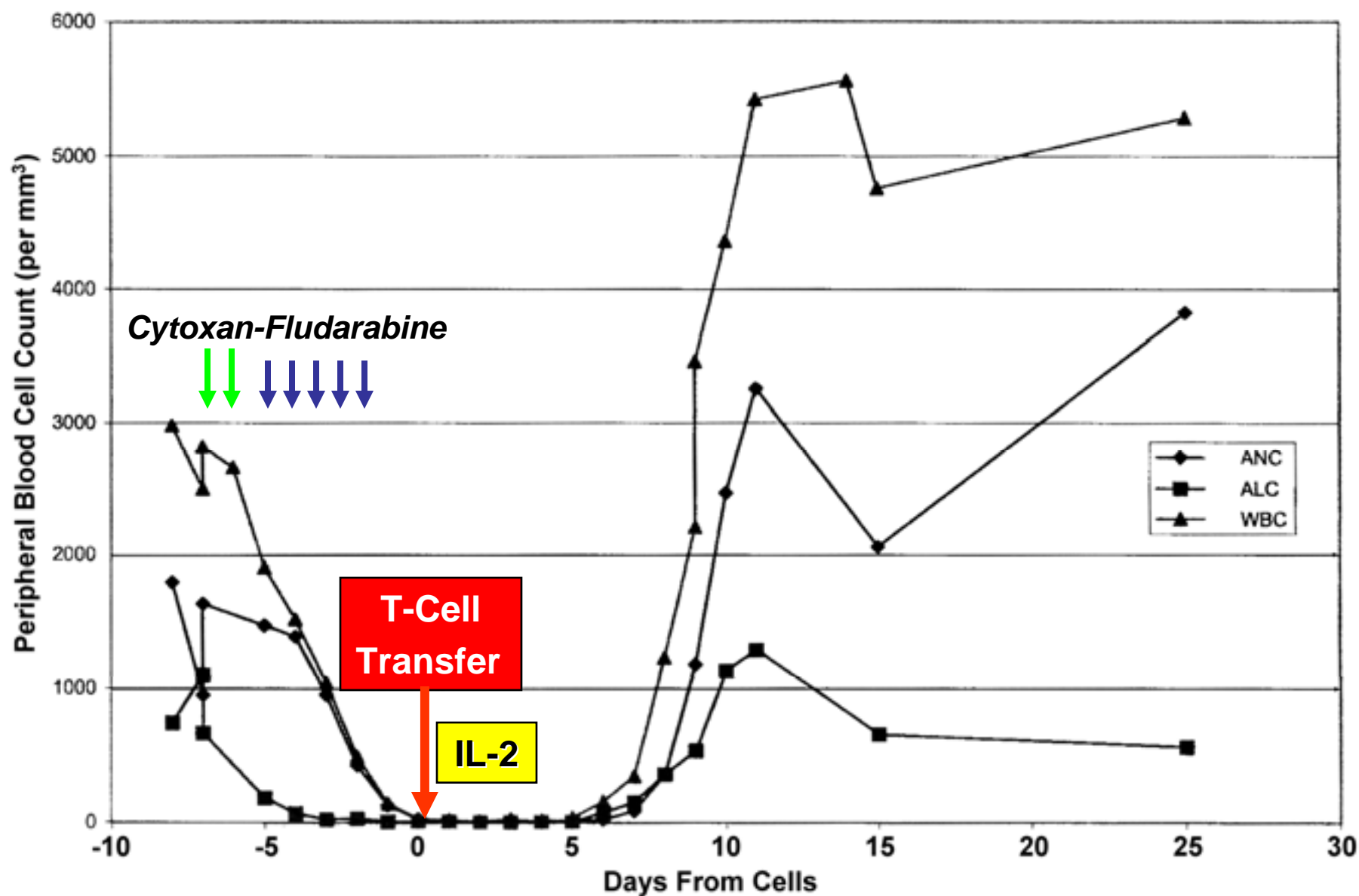
Target Cells

[illegible]

Host Lymphodepletion?

- Animal models demonstrate that transferred lymphocytes proliferate better in an immunodepleted recipient (homeostatic proliferation)
- T-regulatory cells can impede tumor rejection and are reduced by host lymphodepletion prior to adoptive transfer (Antony et al, JI 2005)

Cyclophosphamide + Fludarabine Preparative Regimen

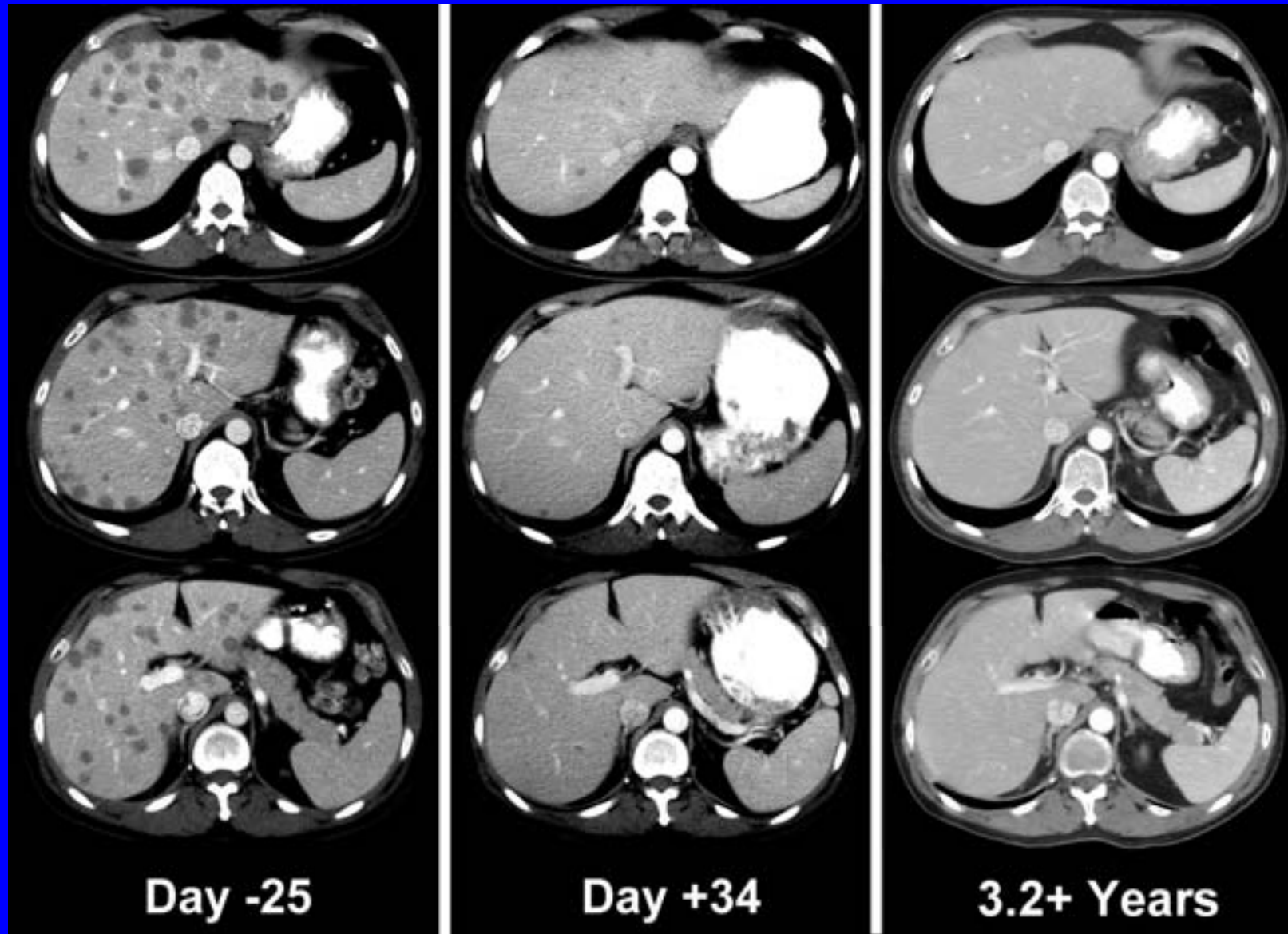


Clinical Trial: Cy-Flu, TIL and IL-2

Characteristics	No.	%
Total patients	35	100
Sex		
Male	21	60
Female	14	40
Age, years		
≤40	10	29
41-50	13	37
51-60	10	29
61-70	2	6
Performance status		
0	29	83
1	6	17
Prior treatments		
Surgery	35	100
Chemotherapy	18	51
Radiotherapy	11	31
Immunotherapy	35	100
Any, ≥ 2	35	100
Any, ≥ 3	22	63

Dudley et al, JCO 2005

Chemotherapy, TIL Transfer and IL-2



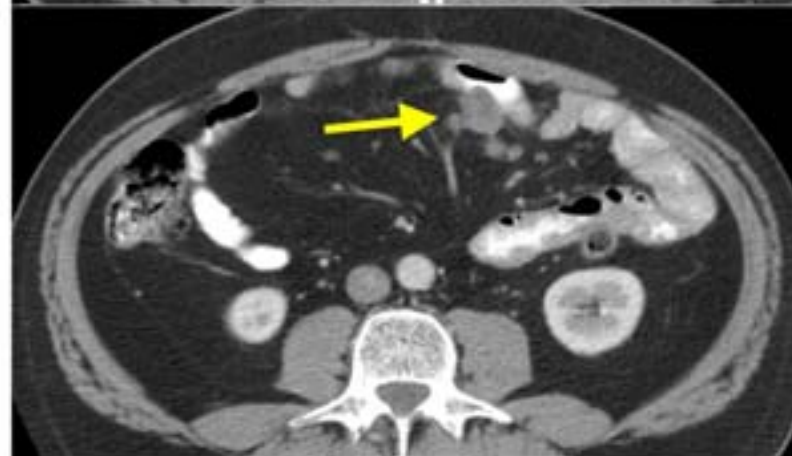
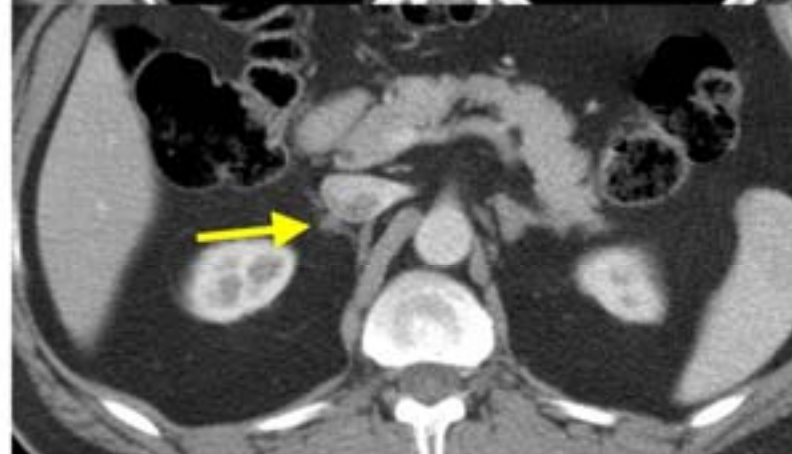
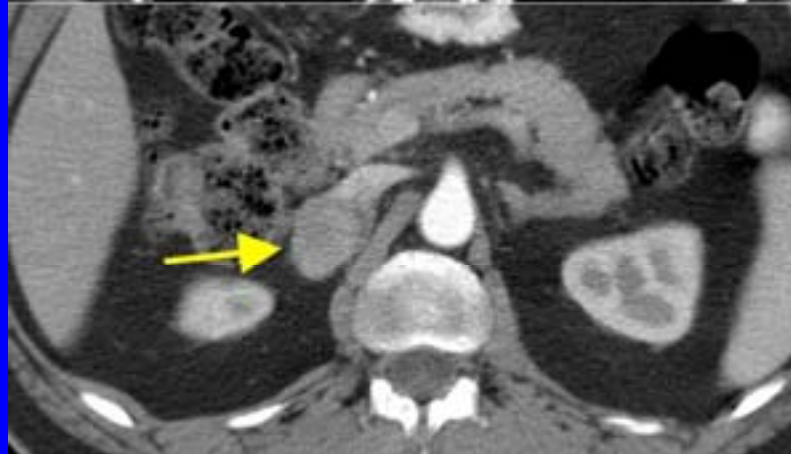
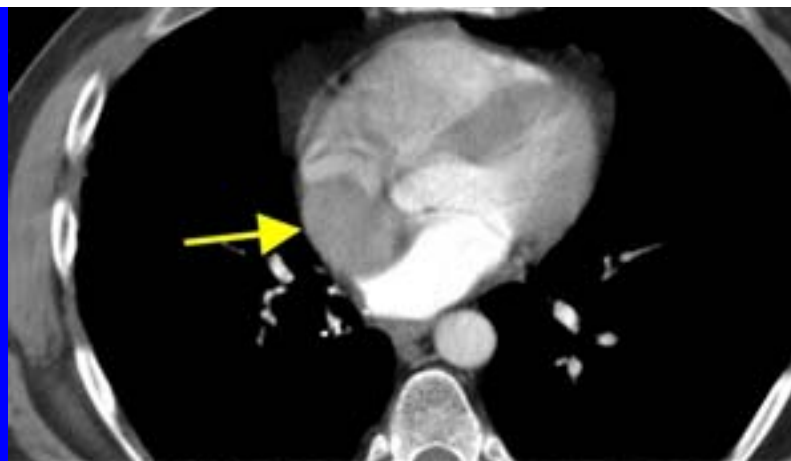
Metastatic Melanoma

Pre-Treatment



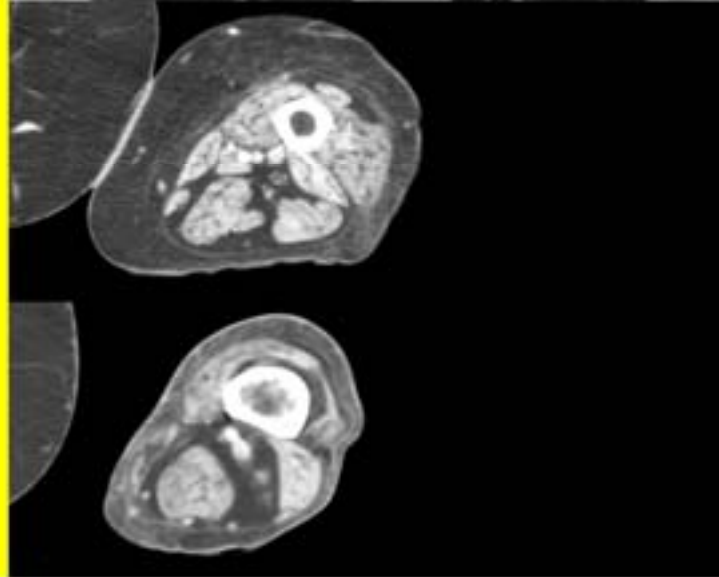
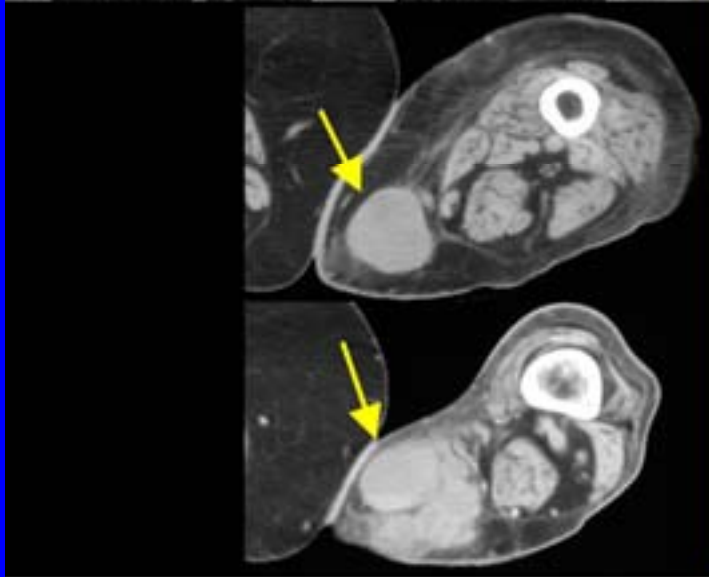
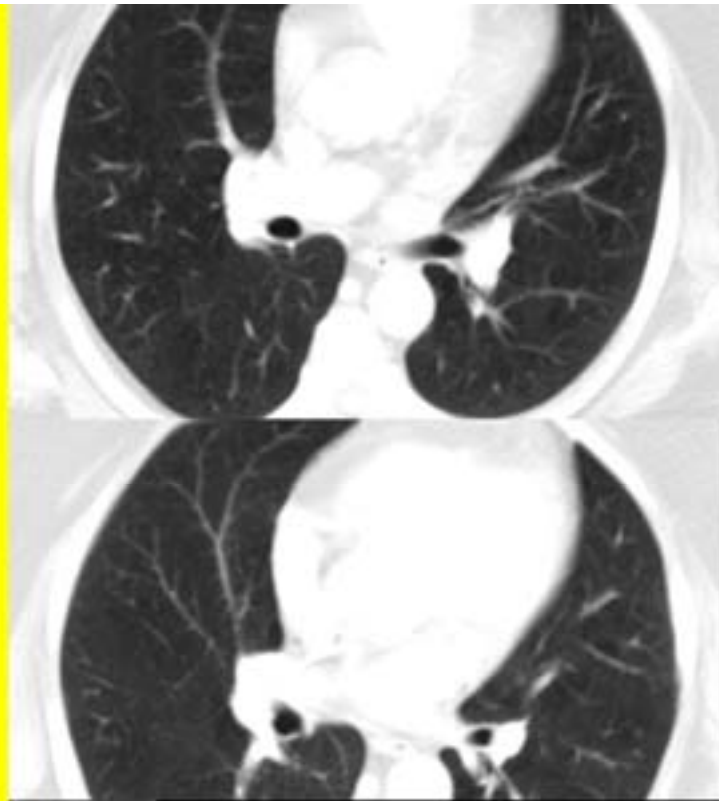
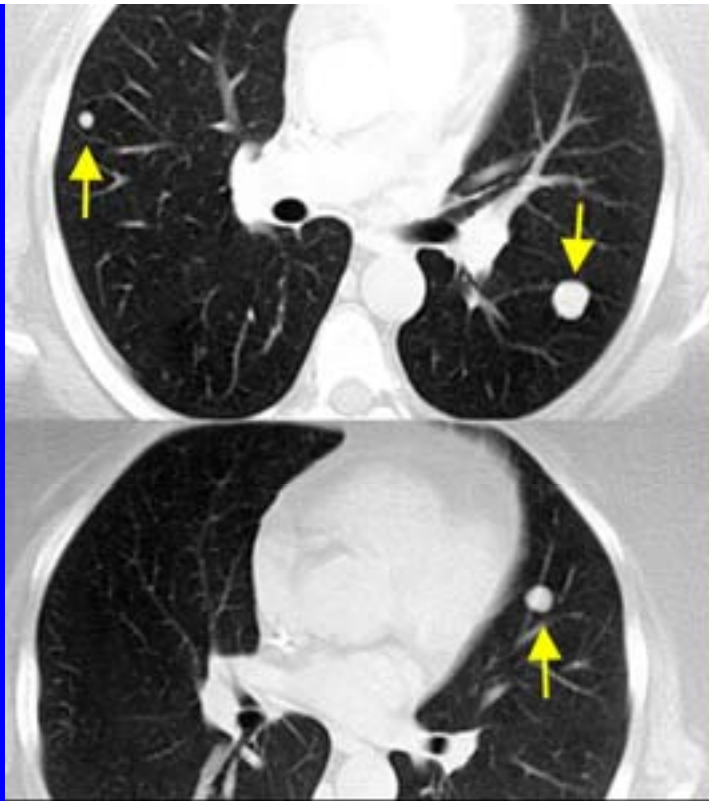
30+ Months





Pre-Treatment

19 Months



Pre-Treatment

38+ Months

Results

- 43 patients treated currently (35 reported)
- 41 had previous IL-2
- Median number of TIL given 5×10^{10}
- Response Rate = 49% (17 PR, 4 CR)

Response Durations: Cy-Flu, TIL and IL-2 (Months)

- PR: 62+, 30+, 29, 28, 18+, 14, 13, 11, 8, 8, 7, 4, 3, 3, 2, 2, 2
- CR: 61+, 56+, 46+, 45+
- Median duration= 13 mo

Persistence of T-Cell Clones After TIL Transfer Into a Lymphodepleted Patient

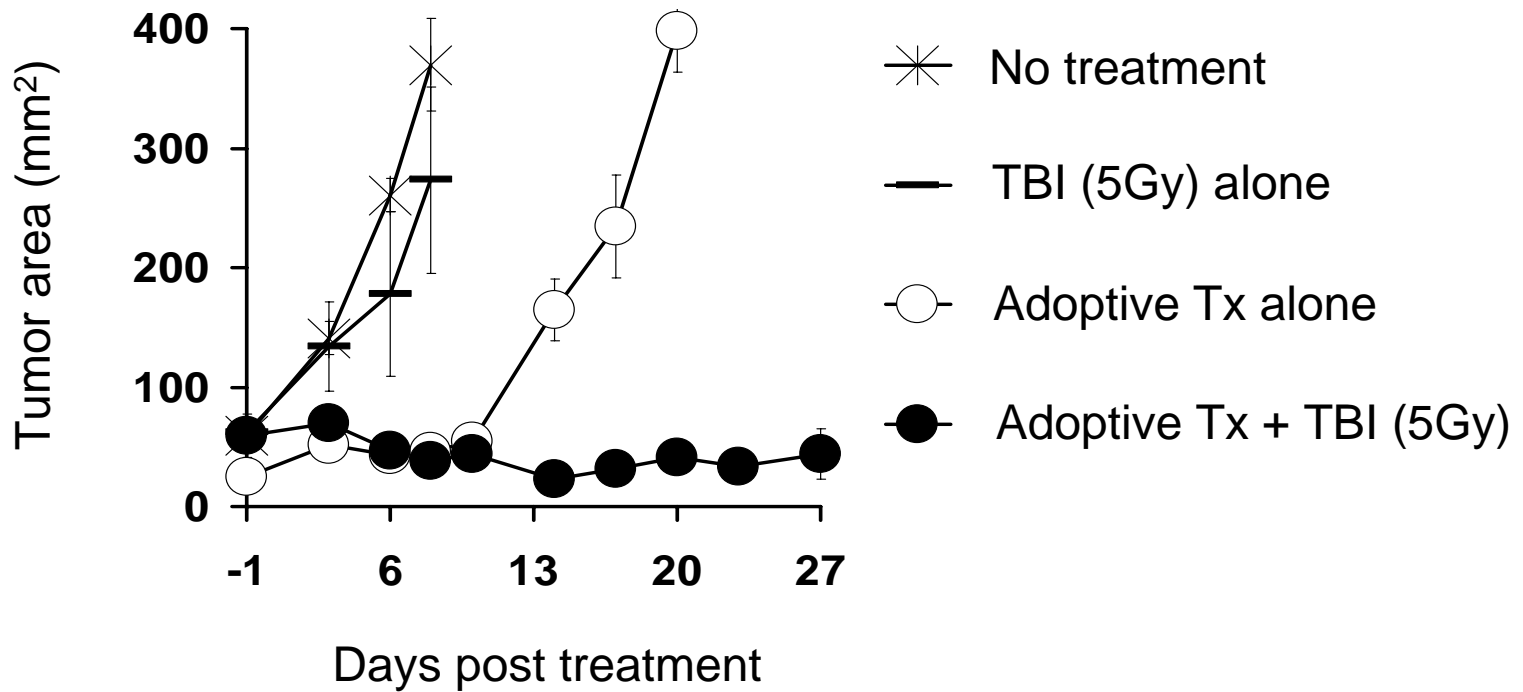
TCR BV <u>Genes</u>	<u>TIL</u>	<u>Day 9</u>	<u>Day 19</u>	<u>Day 46</u>
	(% of lymphocytes)			
BV3	--	--	--	6
BV4-1	--	--	3	--
BV6-4	23	5	--	--
BV6-6	--	--	6	--
BV7-6	--	--	--	3
BV7-9	--	--	--	3
BV10-3	34	69	88	88
BV20-1	21	8	--	--
BV27	5	12	--	--
BV29-1	2	--	--	--
BV30	15	3	--	--
<hr/>				
CD8+tetramer+	89	n.d.	60	n.d.

Augmentation of Adoptive T-Cell Transfer by Host Immunosuppression

Hypotheses Based on Murine Modeling

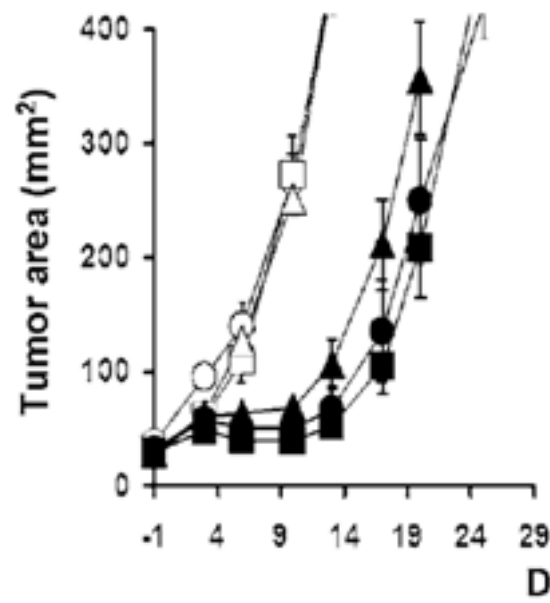
- Elimination of cytokine “sinks”
- Elimination of T reg Cells
- TLR stimulation via enteric organisms

Enhanced Function of Adoptively Transferred pmel-1 T-cells after Total Body Irradiation (TBI)

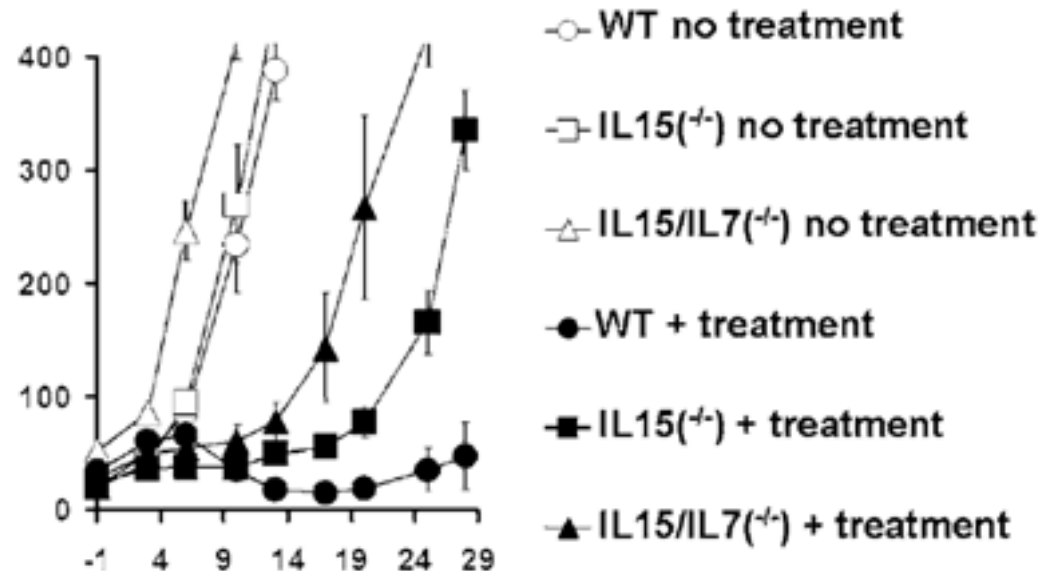


IL-7 and IL-15 Contribute to the Benefit of Host Irradiation

a Nonirradiated host



b Irradiated host



Revised TIL Protocol Adding 200cGy TBI

- Same dose of cyclophosphamide and fludarabine with 200cGy TBI added just prior to cell transfer
- Autologous g-CSF-mobilized, purified CD34+ stem cell support given one day after T-cell transfer

	Cy	Cy								
	Flu	Flu	Flu	Flu	Flu	TBI	TIL	CD34		
							IL-2	IL-2	IL-2	
Day	-6	-5	-4	-3	-2	-1	0	1	2	3



Pre-Treatment

21+ Months

Pretreatment



31 Months





Pre-Treatment



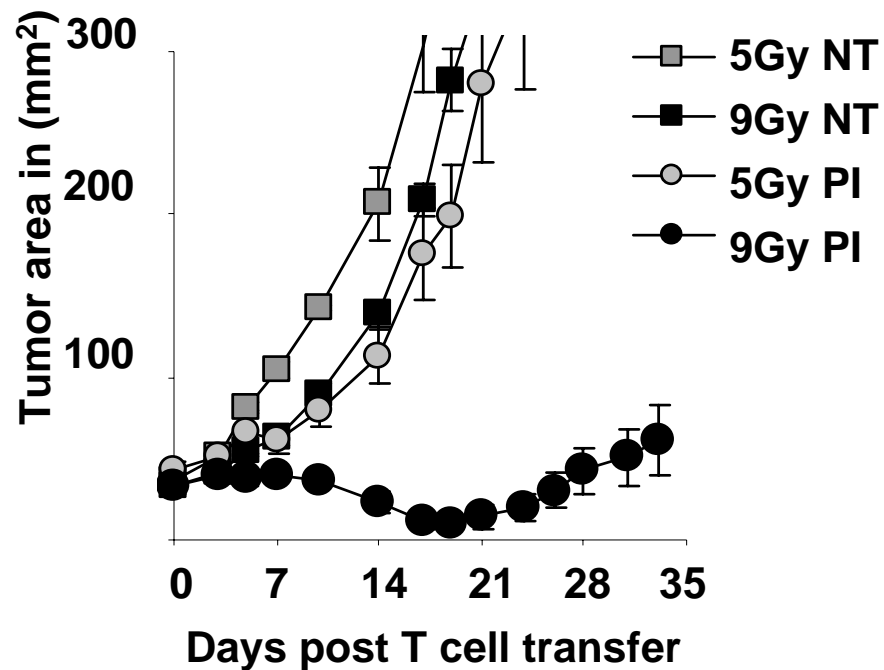
12 Days after Cell Transfer



TIL After Cy+Flu+200cGy TBI

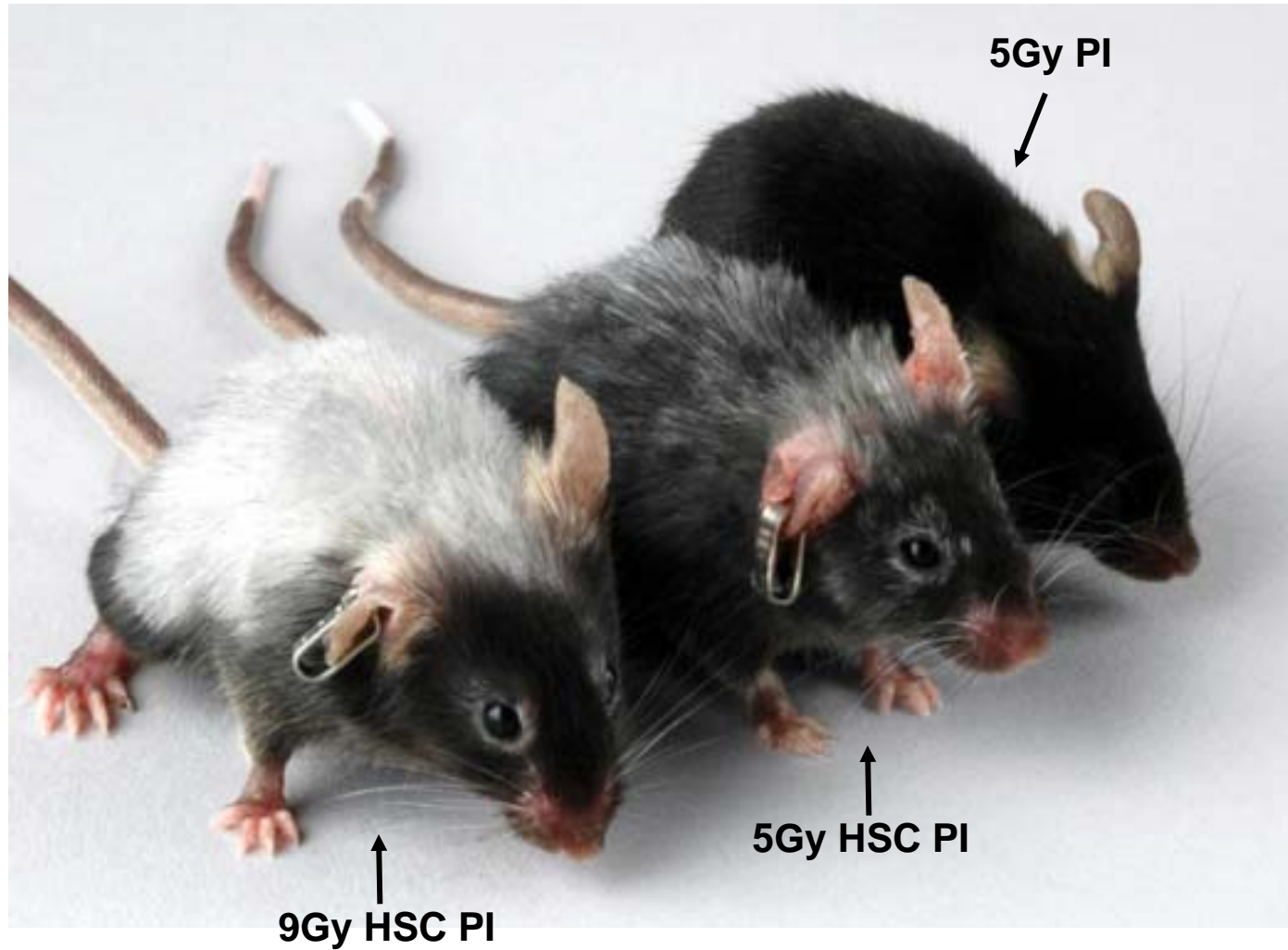
- 25 patients treated; median number of TIL given= 5×10^{10}
- RR= 52% (13/25)
- Response Durations (months):
 - CR: 35+, 23+
 - PR: 31+, 27+, 21+, 14, 10, 6,
5, 5, 4, 3, 3

900 cGy TBI + Hematopoietic stem cells (HSC)
significantly improves tumor response in the
absence of a vaccination



P: 1e6 1 week cultured pmel-1 transgenic T cells;
I: IL-2 (100K CU/bid x 3d).
9Gy irradiated animal received a HSC transplant

Induction of profound vitiligo after 9Gy TBI and HSC transplantation in the absence of vaccination



TIL with Cy+Flu+1200cGy TBI

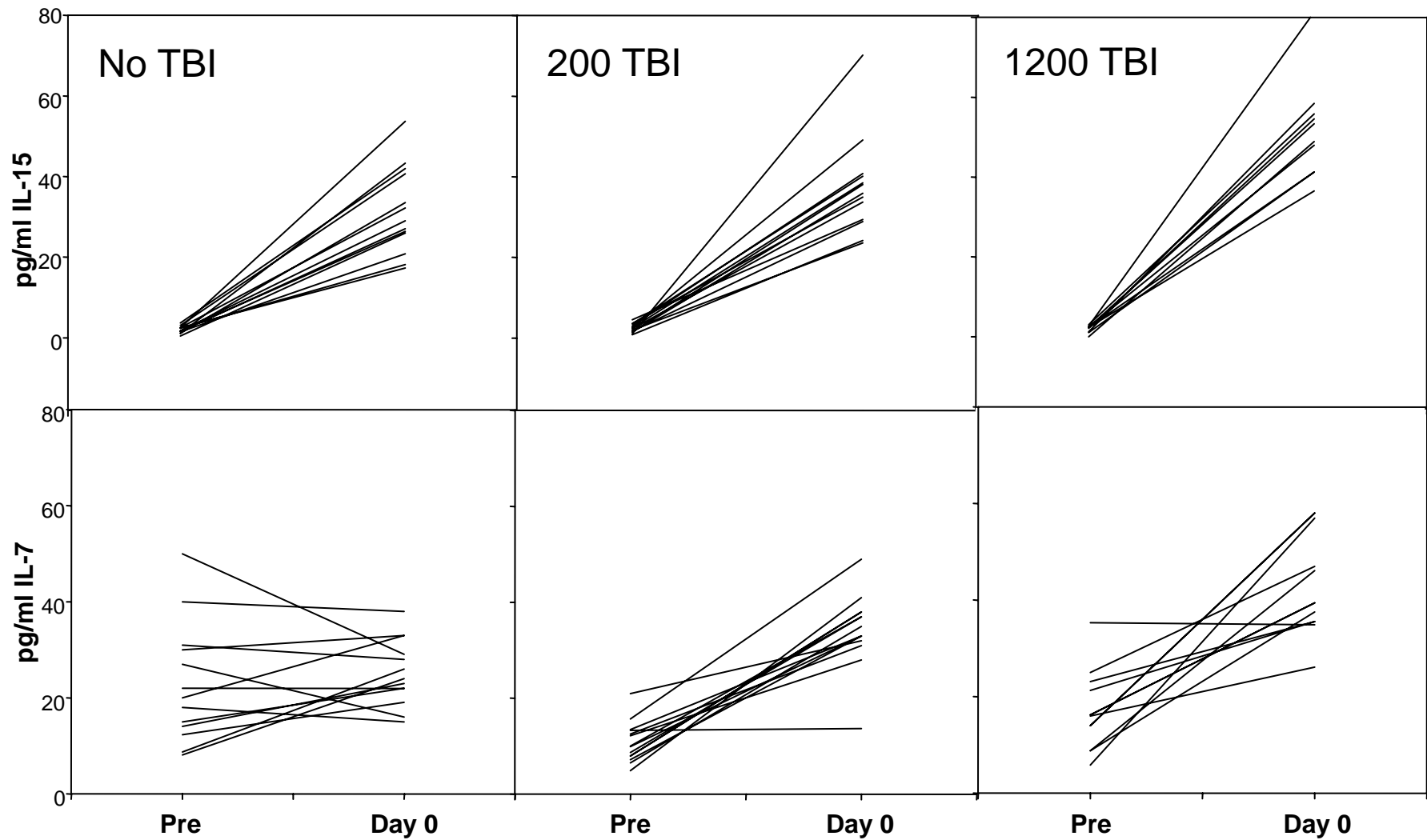
Cy	Cy										
Flu	Flu	Flu	Flu	Flu							
					TBI	TBI	TBI				
								TIL	CD34		
								IL-2	IL-2	IL-2	

Day	-7	-6	-5	-4	-3	-2	-1	0	1	2	3
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Grade 3/4 Toxicities in TIL/TBI Protocols

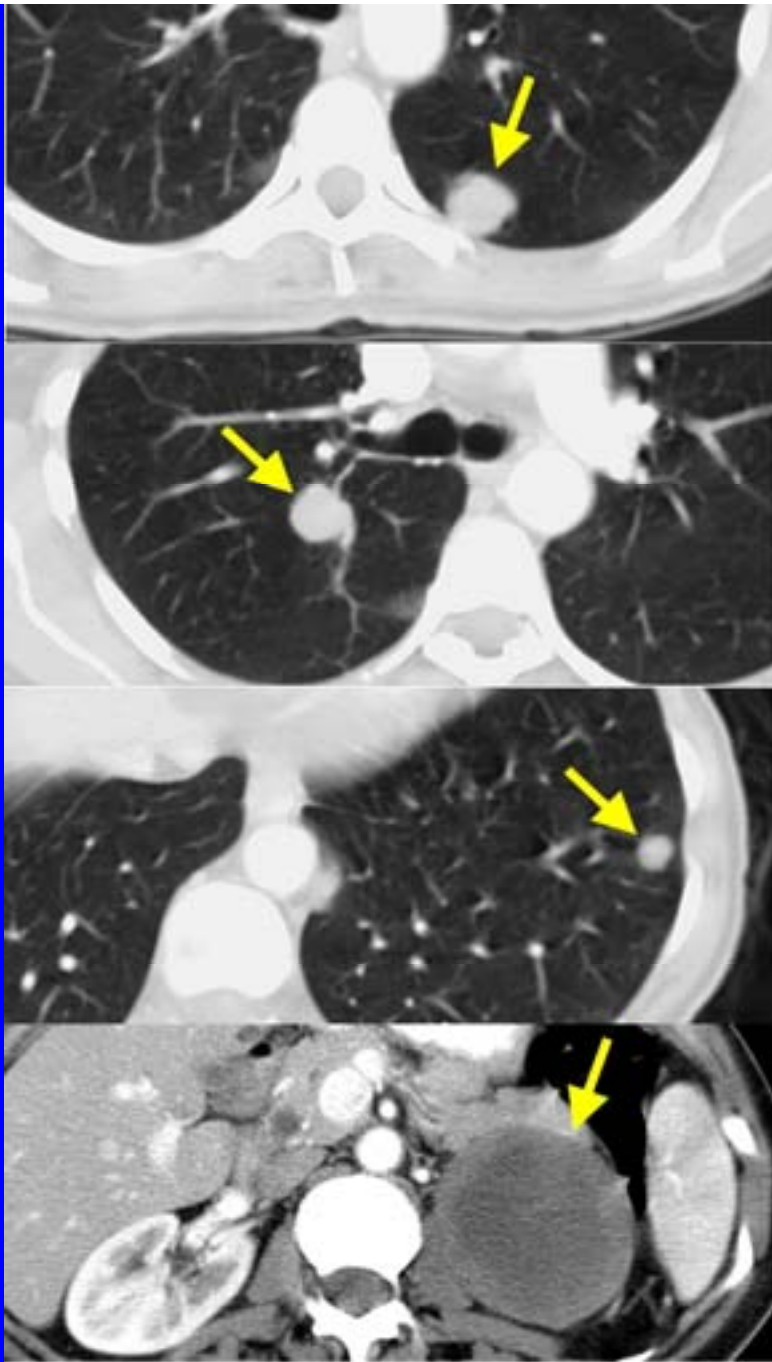
	200 TBI (n = 25)		1200 TBI (n = 18)	
	Grade 3	Grade 4	Grade 3	Grade 4
	(number of patients)			
Febrile neutropenia	12	0	11	0
Sepsis	2	0	3	0
Pneumonia	3	0	0	0
PTT elevation	6	0	4	0
Hypoalbuminemia	7	0	1	0
ALT/AST increase	2	0	2	0
Alkaline p'tase increase	0	0	11	0
Hyperbilirubinemia	1	0	1	0
Hypocalcemia	3	1	4	1
Hypomagnesemia	2	0	0	0
Hypophosphatemia	4	0	7	0
Hyponatremia	3	0	4	0
Hyperuricemia	0	1	0	2
Somnolence	2	0	4	0
Dyspnea	7	1	3	0
Renal failure	0	1	2	0
Autoimmunity	0	0	1	0
Thrombotic microangiopathy	0	0	1	0
Desquamating rash	0	0	2	0
Death		1		

Impact of Lymphodepletion on Serum Levels Of IL-15 and IL-7



TIL with Cy+Flu+1200cGy TBI : (Preliminary Results)

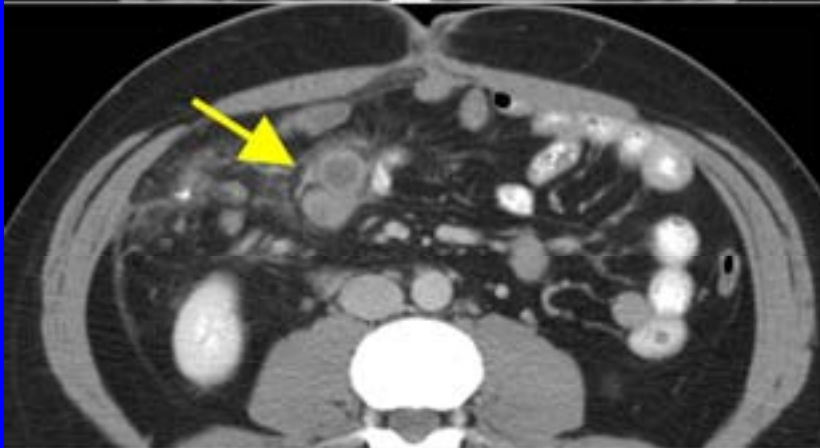
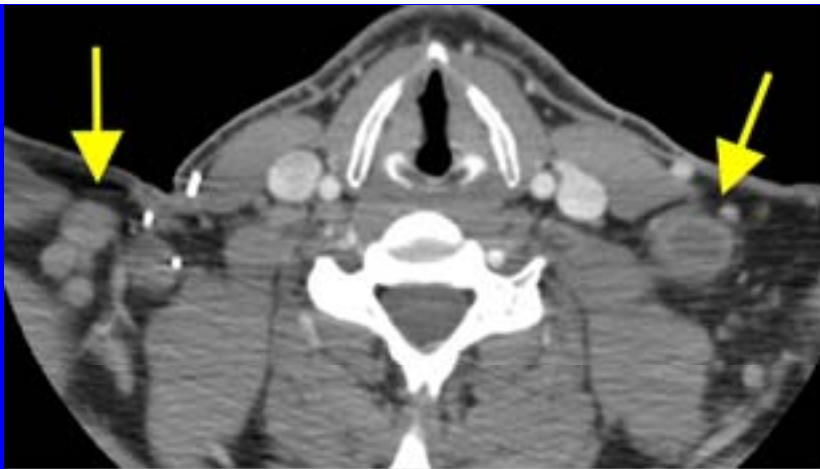
- 25 patients treated
- RR= 72% (18/25)
- Response Durations (months):
 - CR: 15+, 11+
 - PR: 13+, 12+, 11+, 8+, 7, 6, 6, 5+,
5+, 5+, 4+, 4+, 4, 3+, 3, 2+



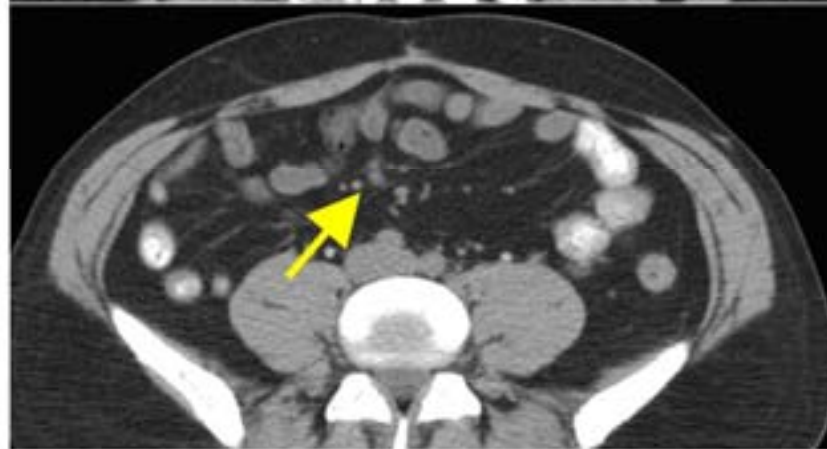
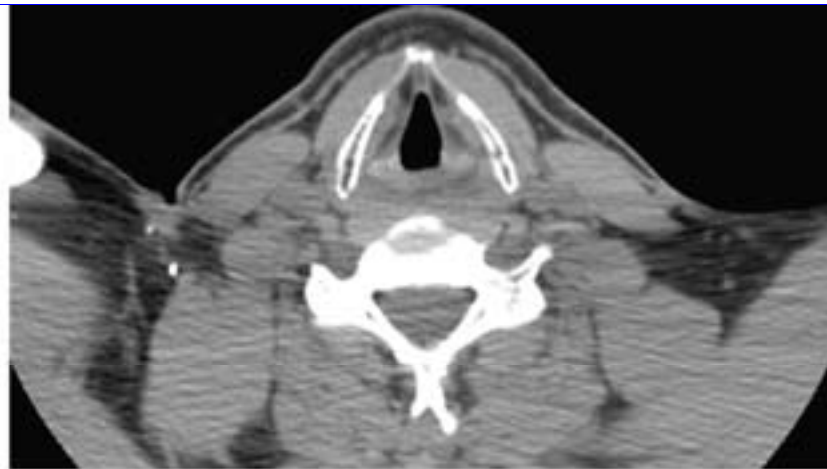
Baseline



11 Months

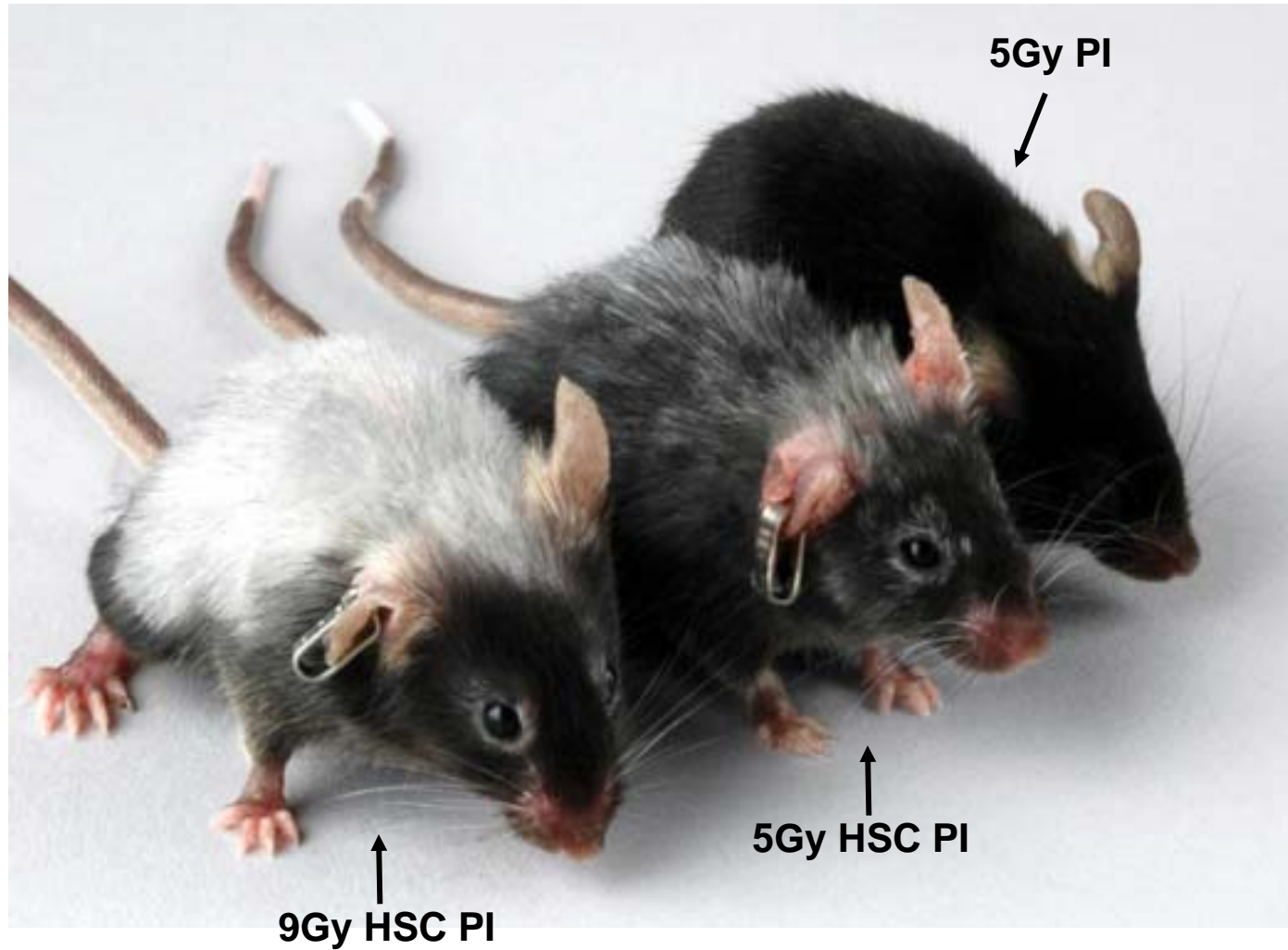


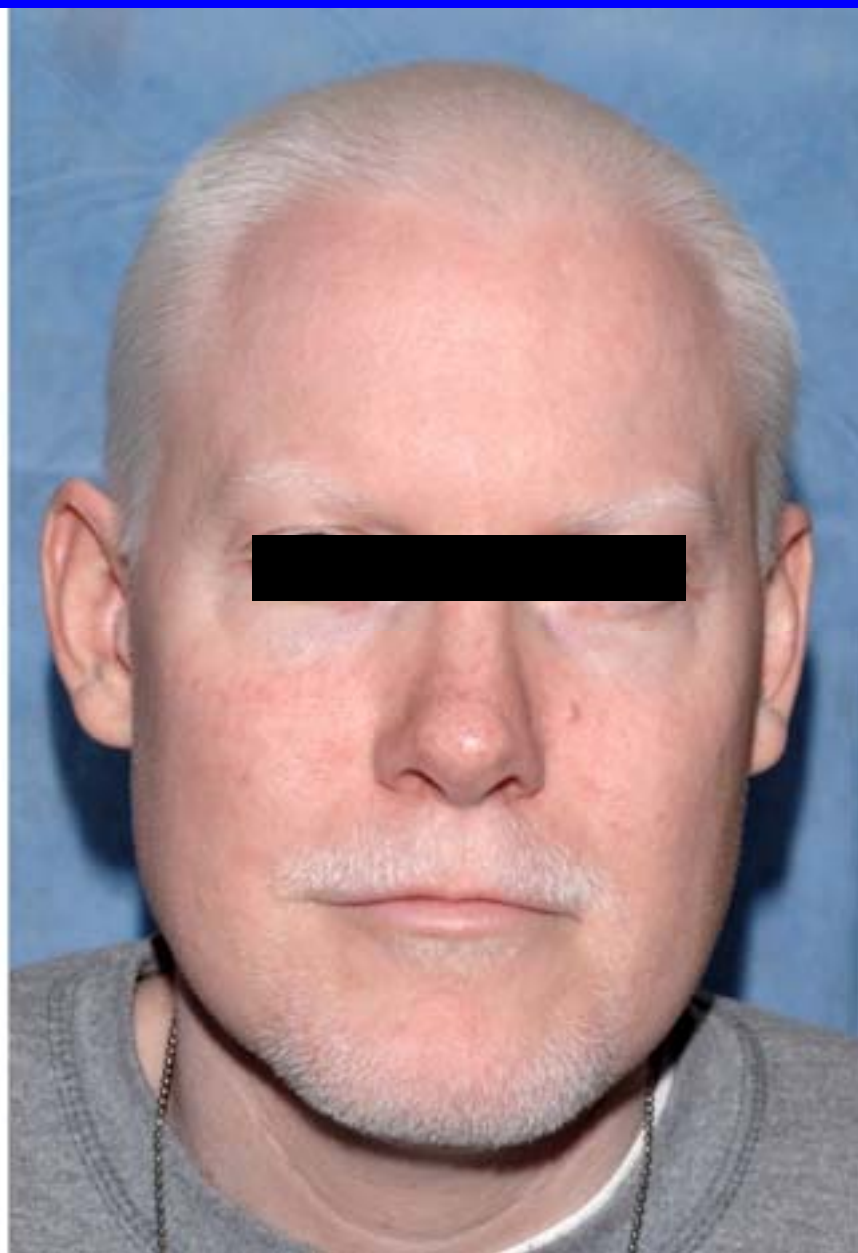
Pre-Treatment

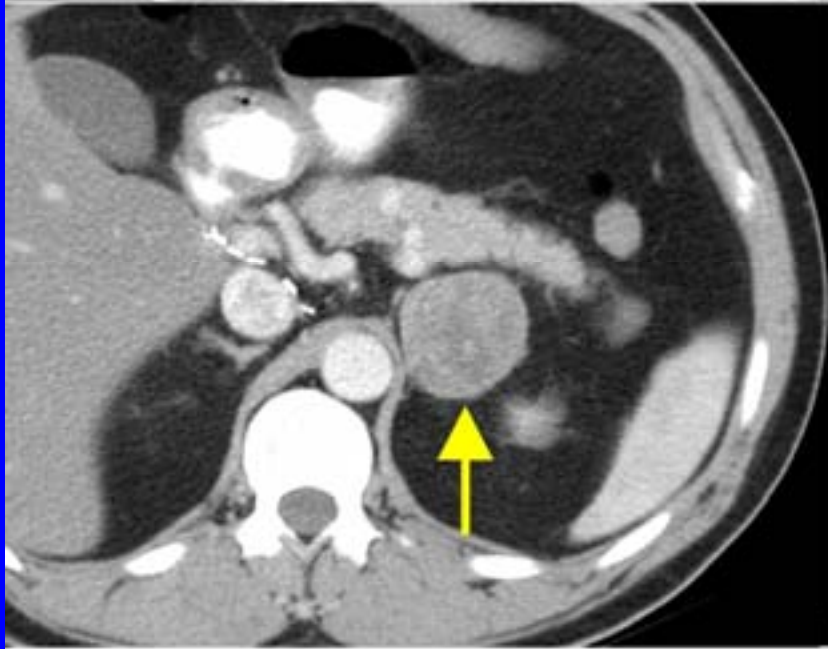
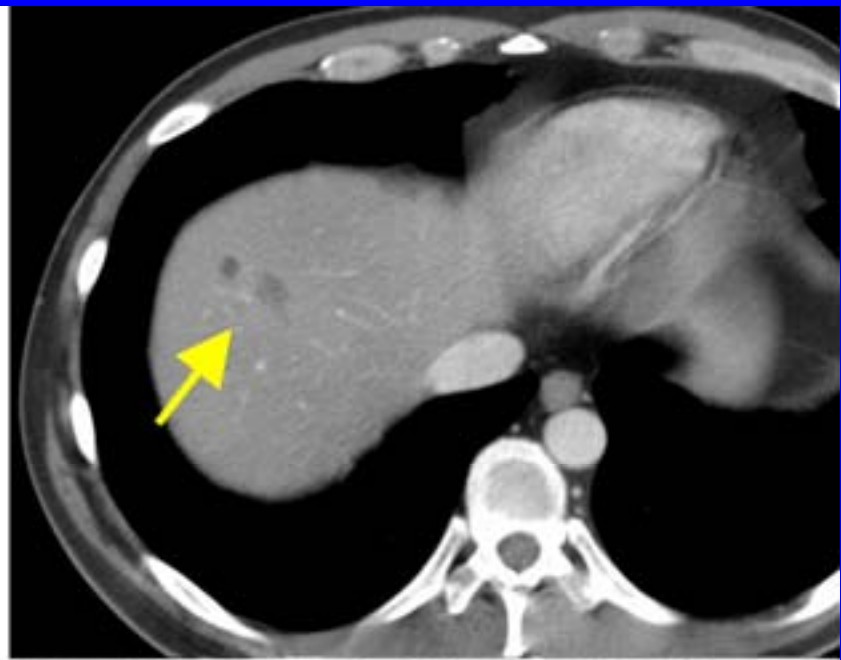
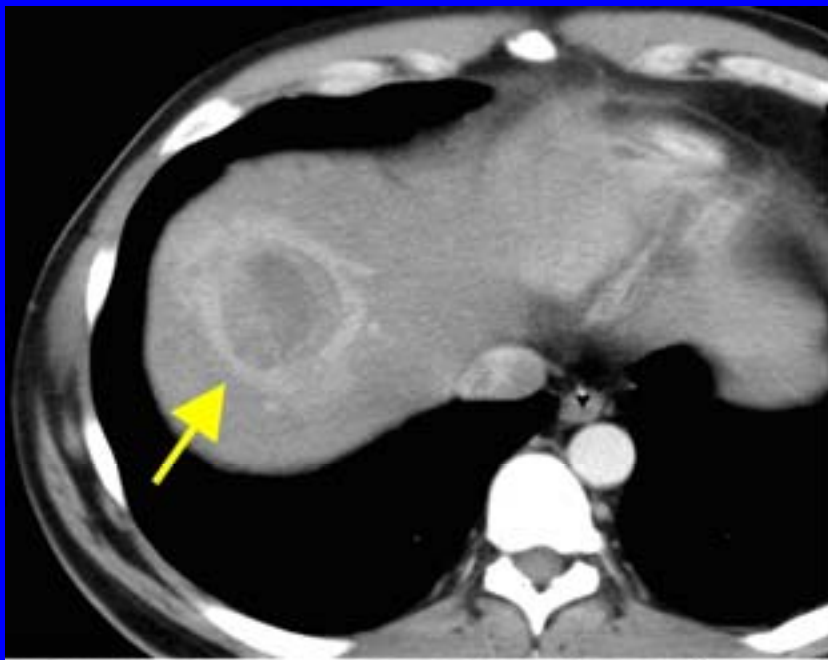


15+ Months

Induction of profound vitiligo after 9Gy TBI and HSC transplantation in the absence of vaccination

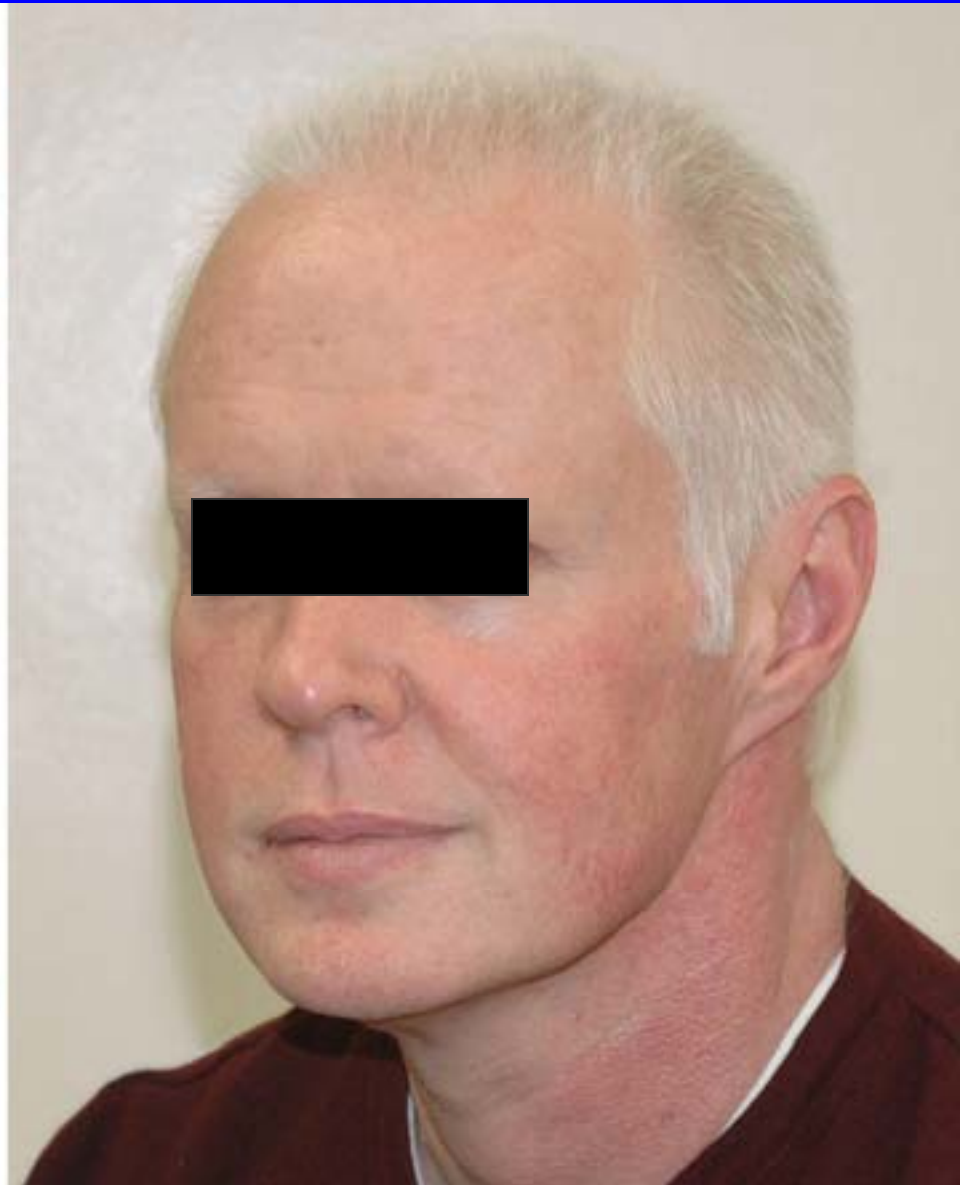




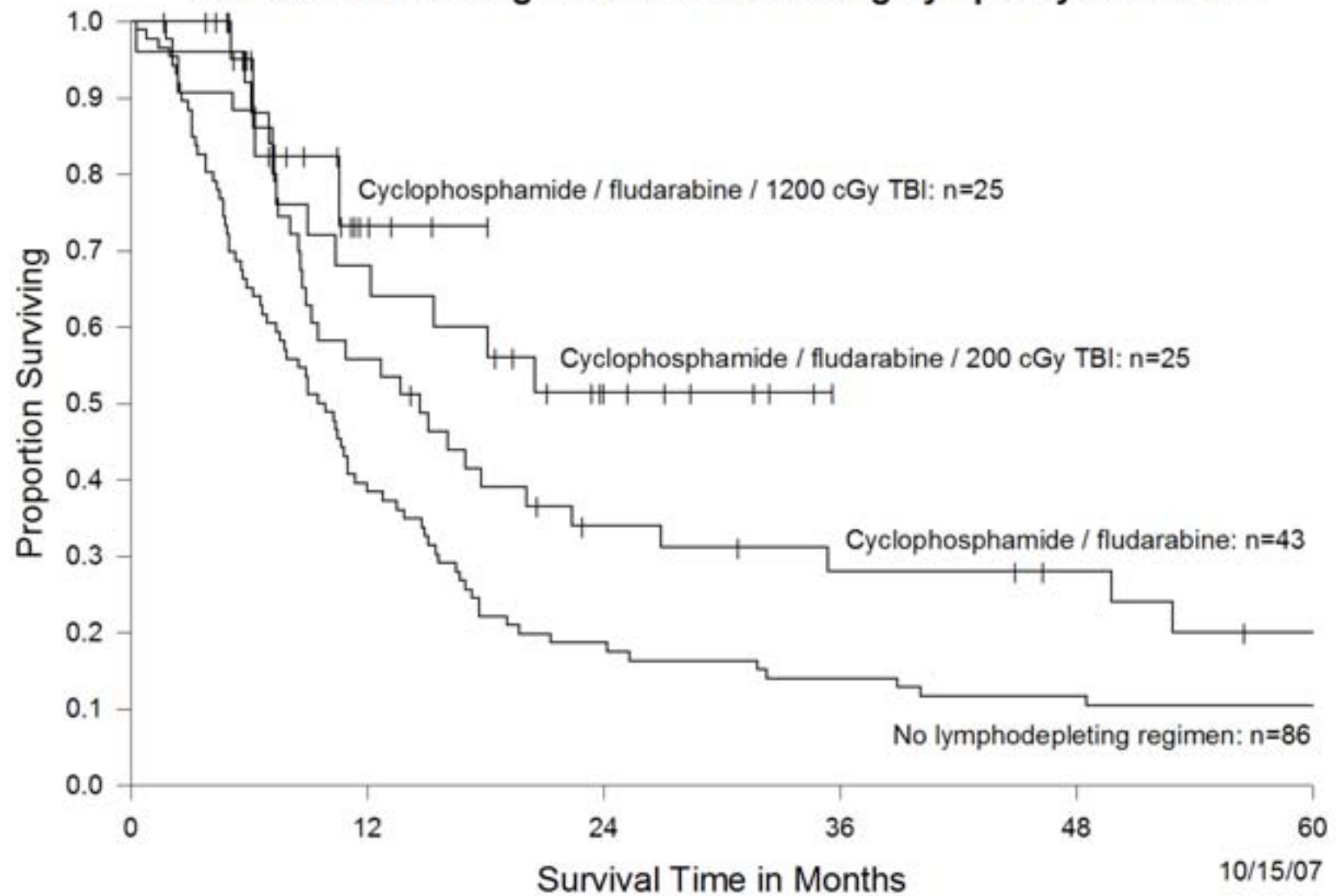


Pre-Treatment

11 Months



**Survival of Patients with Metastatic Melanoma
Treated with Autologous Tumor Infiltrating Lymphocytes and IL-2**



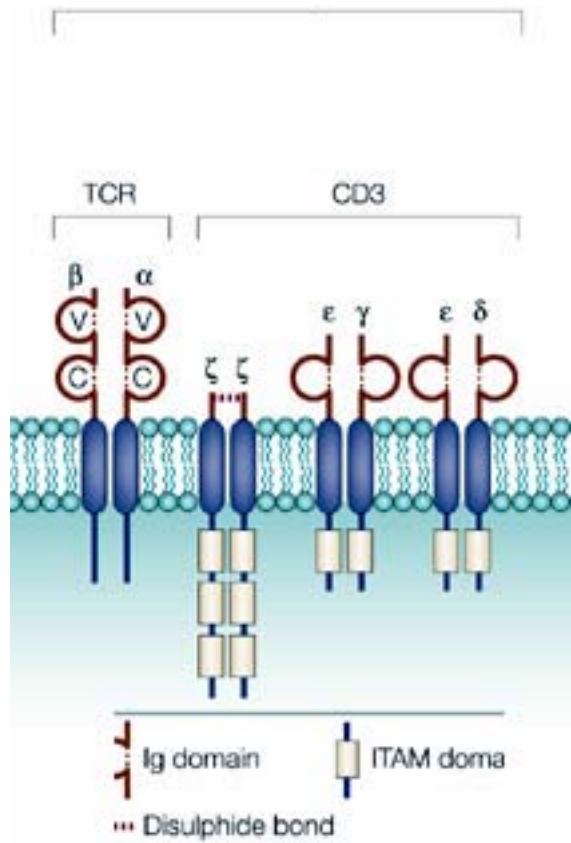
Problems with TIL Therapy

- Some patients cannot grow reactive TIL
- Effective TIL can only be found consistently in melanoma
- Every treatment must be individualized

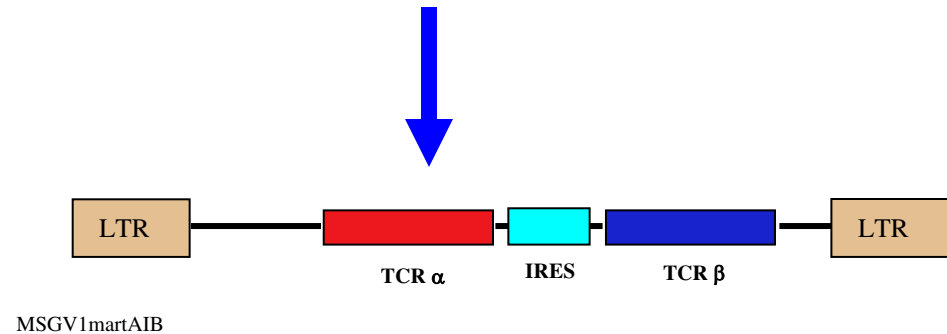
Q: Can effective tumor-reactive T-cells be genetically constructed?

TCR Gene Therapy Trial

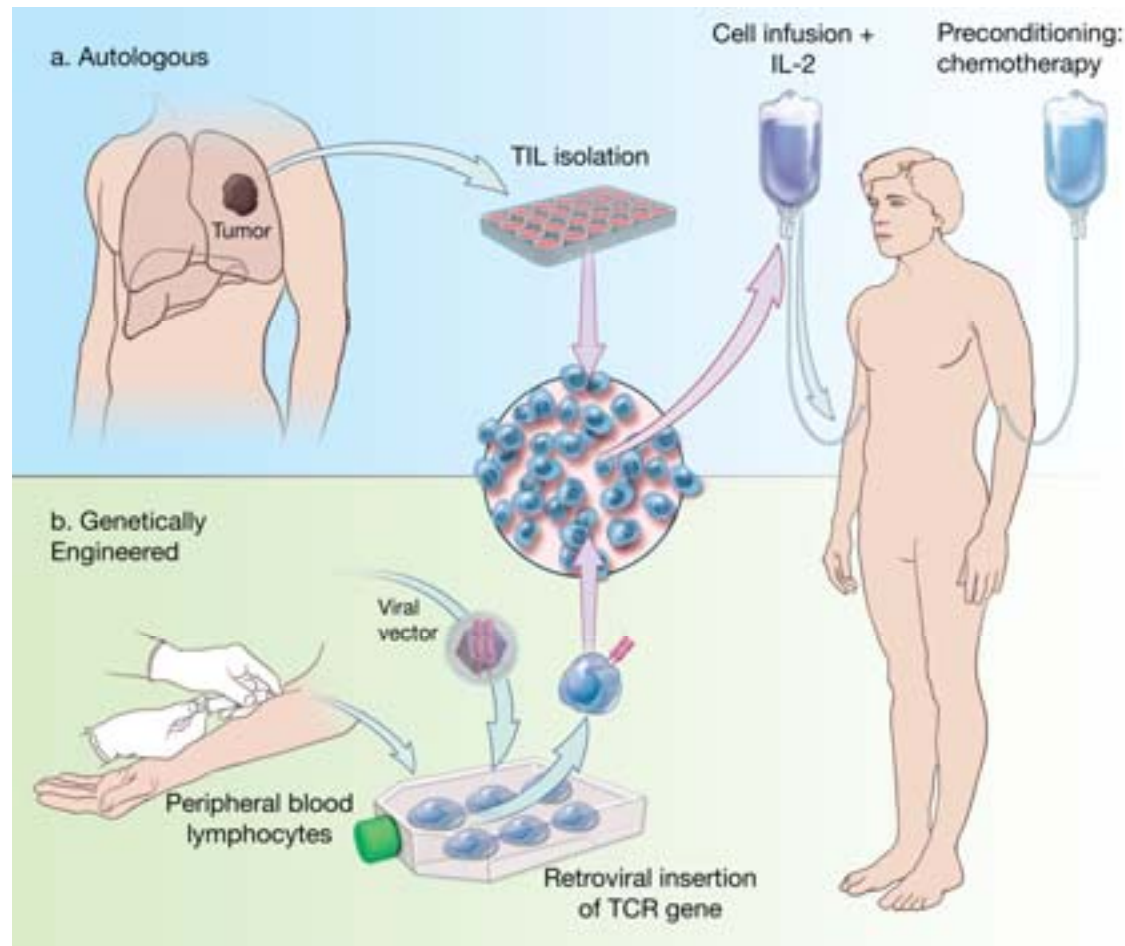
The T-Cell Receptor Complex



Isolate and clone TCR α /b chains into retroviral vector



TCR Gene Therapy Protocol Schema



**Isolate PBL
and stimulate
with OKT-3**

**Day 2: transduce PBL
with retroviral vector
encoding TCR
and culture in IL-2**

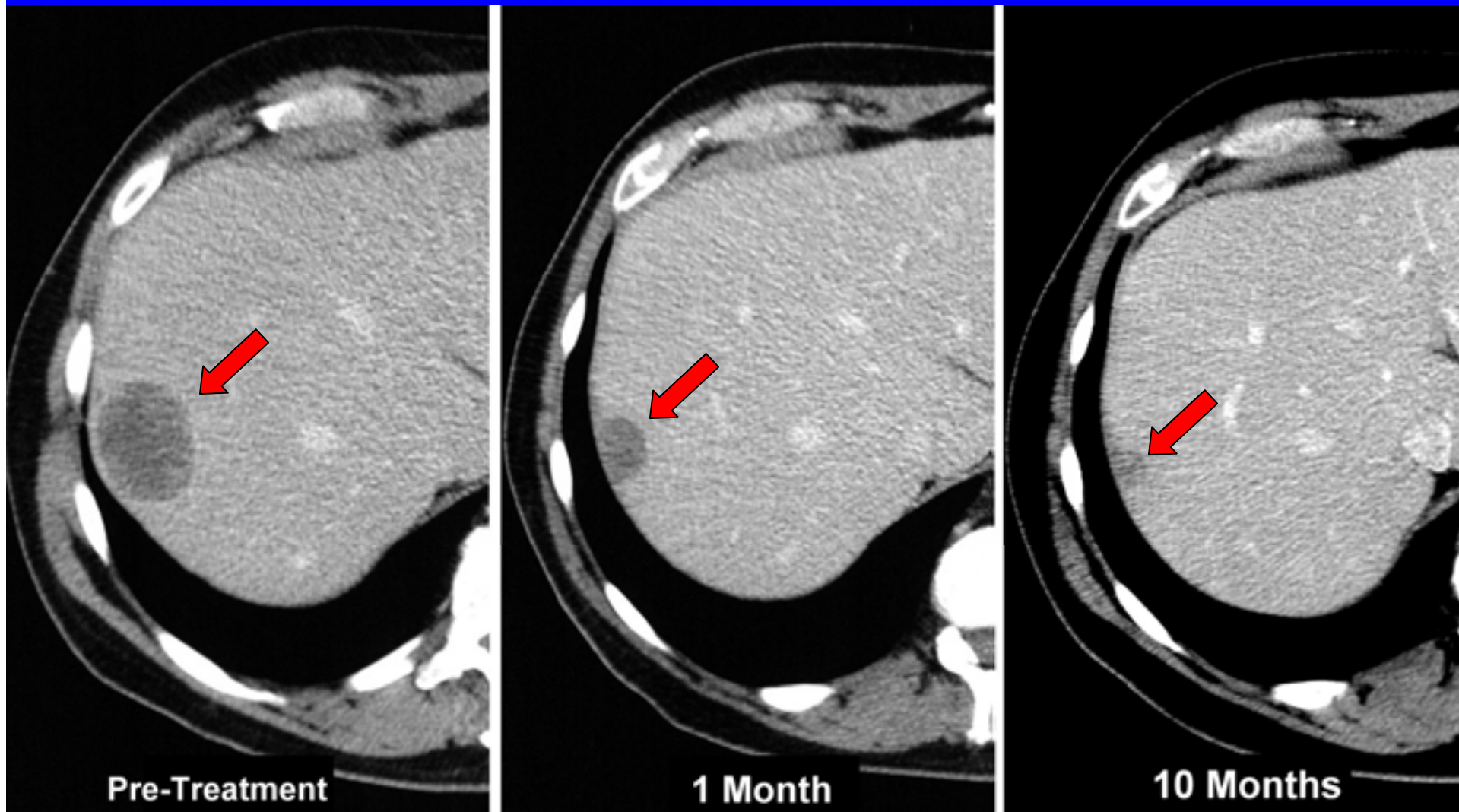
**Infuse transduced cells
after pt. lymphodepletion
instead of TIL**

Cancer Regression in Patients After Transfer of Genetically Engineered Lymphocytes

Richard A. Morgan, Mark E. Dudley, John R. Wunderlich, Marybeth S. Hughes, James C. Yang, Richard M. Sherry, Richard E. Royal, Suzanne L. Topalian, Udai S. Kammula, Nicholas P. Restifo, Zhili Zheng, Azam Nahvi, Christiaan R. de Vries, Linda J. Rogers-Freezer, Sharon A. Mavroukakis, Steven A. Rosenberg*

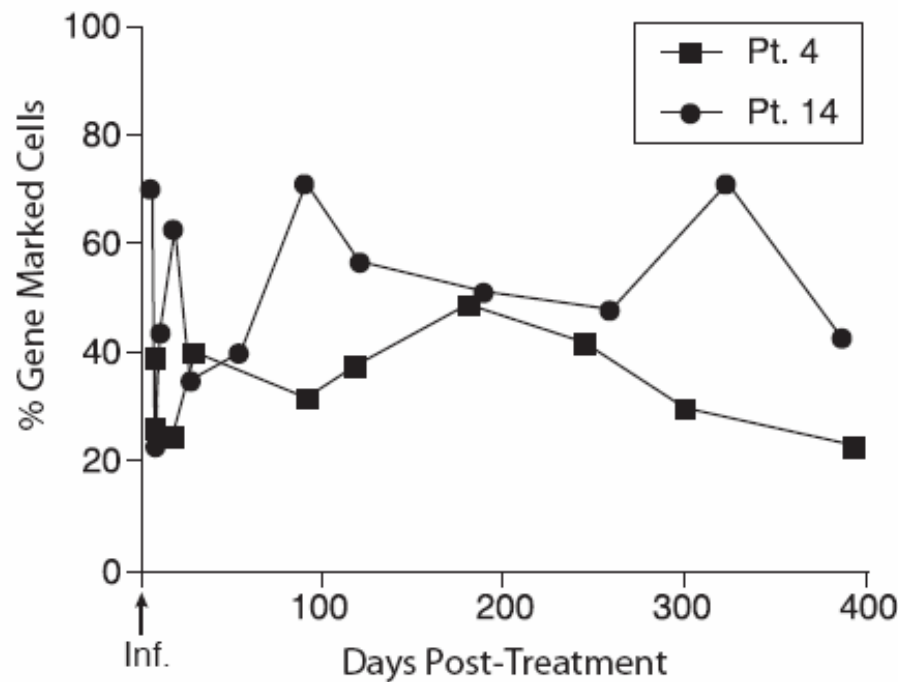
Through the adoptive transfer of lymphocytes after host immunodepletion, it is possible to mediate objective cancer regression in human patients with metastatic melanoma. However, the generation of tumor-specific T cells in this mode of immunotherapy is often limiting. Here we report the ability to specifically confer tumor recognition by autologous lymphocytes from peripheral blood by using a retrovirus that encodes a T cell receptor. Adoptive transfer of these transduced cells in 15 patients resulted in durable engraftment at levels exceeding 10% of peripheral blood lymphocytes for at least 2 months after the infusion. We observed high sustained levels of circulating, engineered cells at 1 year after infusion in two patients who both demonstrated objective regression of metastatic melanoma lesions. This study suggests the therapeutic potential of genetically engineered cells for the biologic therapy of cancer.

Tumor Regression After Receiving TCR-Transduced T Cells

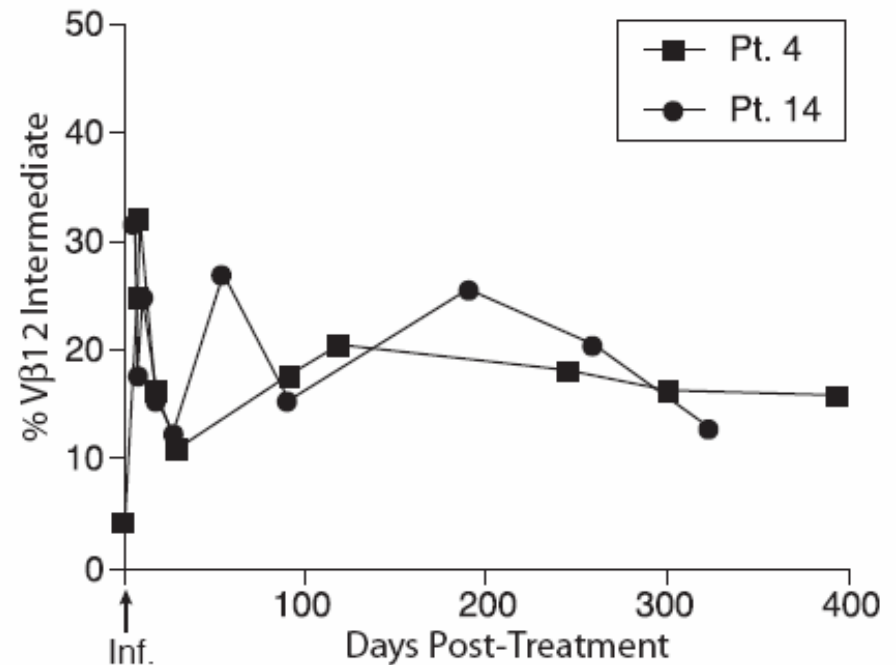




Long-Term Persistence of TCR Transduced Cells in Responding Patients



Quantitative PCR (TaqMan)

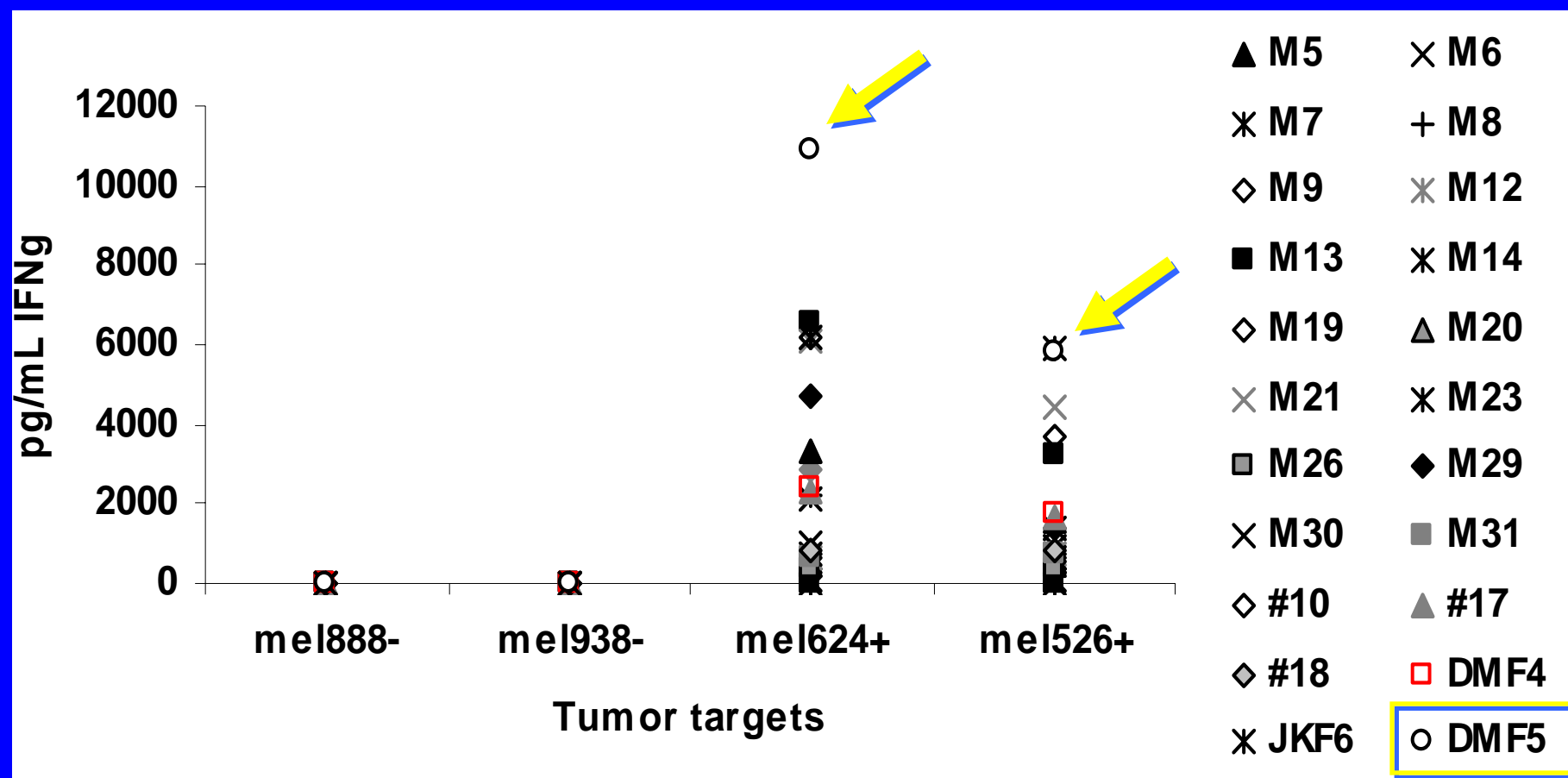


FACS for Vb12

Second Generation TCRs: Higher Avidity

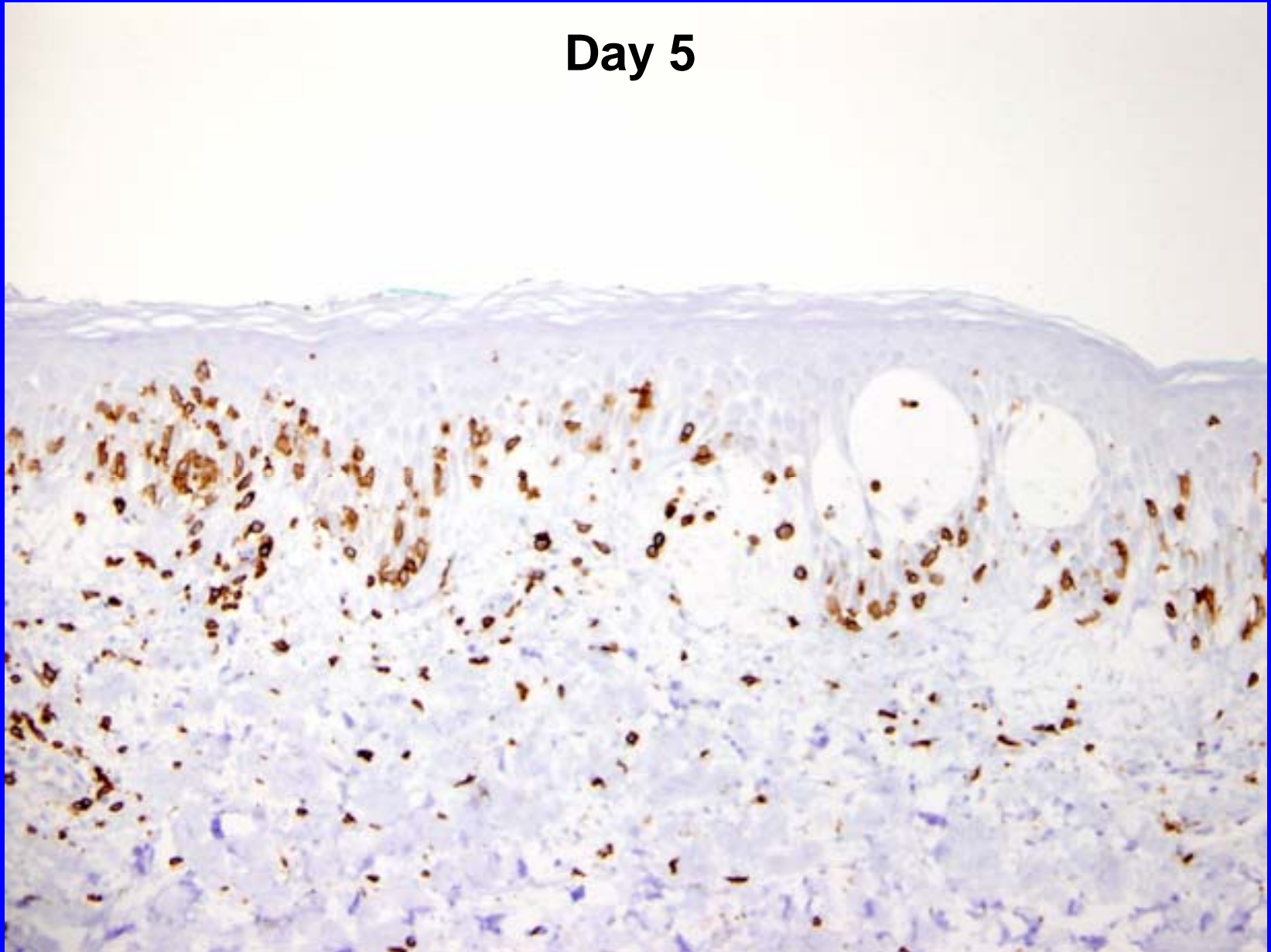
- The MART-1-reactive TCR selected for this study was from a patient who responded to TIL transfer after Cy-Flu and was a clone that persisted in vivo
- Extensive cloning of other MART-1-reactive TIL showed a wide range of TCR avidities

Tumor Recognition of MART-1-Reactive TIL clones: IFN Release



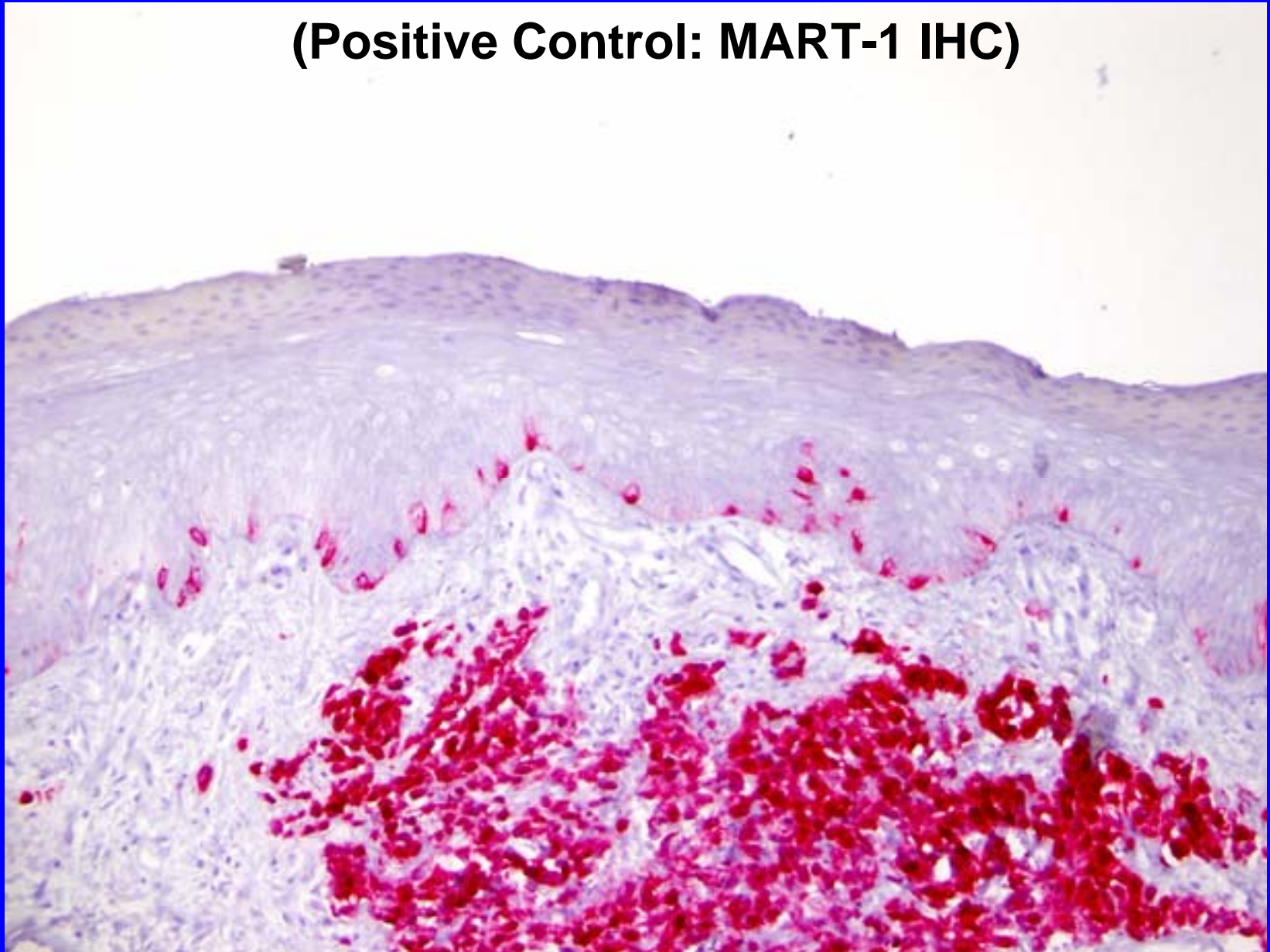
RNA Electroporation of PBL

Day 5

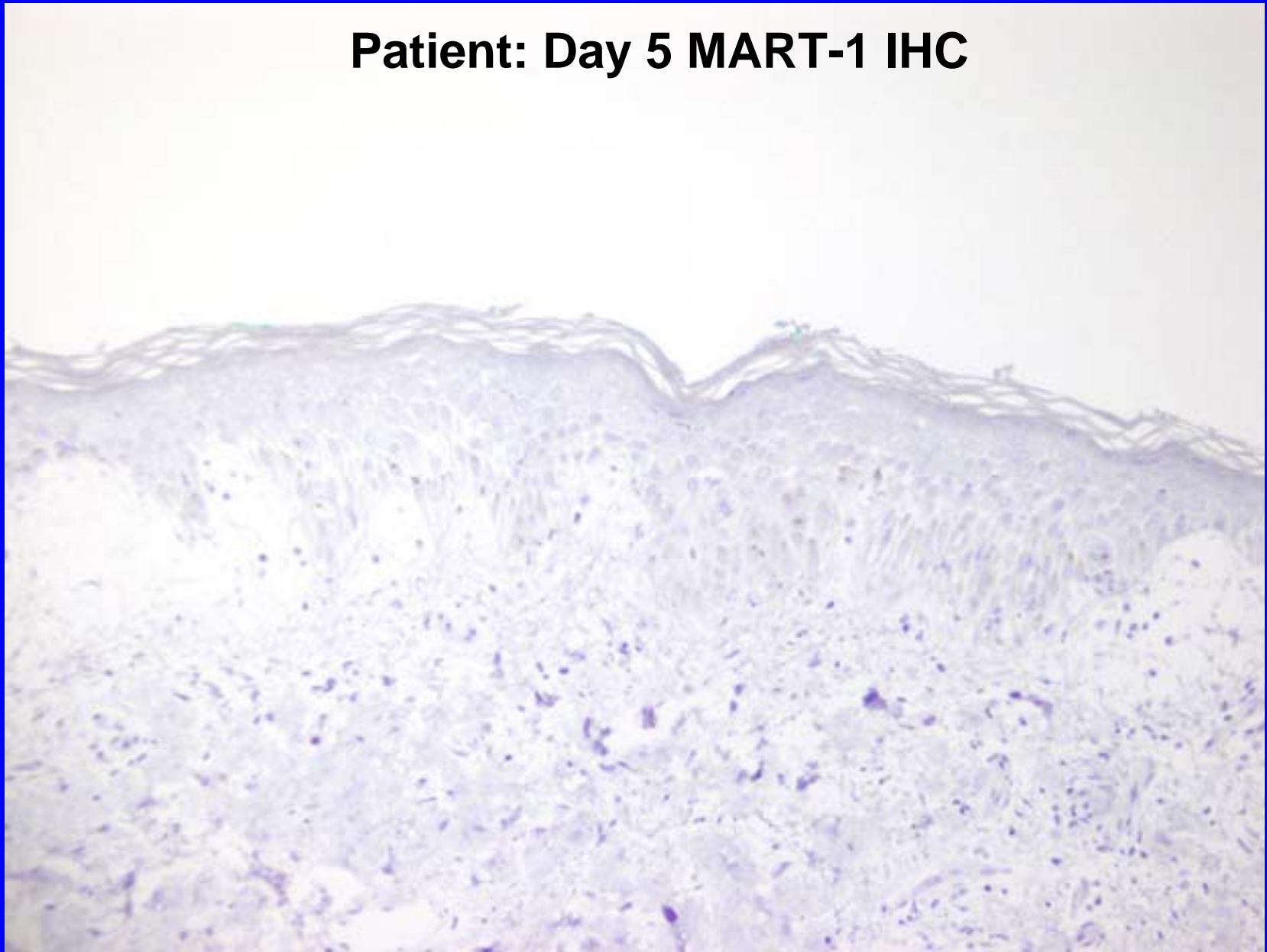


CD8 positive cells

(Positive Control: MART-1 IHC)



Patient: Day 5 MART-1 IHC



F5 High-Avidity Anti-MART1 TCR

D8



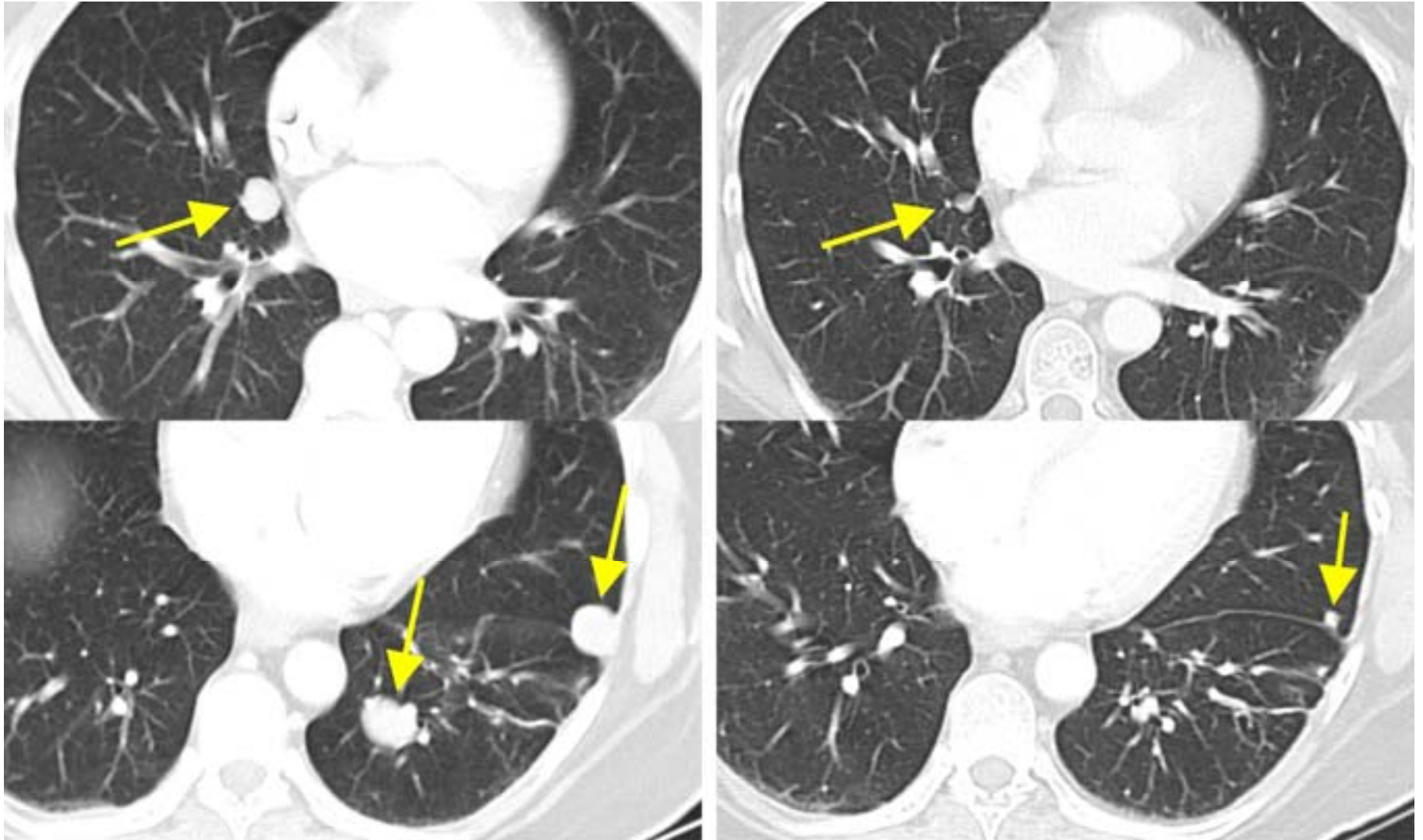
D12



D57



F5 High-Avidity Anti-MART1 TCR



2 Month Follow-Up

Conclusions

- Transfer of tumor-reactive T-cells after recipient lymphodepletion can cause dramatic regressions of metastatic melanoma in $\geq 50\%$ of patients, with some complete and durable responses
- The principle that tumor-reactive T-cells generated by introducing the genes encoding tumor-reactive TCRs can cause tumor regression has been shown
- Genetic construction of tumor-reactive T-cells has potential for significant improvement and can allow novel immunomodulatory strategies that may greatly enhance the efficacy of T-cell transfer

Future Directions

- New gene therapy vectors and methods are being developed to improve TCR expression in PBL
 - Lente viruses
 - Transposons
 - 2A peptides
 - CDR2/CDR3 modified receptors
- Receptors that recognize antigens on non-melanoma tumors have been developed to treat more common cancers with T-cell transfer
- Adding vaccines and alternative in vivo cytokine support (eg. IL-7 or IL-15)
- Manipulating T-cell function with gene therapy (eg. Mir 181a, zeta chain, CD8)

Acknowledgements:

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- Marybeth Hughes
- Udai Kammula
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