

Targeting Tumor Antigens by Redirecting T cells using Bispecific Antibodies



Lawrence G. Lum, MD, DSc

Professor of Medicine
Director of Translational Research
Director of Immunotherapy
Director of Stem Cell Laboratory

**Roger Williams Hospital
Providence, RI
Boston University School of Medicine
Boston, MA**

Clinical Problem

- Patients with metastatic breast cancer, hormone refractory prostate cancer, and other cancers have limited clinical options.
 - Chemotherapy, irradiation, or high dose chemotherapy have become dose-limiting.
 - New non-toxic strategies are needed to provide an anti-tumor effect without enhancing treatment toxicities.
-

Perspective

Earlier Talks

- **Humoral Immunity** and Antibodies – Paul Sondel
- **Monoclonal Antibodies** in Cancer Therapy – Ralph Schwall

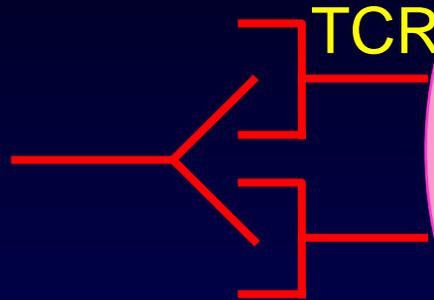
Later Talks

- **Cytokines** for Cancer Therapy – Jan Dutcher
 - **Cellular** Therapies – Robert Dillman
 - Critical Factors that Limit Success –Soldano Ferrone
-

Activated T Cells (ATC)

Signal 1

**Binding of OKT3
Activates the T cells**



Signal 2

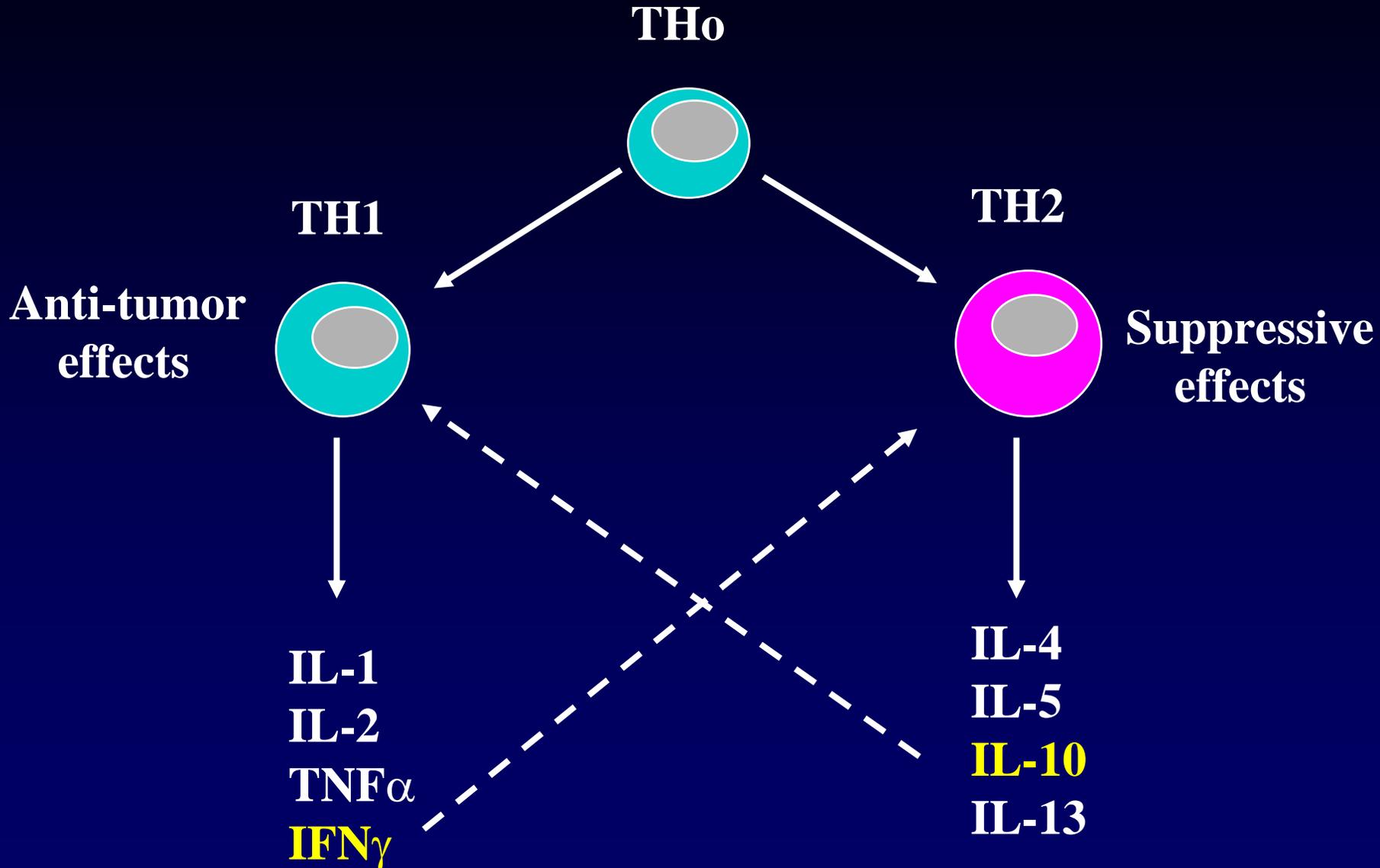
**IL-2 or
Anti-CD28
Keeps T cells alive**

Grow and Divide

Produce Cytokines/Chemokines

Directly Kill Tumor Cells

A Balancing Act for Anti-tumor Effects



Definitions

ATC: Activated T cells produced by anti-CD3 activation and culture in low dose IL-2 for 6-14 days.

BiAb: Consists of two mAbs produced by chemical, genetic, or hybridoma technology with 2 specificities (could be single chain fragment variable regions, scFVs).

Armed ATC: ATC with a BiAb that binds to CD3 on T cells and to a TAA on the tumor (artificial TCR).

Combination of Cellular and Humoral Therapeutic Strategy

- The specificity of monoclonal antibodies

AND

- Non MHC restricted cytotoxicity mediated by T cells, NK cells, or other effector cells
-

Preclinical Studies Using BiAbs

mAb	Target
Anti-Tenascin	Glioma
Anti-Glioma	Human Glioma
Anti-CD13	AML
Anti-MUC1	Bile Duct CA
Anti-EpCAM	Epithelial Cell Adhesion on AdenoCA
OC/TR	Folate Receptor on ovarian CA
Anti-kDal K29	Renal cell carcinoma
Anti-G250	Renal cell carcinoma
OKT9	Anti-transferrin receptor
Anti-AMOC-31	40 kDa membrane glycoprotein on carcinomas

mAb	Target
Anti-idiotypic	BCL1 lymphoma
Anti-CD19	Leukemic B cells
Anti-tumor (Fab)2	Retargeting TIL
Anti-CEA	Carcinoembryonic antigen
Anti-Her2	Her2 on RCC, colon, breast
Anti-CD20	NHL
Anti-PSA	Prostate specific antigen
Anti-CA19-9	Carcinomas
Anti-HLA-DR beta chains	B cells
Anti-EGFR	Glioma, neuroblastoma, colon, pancreatic, lung

BiAb Trials

- SHR-1: Anti-CD3 x **anti-CD19** quadroma for NHL; 10 mcg -5 mg. No adverse effects except thrombocytopenia.
 - BIS-1: anti-CD3 x **anti-EGP-2** for epithelial carcinoma-associated transmembrane glycoprotein; MTD of 5 mcg/kg; induced high levels of TNF and IFN; dyspnea, vasoconstriction and fever without anti-tumor effect.
 - 2B1: anti-CD16 x **anti-HER2** quadroma for Her2+ tumors; DLT were fever, chills, N/V, and leucopenia; HAMA in 14 of 15; MTD was 2.5 mg/m².
-

BiAb Trials

- HRS-3/A9: anti-FCR3 x **anti-CD20** for HD of B cell malignancy; MTD not reached at 64 mg/m²/dose
 - MDX-H210: anti-CD64 x **anti-Her2** for breast, ovarian, prostate CA; doses ranged from 1-40 mg/m² without DLT
 - MDX-447: anti-CD64 x **anti-EGFR** for renal and head and neck cancer; Hypotension DLT, doses up to 40 mg/m²
 - H22x Ki-4: anti-CD64 x **anti-CD30** for Hodgkin's Disease, doses up to 20 mg/m²
 - Common thread: deletion of Fc portions improved toxicity profiles
-

Trials Using BiAb Armed T cells

- Nitta, 1990, anti-CD3 x **anti-glioma** armed lymphocytes; 4 of 10 pts had tumor regression.
 - Lamers, 1992, ATC armed with anti-CD3 x **anti-Mov28** were used to treat ovarian Ca.
 - Canevari, 1995, ATC with anti-CD3 x **anti-folate receptor** given intraperitoneal with IL-2 resulted in tumor regression in advanced ovarian; 7 of 26 (4 CRs, 3 PRs).
 - These early studies provided the impetus to develop engineered T-bodies and molecular engineering of BiAbs for targeting TAA.
-

Begin with the “End” In Mind

STRATEGY:

Make T Cells better killers by redirecting or focusing their non-MHC restricted cytotoxicity on TAAs

MEANS: Arm T cells with BiAbs directed at TAAs

GOALS:

1. Improve tumor lysis
 2. Immunize the patient by inducing specific CTL and humoral anti-tumor responses.
 3. Induce remissions with persistent anti-tumor immunity.
-

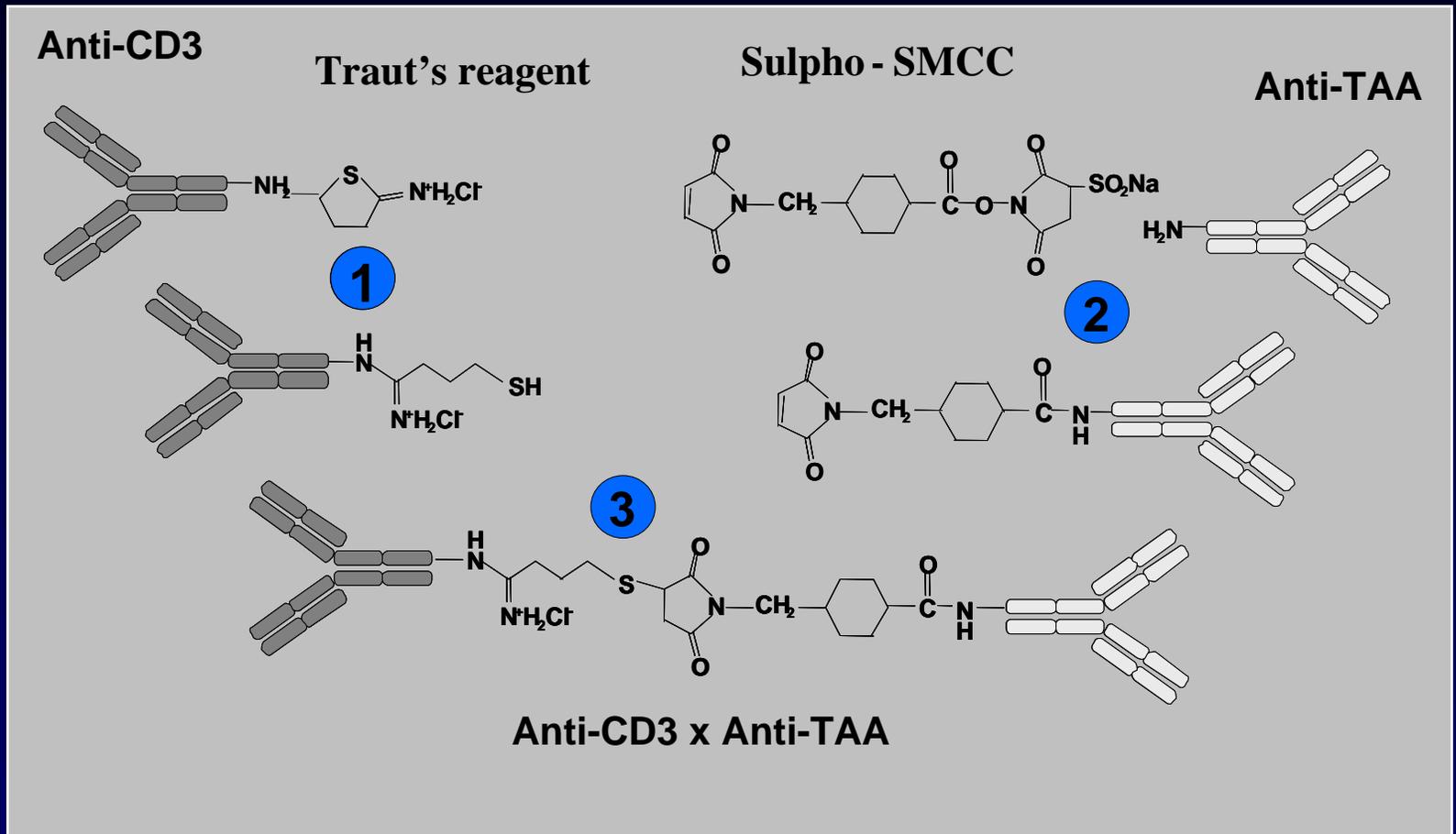
Phase I/II Studies of Ex vivo Expanded T cells

1. Phase I up to 40 billion ATC given on days 1, 4, 7, and 11 for a total of **160 billion ATC** after Cytoxan/TBI and **PBSCT for hematologic malignancies** without adverse effects.
 2. Phase I up to **80 billion** anti-CD3/anti-CD28 coactivated T cells given in 8 doses to patients with **solid tumors** with IL-2. Safe with no dose limiting toxicities (J Immunotherapy 5:408, 2001) as well as with low dose Cytoxan.
 3. Phase II **210–310 billion ATC**. 10 billion ATC given 3 times/wk for 3 weeks and then 20 billion/week for 6 weeks after **PBSCT for stage IV breast cancer**. 70% OS and 50% PFS at 32 months without regimen or cell-based adverse effects. (Autologous Blood and Marrow Transplant 10th Proc, pp95, 2001).
-

Development of Bispecific Antibodies

- Produced by chemical heteroconjugation of existing mAbs, recombinant DNA technology, or a combination thereof.
 - A variety of design formats allow for: 1) different sizes to allow tissue penetration; 2) enhanced specificity; 3) increased affinity to effector cells.
 - Applications: targeting effector cells, targeting toxins, drugs, prodrugs, enzymes, DNA, anti-vascular agents, gene therapy vectors, radionuclides, and others (use your imagination)
-

BiAb Production by Chemical Heterconjugation

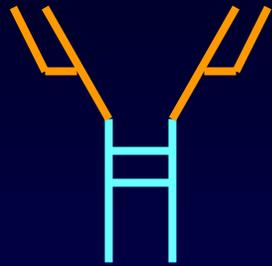


Targeted Killing by T cells with BiAbs



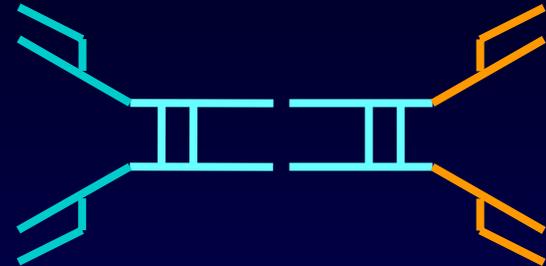
Anti-CD3

+

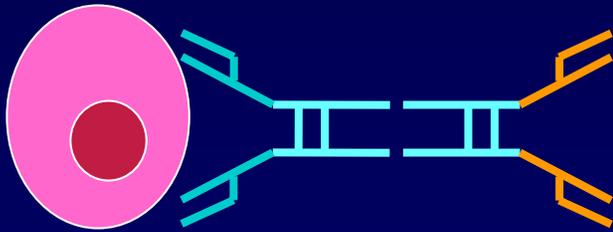


Anti-TAA

=

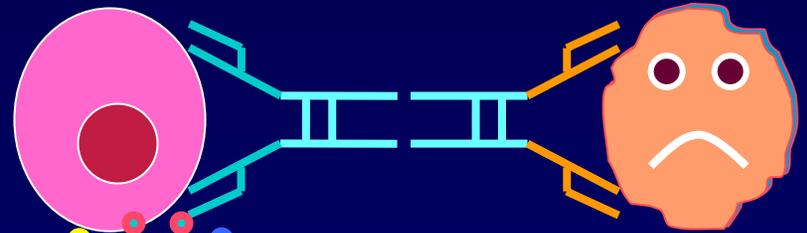


Anti-CD3 x Anti-TAA



Armed T Cell

→



T Cell

**Tumor
Lysis**

Production of Armed T cells

PBMC from Pheresis



OKT3 (20 ng/ml) + 100 IU/ml of IL-2



ATC are split every other day



Harvest, Arm with BiAb and Cryopreserve after 10-14d



**Quality Control (Bacteria, fungal,
and Mycoplasma stain) 7 days**



Testing for cytotoxicity and cytokine production

Characteristics of Armed ATC

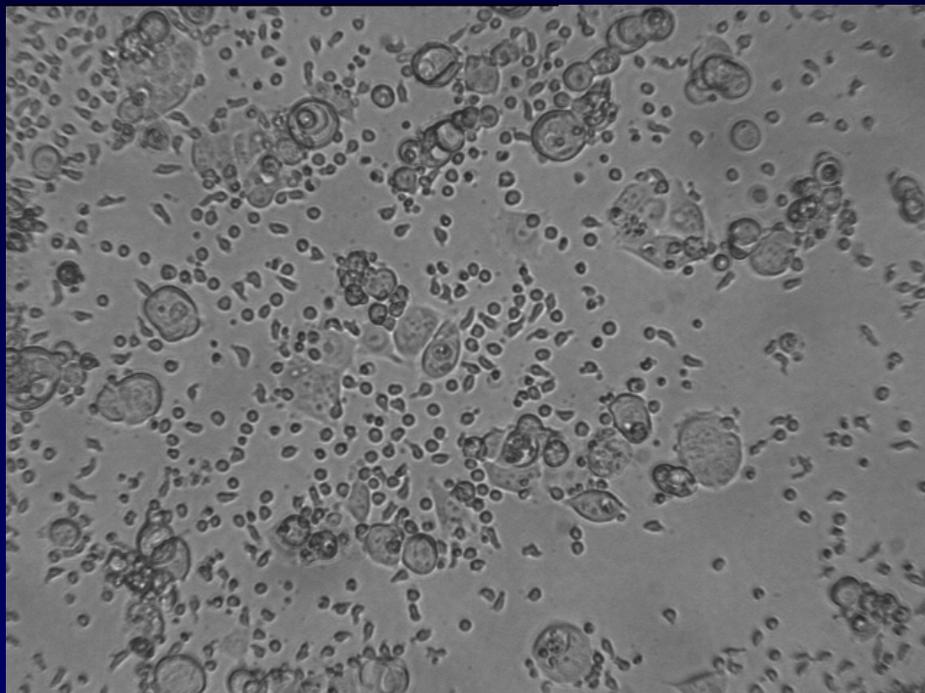
1. Exhibit non-MHC restricted cytotoxicity (“promiscuous killers”).
 2. Secrete IL-2, IFN γ , TNF α , GM-CSF, MIP-1, and RANTES after antibody receptor binding.
 3. >95% CD3+ cells, 60-80% CD8 cells, 20-40% CD4 cells, and <5% CD56+ cells.
 4. Patient CD3 cells expand up to 30 fold in 14 days.
-

Preclinical Questions

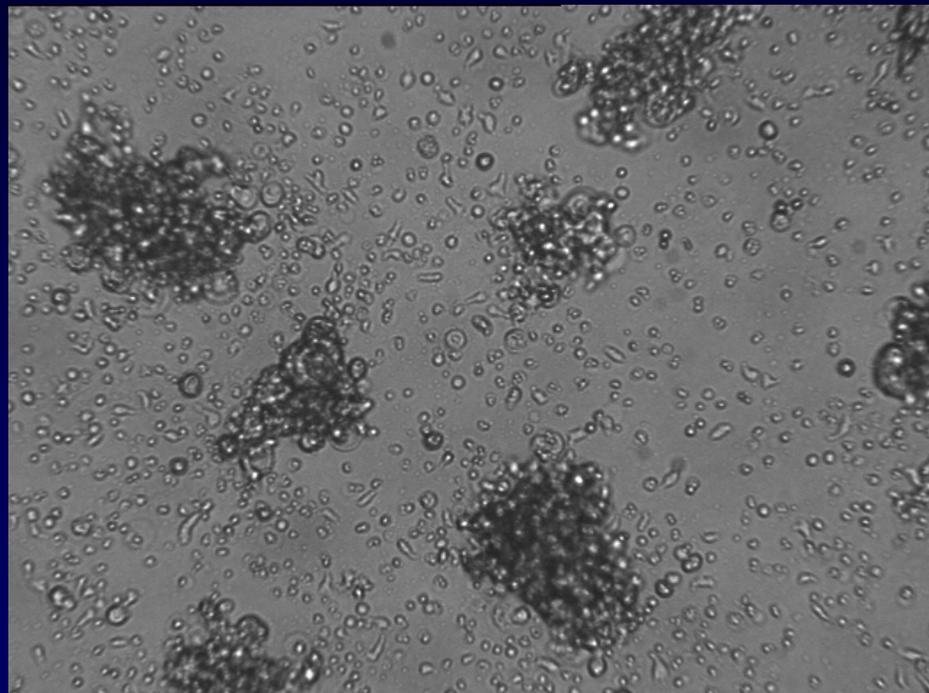
- **How long will the BiAb remain on ATC?**
 - **How long will armed ATC kill?**
 - **Will binding to tumor trigger cytokine secretion?**
 - **How many times will armed ATC kill?**
 - **How long can armed ATC be detected in patients?**
-

Targeting and Killing

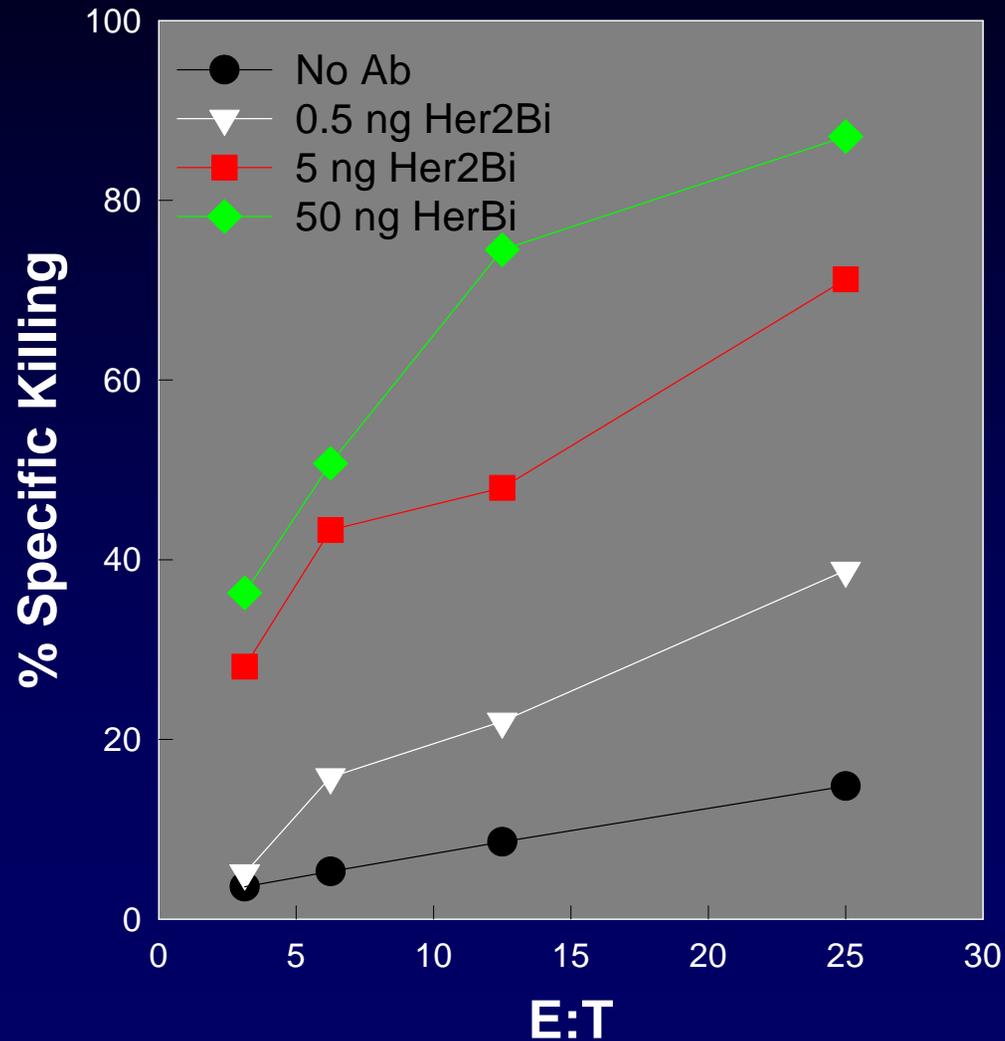
Unarmed



Armed

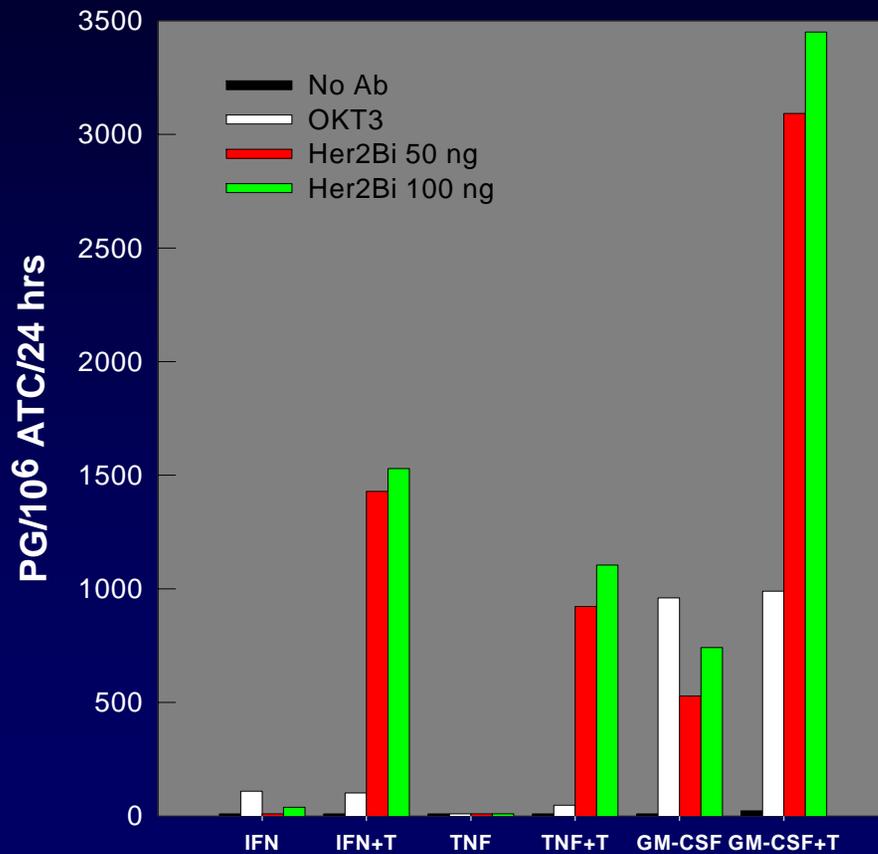


Killing of MCF-7 Cells by Armed T Cells

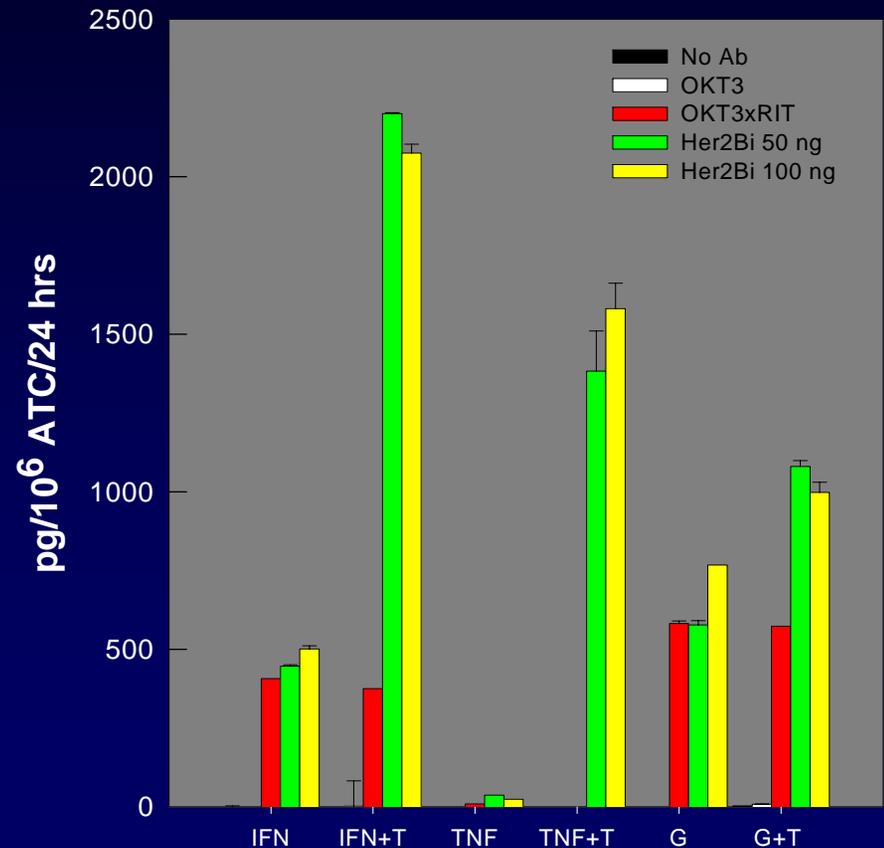


Cytokine Production by Armed T Cells Exposed to SK-BR3

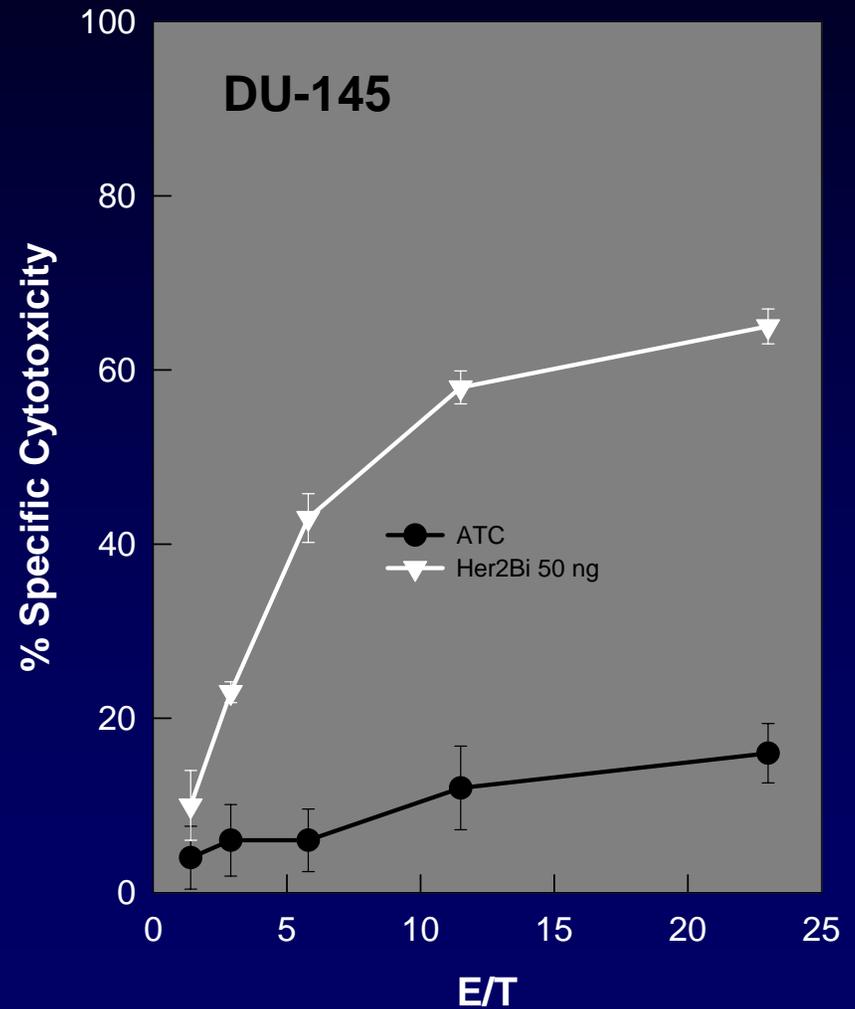
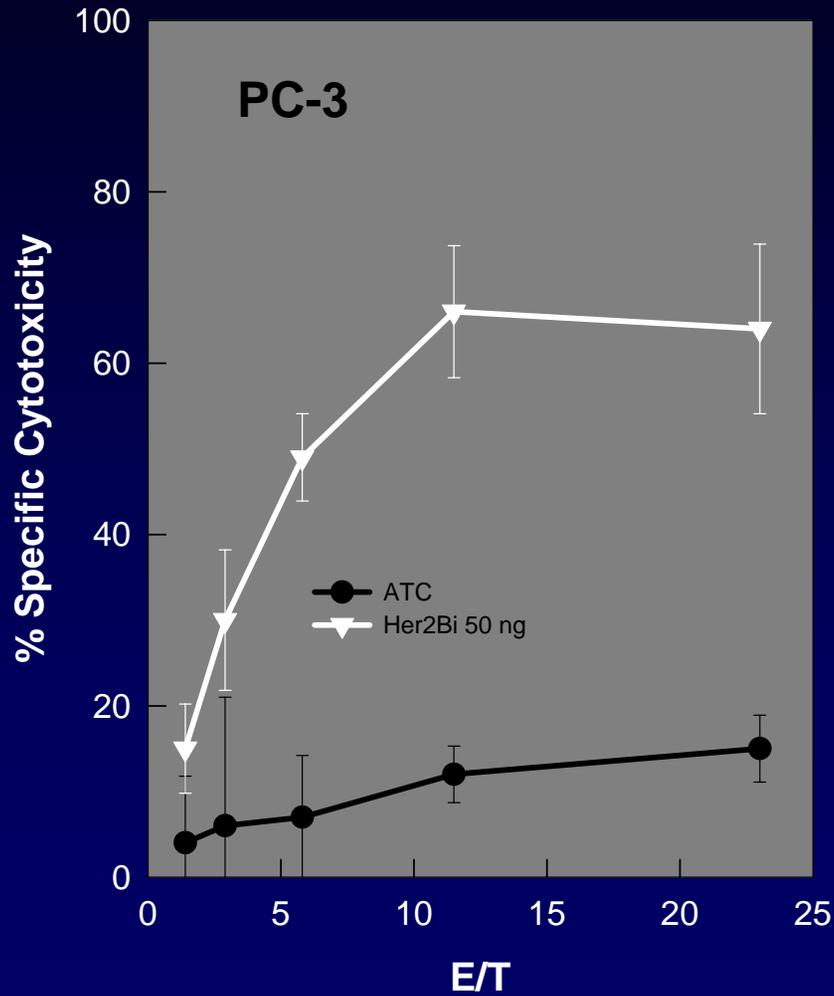
Normal



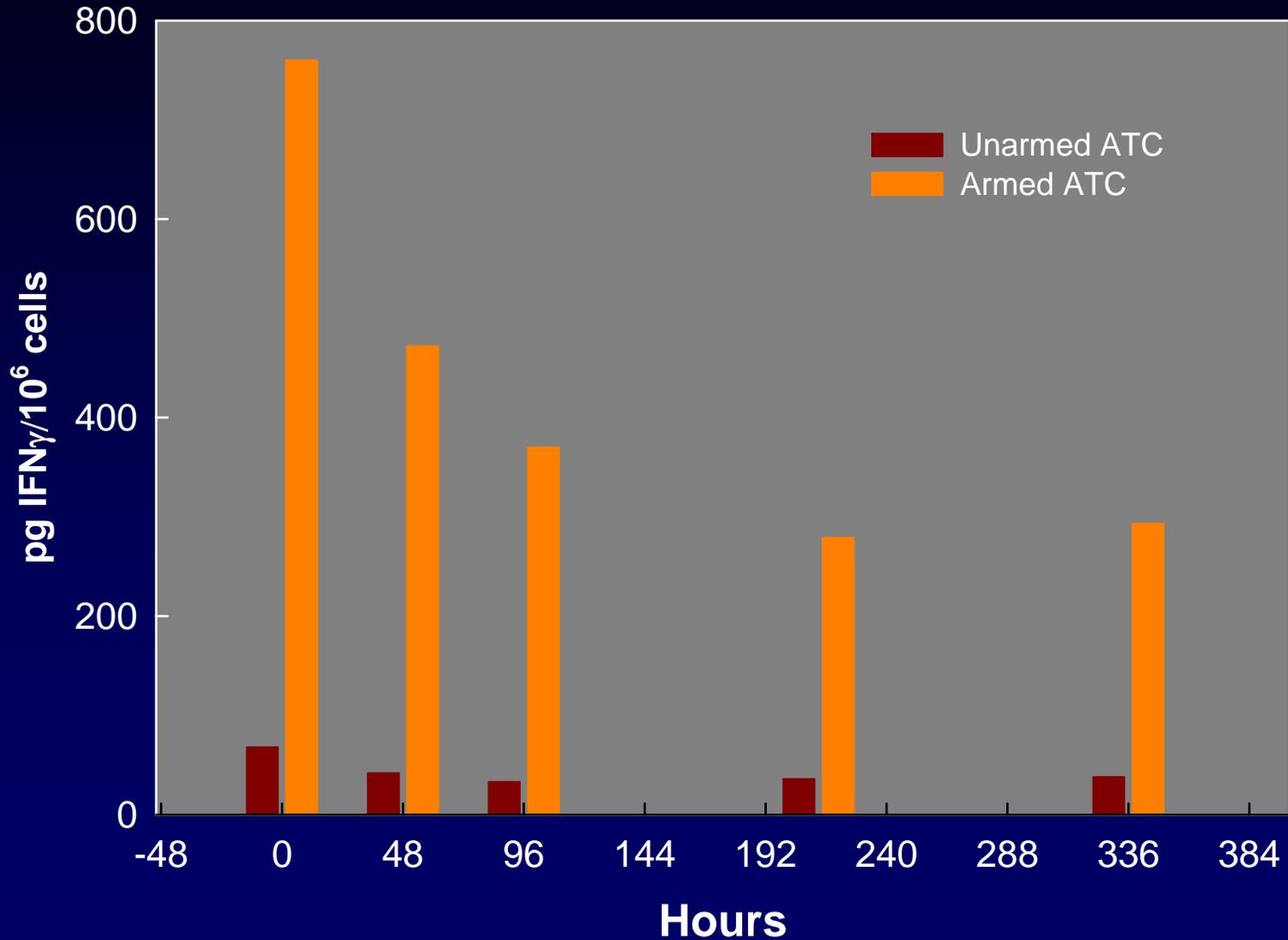
Patient



Cytotoxicity Directed at Prostate Cancer Lines

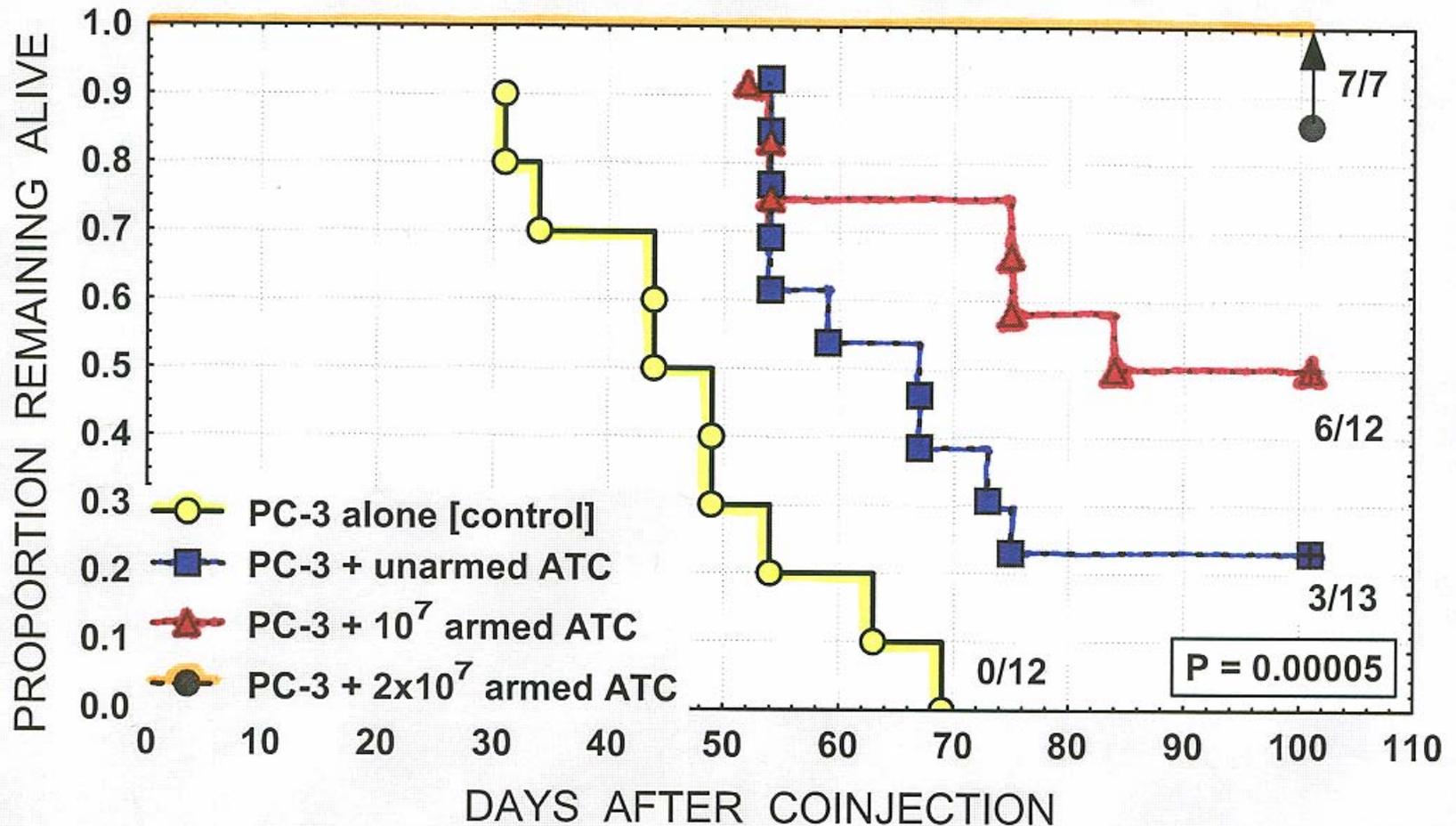


IFN γ Secretion upon Repeated SK-BR-3 Restimulation

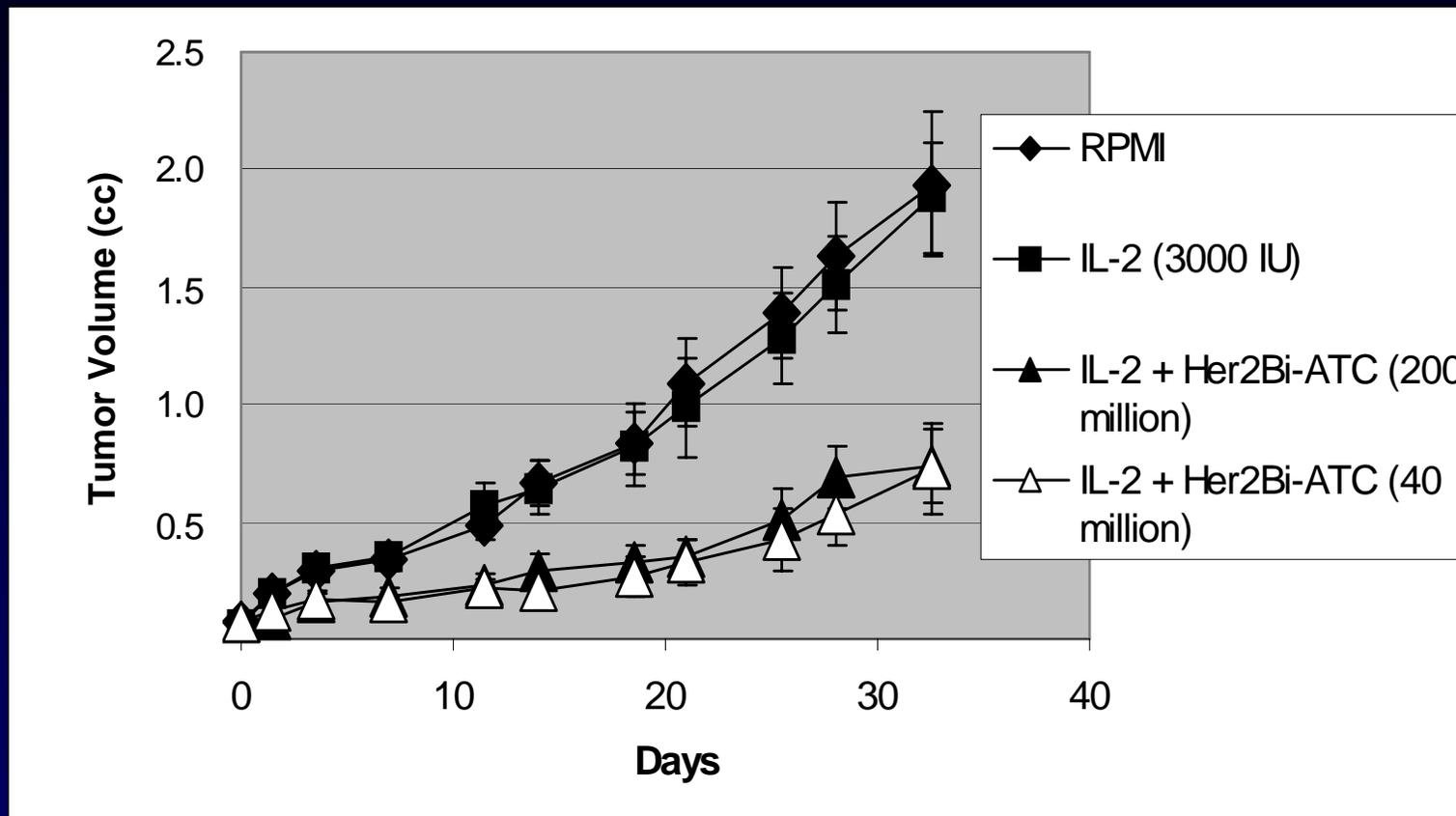


Prevention of Prostate Cancer

COINJECTION OF PC-3 PROSTATE CA CELLS + ACTIVATED T CELLS
KAPLAN-MEIER PLOTS

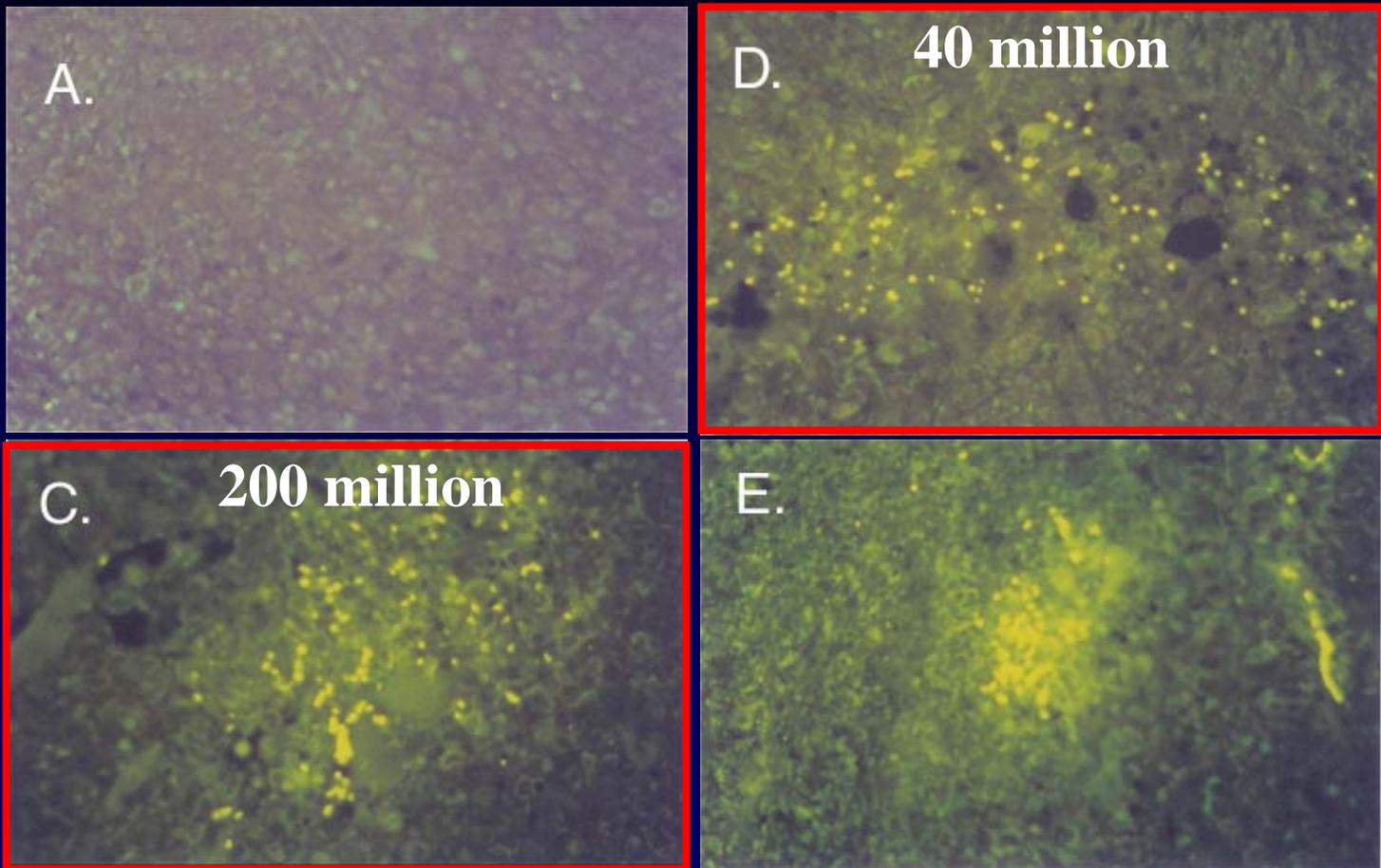


IV Treatment Delays PC-3 Xenografts



PC-3 tumor cells (10^7) were implanted SC in flanks of SCID-Beige mice. 7 days later when tumors were ~ 0.05 cc, treatments were started once/week x 4 weeks and tumor growth monitored. Results are from 2 experiments (n=10 mice/group).

Trafficking of Her2Bi Armed ATC in Beige/SCID



A RPMI/ IL-2 IV; C) IL-2 + armed ATC (2×10^8 cells) IV; D) IL-2 + armed-ATC (4×10^7 cells) IV; or E) IL-2 (3000 IU) + armed ATC (2×10^8 cells)IT. Tumors were excised 18 hr after treatment, formalin fixed, paraffin embedded, sectioned, and stained for human CD3+ cells

Immunotherapy Approaches

Cells	CMV CTL to Prevent Infection	EBV for LPD	MM Specific CTL after Chemo	DLI after Allo-BMT	ATC after HDC+PBSCT for BrCa	Armed ATC
Type	Specific	Specific	Specific	Polyclonal	Polyclonal	Specific
Dose	$10^6/\text{kg}$	$\sim .5 \times 10^9/\text{kg}$	$10^9/\text{kg}$	$10^8/\text{kg}$	$3 \times 10^9/\text{kg}$	$4 \times 10^9/\text{kg}$ ($0.5 \times 10^9/\text{kg}$)
Effect	Prevents CMV Pneumonia	Treatment of LPD	Induce CR in MM	Induce CRs in CML>AML>ALL	Improve PFS?	Decrease Bone Pain, PSA, CA 27-29
Target	Viral	EBV LPD	Solid tumor	Liquid tumors	Solid tumor	Solid tumors

Protocols: FDA and IRB Approved

Breast Cancer

- RWH #356-46: TAC + Armed ATC for Stage II-III BrCa
- RWH #351-46: Armed ATC for Stage IV BrCa (NCI-R01 funded)

Hormone Refractory Prostate Cancer

- RWH #355-46: Armed ATC for HRPC

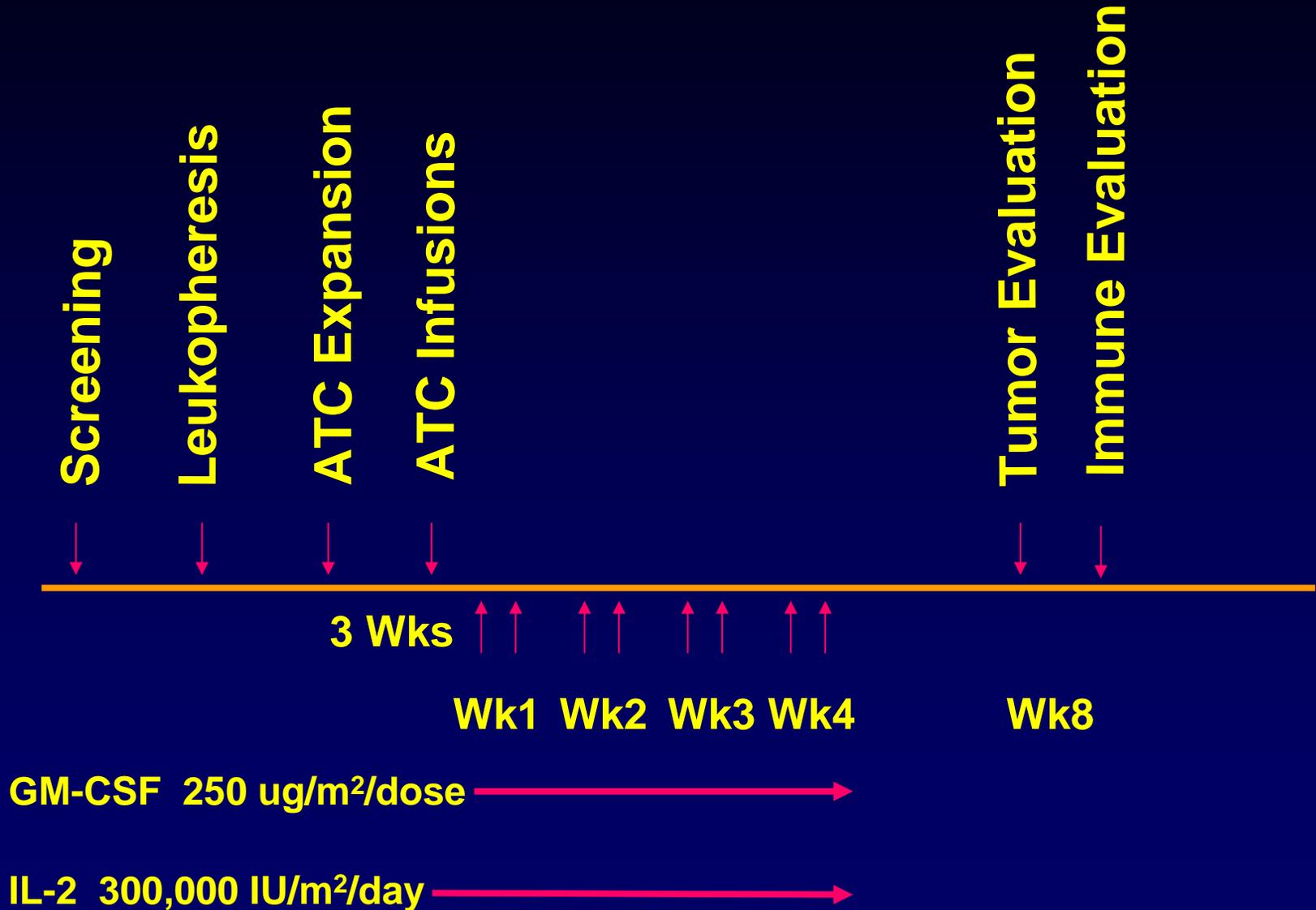
Eligibility:

- Metastatic, measurable, or evaluable sites
- Phase I: Her2/*neu* positive or negative
- Phase II: Her2/*neu* positive
- No active cardiac disease, ECOG PS 0-2, life expectancy >3 months

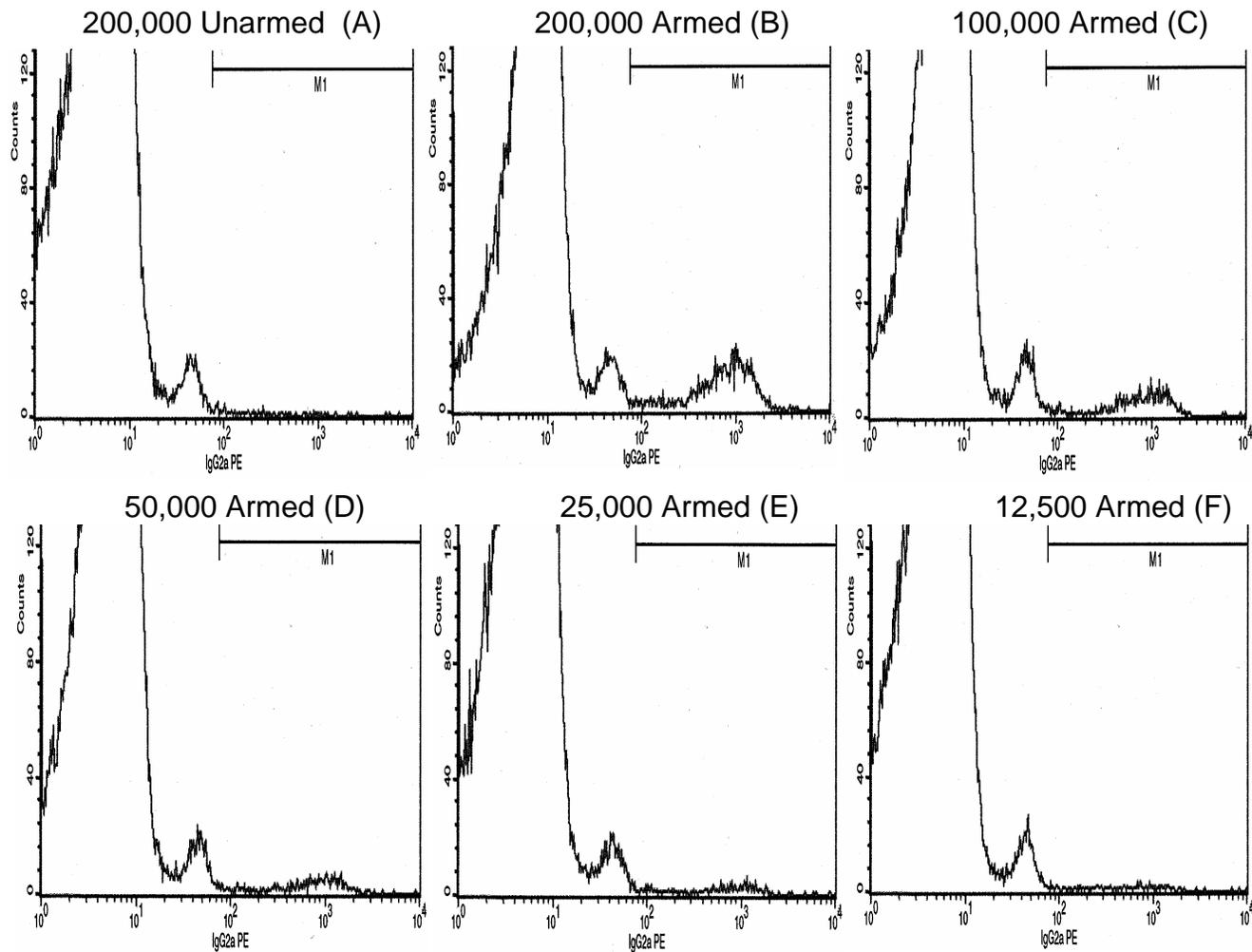
Lymphoma

- RWH #394-46 : Armed ATC targeting CD20 lymphomas after PBSCT
FDA approved 11/02/04 and (Leukemia & Lymphoma Society funded)
-

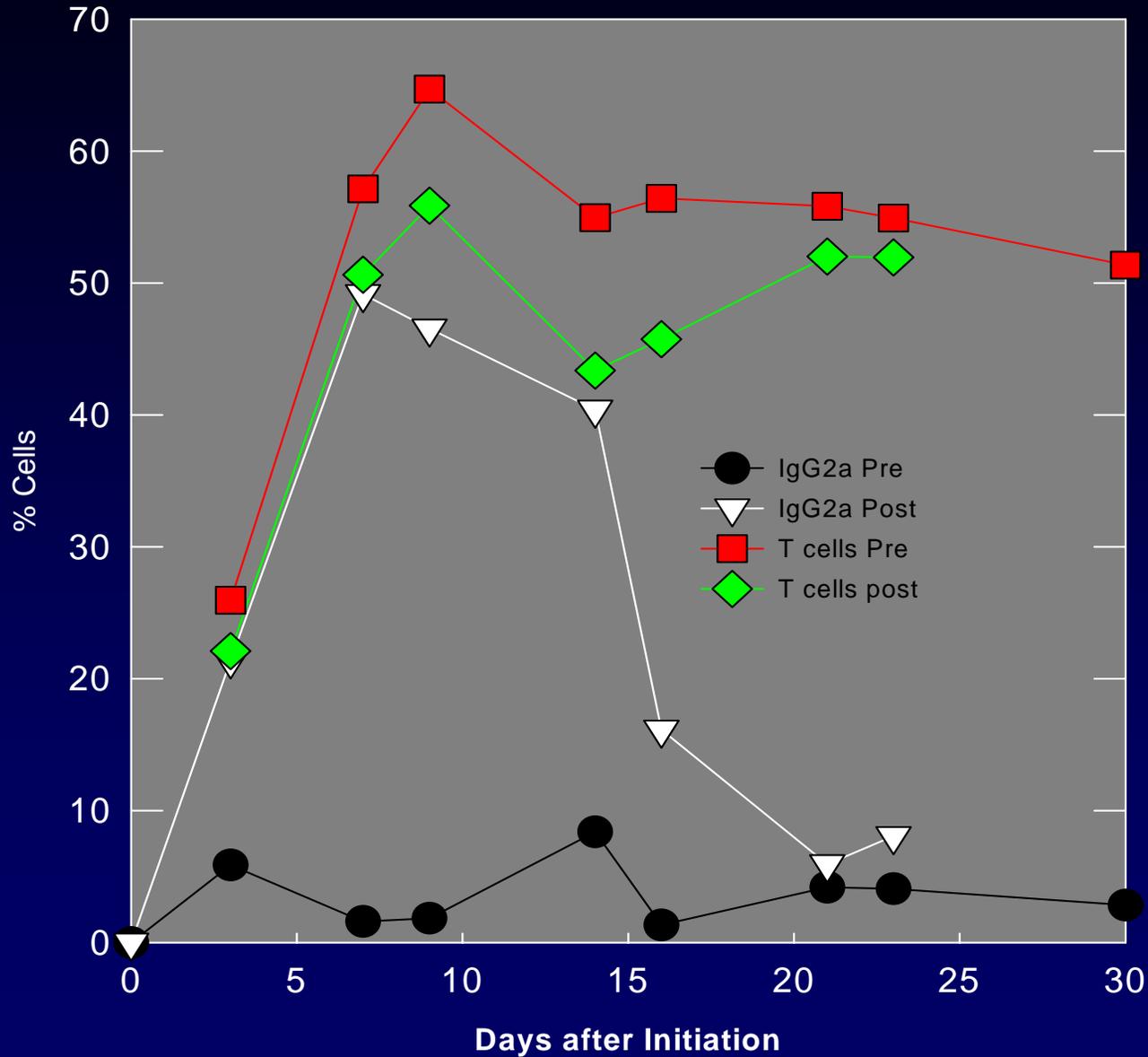
Treatment Schema for Breast and Prostate



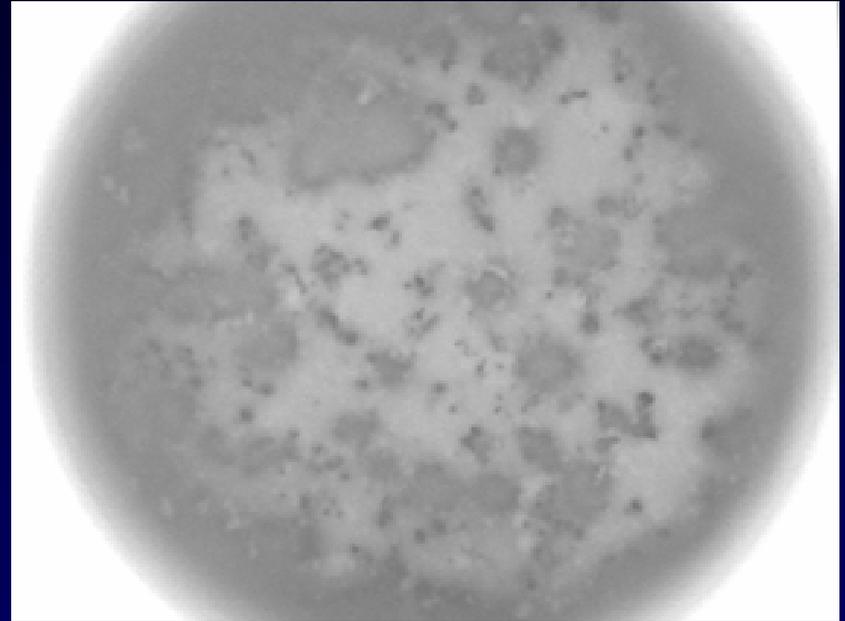
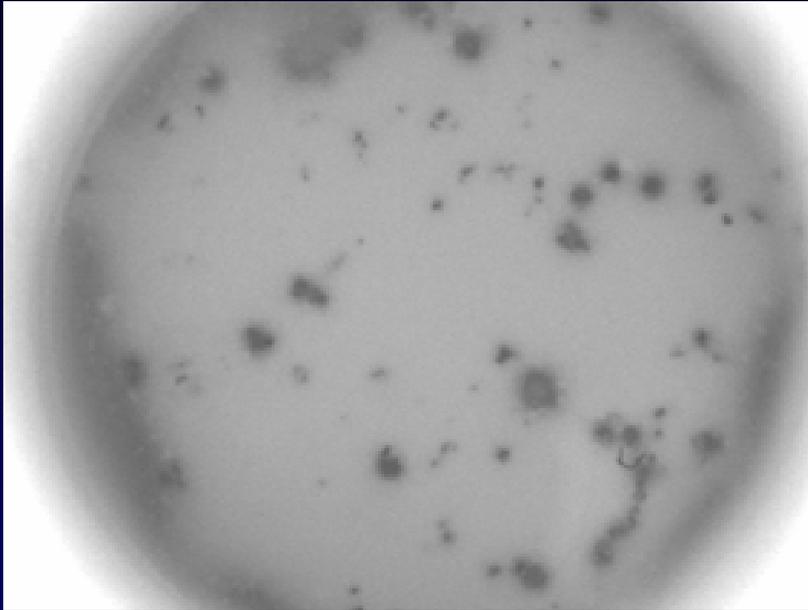
Whole Blood Spiked with HER2Bi ATC



Detection of Armed ATC in Blood

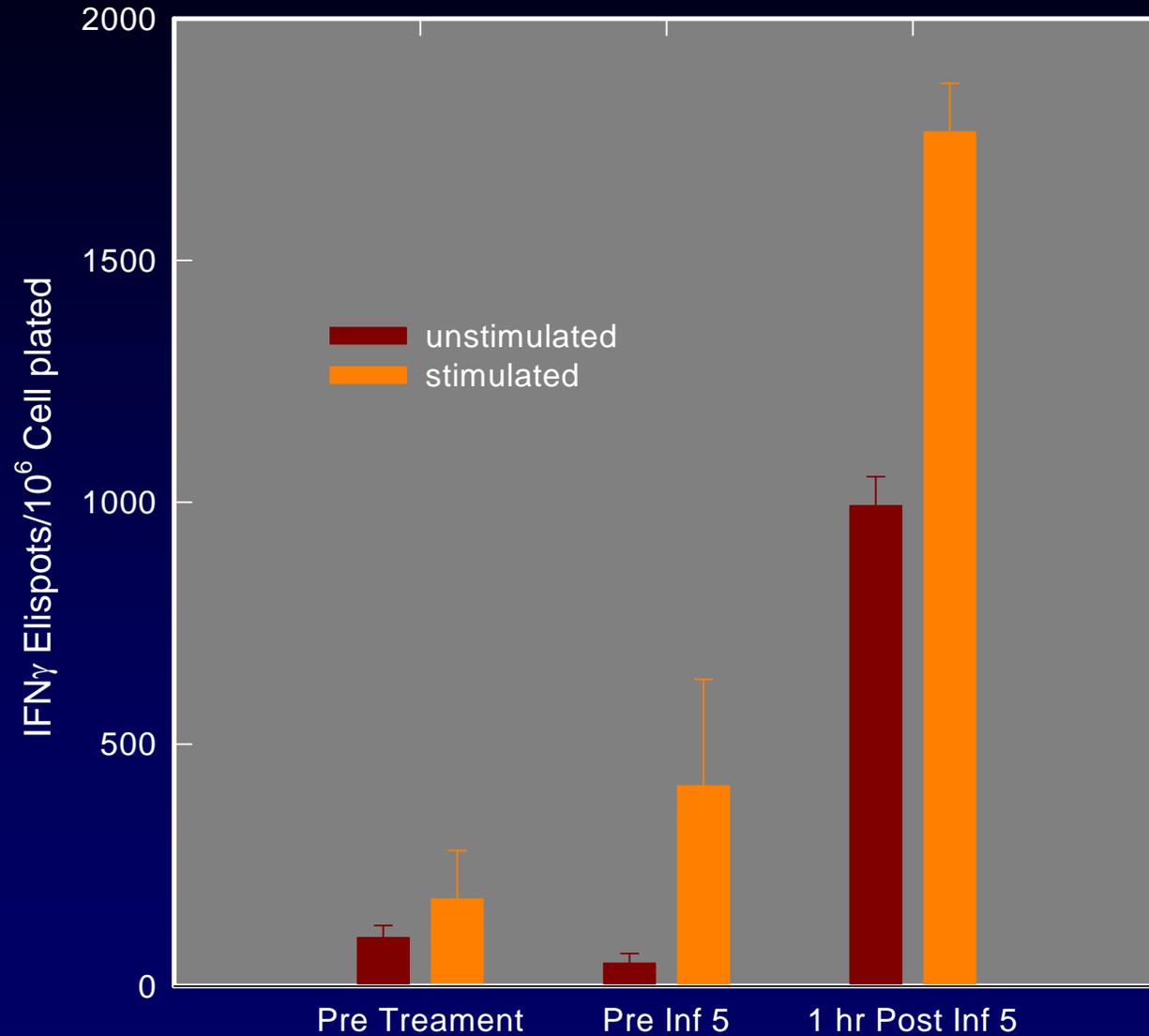


Pre and Post EliSpots

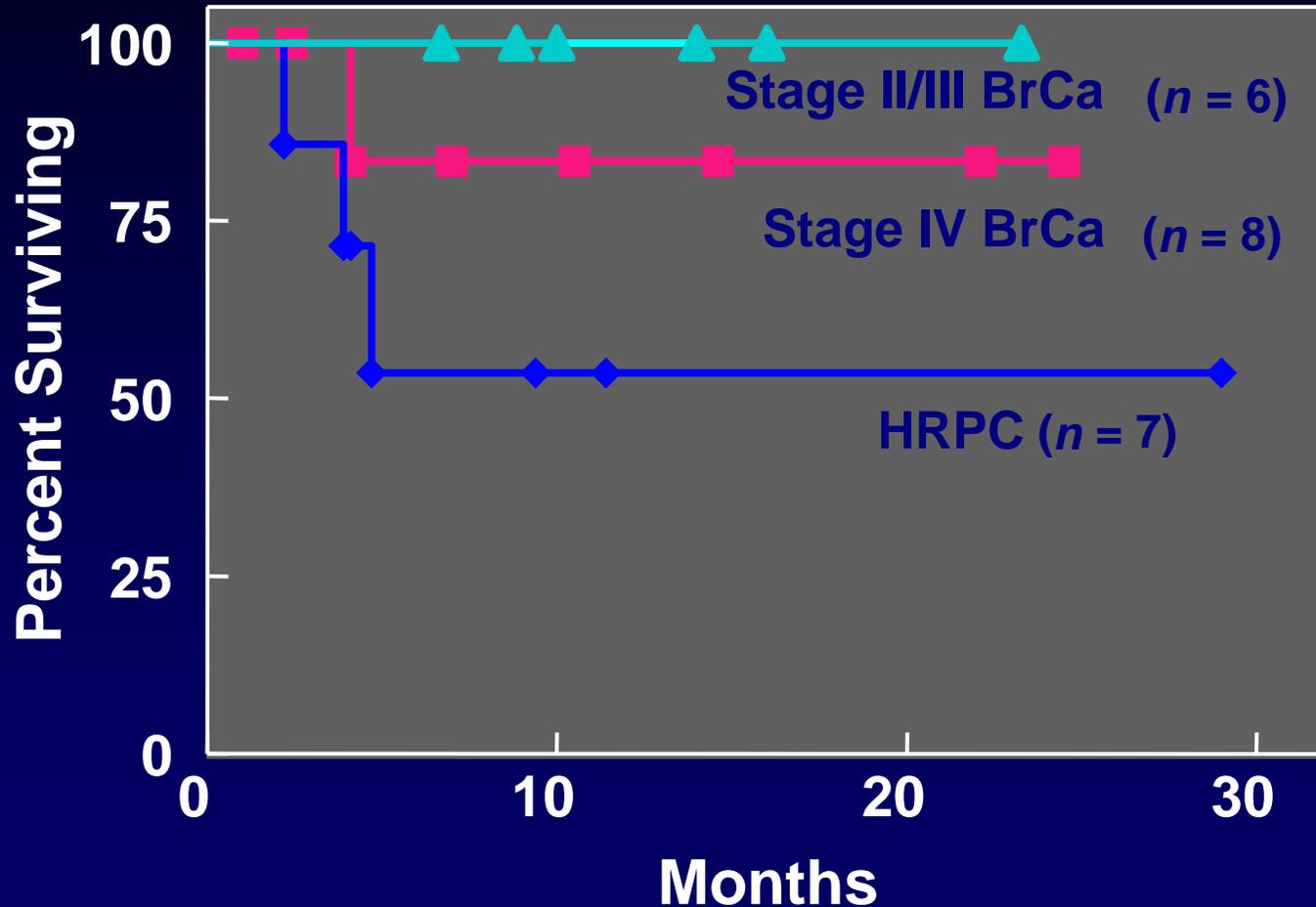


IFN- γ EliSpots with PBMC isolated from a patient before and after the 4th infusion of Her2Bi-armed ATC (5×10^9). Her2/*neu*-specific IFN- γ secretion by T cells was measured by exposing 10^5 PBMC to SK-BR-3 cells at an E:T of 10 for 2 h at 37°C and then the PBMC to an EliSpot plate coated with anti-IFN- γ .

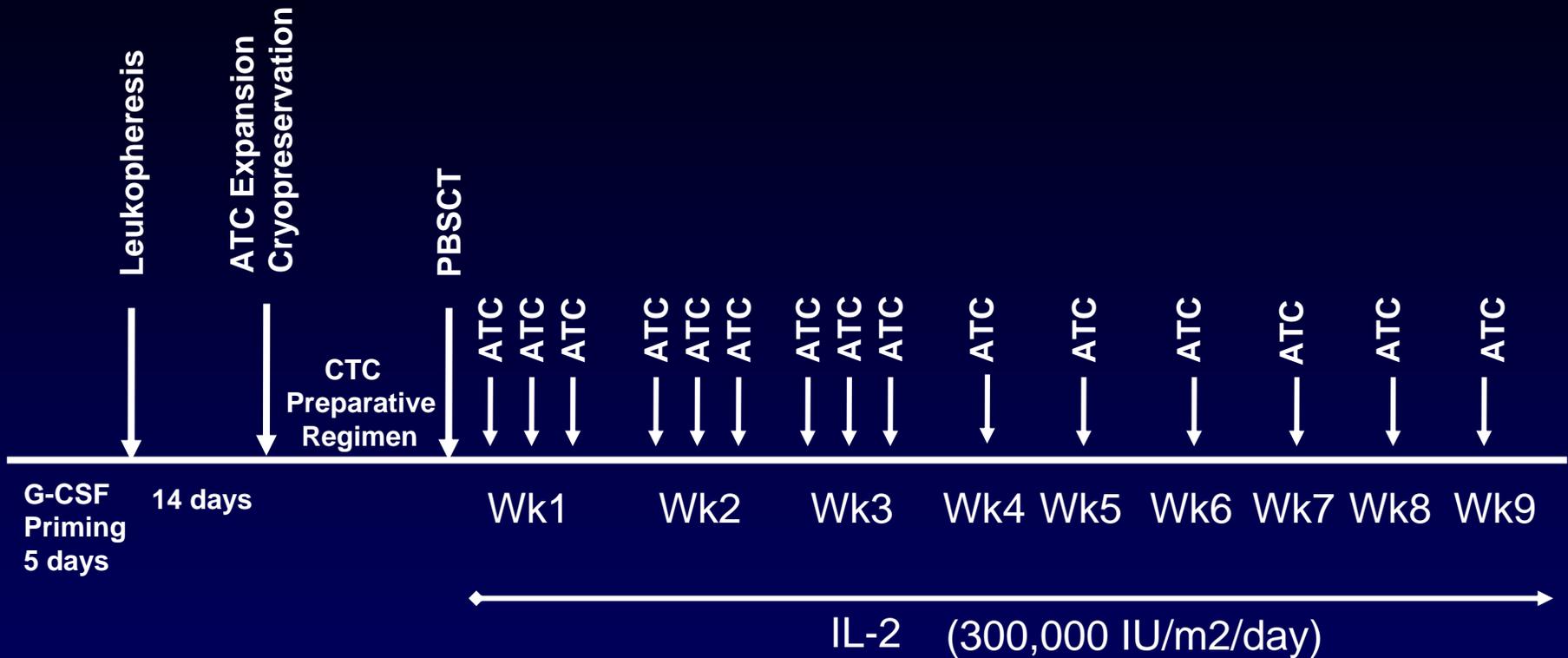
EliSpots from Stage IV BrCa Pt



Overall Survival in Phase I Trials with Her2Bi-armed ATC



RWH# 04-394-46 Infusion of ATC Armed With CD20Bi for CD20+ NHL



- Dose Level 1: 5 billion/infusion Total 75 billion
- Dose Level 2: 10 billion/infusion Total 150 billion
- Dose Level 3: 15 billion/infusion Total 225 billion
- Dose Level 4: 20 billion/infusion Total 300 billion

Armed ATC

- Grow and divide after engaging and killing the tumor
 - Secrete chemokines and cytokines multiple times
 - Bind and kill tumors cells multiple times
 - Survive in vivo > 3 weeks
 - Develop into Ag-specific CTL over 2 weeks in culture
 - Patients infused with armed ATC develop levels of cytokines during their infusion
 - PBMC from patients develop cytotoxicity that persists up to a month after the last infusion
-

Summary for Armed ATC

1. Armed ATC kill multiple times.
 2. Armed ATC proliferate after engaging tumor and do not undergo apoptosis via Fas/FasL or ACID.
 3. Large numbers (320+ billion) of armed ATC can be produced in 2 weeks whereas cloned CTL are time consuming requiring a customized effort.
-

Summary for Armed ATC

- 4. Armed ATC may develop into Ag-specific CTL directed at other TAA **AND** induce endogenous T cells to become cytotoxic.**
 - 5. “Multiple infusional vaccinations” may immunize patients to their autologous tumor.**
 - 6. Significant amounts of cytokines are found in patient serum during and after infusions with a Th1 profile.**
 - 7. There is a strong suggestion that overall survival for metastatic breast and HRPC patients is improved even with small numbers of patients.**
-

Platform Technology

OKT3 + Anti-Her2/*neu*

Breast

Prostate

Eight infusions of armed ATC

OKT3 + Anti-CD20

Lymphoma

ALL?

CLL?

PBSCT + Armed ATC

OKT3 + Anti-EGFR

Colon

Pancreatic

Lung

Glioblastoma

Neuroblastoma

Concepts/Principles

- Bispecific antibodies can be used to target effector cells to tumors.
 - The non-MHC restricted cytotoxicity can be redirected with BiAbs.
 - The targeted cell therapy can be used in combination with cytokine, chemotherapy, or stem cell transplant strategies that make immune space and reduce tumor burdens.
 - The platform allows flexibility for targeting different TAAs by switching BiAbs
-

Immunotherapy Team (A Rhode Island Consortium)

Ritesh Rathore, MD – Clinical Director

Pam Davol, MEd, Senior Research Associate

Wendy Young, RN, OCN – IT Clinical Coordinator

Delia Devito – Administrative Assistant

Technical

Ryan Grabert

Yoni Gall

Tom Tarro

Estie Palushock

James Davis

Jen Jarvis

Coinvestigators/Contributors:

Frank Cummings, MD

Tony Testa, MD

Nick Koutab, PhD

Kathy Radie-Keane, MD

Steven Schiff, MD

Peter Quesenberry, MD

Ray Chaquette, MD

Gerald Colvin, DO

Augusto Zabbo, MD

Cheru Taneja, MD

Steven Cohen, MD

Bob Legare, MD

Tony Mega, MD

Abby Maizel, MD,

John Pryzgoda, MD

Mehrdad Abedi, MD

Facilities:

1. Specialized ultra-clean rooms that are FDA approved for producing T cells and bispecific antibodies for clinical trials.
2. Seamless clinical coordination between institutions/practices
3. Immunotherapy clinic that is staffed with experienced nurses.