
Identification of antigens recognized by T cells

Victor H. Engelhard, PhD

Carter Immunology Center

Department of Microbiology, Immunology, and
Cancer Biology

University of Virginia School of Medicine

vhe@virginia.edu

Disclosures

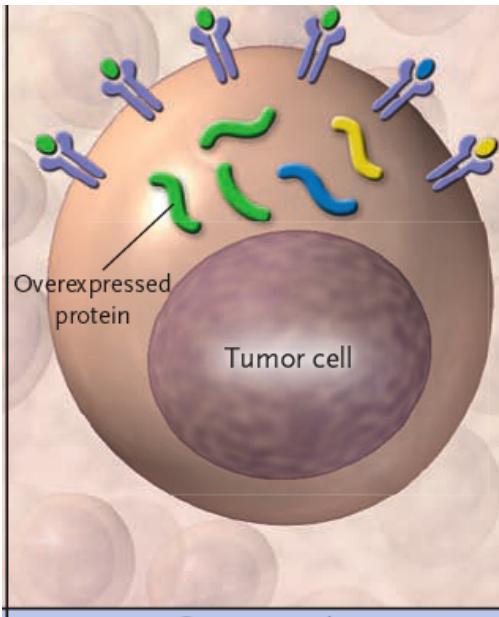
Agenus Inc.

Consultant

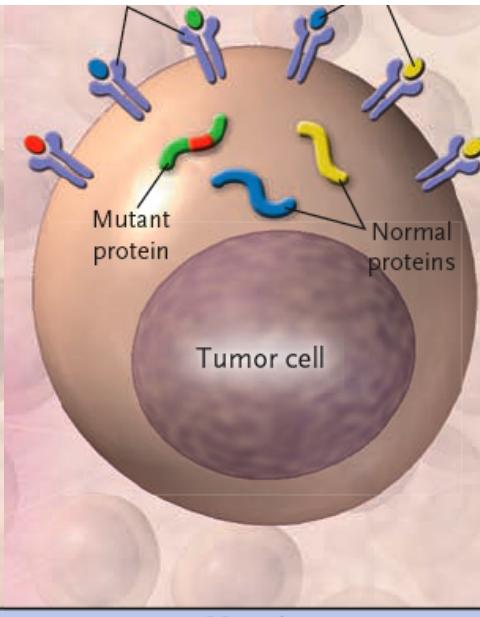
Shareholder

Characteristics of identified tumor antigens

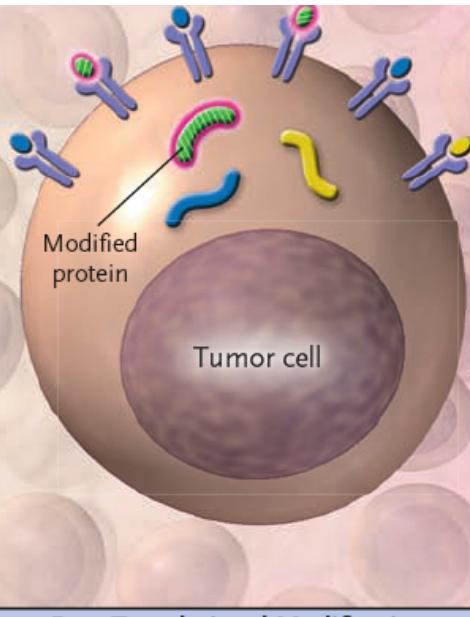
Tissue-specific,
cancer-testis antigens



Patient-specific
neoantigens



Phosphopeptides



Overexpression

Mutation

Post-Translational Modification

General approaches to antigen discovery

- Direct methodology
 - Isolate cancer reactive T cells from patients
 - Identify MHC and peptide that comprise the antigen recognized
- Reverse methodology
 - Identify genes, proteins, or peptides of interest expressed in/on cancer cells
 - Establish whether there are T cells in patients that recognize them

cDNA library approach

T. Boon/S. Rosenberg

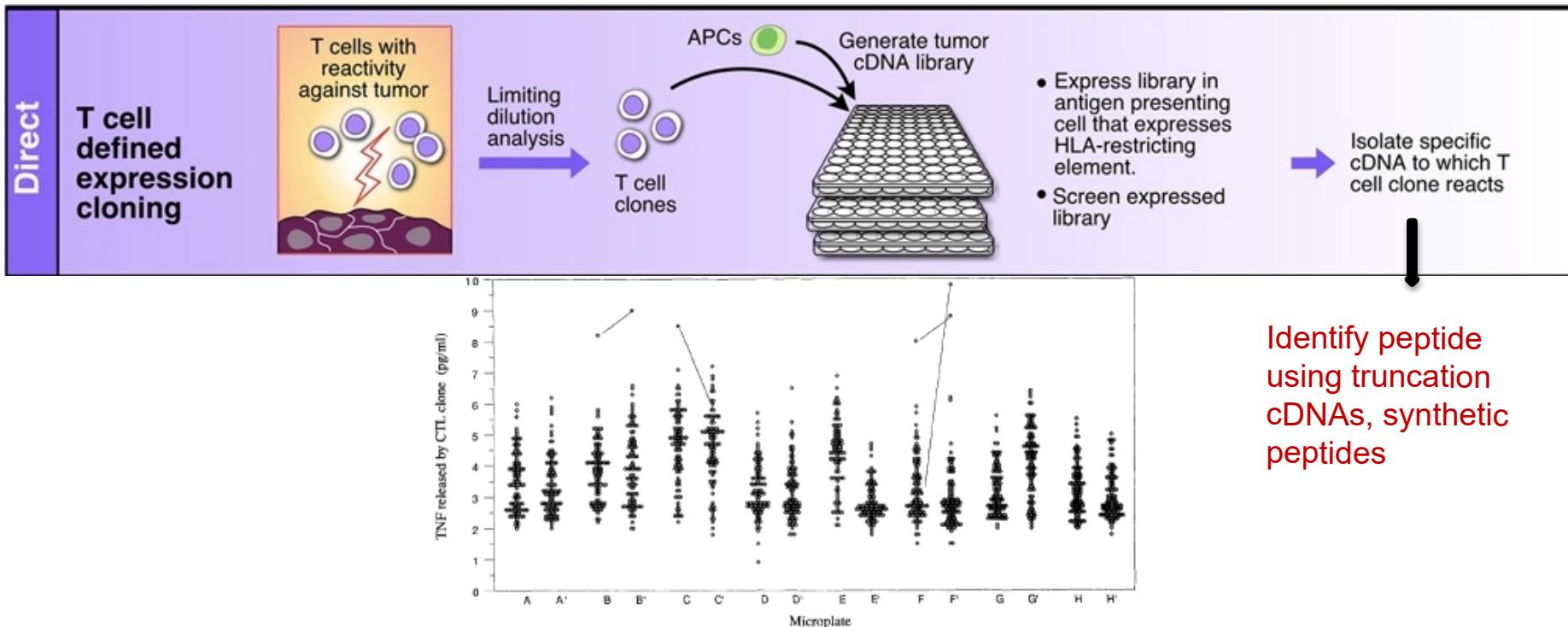


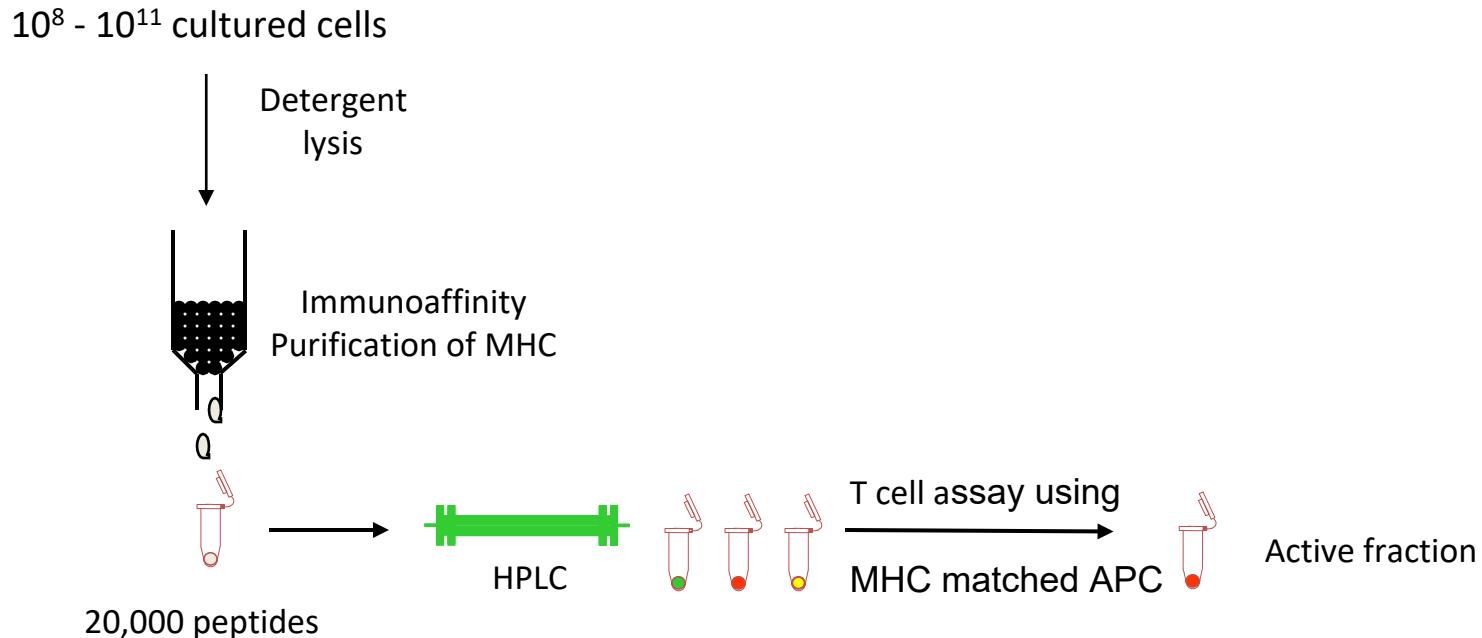
Figure 2. Stimulation of CTL 1/95 by COS cells cotransfected with HLA-A2 and pools of approximately 25 different cDNAs obtained from melanoma cell SK29-MEL. Each symbol represents the TNF content of one individual microculture, and the duplicate microcultures are named A and A', B and B', etc. Duplicates of seemingly positive microcultures are connected with a line. Each pool of cDNAs was transfected into duplicate microcultures. Both the HLA-A2 gene and the cDNAs were cloned into expression vector pcDNA1/Amp. They were cotransfected with DEAE-Dextran into subconfluent COS cells. 48 h after transfection, CTL 1/95 (2,000 cells per well) was added. The culture supernatants were harvested 1 d later and tested on W13 cells for their TNF content.

Brusic & Wu Frontiers Biosci 17:635 (2012)

Coulie et al J Exp Med 180:35 (1994)

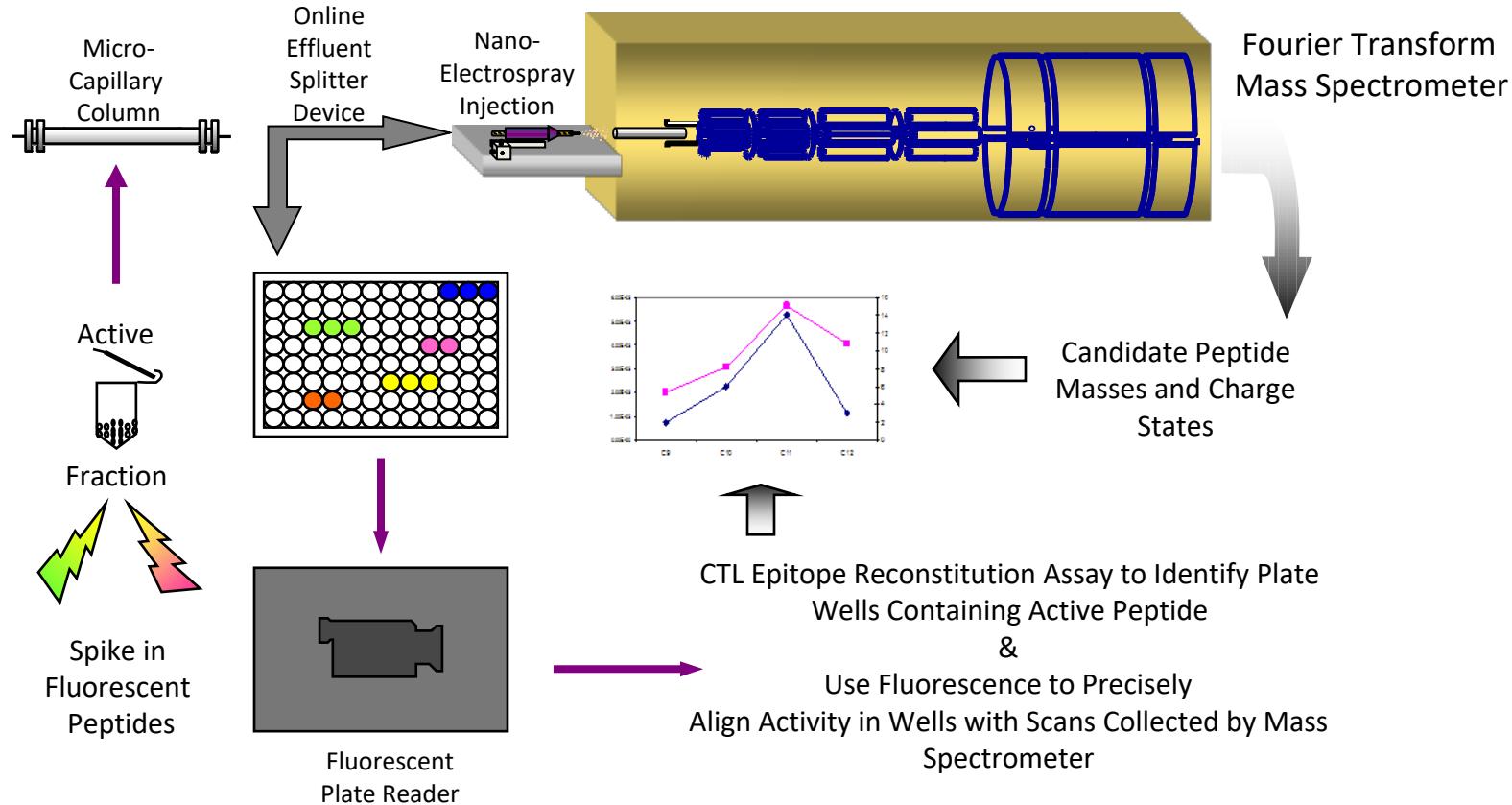
Mass spectrometry approach

V. Engelhard/D. Hunt

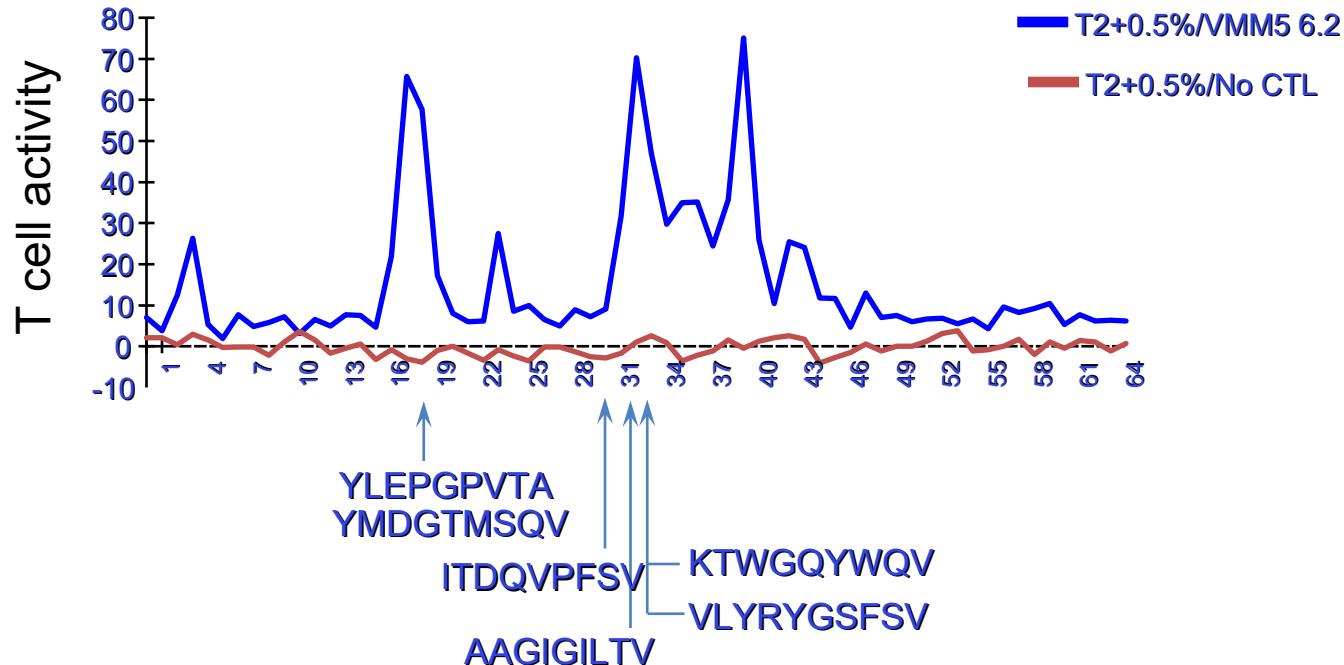


Mass spectrometry approach

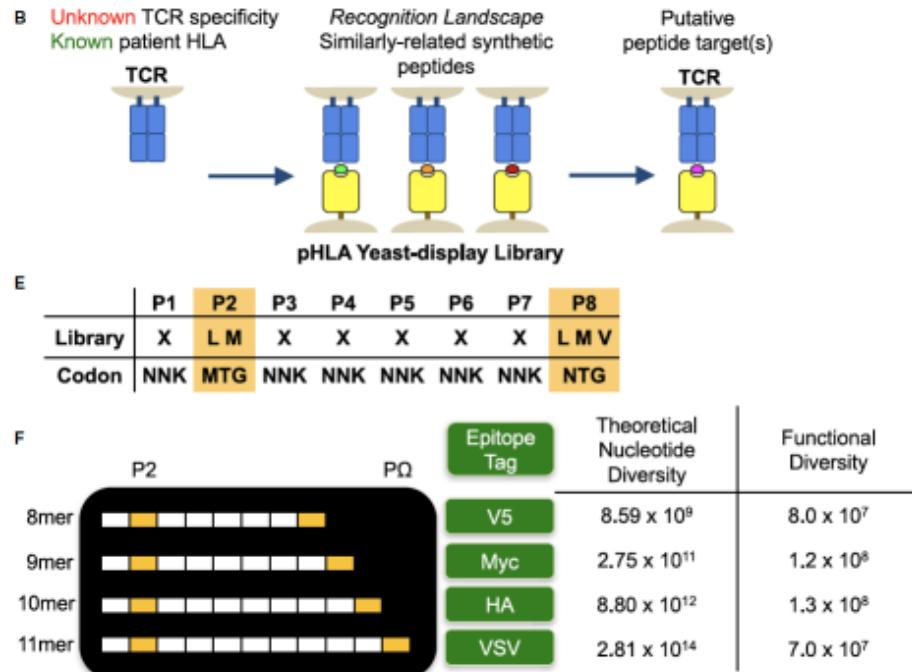
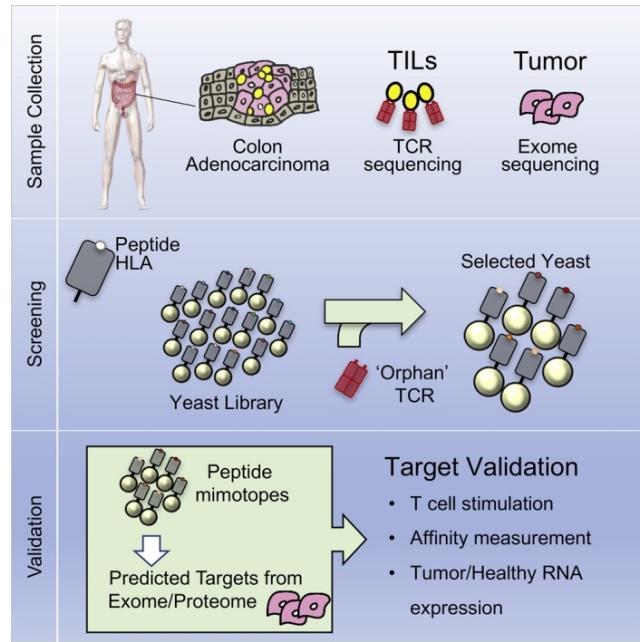
V. Engelhard/D. Hunt



Antigens identified among naturally processed melanoma peptides



Approach to identification of antigens recognized by “orphan TCR” from TIL

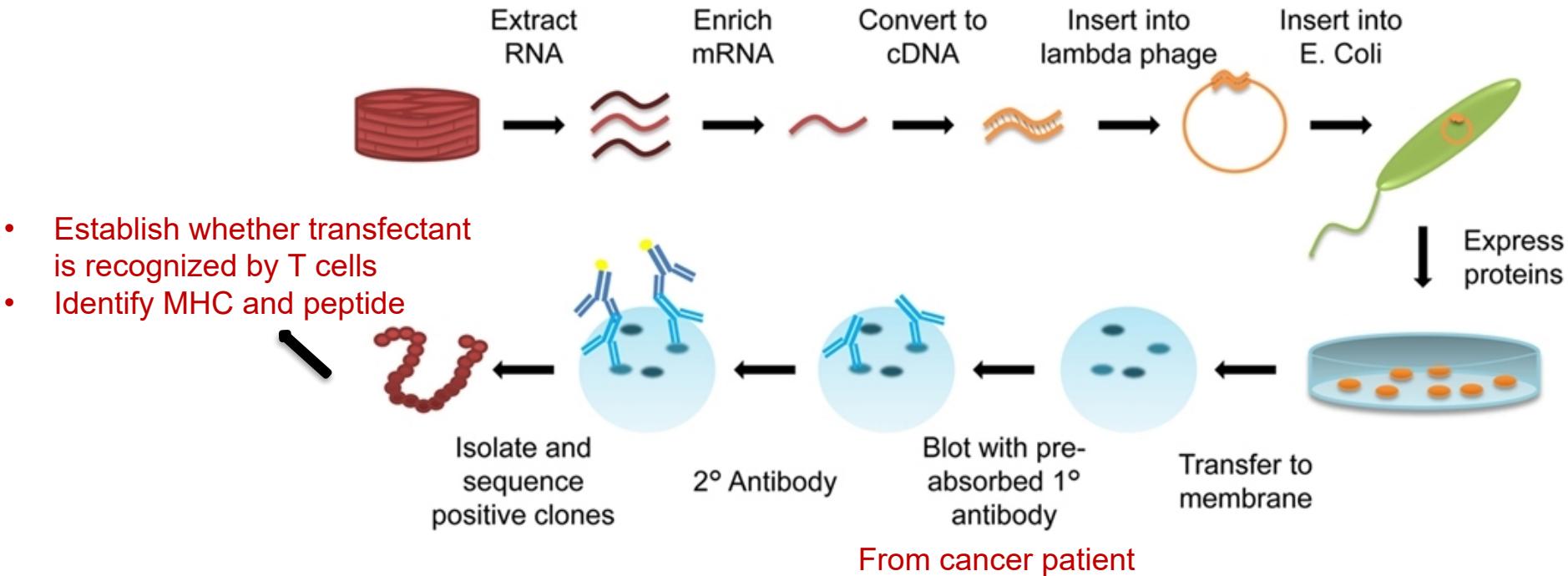


Indirect Methods

Every method needs a hook.....

SEREX

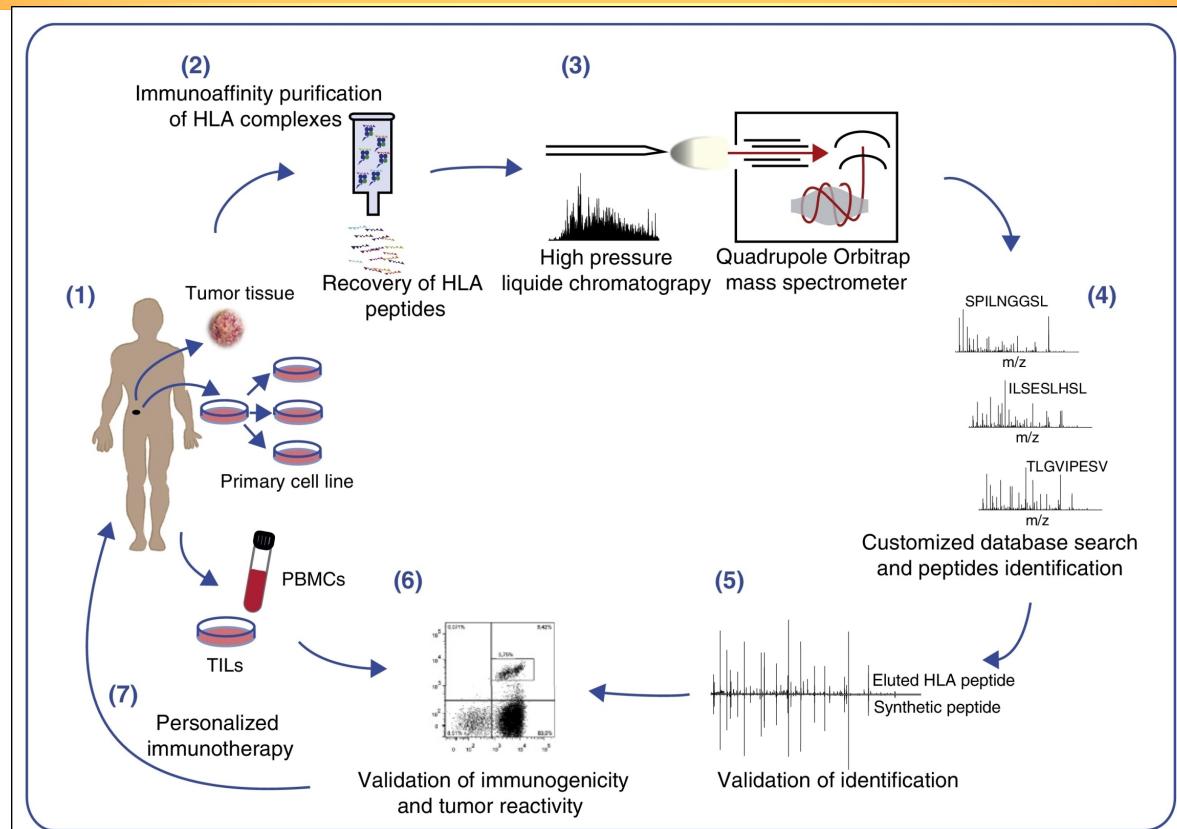
L. J. Old



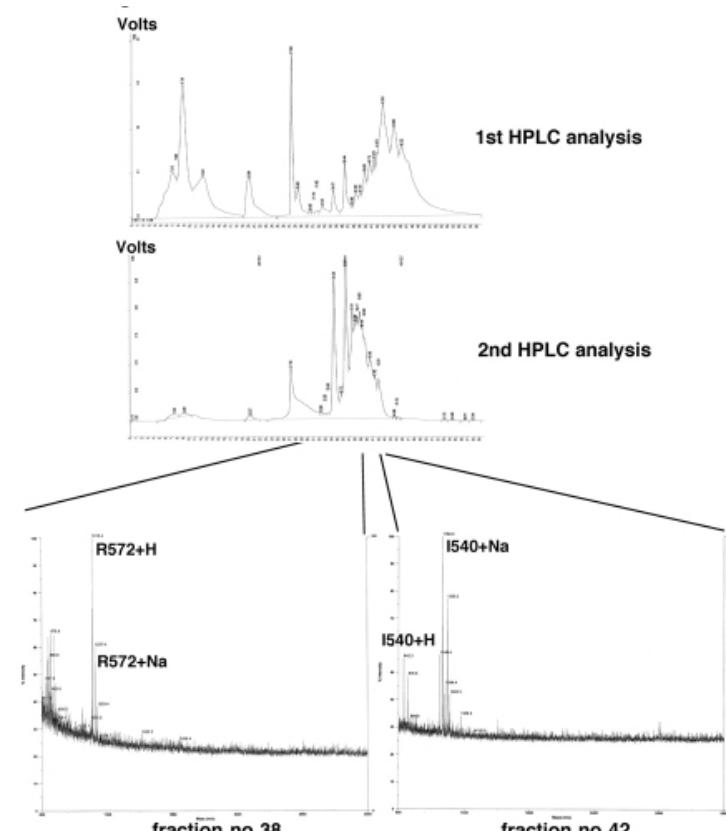
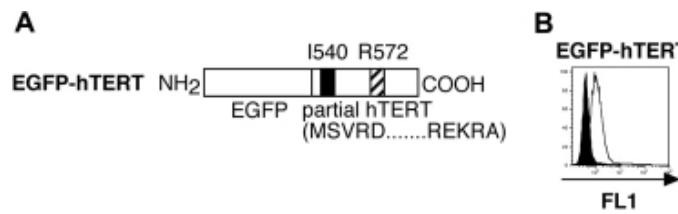
Success!: NY-ESO1

Gates et al. Front Immunol 8: 1332 (2017)

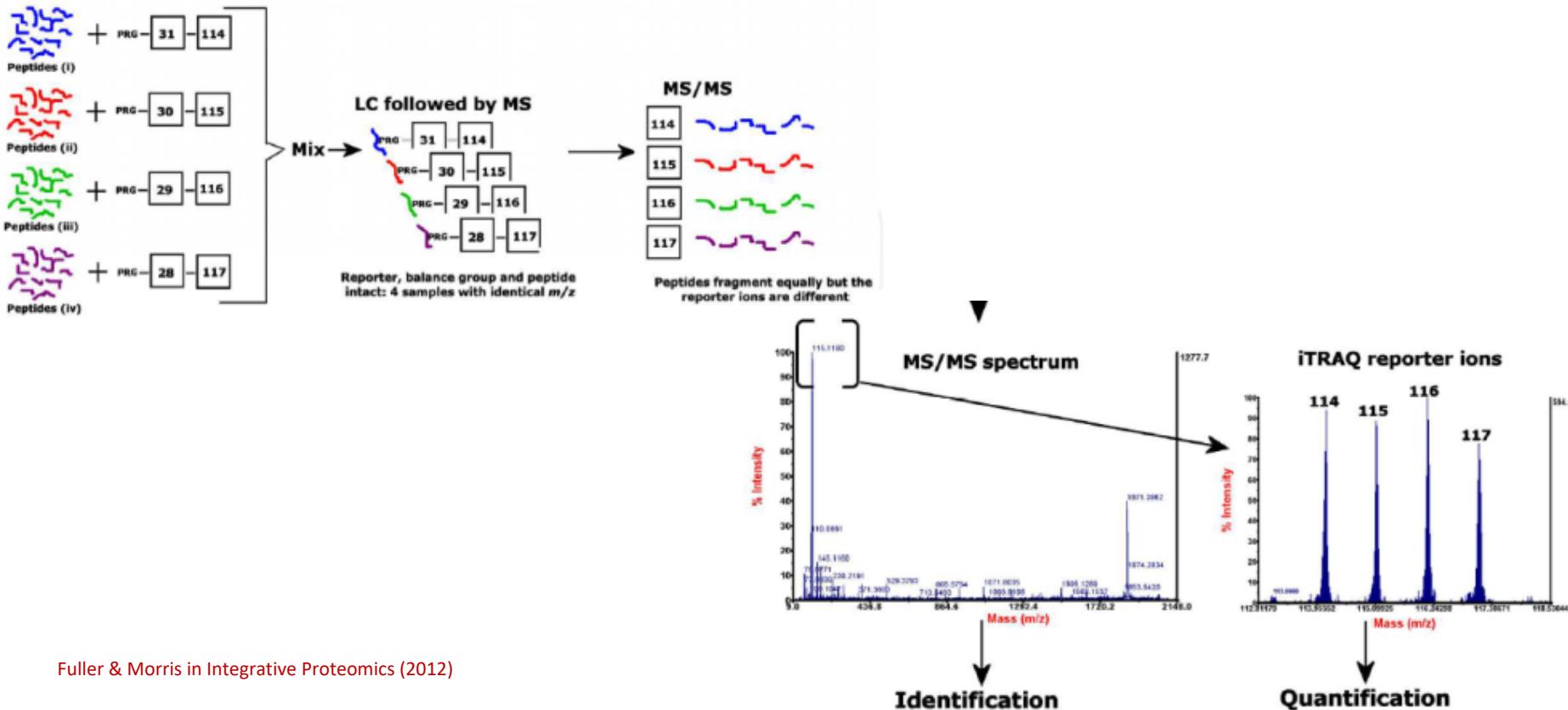
Mass spectrometry for general MHC peptide analysis and potential antigen identification



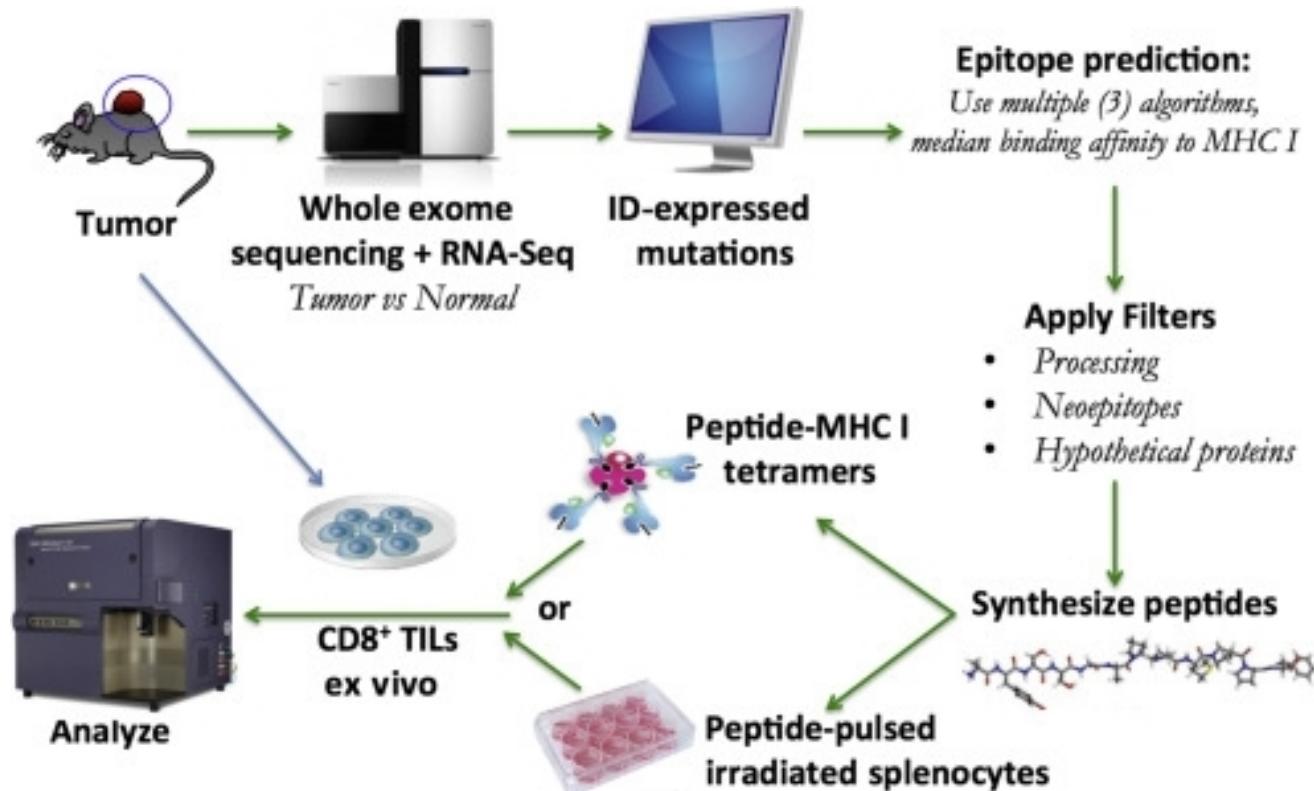
Targeted mass spectrometry: Finding what you are looking for



iTRAQ enables quantitative comparison of the same peptide in different samples

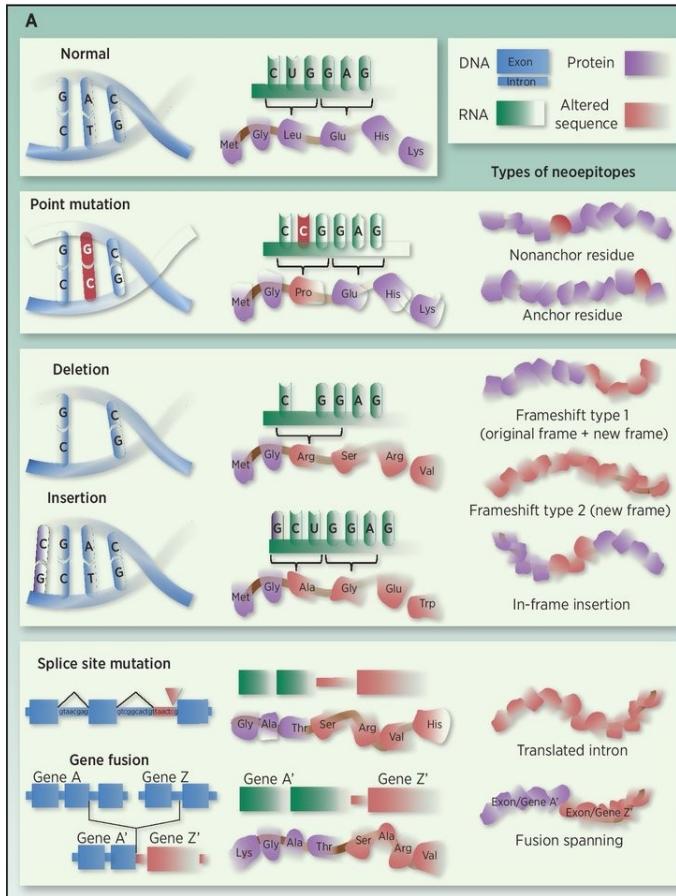


Rapid neoantigen identification enabled by NGS



Sources of NeoAgs

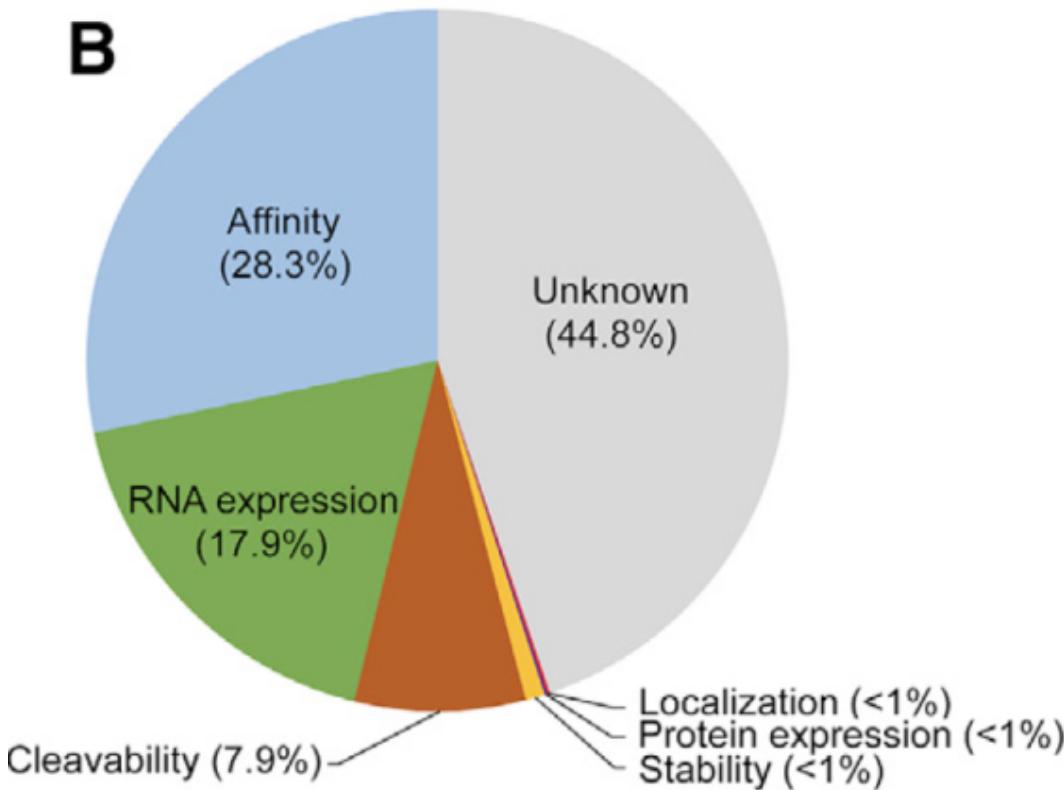
Missense



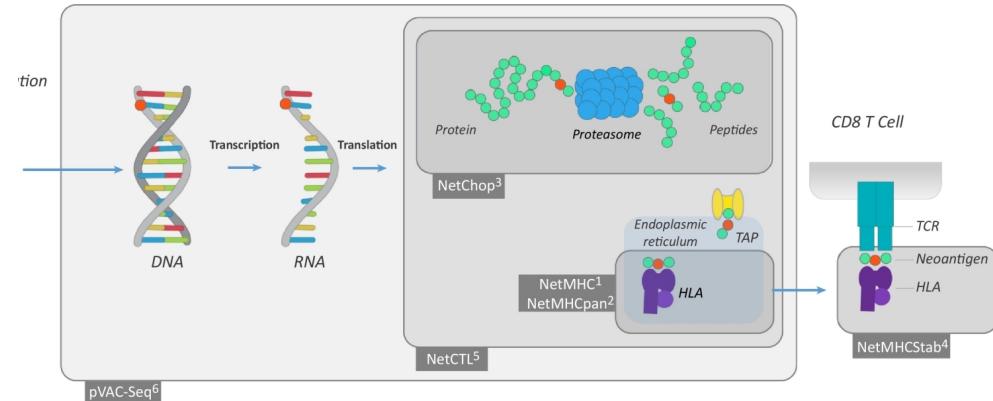
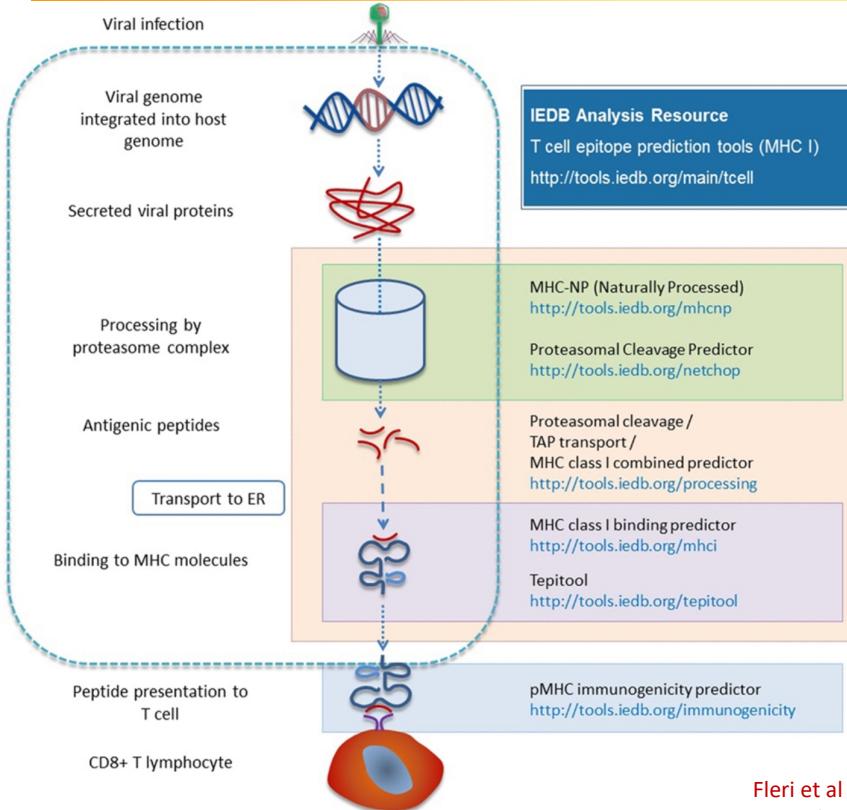
Frameshift

Splice variants

Analysis of factors predicting display of identified MHC-I associated peptides



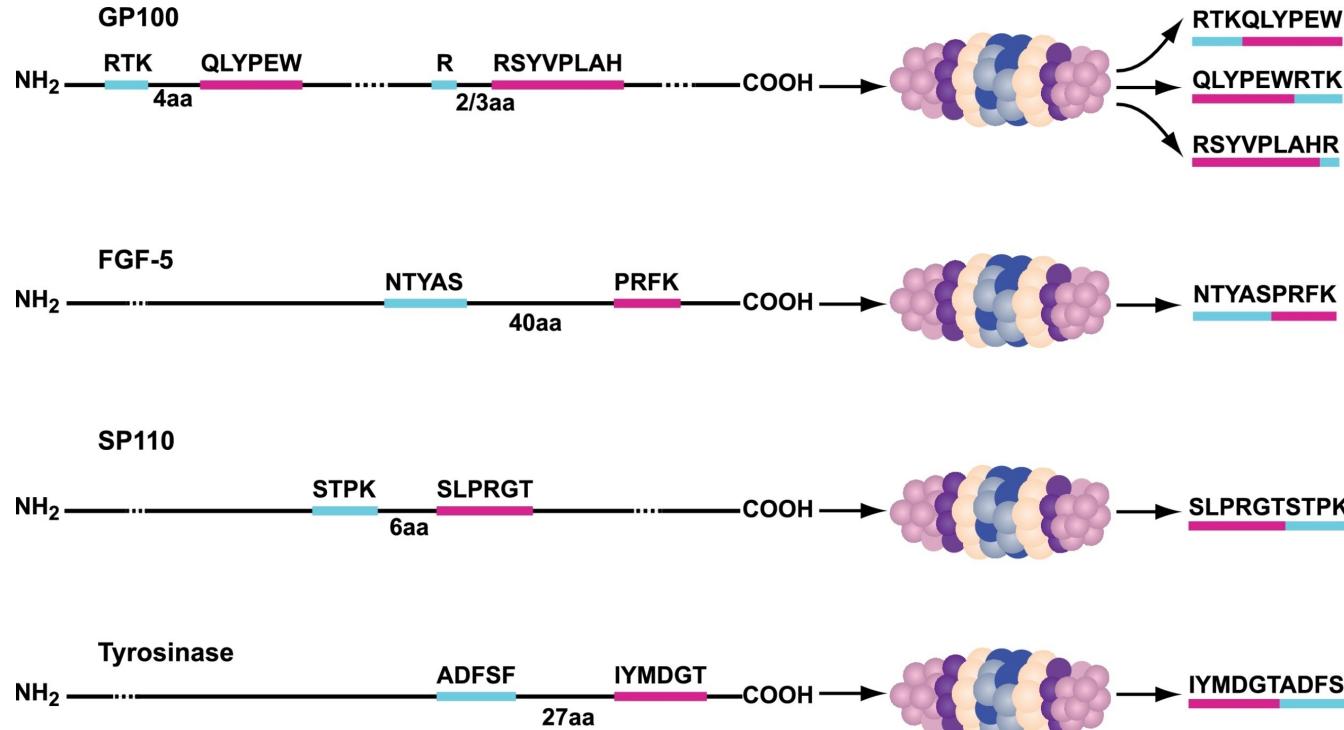
Software tools to predict whether point mutations will lead to MHC-I presented peptides



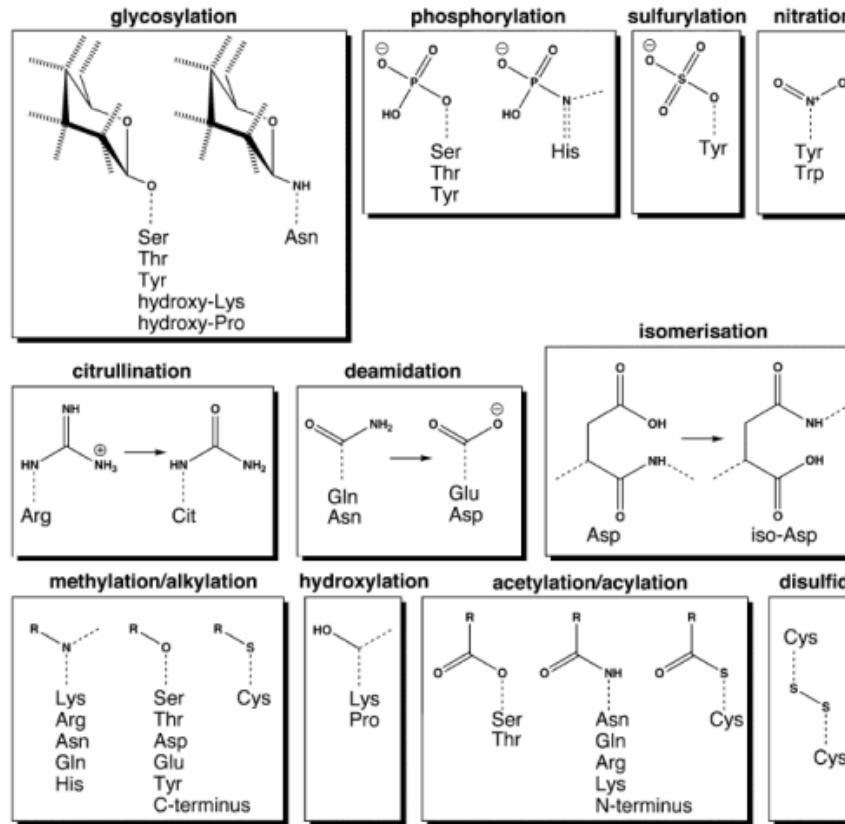
Ref. #	Tool	[Refs]	Elements of TSNA biology modeled
1	NetMHC	49	HLA binding
2	NetMHCpan	93	HLA binding across more HLA alleles
3	NetChop	94	Proteasomal peptide processing
4	NetMHCstab	54	HLA/peptide complex stability
5	NetCTL	53	Proteasomal processing, TAP transport, HLA binding
6	pVAC-Seq	61	DNA mutation, RNA expression, HLA binding

Trends in Immunology

Proteasomal splicing during protein degradation creates new antigens



Post-translational modifications of proteins can survive to be presented on MHC peptides



MHC-associated phosphorylated peptides *a better class of tumor antigen*

