Microenvironmental metabolites shaping T cell fate and function in cancer

SITC Tumor Microenvironment Workshop April 22, 2022

UPMC | HILLMAN CANCER CENTER

Greg M. Delgoffe, Ph.D



Disclosures

Consulting/SAB: BlueSphere Bio, Kalivir, Century Therapeutics, Nanna Therapeutics, Astellas, RemplirBio, and Novasenta

- **Research Support:** Kalivir, Century Therapeutics, TCR2, bluebird bio, Nanna Therapeutics, Pfizer, and Novasenta
- Clinical Trials: Merck, Pfizer, BMS, and Novasenta
- Founder and Advisor, Equity: RemplirBio, Novasenta





delgoffe-lab.com

How does the tumor microenvironment's metabolic landscape affect immunotherapy response?



- The tumor microenvironment engages in some very immunosuppressive functions
 - Altering stromal cell function to support tumor growth
 - Changing angiogenesis patterns
 - Providing chronic antigen stimulation
 - Recruitment of immunosuppressive cell types
- However, a common phenotype of cancer is that it is **hungry**
- The tumor microenvironment, driven by the metabolic derangement of tumor cells, generates a distinct metabolic landscape, creating both a sink for essential nutrients as well as a buildup of toxic byproducts
 - Tumors increase oxygen consumption, which creates local zones of hypoxia and production of high concentrations of reactive oxygen species
 - Tumors also become highly glycolytic, creating both a hypoglycemic interstitia but also generating high concentrations of lactic acid
- A tumor cell's 'appetite' may be variable in the patient population!

delgoffe-lab.com

Tumor cell (oxidative) metabolism is associated with immunotherapy response

- We performed metabolic profiling of melanoma patient tumor cells directly from biopsies prior to anti-PD1
- Tumor cell metabolism was highly variable in IO-naïve patients
- Patients with highly oxidative tumor cells were more likely to fail anti-PD1 therapy
- Immune activation and function are extremely energetically demanding!



What is the metabolic environment in cancer?





Watson and Delgoffe, JCI 2022

delgoffe-lab.com

How can you isolate how metabolism specifically affects anti-cancer immunity?

- Not only can metabolism act on multiple phases of the immune response, but there are also dozens of other signals T cells endure simultaneously within the tumor microenvironment
 - Oncogenic signals
 - Inflammatory vs suppressive cytokines
 - Physical barriers
 - Immunologic cell-surface signals
- Can we ask how the specific metabolism of the tumor microenvironment acts upon different phases of T cell immunity?

Tumor interstitial fluid: the milieu of the tumor microenvironment

- What do immune cells see metabolically in cancer?
- Enter Alex Muir
 - Metabolic regulation of cancer progression, notably in KPC mice Kras^{G12D}p53^{f/f}Pdx1^{Cre}
 - low response to immune checkpoint blockade
 - Highly metabolically deregulated
 - Developed a cost-effective means to generate metabolite libraries for quantitation of metabolomics experiments
- He published a sweeping metabolomic characterization of tumor interstitial fluid (Sullivan et al, *eLife* 2019)





TIFM (Tumor Interstitial Fluid Media)

(Sullivan et al. eLife

delgoffe-lab.com

How do T cells respond acutely to TIF media?



Culture in the TIFM arrests proliferation



delgoffe-lab.com

Culture in the TIFM promotes dysfunction



What's missing in TIF media that T cells need?



delgoffe-lab.com

Arginine supplementation of TIFM restores T cell proliferation



delgoffe-lab.com

Arginine supplementation of TIFM fails to rescue cytokine production



delgoffe-lab.com

It's not always about what's missing...



- The immunometabolic 'party line' currently is all about a land grab: tumor cells and T cells are competing for the same nutrients
- A lesser appreciated aspect of the tumor microenvironment is the build-up of metabolic byproducts – as tumors are poorly perfused, these potentially toxic wastes can also act as dominant immunoregulatory molecules
- We are learning these metabolites may be the true culprit in immunoregulation within the TME and beyond...

Are there immunosuppressive metabolites present in TIF which *cause* dysfunction?





delgoffe-lab.com @De

Phosphoethanolamine: a novel oncometabolite?

- pEtn is a commonly upregulated metabolite in TIFs
- pEtn is the head group of phosphatidylethanolamine, a membrane phospholipid
- PE is the second most abundant phospholipid in mammalian cells
 - PC/PE in mice (3:1)
 - PC/PE in flies (1:3)
- pEtn contributes to PE synthesis through the Kennedy pathway
 pEtn



Phosphatidylethanolamine(PE)



delgoffe-lab.com

Phosphoethanolamine (pEtn) supports T cell proliferation



delgoffe-lab.com

Despite stimulating proliferation, pEtn severely compromises CD8 T cell cytokine production



delgoffe-lab.com

Exposure to pEtn for one day is enough to cause dysfunction



Is the effect driven by pEtn or the Kennedy pathway entirely?

 Choline and phosphocholine in the choline-arm of Kennedy pathway are also enriched in the TIF



2019;8:e44235)



delgoffe-lab.com @DelgoffeLab

Phosphotidylcholine (PC) precursors do not cause CD8 T cell dysfunction

- Repeated experiments with RPMI media cultured with choline or phosphocholine
- The PC arm of the Kennedy pathway does not induce T cell dysfunction



delgoffe-lab.com

Both precursor and product of pEtn metabolism <u>can</u> induce T cell dysfunction

- Our data highlight the importance of the ethanolamine arm of the Kennedy pathway
- While Etn and CDP-Etn are not enriched in TIF, we asked if a similar treatment regimen would induce T cell dysfunction





Thus, it is not pEtn itself but rather **flux through the ethanolamine arm of the Kennedy pathway** that induces T cell dysfunction





RNAseq identifies some interesting immune or metabolic hits, but no smoking gun

- Performed RNAseq on RPMI or pEtn-expanded T cells
- While these cells had some hints of a dysfunctional status (*Pdcd1, Lag3, Tigit*), the data were certainly not consistent with exhaustion or anergy
- No strong (any?) hits with GSEA/GO analysis
- We had to go beyond transcriptomes to understand this cell-intrinsic immune regulation

Phosphoethanolamine supplementation dramatically shifts the PE/PC balance of T cells



Stacy Wendell

delgoffe-lab.com

pEtn treatment shifts the entire metabolome of T cells



- Global metabolomics profiling revealed pEtn supplementation drove massive ethanolamine flux into membrane and second messenger phospholipids
- The result was a striking reprogramming of the cells' carbohydrate metabolism, and suppression of mitochondrial biogenesis, which we've previously associated with T cell dysfunction



How much of the "TIFM" phenotype is pEtn?

- Formulated TIFM without pEtn
- T cells cultured in TIFM still possessed a proliferative defect
- However, T cells cultured in TIFM retain the cytokine production of control cells
- Not a complete rescue, so lots left to do!



The dysfunction induced by pEtn culture is at the level of TCR and/or CD28 signaling



delgoffe-lab.com

Do pEtn treated cells have defects in TCR signaling?



delgoffe-lab.com 🧔

How does pEtn flux reduce distal but not proximal TCR signaling?



Heightened flux through the Kennedy pathway depletes DAG, a critical second messenger!

wpergoffeLab

Restoration of DAG can rescue pEtninduced T cell dysfunction



delgoffe-lab.com @DelgoffeLab

Conclusions

- Media based on tumor interstitial fluid (TIF) allows for a direct view of tumor microenvironment metabolism's role on T cell function and fate
- TIFM induces persistent, heritable dysfunction in CD8+ T cells
- TIFM confirmed the importance of arginine in T cell proliferation, but not T cell dysfunction
- TIFM identified a novel oncometabolite, phosphoethanolamine, which drives flux through the ethanolamine arm of the Kennedy pathway, producing PEs
- pEtn treatment paradoxically drives T cell proliferation but inhibits cytokine production
- pEtn induces a distinct form of T cell dysfunction induced through amplified (pathologic?) metabolic flux that depletes critical second messengers, like DAG



The Delgoffe Lab Microenvironment

Dayana Rivadeneira, Ph.D Ronal Peralta Andrew Frisch Rachel Cumberland Mary Philbin Supriya Joshi, Ph.D *Nicole Scharping, Ph.D Ashley Menk Yiyang Wang* Paolo Vignali Kristin DePeaux Dinos Lontos, MD **Yupeng Wang**

Jessica Jana Alok Kumar, Ph.D Drew Wilfahrt, Ph.D Bingxian Xie, Ph.D *McLane Watson, Ph.D*

Key Collaborators

Alex Muir, Ph.D Yana Najjar, MD Dan Zandberg, MD Stacy Wendell, PhD Jeff Rothstein, PhD Brett Morrison, MD PhD

Amanda Poholek, PhD Rhodes Ford Bob Ferris MD PhD

Bob Ferris, MD PhD

Abigail Overacre-Delgoffe, PhD Tim Hand, PhD Acknowledgments

Center for Biologic Imaging UPMC Hillman Flow Cytometry/Animal Facilities **Patients and their families**

Funding sources:

NIH Director's New Innovator Award (DP2AI136598)
Sidney Kimmel Foundation
Stand Up 2 Cancer Innovative Research Grant
Melanoma and Head and Neck Cancer SPOREs
Alliance for Cancer Gene Therapy
DoD Team Science Award (CA170483P1)
R21AI135367-01
Melanoma Research Alliance
Cancer Research Institute Lloyd J. Old STAR Award
Mark Foundation for Cancer Research Emerging Leader













@ DelgoffeLab

delgoffe-lab.com