



Memorial Sloan Kettering
Cancer Center

Targeting Tumor Metabolism to Overcome Resistance to Immune Checkpoint Blockade

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Immuno-Oncology Service
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and Swim Across America lab at MSK
Prof. of Immunol. Research, Weill Cornell

**LUDWIG
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SITC 2022 Interim
Workshop April 21 – 22,
2022 | San Diego, CA



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ACROSS AMERICA
★ MAKING WAVES TO FIGHT CANCER ★

Disclosures

- IMVAQ therapeutics co-founder
- Advisory board immunos therapeutics, Algerian Research Agency in Health and Life Science
- Consulting for Pfizer, Daichii, Immunogenesis, Kowa.
- Inventor on a patent applications related to work on Oncolytic Viral therapy, Alpha Virus Based Vaccine, Neo Antigen Modeling, CD40, GITR, OX40, PD-1 and CTLA-4.

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Bristol-Myers Squibb

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Peregrine Pharmaceuticals, Inc.

Adaptive Biotechnologies

Leap Therapeutics, Inc.

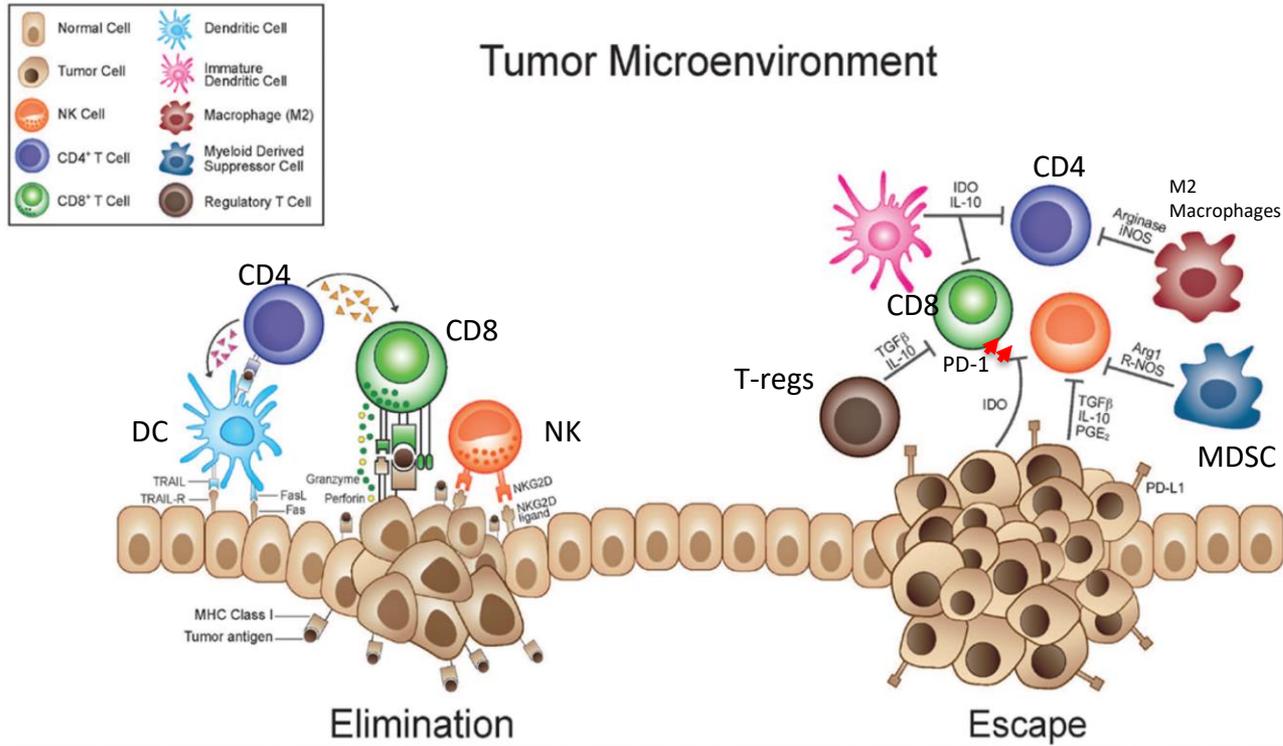
Aprea.

Roche-Boehringer

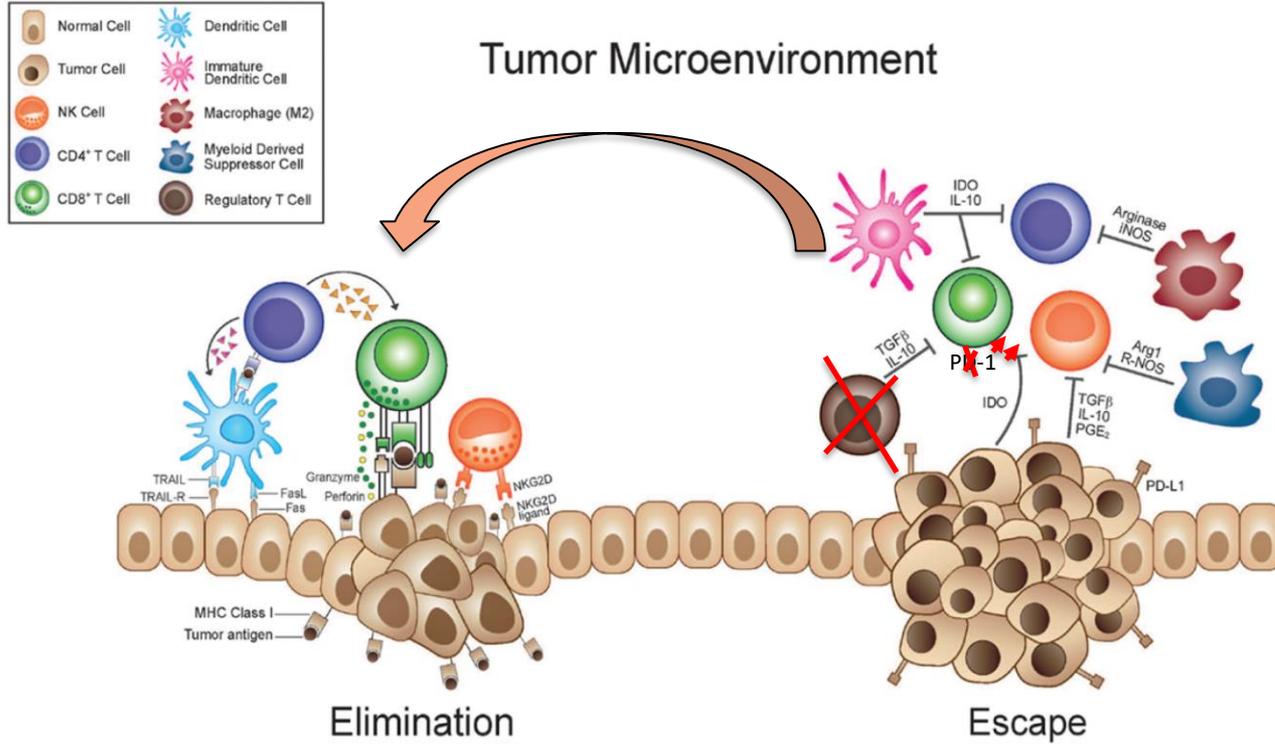


Immunoediting

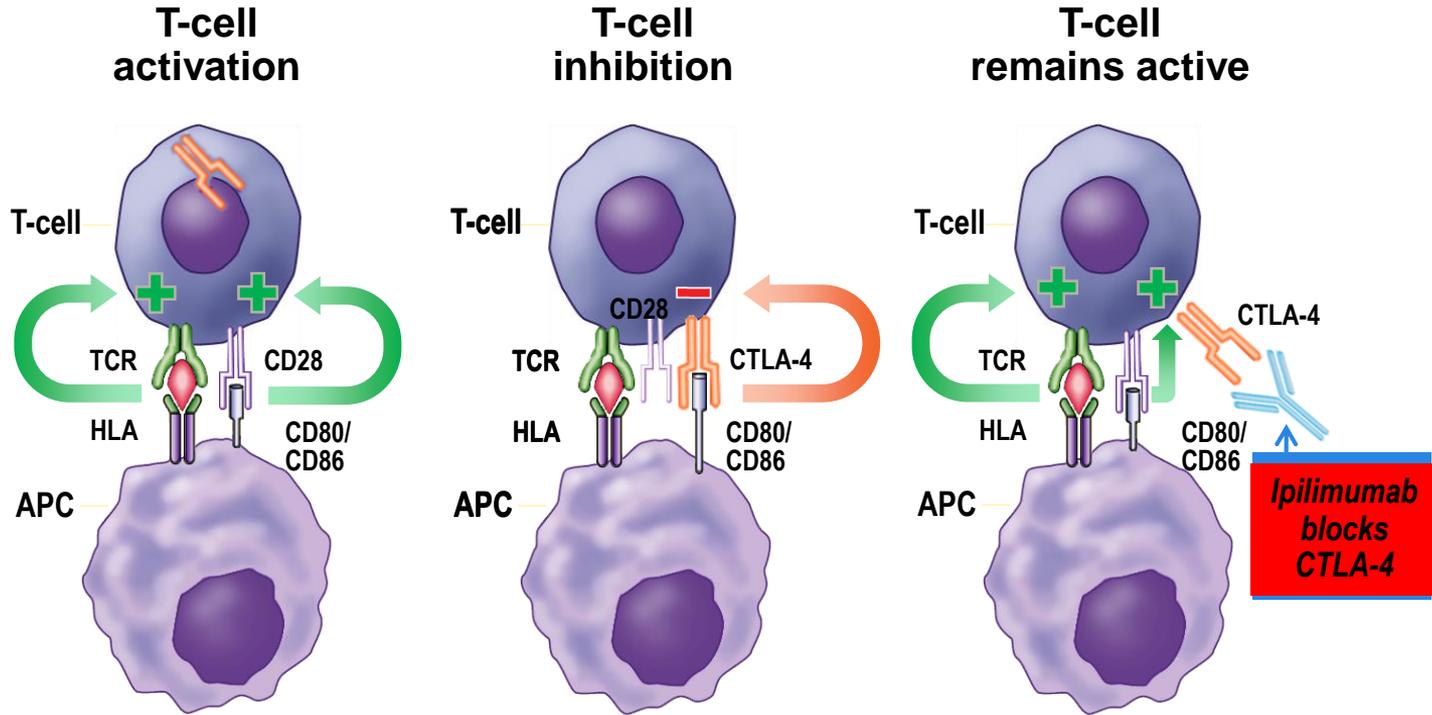
Immune Suppressive Microenvironment



Immune Suppressive Microenvironment



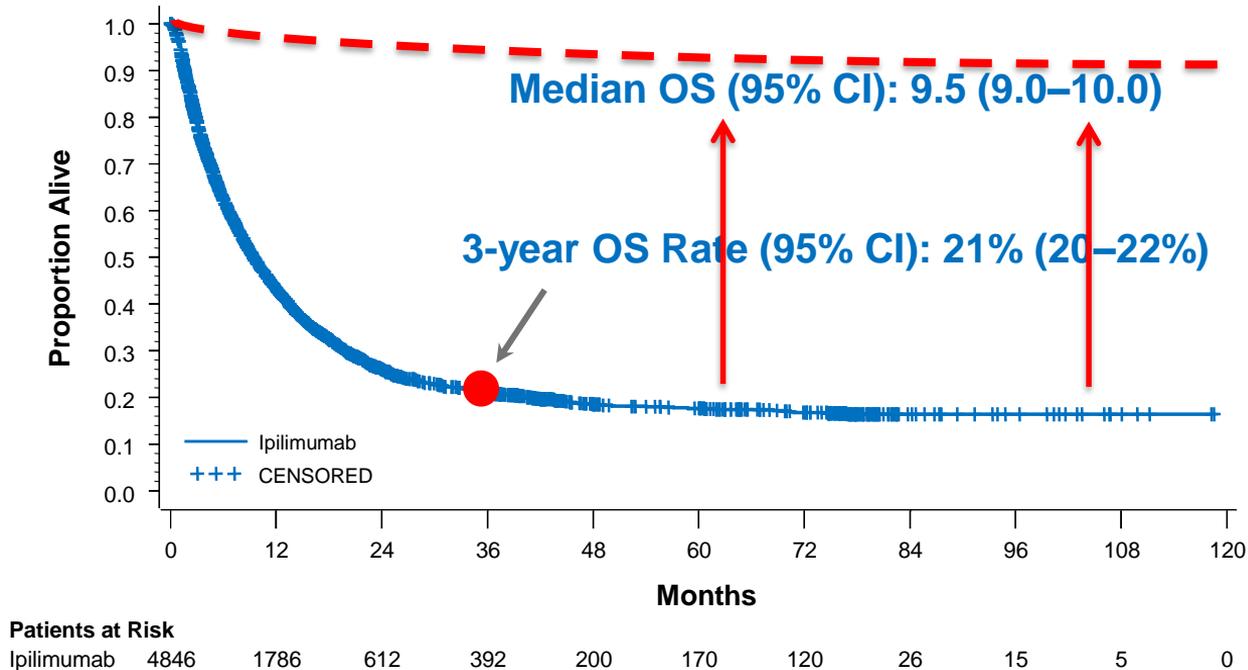
Ipilimumab Augments T-Cell Activation and Proliferation



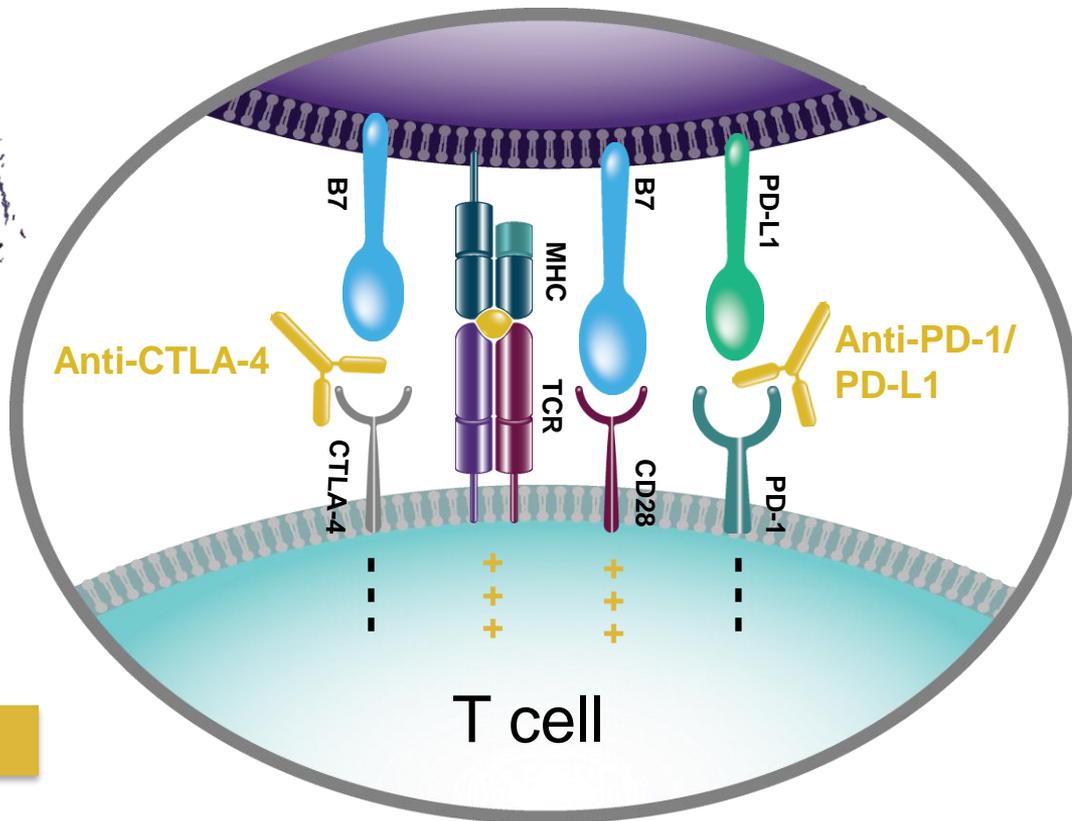
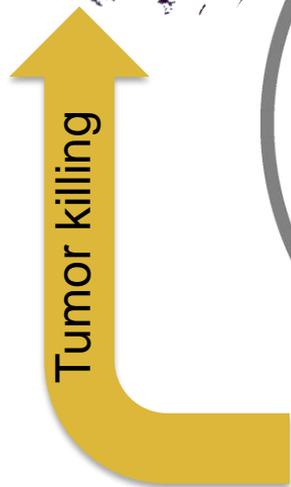
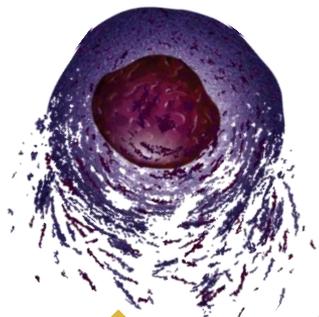
Adapted from O'Day et al. Plenary session presentation, abstract #4, ASCO 2010.

Immune checkpoint blockade is effective in a limited fraction of patients

**Ipilimumab Long Term Pooled Survival Analysis:
4846 Patients**



Inhibition of negative immune regulation



FDA approved anti-CTLA-4
Ipilimumab

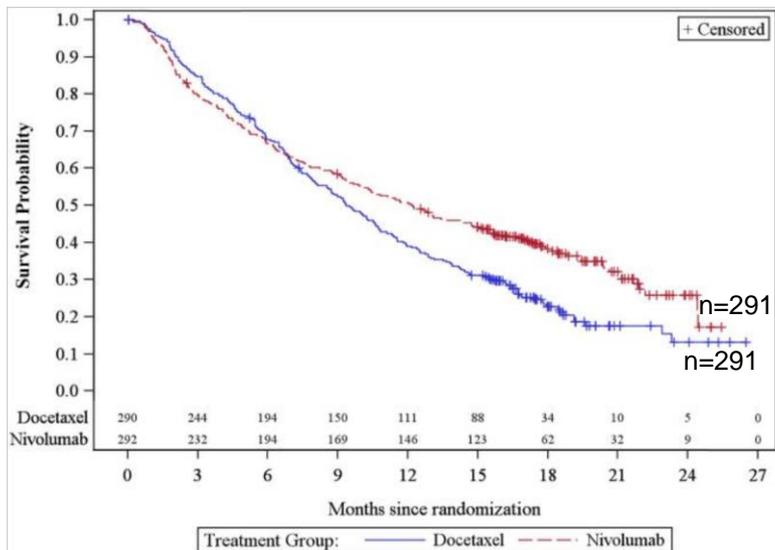
FDA approved anti-PD-1
Nivolumab
Pembrolizumab
Cemiplimab

FDA approved anti-PD-L1
Atezolizumab
Avelumab
Durvalumab



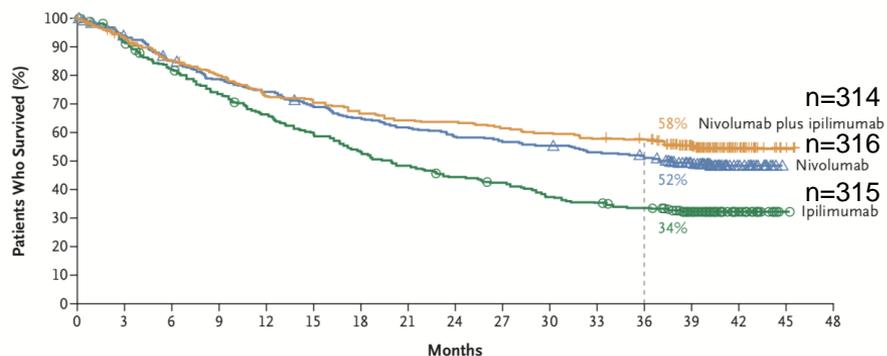
Immune checkpoint blockade is effective in a limited fraction of patients

α PD-1 in advanced NSCLC



CheckMate 057; Kazandjian et al., The Oncologist 2016

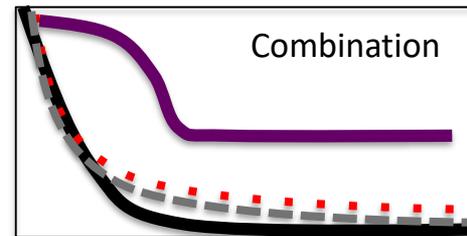
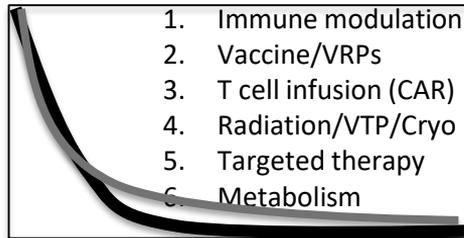
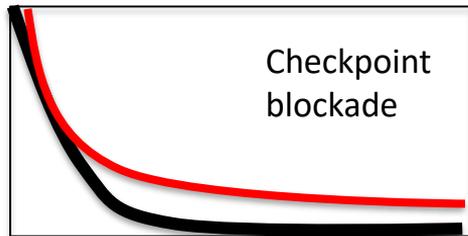
α CTLA-4 \pm α PD-1 in advanced melanoma



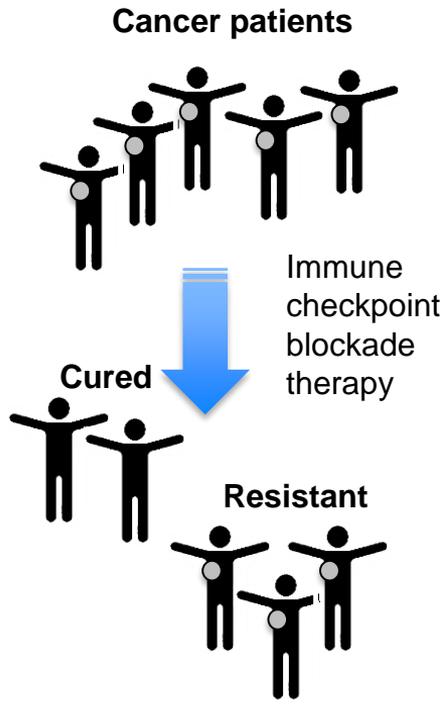
CheckMate 067
 Wolchok et al., NEJM 2017

Rationale for Combination with other therapies:

- Use other means to enhance tumor recognition
- Strategy to address low response rates of checkpoint blockade



Lab “focuses” on overcoming major mechanisms of resistance to anti-tumor immunity



Tumor intrinsic resistance
LETTER
 doi:10.1058/nature16404

Melanoma-intrinsic β -catenin signalling prevents anti-tumour immunity
 Stefan Spranger¹, Rhyee Ibar² & Thomas F. Gajewski^{1,3}

Article Cell

Loss of IFN- γ Pathway Genes in Tumor Cells as a Mechanism of Resistance to Anti-CTLA-4 Therapy
 Jianjun Gao,^{1,2} Lewis Zhichang Shi,^{1,2} Hao Zhao,^{1,2} Jianfeng Chen,¹ Liangwen Xiong,¹ Qiuming He,¹ Tenghui Chen,¹ Jason Rozak,¹ Chantale Benatchez,¹ Scott E. Woodman,¹ Pei-Ling Chen,¹ Patrick Hwu,¹ James P. Allison,¹

Article Cell

Genomic and Transcriptomic Features of Response to Anti-PD-1 Therapy in Metastatic Melanoma
 Willy Hugo,^{1,2} Jesse M. Zaretsky,^{1,2,3} Lu Sun,^{1,2} Chunyang Song,^{1,2} Blanca Hornet Moreno,¹ Siwen Hu-Lieskovan,¹ Beata Berent-Maoz,¹ Jia Pang,¹ Bartosz Chmielowski,¹ Geisuo Cheny,¹ Elizabeth Seig,¹ Shirley Lorenz,¹ Xiangji Kong,^{1,2} Mark C. Kelley,¹ Jeffrey A. Sosman,¹ Douglas B. Johnson,¹ Antoni Ribas,^{1,2,3,4,5,6} and Roger S. Lo,^{1,2,5,6*}

The NEW ENGLAND JOURNAL of MEDICINE
 ESTABLISHED IN 1812 SEPTEMBER 1, 2016 VOL. 375 NO. 9

Mutations Associated with Acquired Resistance to PD-1 Blockade in Melanoma
 Jesse M. Zaretsky, B.S., Angel Garcia-Diaz, Ph.D., Daniel S. Shin, M.D., Helena Escuin-Ordinas, Ph.D., Willy Hugo, Ph.D., Siwen Hu-Lieskovan, M.D., Ph.D., David Y. Tormason, M.D., Gabriel Abad-Rodriguez, M.Sc., Saleem Sandoval, Ph.D., Lucas Barthly, M.Sc., Justin Saco, B.S., Blanca Hornet Moreno, M.D., Riccardo Mezzadra, M.Sc., Bartosz Chmielowski, M.D., Ph.D., Kathleen Ruchalski, M.D., I. Peter Shintaku, Ph.D., Phillip J. Sanchez, Ph.D., Cristina Paig-Sava, Ph.D., Grace Cherry, R.N., N.P., Elizabeth Seig, B.A., Xiangji Kong, M.Sc., Jia Pang, B.S., Beata Berent-Maoz, Ph.D., Begolia Comin-Andujk, Ph.D., Thomas G. Graeber, Ph.D., Paul C. Tumeq, M.D., Ton N.M. Schumacher, Ph.D., Roger S. Lo, M.D., Ph.D., and Antoni Ribas, M.D., Ph.D.

Immune-mediated resistance

Cancer Immun Immunother (2014) 63:247–257
 DOI 10.1007/s00262-013-1508-5

ORIGINAL ARTICLE

Frequencies of circulating MDSC correlate with clinical outcome of melanoma patients treated with ipilimumab
 Christiane Meyer · Laurie Cagnon · Carla M. Costa-Nunes · Petra Baumgartner · Nicole Montandon · Loredana Leyvraz · Olivier Michelin · Emanuela Romano · Daniel E. Speiser

Clin Transl Oncol (2016) 18:251–258
 DOI 10.1007/s12094-015-1373-0

REVIEW ARTICLE

Tumor-associated macrophages in cancers
 W. Hu^{1,2,3}, X. Li^{1,2,3}, C. Zhano¹, Y. Vano¹, I. Hano^{2,3}, C. Wu^{1,2,3}

Research article

CTLA4 blockade and GM-CSF combination immunotherapy alters the intratumor balance of effector and regulatory T cells
 Sergio A. Quezada, Karl S. Peggs, Michael A. Curran, and James P. Allison

JEM Article

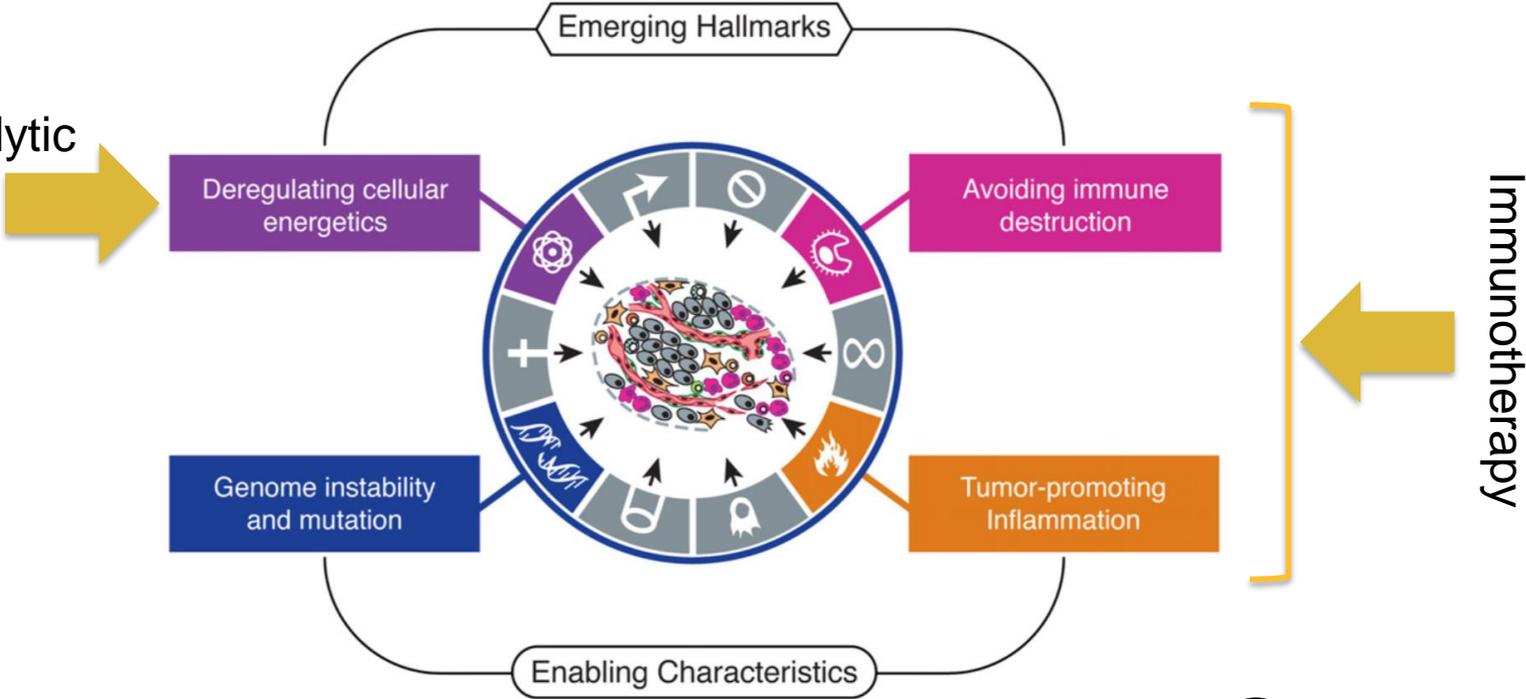
Indoleamine 2,3-dioxygenase is a critical resistance mechanism in antitumor T cell immunotherapy targeting CTLA-4
 Rikke B. Holmggaard,^{1,2} Dmitriy Zamarin,^{1,2,3} David H. Munn,⁴ Jedd D. Wolchok,^{2,3,5,6} and James P. Allison^{1,7}



Hypothesis – Inhibition of tumor glycolysis to overcome resistance to immunotherapy

1. Cellular energy metabolism reprogramming is a critical hallmark of cancer

Inhibition of tumor glycolytic metabolism



Hypothesis – Inhibition of tumor glycolysis to overcome resistance to immunotherapy

3. Immune checkpoints and co-stimulatory molecules regulate T cell metabolism

Immunity, Vol. 16, 769–777, June, 2002, Copyright ©2002 by Cell Press

The CD28 Signaling Pathway Regulates Glucose Metabolism Mitochondrial Priming by CD28

Kenneth A. Frauwirth,^{1,2,5} James L. Riley,^{1,2,5} Marian H. Harris,^{1,2} Richard V. Parry,^{1,3} Jeffrey C. Rathmell,^{1,2} David R. Plas,^{1,2} Rebecca L. Elstrom,¹ Carl H. June,^{1,2} and Craig B. Thompson^{1,2,4}

Ramon I. Klein Geltink,¹ David O'Sullivan,¹ Mauro Corrado,¹ Anna Bremser,^{2,3} Michael D. Buck,¹ Joerg M. Buescher,¹ Elke Firat,⁴ Xuekai Zhu,⁵ Gabriele Niedermann,^{1,6} George Caputa,¹ Beth Kelly,¹ Ursula Warthorst,² Anne Rensing-Ehl,² Ryan L. Kyle,¹ Lana Vandersarren,^{7,8} Jonathan D. Curtis,¹ Annette E. Patterson,¹ Simon Lawless,¹ Katarzyna Grzes,¹ Jing Qiu,¹ David E. Sanin,¹ Oliver Kretz,^{9,10} Tobias B. Huber,^{10,11,12} Sophia Iancu,^{7,8} Bot M. Lambrecht,^{7,8} Angelika S. Rambold,^{2,3} Edward J. Pearce,^{1,13} and Erika L. Pearce^{1,14,*} Cell 171, 385–397, October 5, 2017 © 2017 Elsevier Inc.



ARTICLE

Received 13 Aug 2014 | Accepted 19 Feb 2015 | Published 26 Mar 2015

DOI: 10.1038/ncomms7692

OPEN

PD-1 alters T-cell metabolic reprogramming by inhibiting glycolysis and promoting lipolysis and fatty acid oxidation

Nikolaos Patsoukis^{1,2,3}, Kankana Bardhan^{1,2,3}, Pranam Chatterjee^{1,2,3}, Duygu Sari^{1,2,3}, Bianling Liu^{1,2,3}, Lauren N. Bell⁴, Edward D. Karoly⁴, Gordon J. Freeman⁵, Victoria Petkova^{1,2,3}, Pankaj Seth^{2,3,6}, Lequn Li^{1,2,3} & Vassiliki A. Boussiotis^{1,2,3}

4-1BB signaling activates glucose and fatty acid metabolism to enhance CD8⁺ T cell proliferation

Beom K Choi^{1,6}, Do Y Lee^{1,2,6}, Don G Lee¹, Young H Kim³, Seon-Hee Kim¹, Ho S Oh¹, Chungyong Han¹ and Byoung S Kwon^{1,4,5}

Research Article

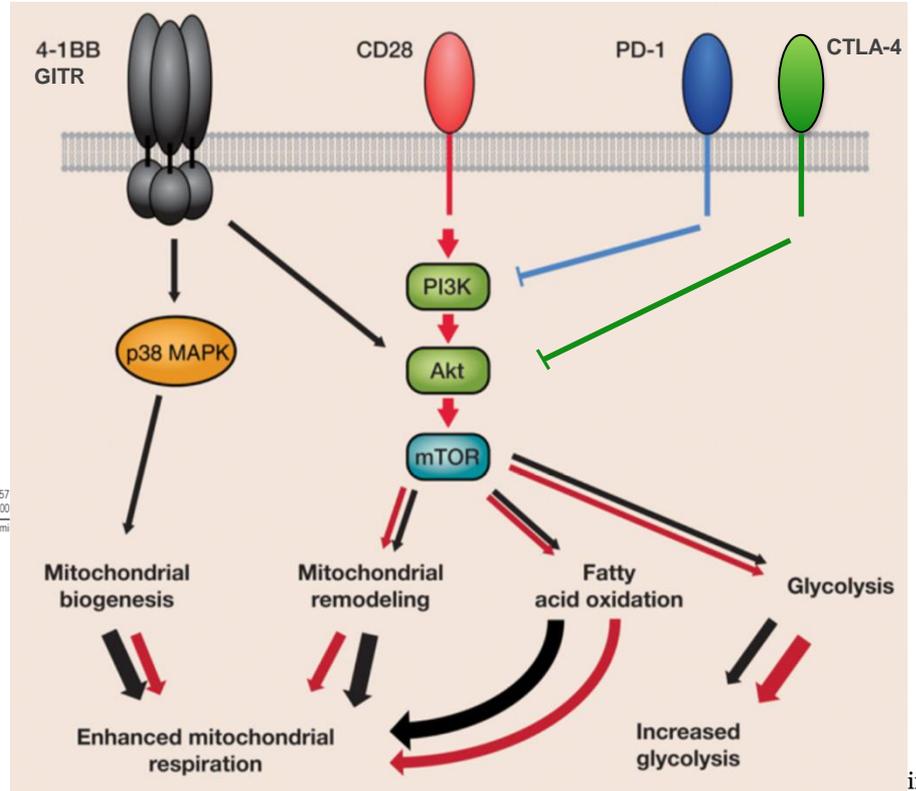
Cancer Immunology Research (2017) 14, 748–757
© 2017 CSI and USTC All rights reserved 1672-7681/17 \$32.00
www.nature.com/cimr

Cancer Immunology Research



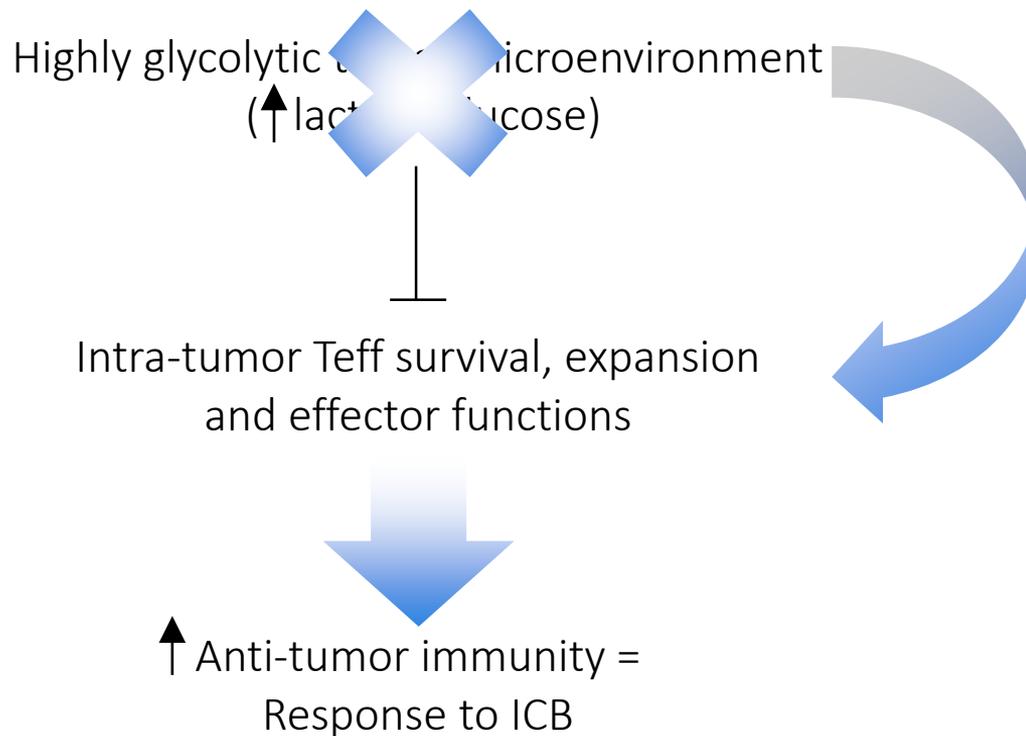
GITR Agonism Enhances Cellular Metabolism to Support CD8⁺ T-cell Proliferation and Effector Cytokine Production in a Mouse Tumor Model

Simran S. Sabharwal¹, David B. Rosen¹, Jeff Grein¹, Dana Tedesco¹, Barbara Joyce-Shaikh¹, Roanna Ueda¹, Marie Semana², Michele Bauer², Kathy Bang², Christopher Stevenson², Daniel J. Cua¹, and Luis A. Zúñiga¹ Cancer Immunol Res; 6(10) October 2018



Adapted from Teijeira A. et al, CIR 2019

Working Hypothesis

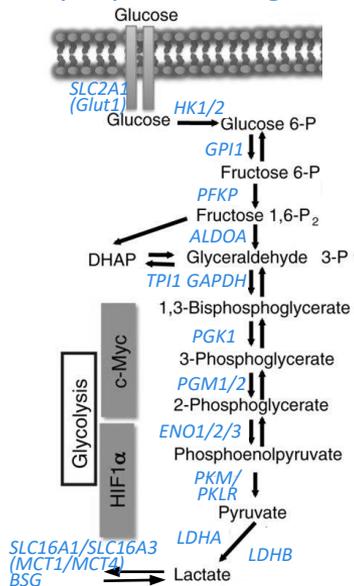


Aims

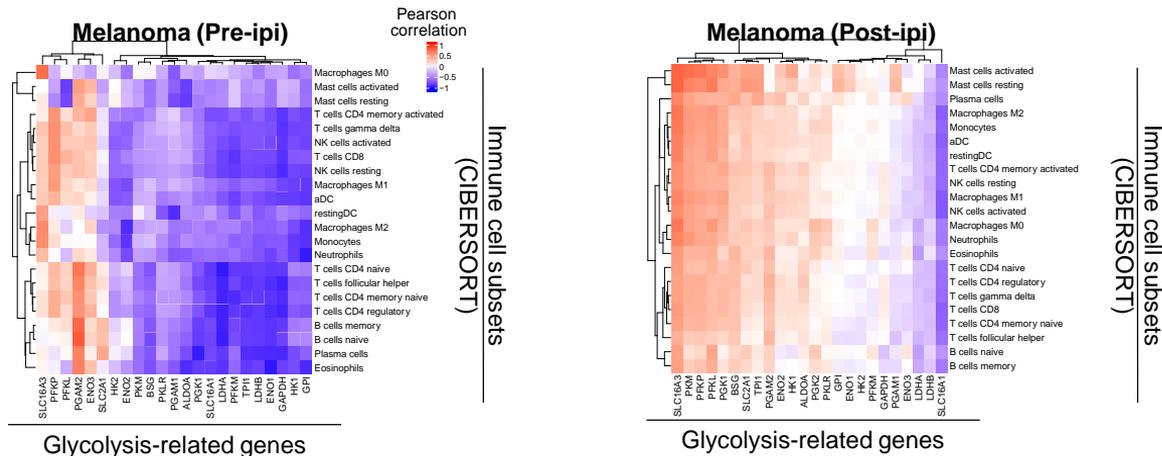
1. Determine how **tumor glycolysis impacts on ICB** activity;
2. Identify the **immune cell types** that are potentiated the most by ICB when tumor glycolysis is hampered;
3. Define the **mechanism underlying** these effects.
4. Modulate glycolysis **pharmacologically**.

Glycolysis vs. immune infiltration in human tumors before and after immune checkpoint blockade

Glycolysis-related genes



Heterogeneous tumor glycolysis



Zappasodi et al., *Nature* 2021

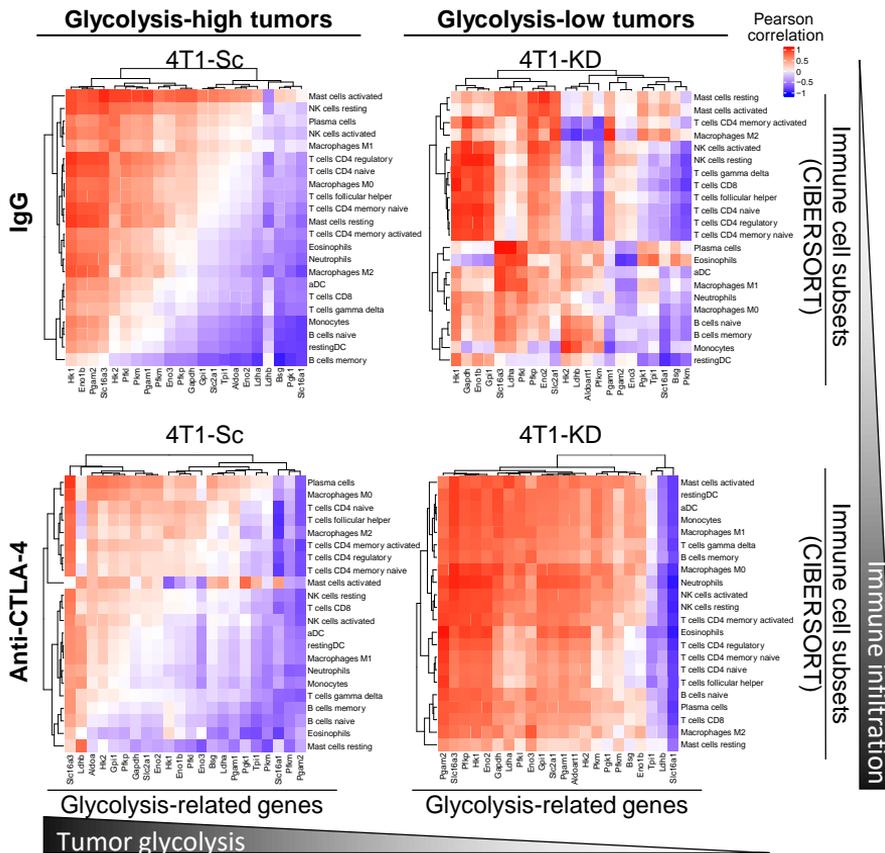
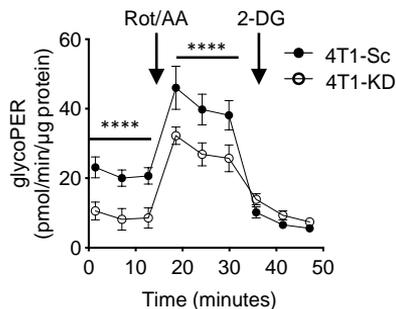
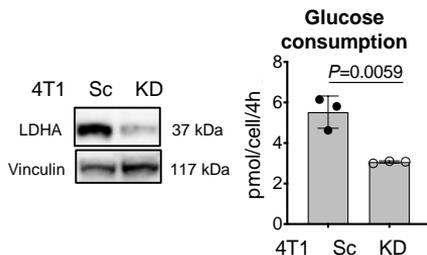
- Immune infiltration and glucose catabolism genes are mutually exclusive;
- CTLA-4 blockade facilitates immune infiltration;
- however, expression of key glycolytic genes and immune infiltrate remain negatively correlated after ipilimumab:

➤ **Does CTLA-4 blockade work better in glycolysis-low tumors?**

Glycolysis vs. immune infiltration in murine tumors before and after immune checkpoint blockade

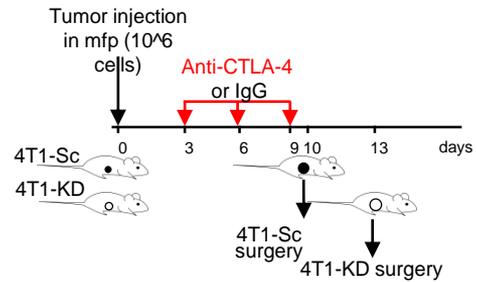
Model System: LDHA knock down in aggressive and immunotherapy resistant mouse tumor models

4T1 mammary carcinoma

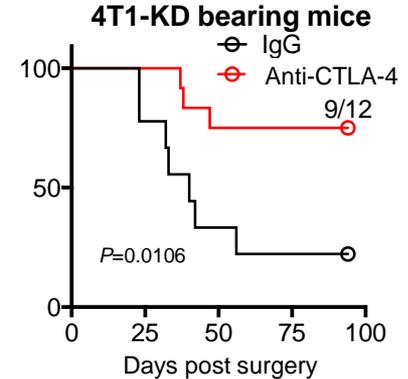
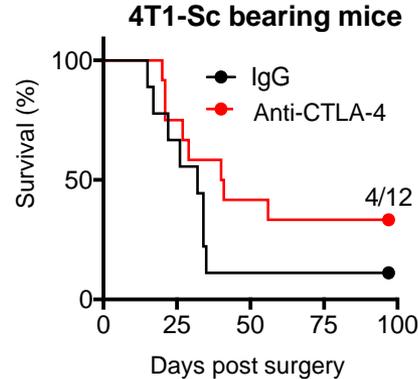


Improved responses of glycolysis-defective tumors to CTLA-4 blockade

4T1 mammary carcinoma (neoadjuvant ICB)



Zappasodi et al., *Nature* 2021



- Increased efficacy and long-lasting anti-tumor immunity of ICB with anti-CTLA-4 in glycolysis-defective LDHA-KD tumors.
- Activity of PD-1 blockade is not substantially improved in the same experimental settings.

Aims

1. Determine how **tumor glycolysis impacts on ICB** activity;
2. Identify the **immune cell types** that are potentiated the most by ICB when tumor glycolysis is hampered;
3. Define the **mechanism underlying** these effects.
4. Modulate glycolysis pharmacologically.

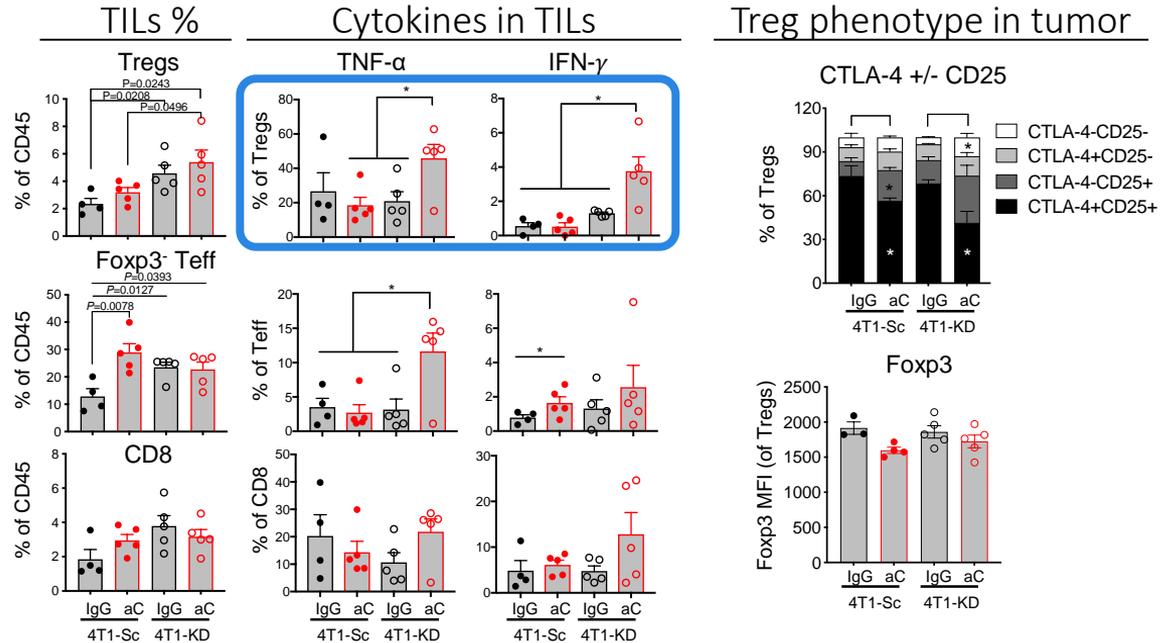
CTLA-4 blockade drives in Treg instability in glycolysis-defective tumors

- Changes in frequency of TIL subsets: NO

✓ Changes in Treg function: YES*

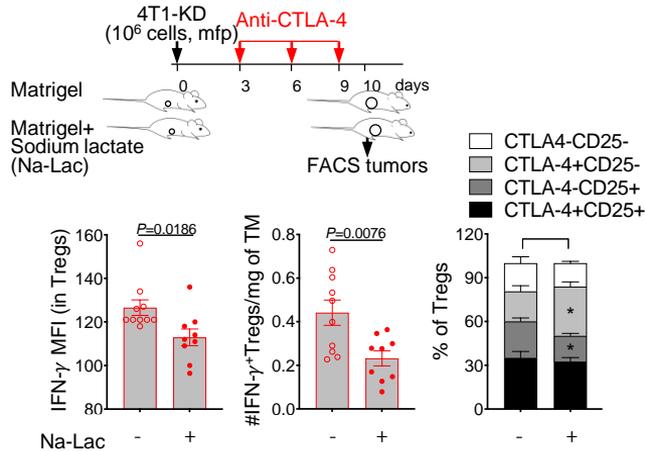
✓ Changes in Treg phenotype: YES*

*Similar results with B16-KD vs. B16-Sc

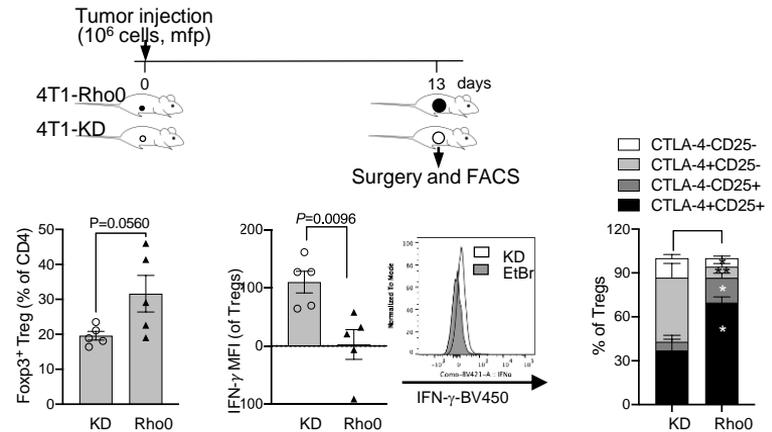


Does the local lactate:glucose ratio play a role?

Supplying lactate in the KD TME



Maximizing tumor glycolysis (4T1-Rho0)



Zappasodi et al., *Nature* 2021

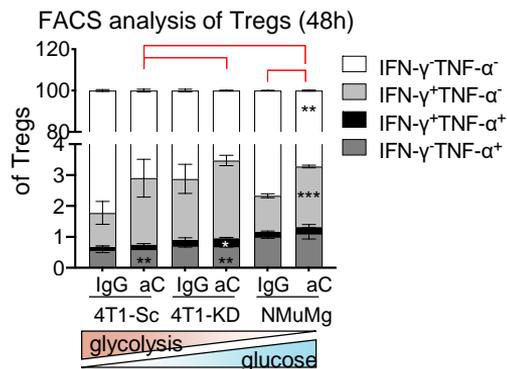
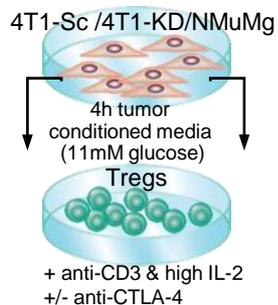
- Increasing lactate:glucose ratio or boosting tumor glycolysis potentiates Treg stability;
- Loss of Treg stability induced by anti-CTLA-4 in glycolysis-defective tumors is dependent on local lactate:glucose ratio.

Aims

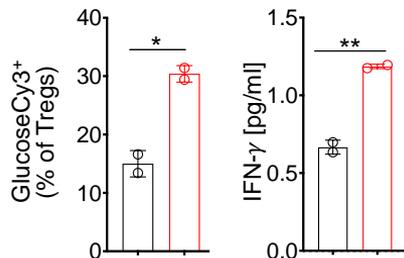
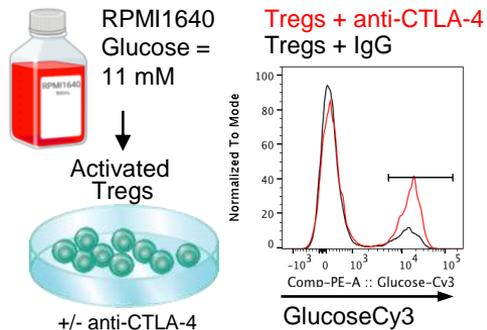
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3. Define the **mechanism underlying** these effects.
4. Modulate glycolysis **pharmacologically**.

Anti-CTLA-4 promotes Treg glucose utilization and IFN- γ production associated with reduced Treg suppression

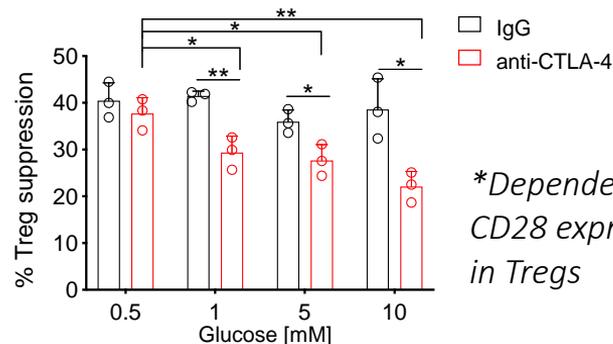
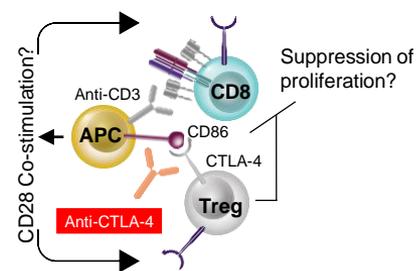
Tregs in tumor conditioned media \pm anti-CTLA-4



Tregs cultured alone \pm anti-CTLA-4



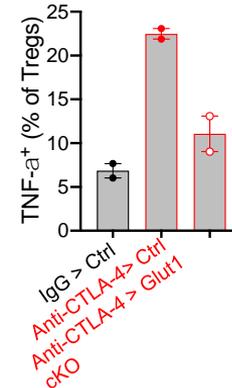
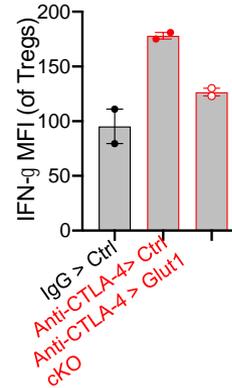
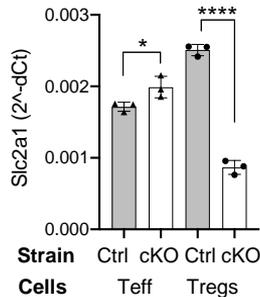
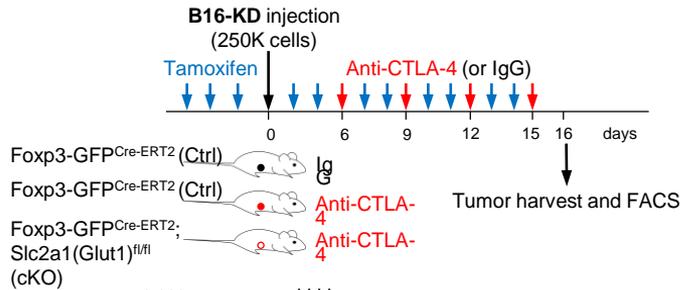
Treg suppression assay in increasing [glucose] \pm anti-CTLA-4



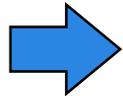
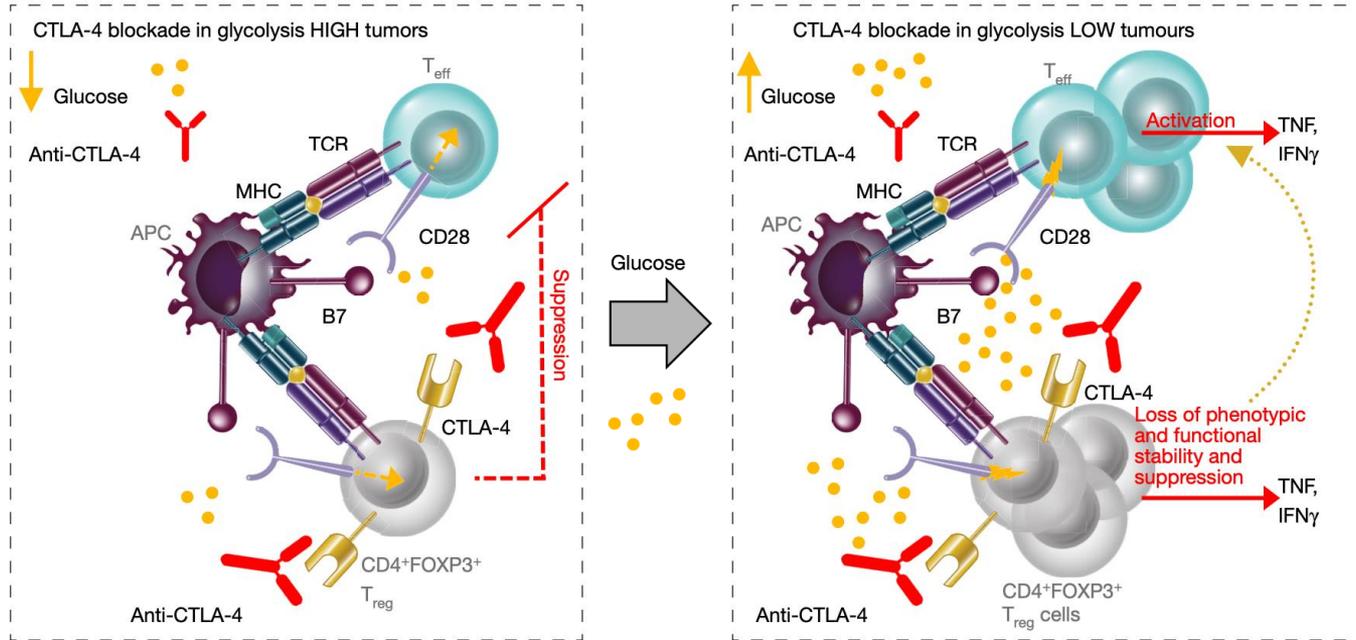
**Dependent on CD28 expression in Tregs*

Loss of Treg stability upon anti-CTLA-4 is dependent on Treg capacity to metabolize glucose *in vivo*

LDHA-KD tumors (↑glucose:lactate) → Glut1 cKO mice (no glucose uptake in Tregs) → Anti-CTLA-4 (or IgG) → IFN- γ , TNF- α production by Tregs?



Model & Perspectives



CTLA-4 blockade may be best exploited to treat glycolysis-low tumors and/or in combination with inhibitors of tumor glycolysis.

Aims

1. Determine how **tumor glycolysis impacts on ICB** activity;
2. Identify the **immune cell types** that are potentiated the most by ICB when tumor glycolysis is hampered;
3. Define the **mechanism underlying** these effects.
4. Modulate glycolysis **pharmacologically**.

Pharmacological Modulation of Metabolism to Improve ICB

- Effects of LDH pharmacologic inhibition
- Explore the effects Lactate transport



Pharmacological Modulation of Metabolism to Improve ICB

- Effects of LDH pharmacologic inhibition
- Explore the effects Lactate transport

Mice bearing B16 melanoma tumors have higher serum LDH and lactate levels than non-tumor bearing mice

No tumor



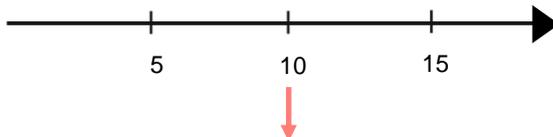
vs

Tumor



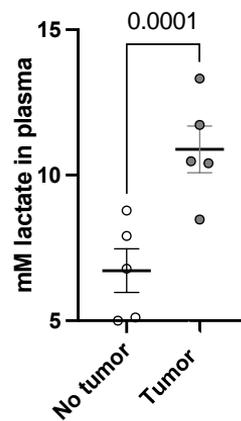
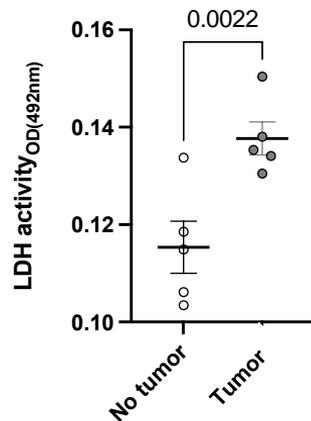
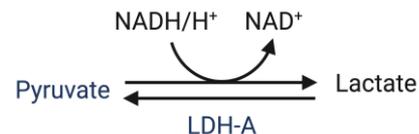
B16
(2×10^5)

Days since tumor implantation

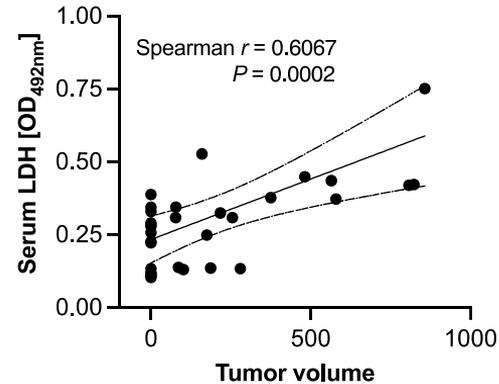
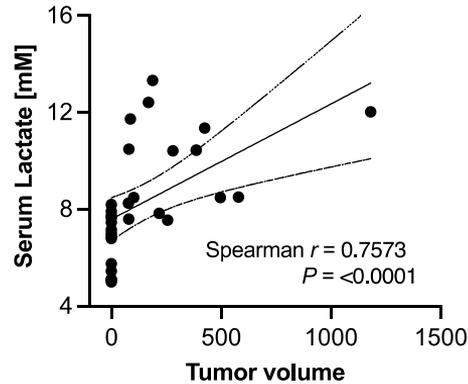
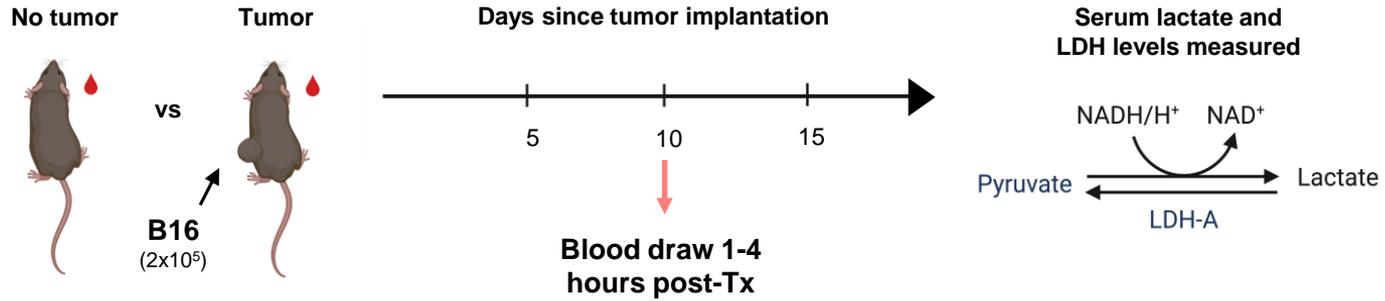


Blood draw 1-4
hours post-Tx

Serum lactate and
LDH levels measured



Serum lactate and LDH levels correlate with B16 melanoma tumor burden

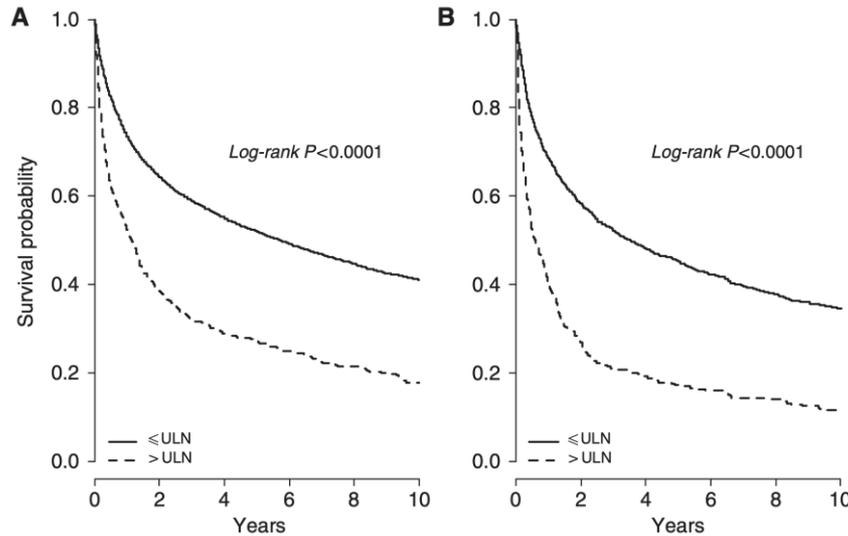


High serum LDH levels are a negative prognostic factor for many cancers

Patients with serum LDH levels above baseline prior to diagnosis have a lower survival probability

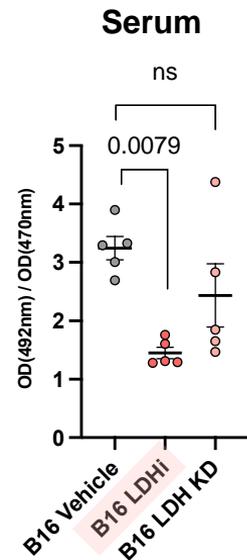
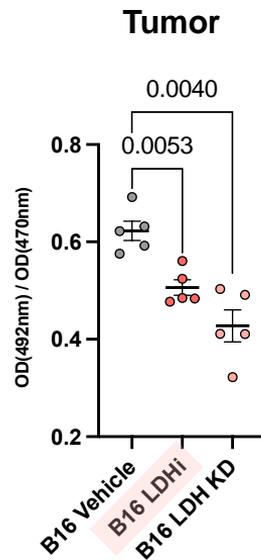
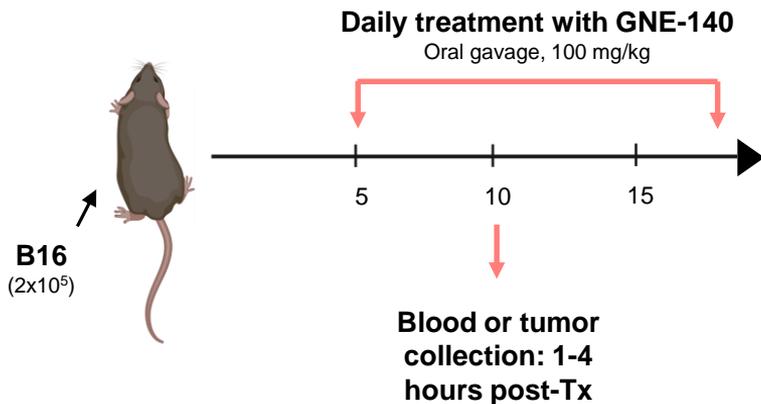
Serum lactate dehydrogenase and survival following cancer diagnosis

Wahyu Wulaningsih^{*,1}, Lars Holmberg^{1,2,3}, Hans Garmo^{1,3}, Håkan Malmstrom⁴, Mats Lambe^{3,5}, Niklas Hammar^{4,6}, Göran Walldius⁷, Ingmar Jungner⁸, Tony Ng⁹ and Mieke Van Hemelrijck^{1,4}

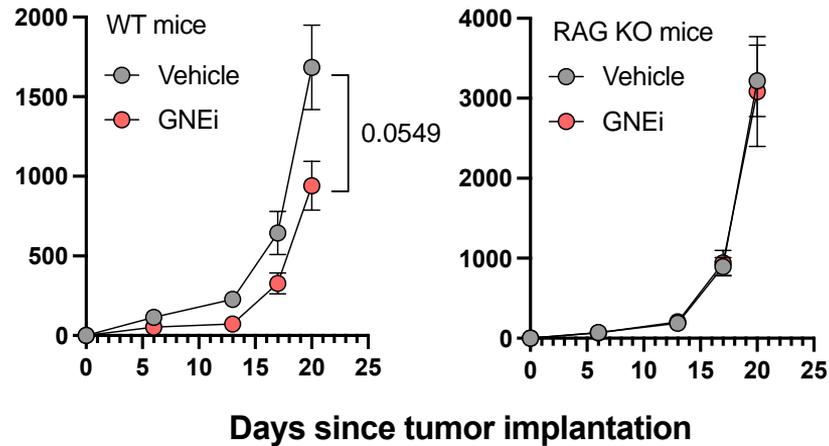
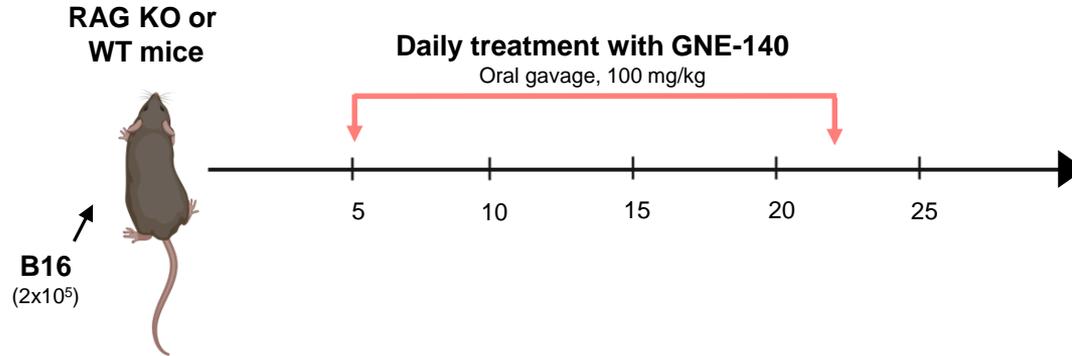


Kaplan-Meier curves for 10-year overall survival following cancer diagnosis by serum LDH (above or below baseline levels) measured within (A) 3 years before diagnosis and (B) 3 months before diagnosis.

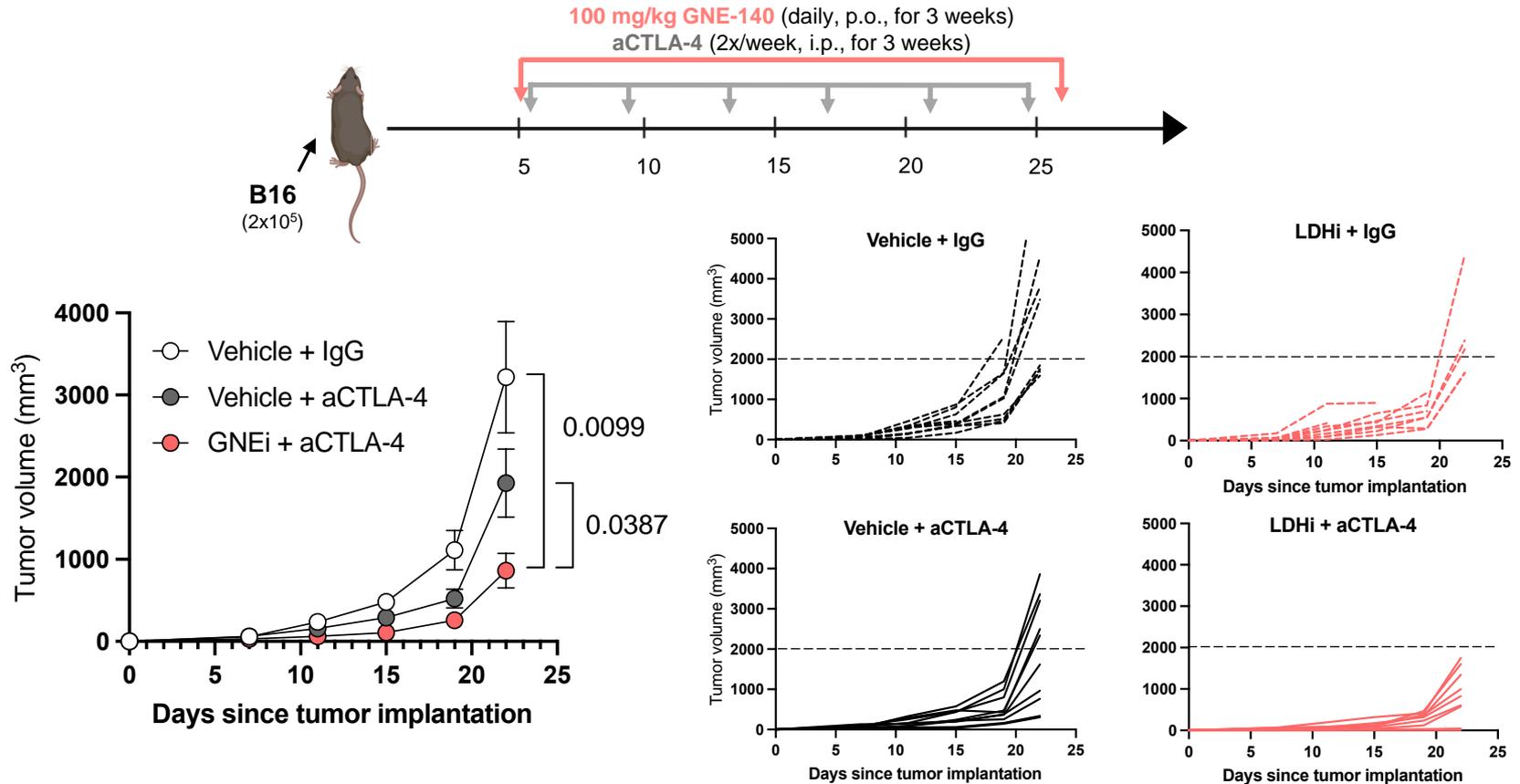
In vivo: When administered daily, GNE-140 (LDHi) reduces LDH activity within the tumor and in the periphery
Mice bearing LDHA KD tumors have low tumor LDH but not significantly lower serum LDH



Adaptive immunity is required for the anti-tumor effect of LDHi

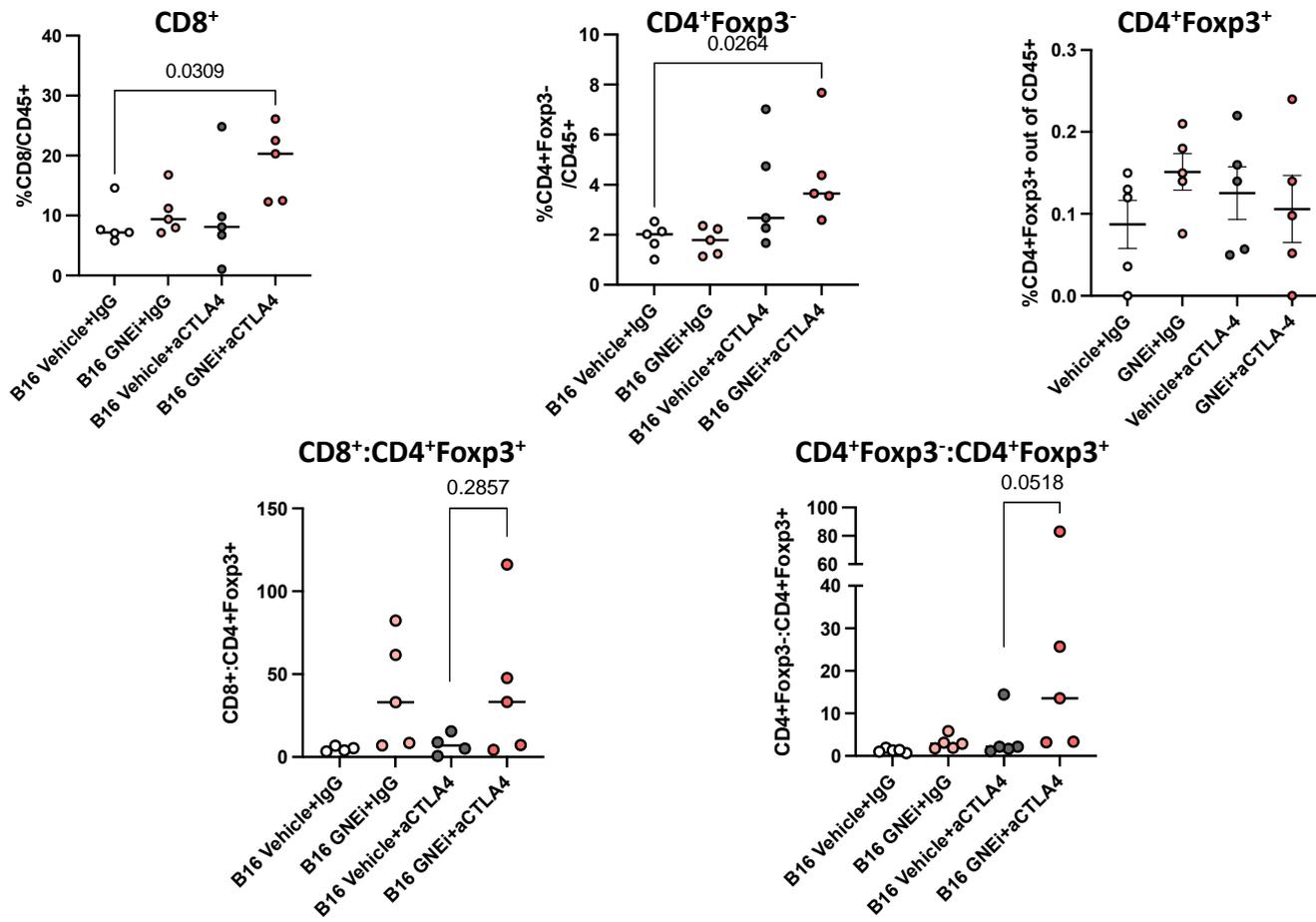


Treatment with LDHi sensitizes B16 melanoma to CTLA-4 blockade



CTLA-4 blockade combined with LDH inhibition leads to increased CD8⁺ and CD4⁺ effector infiltration without increasing Treg infiltration

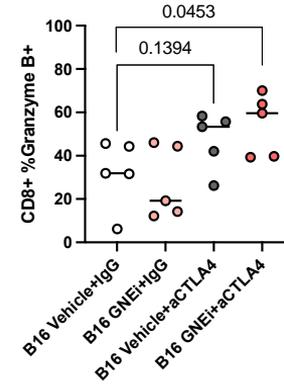
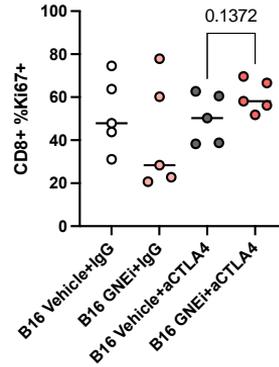
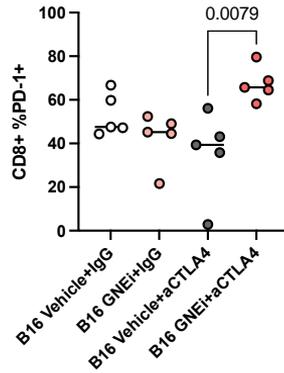
Immune infiltrate was examined after the 3rd anti-CTLA-4 administration, 2 weeks post-tumor implantation



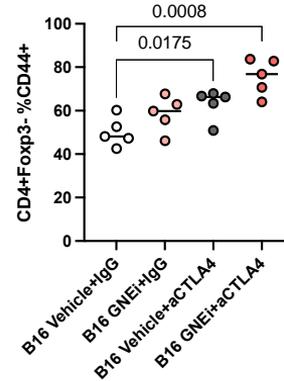
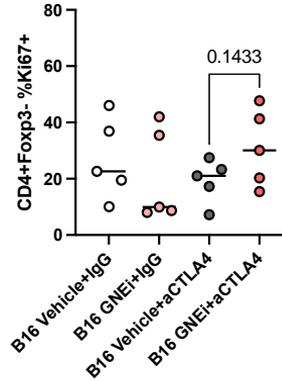
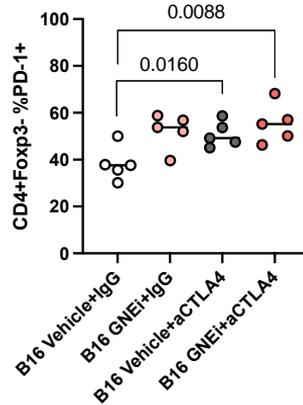
LDHi combined with CTLA-4 blockade favors CD8⁺ and CD4⁺ effector activation

LDHi may be preferentially targeting tumor cells that overexpress LDH, alleviating competition for glucose in the tumor microenvironment

CD8⁺ phenotypes

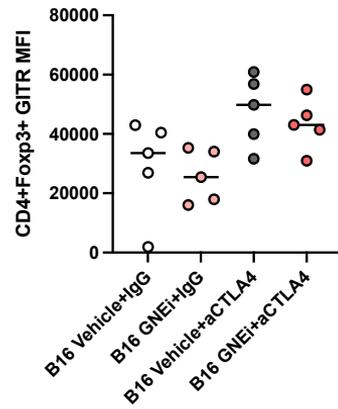
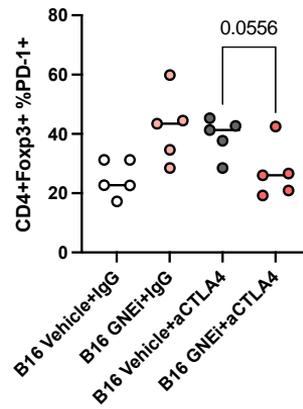
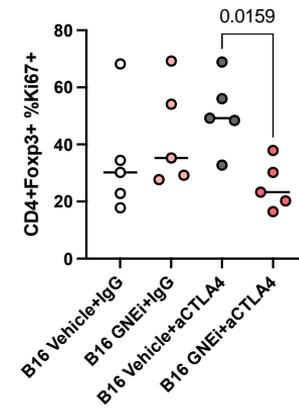
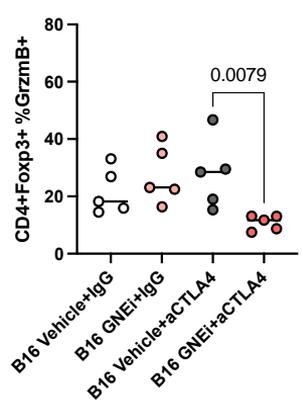
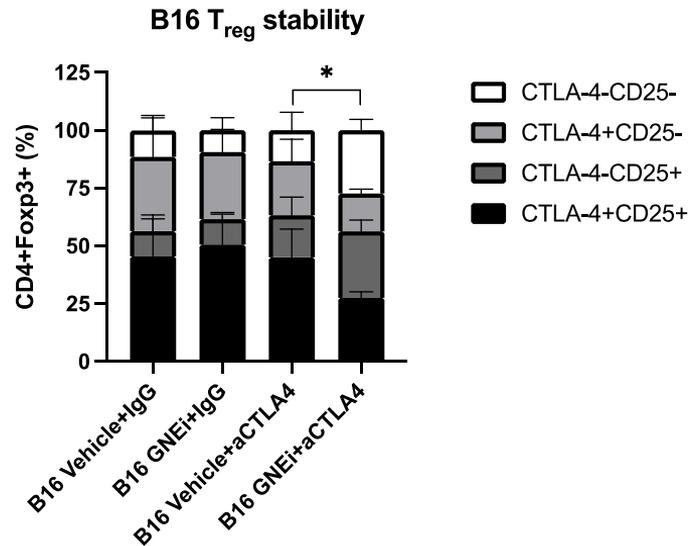


CD4⁺Foxp3⁻ phenotypes



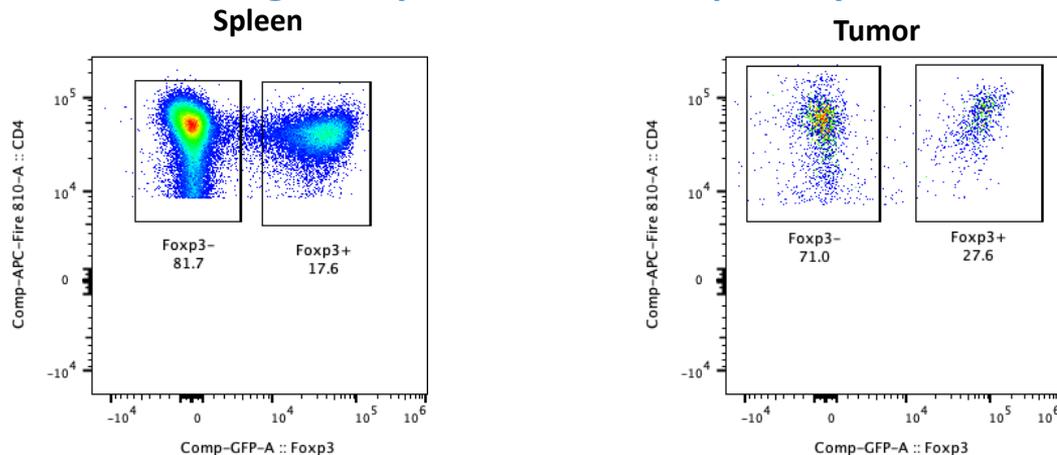
Similarly to when LDH is downregulated within the tumor, LDH inhibition leads to loss of Treg stability in the setting of CTLA-4 blockade

We had previously shown that LDHA-deficient Tregs displayed poor expansion and proliferation potential post-treatment with CTLA-4 blockade
CD4⁺Foxp3⁺ phenotypes

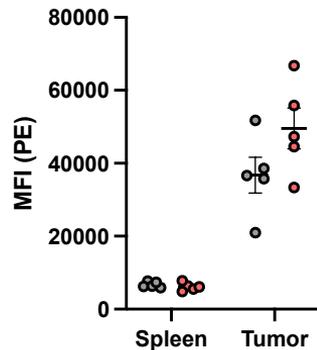


Treatment with LDHi increases glucose uptake capacity of tumor-infiltrating T cells, including Foxp3⁺ regulatory T cells, but not splenocytes

Foxp3 GFP mice:

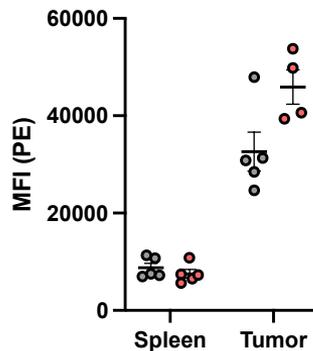


**CD8⁺
Glucose Cy3 MFI**



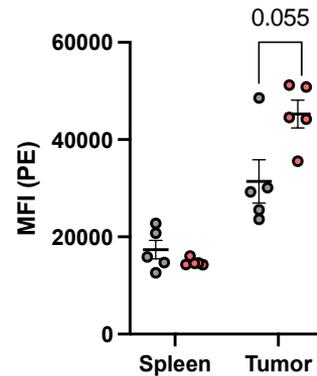
● Vehicle
● LDHi

**CD4⁺Foxp3⁺
Glucose Cy3 MFI**



● Vehicle
● LDHi

**CD4⁺Foxp3⁺
Glucose Cy3 MFI**



● Vehicle
● LDHi

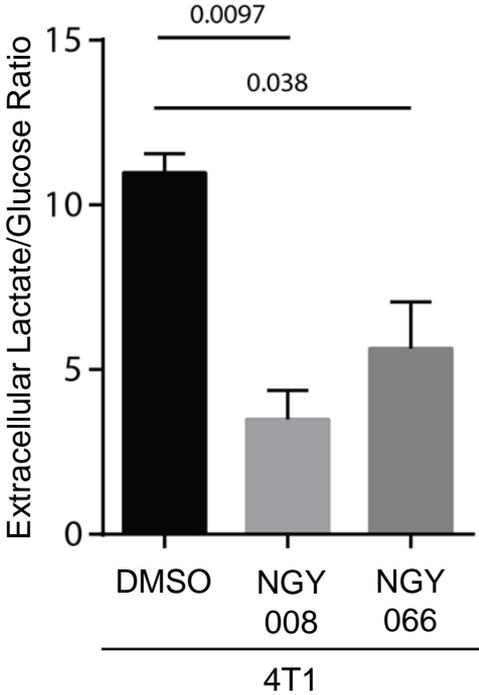
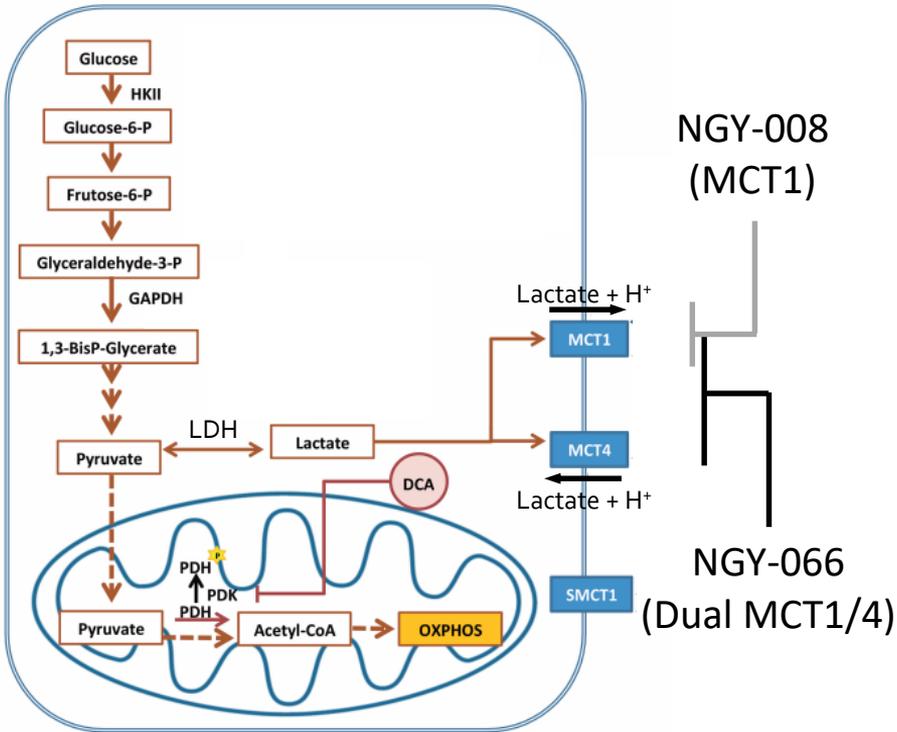
Conclusions

- Serum lactate and LDH levels correlate with B16 melanoma tumor burden, and patients with serum LDH levels above baseline prior to diagnosis have a lower survival probability
- LDH inhibitor GNE-140 reduces tumor cell glycolysis *in vitro*, lowers tumor and serum LDH in mice, and delays B16 melanoma growth in immunocompetent but not immunodeficient mice
- Treatment with LDHi alongside CTLA-4 blockade delays tumor growth more significantly than immunotherapy alone
- CTLA-4 blockade combined with LDH inhibition leads to increased CD8⁺ and CD4⁺ effector infiltration and activation, while resulting in functional destabilization of regulatory T cells
- Upon treatment with LDHi *in vivo*, tumor-infiltrating CD8⁺, CD4⁺Foxp3⁻, and CD4⁺Foxp3⁺ display a higher capacity for glucose uptake, while splenocyte glucose uptake remains unchanged, indicating that LDHi is relatively tumor-specific, due to the tumor's over-reliance on glycolysis and overexpression of LDH

Pharmacological Modulation of Metabolism to Improve ICB

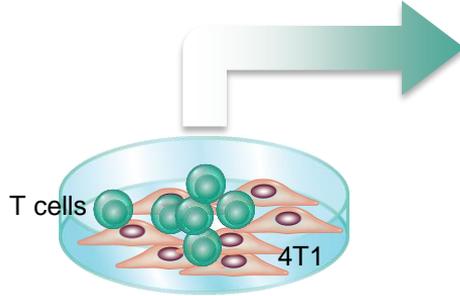
- Effects of LDH pharmacologic inhibition
- Explore the effects Lactate transport

Inhibition of MCTs reduces tumor lactate production



Inhibition of MCTs favors T-cell activation in tumor-T-cell co-cultures

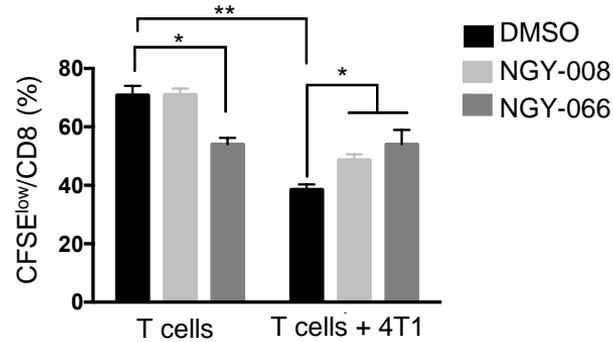
CFSE-labeled T cells



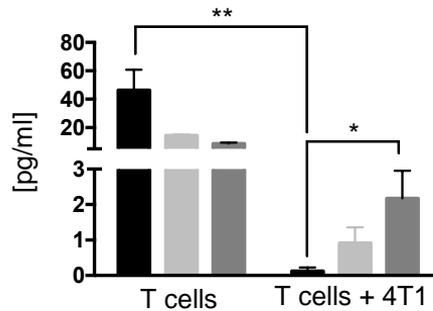
+PHA or anti-CD3/CD28
+vehicle/008/066

Culture supernatants

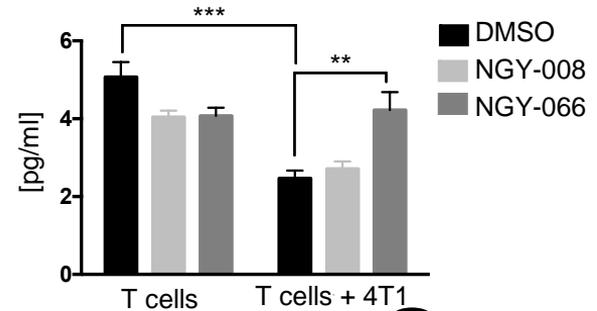
Proliferation



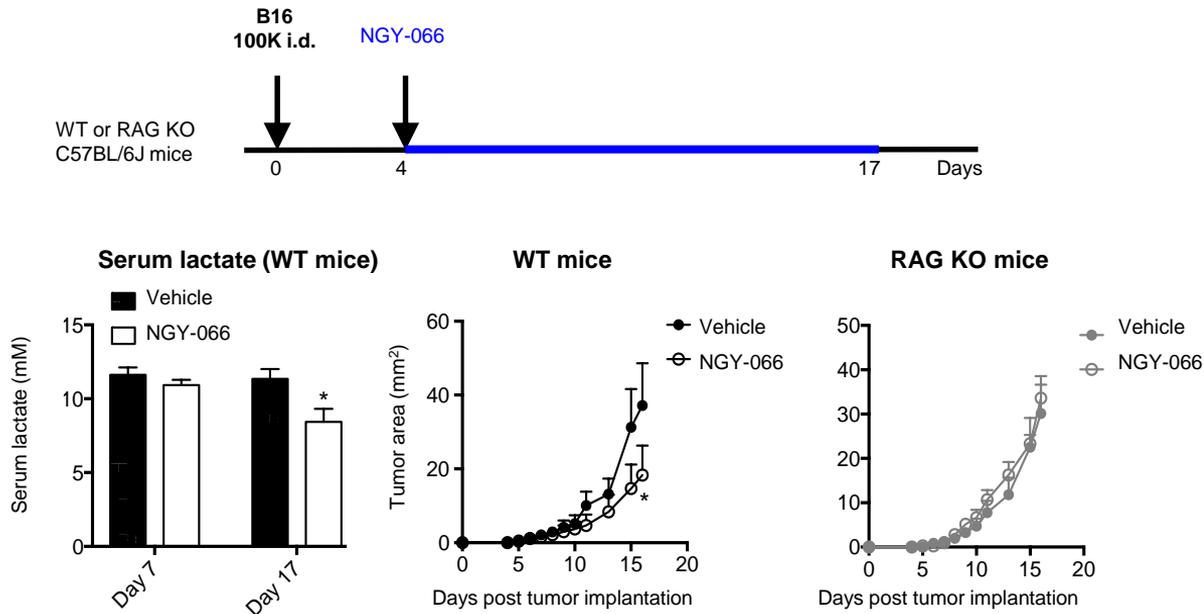
IFN- γ



TNF- α



Immune mediated effects of pharmacologic modulation of tumor lactate metabolism *in vivo*



Conclusions

1. Co-culture of T cells with 4T1 cells significantly limits their immune effector functions
2. Blocking lactate transporters *in vitro* improves T-cell functions
3. Inhibition of lactate transport *in vivo* delays tumor progression in a manner that is dependent on adaptive immunity



Explore MCT expression patterns in different model systems



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