

# What's Next for Cancer Immunotherapy?

**Bernard A. Fox, PhD**

Harder Family Chair for Cancer Research  
Member and Chief,  
Laboratory of Molecular and Tumor Immunology  
Earle A. Chiles Research Institute  
Robert W. Franz Cancer Center  
Providence Cancer Institute

Founder and CEO, UbiVac

Adjunct Graduate Faculty,  
Dept. Molec. Micro and Immunology, and  
Knight Cancer Institute, OHSU  
Portland, Oregon , USA



@BernardAFox

e  
pi

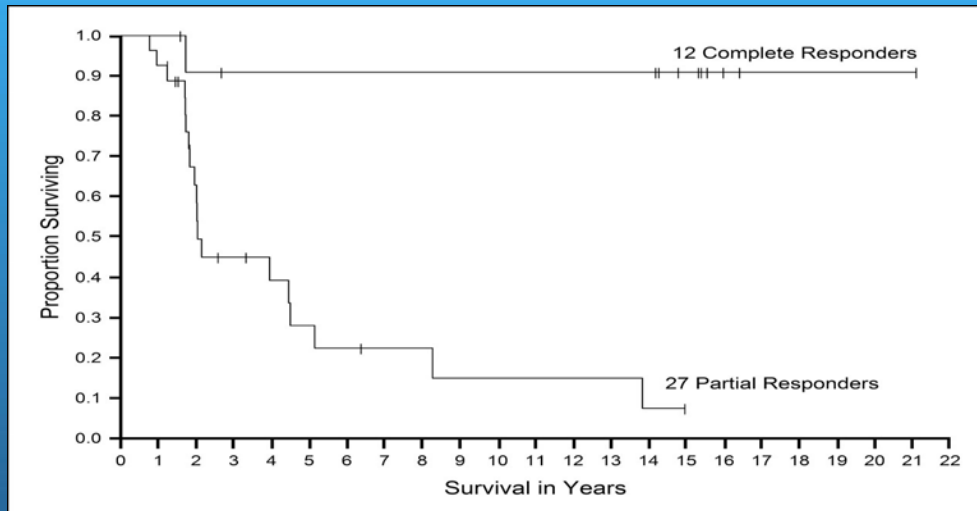
## ***Bernard A. Fox, PhD – COI Disclosures***

- IP Rights: UbiVac
- Consulting Fees: Turnstone Bio, PrimeVax, UltiVue, AstraZeneca, Boehringer Ingelheim
- Contracted Research: MacroGenics, OncoSec, BMS, Akoya, Nanostring, Incyte, Shimadzu, Viralytics/Merck
- Ownership interest *greater* than 5%: UbiVac

April 2021



# Immunotherapy can Cure Patients of Their Cancer!

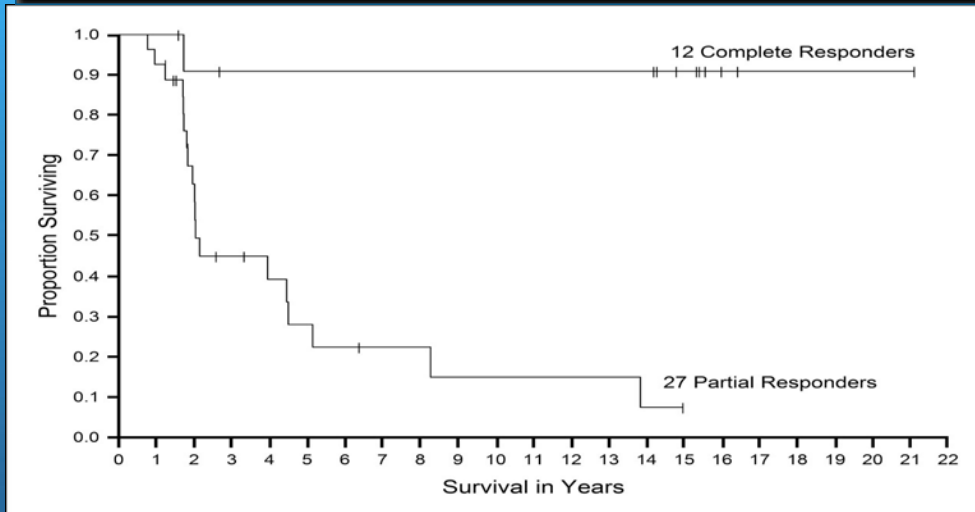


Smith F O et al. Clin Cancer Res 2008;14:5610-5618



# Immunotherapy can Cure Patients of Their Cancer!

- Certainly IL-2 can “Cure”



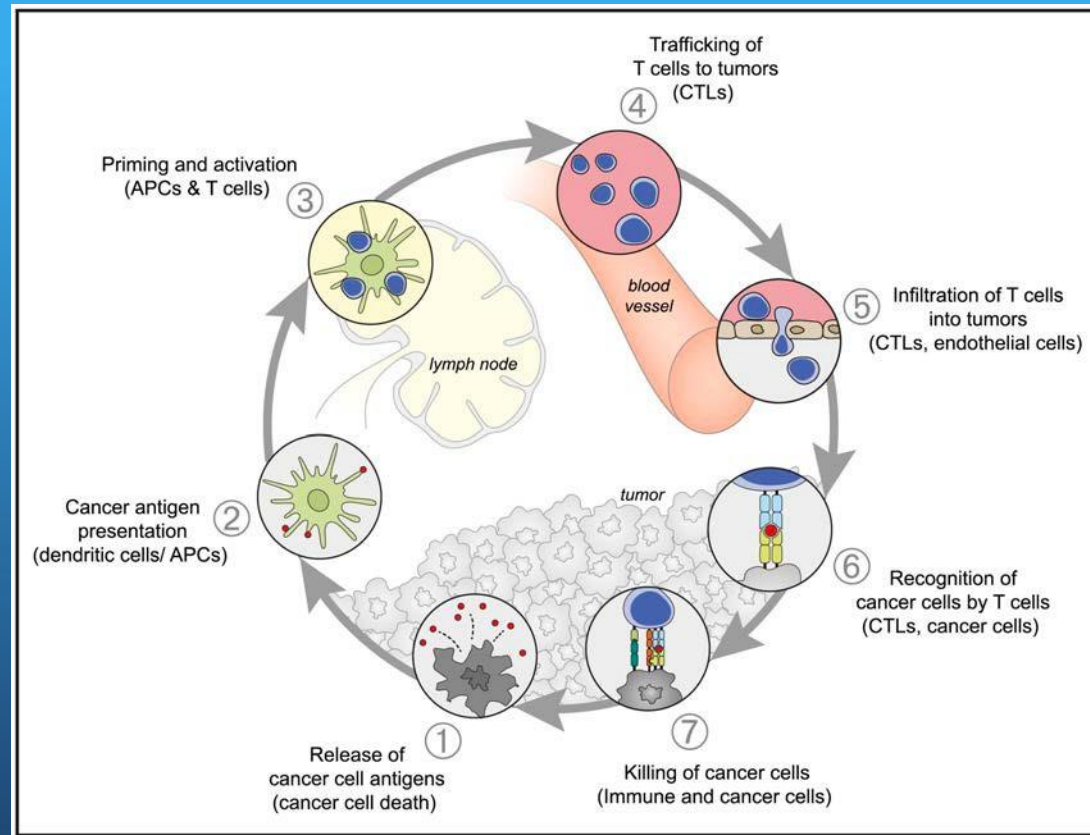
Smith F O et al. Clin Cancer Res 2008;14:5610-5618

In 2021 we still do not know:

- Why these patients were “cured”
- If other immunotherapies will cure



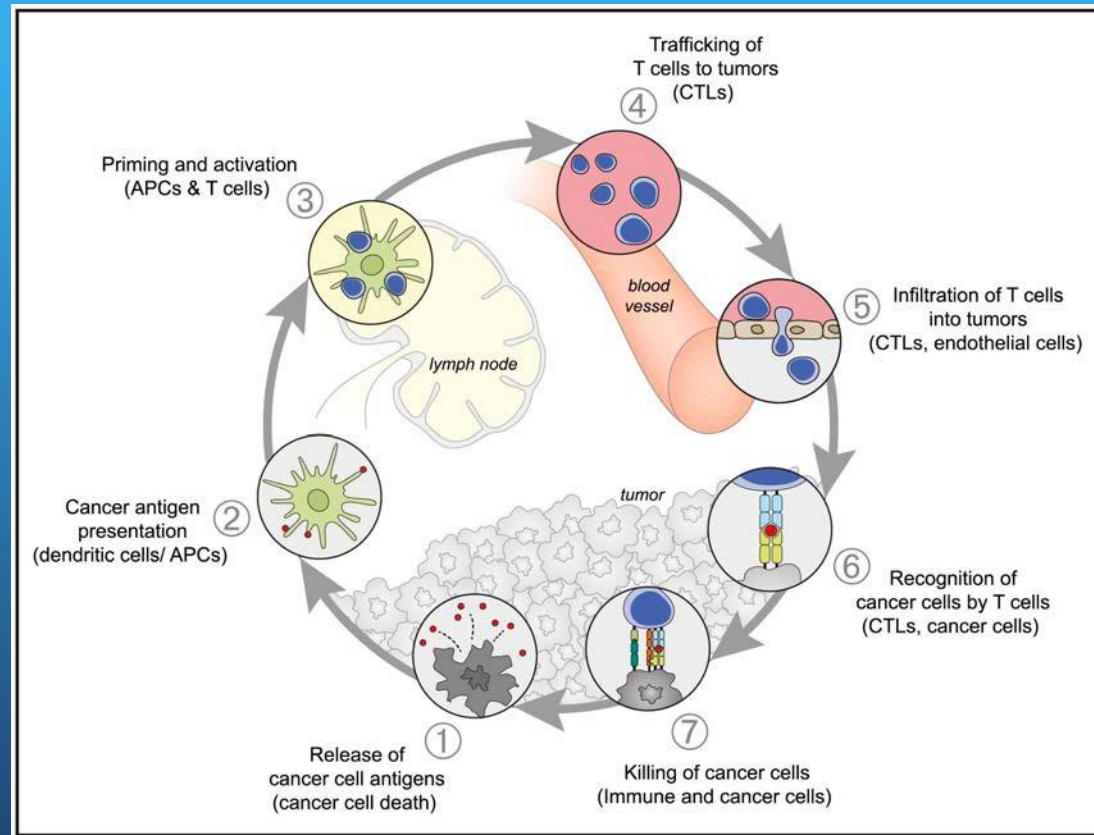
# Cancer Immunity Cycle



## Hypothesis:

T cell immune responses that effectively recognize all cancer cells will **CURE** patients of cancer

# T Cell Centric View



## Hypothesis:

T cell immune responses that effectively recognize all cancer cells will **CURE** patients of cancer

NK cells?

B cells?

Innate effectors?



# B cells to the forefront of immunotherapy

Tullia C. Bruno

Nature | Vol 577 | 23 January 2020

Three studies reveal that the presence in tumours of two key immune components – B cells and tertiary lymphoid structures – is associated with favourable outcomes when individuals undergo immunotherapy. See p.549, p.556 & p.561

## Biomarkers:

- Prognostic significance
- Marker of immune response

## Article

# B cells and tertiary lymphoid structures promote immunotherapy response

<https://doi.org/10.1038/s41586-019-1922-8>

Received: 5 February 2019

Accepted: 4 December 2019

Published online: 15 January 2020

Beth A. Helmink<sup>1,2,4\*</sup>, Sangeetha M. Reddy<sup>3,2,4</sup>, Jianjun Gao<sup>3,2,4</sup>, Shaojun Zhang<sup>4,2,4</sup>, Rafet Basar<sup>4,2,4</sup>, Rohit Thakur<sup>1</sup>, Keren Yizhak<sup>4</sup>, Moshe Sade-Feldman<sup>4,2</sup>, Jorge Blando<sup>4</sup>, Guangchun Han<sup>4</sup>, Vancheswaran Gopalakrishnan<sup>1</sup>, Yuanxin Xi<sup>4</sup>, Hao Zhao<sup>4</sup>, Rodabe N. Amaria<sup>10</sup>, Husseln A. Tawbi<sup>10</sup>, Alex P. Cogdill<sup>1</sup>, Wenbin Liu<sup>4</sup>, Valerie S. LeBlau<sup>11</sup>, Fernanda G. Kugeratski<sup>11</sup>, Sapna Patel<sup>10</sup>, Michael A. Davies<sup>10</sup>, Patrick Hwu<sup>10</sup>, Jeffrey E. Lee<sup>1</sup>, Jeffrey E. Gershenwald<sup>1</sup>, Anthony Lucci<sup>1</sup>, Reetakshi Arora<sup>4</sup>, Scott Woodman<sup>10</sup>, Emily Z. Keung<sup>1</sup>, Pierre-Olivier Gaudreau<sup>1</sup>, Alexandre Reuben<sup>10</sup>, Christine N. Spencer<sup>13</sup>, Elizabeth M. Burton<sup>1</sup>, Lauren E. Haydu<sup>1</sup>, Alexander J. Lazar<sup>4,14,15</sup>, Roberta Zappasodi<sup>16</sup>, Courtney W. Hudgens<sup>14</sup>, Deborah A. Ledesma<sup>14</sup>, SuFey Ong<sup>17</sup>, Michael Bailey<sup>17</sup>, Sarah Warren<sup>17</sup>, Disha Rao<sup>18</sup>, Oscar Krijgsman<sup>18</sup>, Elissa A. Rozeman<sup>18</sup>, Daniel Peepers<sup>18</sup>, Christian U. Blank<sup>18</sup>, Ton N. Schumacher<sup>18</sup>, Lisa H. Butterfield<sup>18</sup>, Monika A. Zelazowska<sup>18</sup>, Kevin M. McBride<sup>20</sup>, Raghu Kalluri<sup>11</sup>, James Allison<sup>4</sup>, Florent Petitprez<sup>21,22,23</sup>, Wolf Herman Fridman<sup>21,22</sup>, Catherine Sautès-Fridman<sup>21,22</sup>, Nir Hacohen<sup>4,7</sup>, Katayoun Rezvani<sup>18,25</sup>, Padmanee Sharma<sup>1,8,26</sup>, Michael T.etzlaff<sup>14,15,25</sup>, Linghua Wang<sup>4,25</sup> & Jennifer A. Wargo<sup>1,4,25\*</sup>

## Article

# Tertiary lymphoid structures improve immunotherapy and survival in melanoma

<https://doi.org/10.1038/s41586-019-1914-8>

Received: 5 February 2019

Accepted: 4 December 2019

Published online: 15 January 2020

Rita Cabrita<sup>11,2</sup>, Martin Lauss<sup>11,2</sup>, Adriana Sanna<sup>1</sup>, Marco Donia<sup>2</sup>, Mathilde Skaarup Larsen<sup>3</sup>, Shamik Mitra<sup>1</sup>, Iva Johansson<sup>1</sup>, Bengt Phung<sup>1</sup>, Katja Harbst<sup>1</sup>, Johan Vallon-Christersson<sup>1</sup>, Allison van Scholack<sup>4</sup>, Kristina Lövgren<sup>4</sup>, Sarah Warren<sup>4</sup>, Karin Jirstrom<sup>1</sup>, Håkan Olsson<sup>1</sup>, Kristian Pietras<sup>4</sup>, Christian Ingvar<sup>4</sup>, Karolin Isaksson<sup>4</sup>, Dirk Schadendorf<sup>1</sup>, Henrik Schmidt<sup>4</sup>, Lars Bastholt<sup>4</sup>, Ana Carneiro<sup>11,2</sup>, Jennifer A. Wargo<sup>1</sup>, Inge Marie Svane<sup>2</sup> & Göran Jönsson<sup>1\*</sup>

## B cells are associated with survival and immunotherapy response in sarcoma

Florent Petitprez, Aurélien de Reyniès, Emily Z. Keung, Tom Wei-Wu Chen, Cheng-Ming Sun, Julien Calderaro, Yung-Ming Jeng, Li-Ping Hsiao, Laetitia Lacroix, Antoine Bouguéin, Marco Moreira, Guillaume Lacroix, Ivo Nataro, Julien Adam, Carlo Lucchesi, Yec'han Laizet, Maud Toulmonde, Melissa A. Burgess, Vanessa Bolejack, Denise Reinke, Khalid M. Wani, Wei-Lien Wang, Alexander J. Lazar, Christina L. Roland, Jennifer A. Wargo, Antoine Italiano, Catherine Sautès-Fridman, Hussein A. Tawbi✉ & Wolf H. Fridman✉

# B cells to the forefront of immunotherapy

Tullia C. Bruno

Nature | Vol 577 | 23 January 2020

Three studies reveal that the presence in tumours of two key immune components – B cells and tertiary lymphoid structures – is associated with favourable outcomes when individuals undergo immunotherapy. See p.549, p.556 & p.561

## Biomarkers:

- Prognostic significance
- Marker of immune response

**B Cell Effector Functions  
and Suppressive Functions**

## Article

# B cells and tertiary lymphoid structures promote immunotherapy response

<https://doi.org/10.1038/s41586-019-1922-8>

Received: 5 February 2019

Accepted: 4 December 2019

Published online: 15 January 2020

Beth A. Helmink<sup>1,2,4\*</sup>, Sangeetha M. Reddy<sup>3,2,4</sup>, Jianjun Gao<sup>3,2,4</sup>, Shaojun Zhang<sup>4,2,4</sup>, Rafet Basar<sup>4,2,4</sup>, Rohit Thakur<sup>1</sup>, Keren Yizhak<sup>4</sup>, Moshe Sade-Feldman<sup>4,2</sup>, Jorge Blando<sup>4</sup>, Guangchun Han<sup>4</sup>, Vancheswaran Gopalakrishnan<sup>1</sup>, Yuanxin Xi<sup>4</sup>, Hao Zhao<sup>4</sup>, Rodabe N. Amaral<sup>10</sup>, Husseln A. Tawbi<sup>10</sup>, Alex P. Cogdill<sup>1</sup>, Wenbin Liu<sup>4</sup>, Valerie S. LeBlau<sup>11</sup>, Fernanda G. Kugeratski<sup>11</sup>, Sapna Patel<sup>10</sup>, Michael A. Davies<sup>10</sup>, Patrick Hwu<sup>10</sup>, Jeffrey E. Lee<sup>1</sup>, Jeffrey E. Gershenwald<sup>1</sup>, Anthony Lucci<sup>1</sup>, Reetakshi Arora<sup>4</sup>, Scott Woodman<sup>10</sup>, Emily Z. Keung<sup>1</sup>, Pierre-Olivier Gaudreau<sup>1</sup>, Alexandre Reuben<sup>10</sup>, Christine N. Spencer<sup>13</sup>, Elizabeth M. Burton<sup>1</sup>, Lauren E. Haydu<sup>1</sup>, Alexander J. Lazar<sup>4,14,15</sup>, Roberta Zappasodi<sup>16</sup>, Courtney W. Hudgens<sup>14</sup>, Deborah A. Ledesma<sup>14</sup>, SuFey Ong<sup>17</sup>, Michael Bailey<sup>17</sup>, Sarah Warren<sup>17</sup>, Disha Rao<sup>18</sup>, Oscar Krijgsman<sup>18</sup>, Elisa A. Rozeman<sup>18</sup>, Daniel Peepers<sup>18</sup>, Christian U. Blank<sup>18</sup>, Ton N. Schumacher<sup>18</sup>, Lisa H. Butterfield<sup>18</sup>, Monika A. Zelazowska<sup>18</sup>, Kevin M. McBride<sup>18</sup>, Raghu Kalluri<sup>11</sup>, James Allison<sup>4</sup>, Florent Petitprez<sup>21,22,23</sup>, Wolf Herman Fridman<sup>21,22</sup>, Catherine Sautès-Fridman<sup>21,22</sup>, Nir Hacohen<sup>4,7</sup>, Katayoun Rezvani<sup>18,25</sup>, Padmanee Sharma<sup>18,26</sup>, Michael T.etzlaff<sup>14,15,25</sup>, Linghua Wang<sup>4,25</sup> & Jennifer A. Wargo<sup>4,25\*</sup>

## Article

# Tertiary lymphoid structures improve immunotherapy and survival in melanoma

<https://doi.org/10.1038/s41586-019-1914-8>

Received: 5 February 2019

Accepted: 4 December 2019

Published online: 15 January 2020

Rita Cabrita<sup>11,2</sup>, Martin Lauss<sup>11,2</sup>, Adriana Sanna<sup>1</sup>, Marco Donia<sup>2</sup>, Mathilde Skaarup Larsen<sup>3</sup>, Shamik Mitra<sup>1</sup>, Iva Johansson<sup>1</sup>, Bengt Phung<sup>1</sup>, Katja Harbst<sup>1</sup>, Johan Vallon-Christersson<sup>1</sup>, Allison van Scholack<sup>4</sup>, Kristina Lövgren<sup>4</sup>, Sarah Warren<sup>4</sup>, Karin Jirstrom<sup>1</sup>, Håkan Olsson<sup>1</sup>, Kristian Pietras<sup>4</sup>, Christian Ingvar<sup>4</sup>, Karolin Isaksson<sup>4</sup>, Dirk Schadendorf<sup>1</sup>, Henrik Schmidt<sup>4</sup>, Lars Bastholt<sup>4</sup>, Ana Carneiro<sup>11,2</sup>, Jennifer A. Wargo<sup>18</sup>, Inge Marie Svane<sup>2</sup> & Göran Jönsson<sup>1\*</sup>

## B cells are associated with survival and immunotherapy response in sarcoma

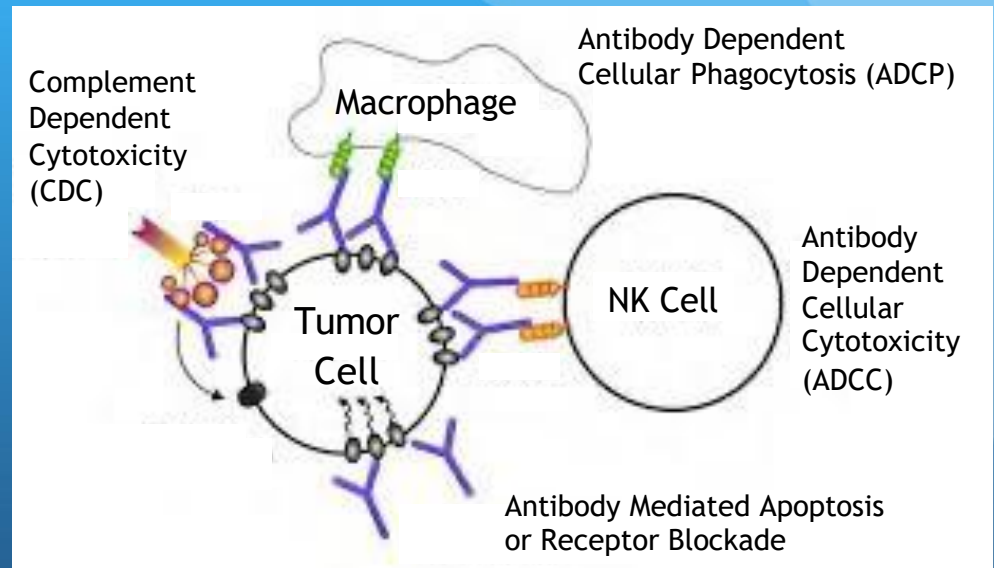
Florent Petitprez, Aurélien de Reyniès, Emily Z. Keung, Tom Wei-Wu Chen, Cheng-Ming Sun, Julien Calderaro, Yung-Ming Jeng, Li-Ping Hsiao, Laetitia Lacroix, Antoine Bouguéin, Marco Moreira, Guillaume Lacroix, Ivo Nataro, Julien Adam, Carlo Lucchesi, Yec'han Laizet, Maud Toulmonde, Melissa A. Burgess, Vanessa Bolejack, Denise Reinke, Khalid M. Wani, Wei-Lien Wang, Alexander J. Lazar, Christina L. Roland, Jennifer A. Wargo, Antoine Italiano, Catherine Sautès-Fridman, Hussein A. Tawbi<sup>✉</sup> & Wolf H. Fridman<sup>✉</sup>



# B Cell Effector Mechanisms Exist!

## B Cell Effector Mechanisms

- × CDC
- × ADCC
  - × Macrophages
  - × NK cells
- × Signaling
  - × Apoptosis
  - × Blocking (Herceptin)
- × Killer B cells (Fas-FasL)
- Other Functions
  - × Augment APC function



- Tao H, Eur J. Immunol 2015 45(4): 999-1009
- Li Q, J. Immunol 2009 183(5): 3195-3203
- Xu G, BBRC 2013 437 (2):287-291
- Neuberger M, JITC 2013 (Supp1 1) : P271

## B Cell Antibody Responses MAY Play a Critical Role in Anti-Cancer Immunity in Humans

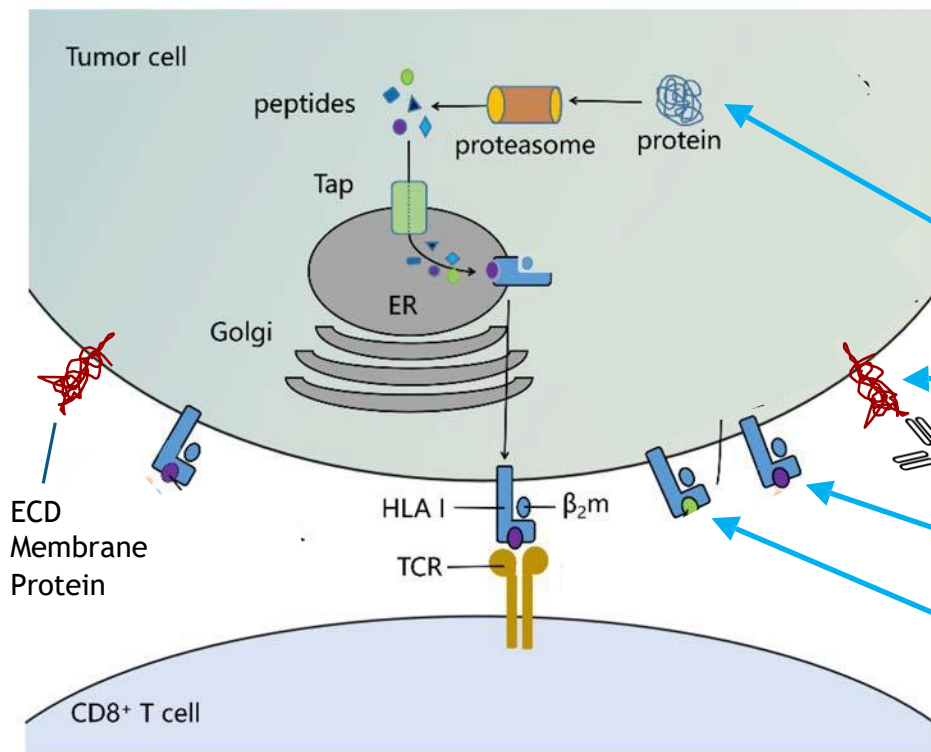
We Don't Know Because we haven't carefully looked using robust technology

Can also be a double-edged sword

- APC and Effector functions
- Suppressive functions

# Immune responses against cancer “Surfaceome”

## - Natural and Synthetic Immunity



Immune responses to proteins whose genes are over-expressed or differentially expressed by cancer

### Antibody responses

- Intracellular proteins
- ECD of transmembrane proteins

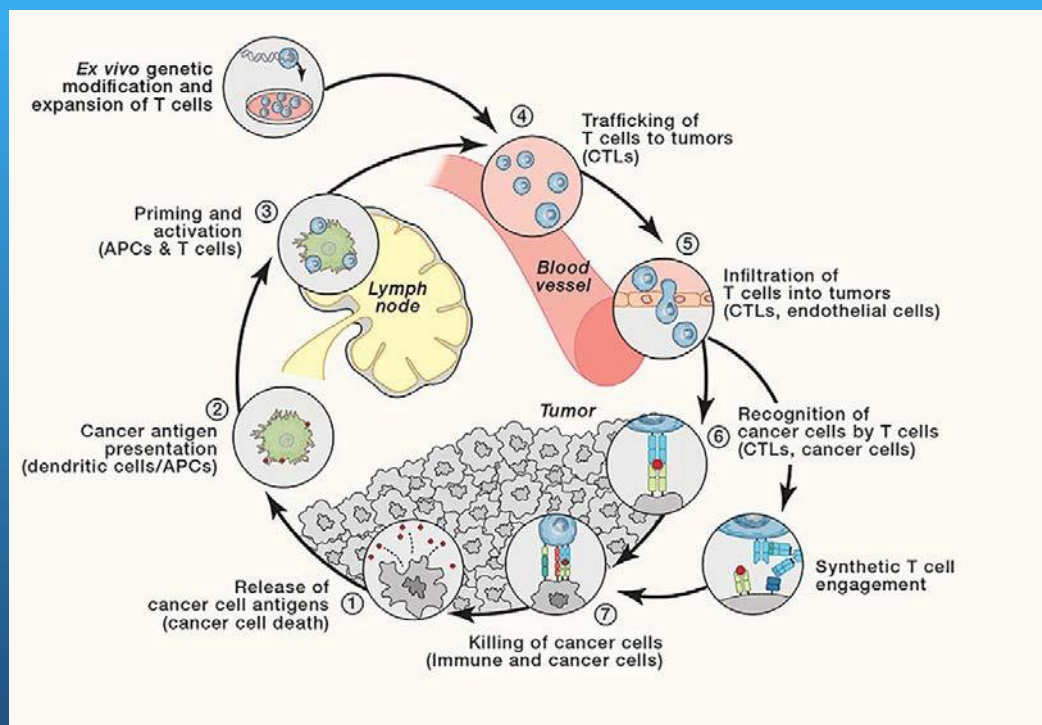
### T cell responses - Peptidome

- Non-mutated shared antigens
- Mutated epitopes

Modified Image from *Int J. Sci* 2019 20, 3912



# Synthetic Immunology



Immunity 52, January 14, 2020

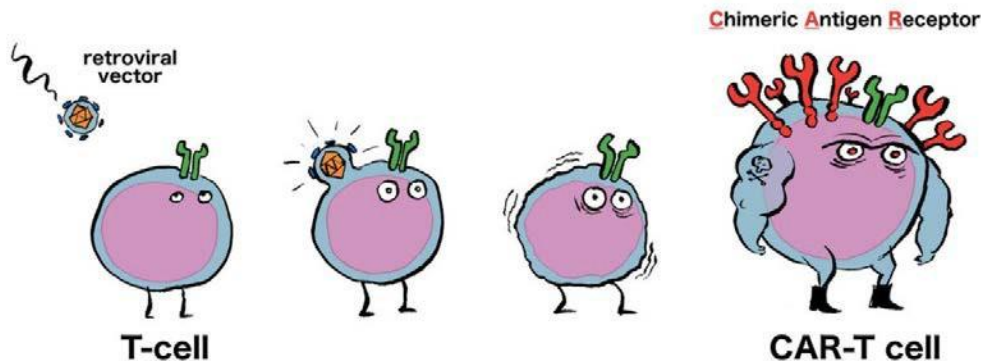
## Identification of surfasome targets:

- New Chimeric Ag receptors (T, NK, other)
- More strategies to target immune cells to cancer
  - DARTs / BiTEs
- Fully synthetic constructs / drugs
  - ADC

# Synthetic Immunology

## - Genetically Engineered Cells

### Generating super-soldiers the production of CAR-T cells

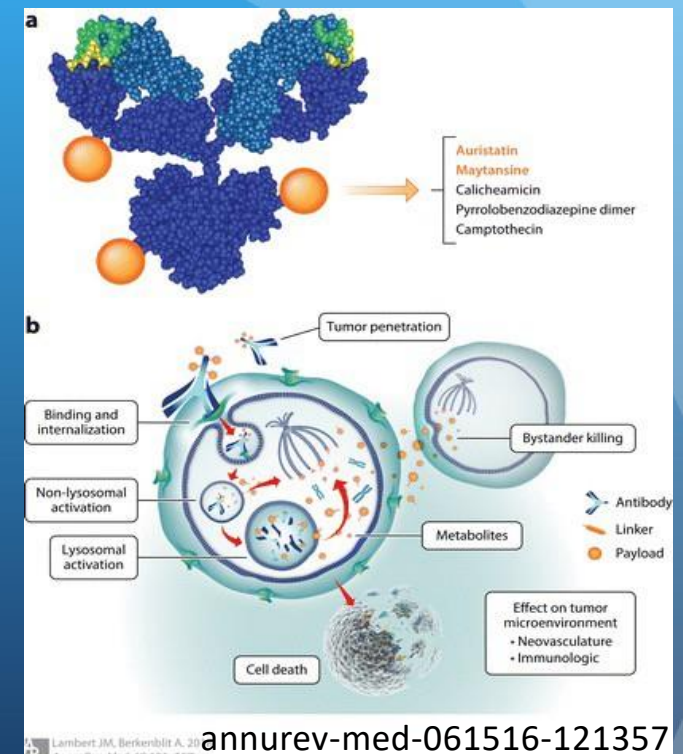
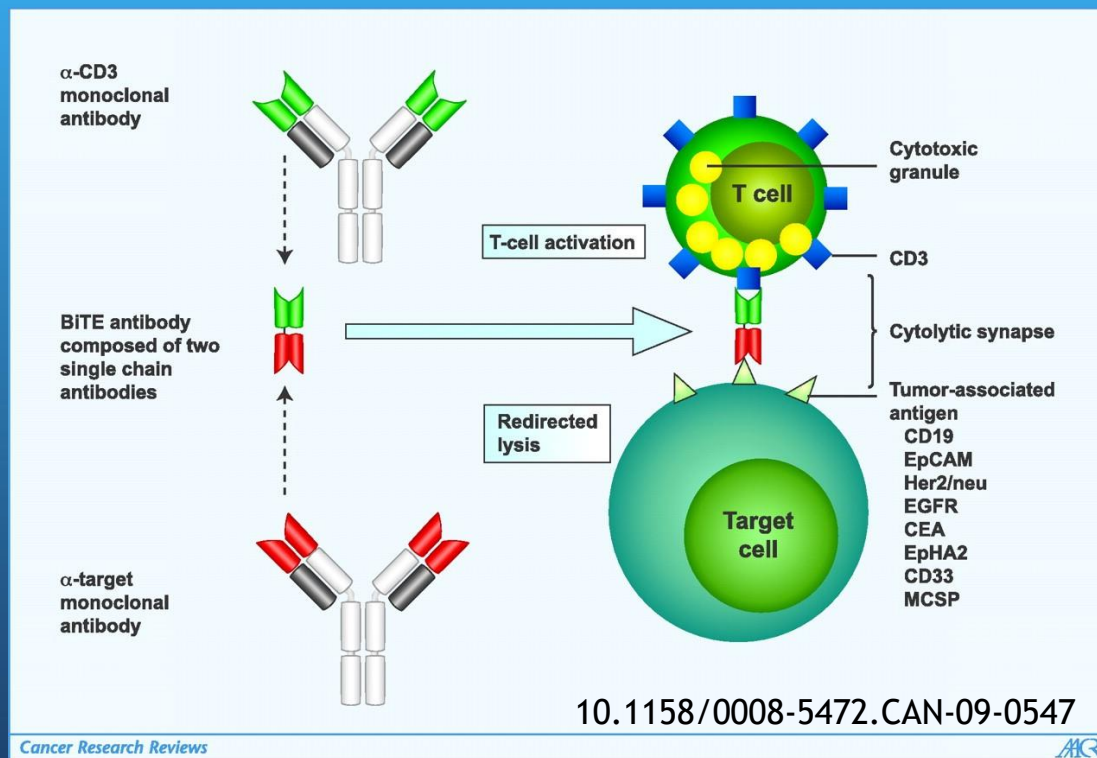


[facebook.com/pedromics](https://facebook.com/pedromics)

- New Chimeric Ag and TCR receptor constructs
  - T & NK / autol & allo
- Augmented effector functions
- Shielded from inhibitory molecules
- Engineered to sense and be protected from low O<sub>2</sub> and pH

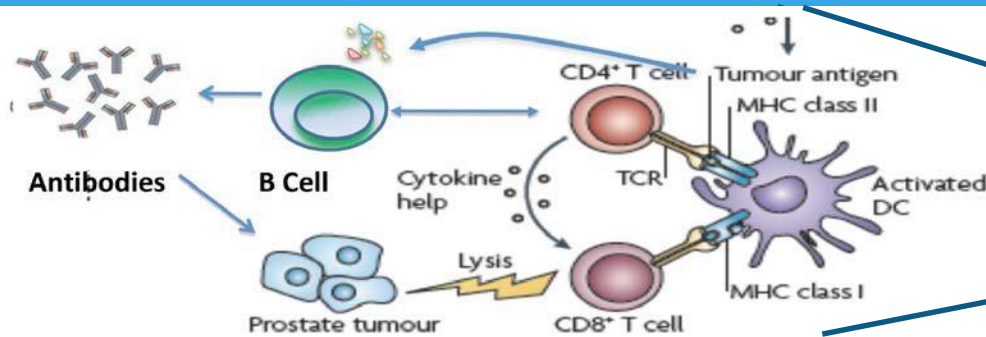
# Synthetic Immunology

## - Targets Immunity or Drug without Gene Therapy



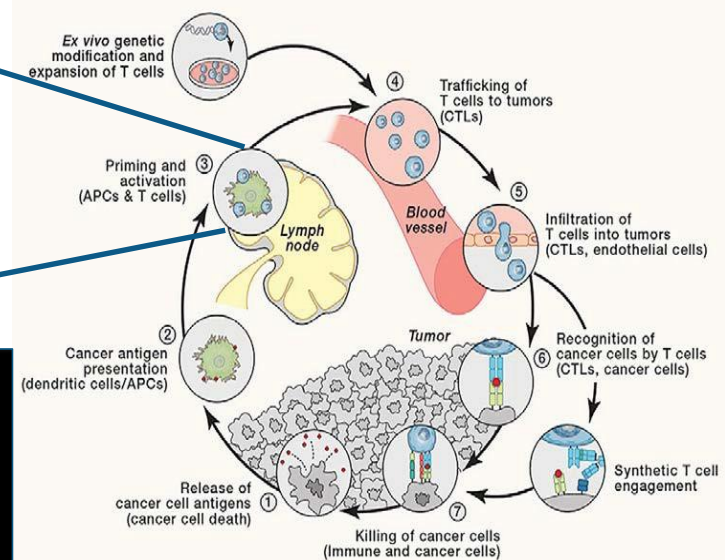


# B Cell Response Reveals Antigens Recognized by CTL



## Coordinated T & B immune response to cancer:

- Kwek S, et al, *J Immunol* 2012
- Tripathi SC, et al, *PNAS* 2016
- Hulett T, et al, *J ImmunoTher Cancer* 2018



Priti S. Hedge and Daniel S. Chen

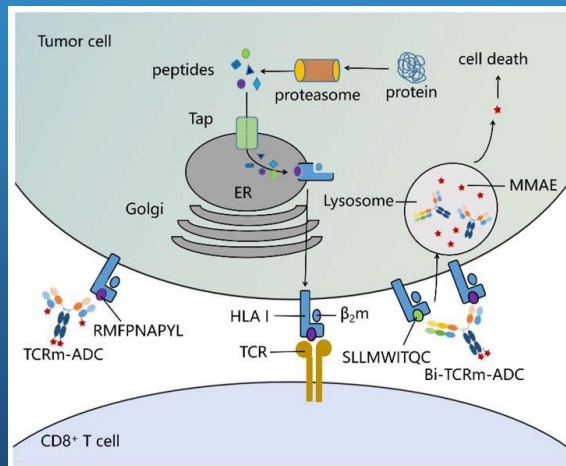
Modified from: Drake C, *Nat Rev Immunol*, 2010  
Goodnow, *Nat. Immunol.* 2010

Immunity 52, January 14, 2020

# Characterized the IgG Profile of Newly Diagnosed Patients with NSCLC - What Were IgG Against?

**Immunoproteasome deficiency is a feature of non-small cell lung cancer with a mesenchymal phenotype and is associated with a poor outcome**

Satyendra C. Tripathi<sup>a</sup>, Haley L. Peters<sup>b</sup>, Ayumu Taguchi<sup>c</sup>, Hiroyuki Katayama<sup>a</sup>, Hong Wang<sup>a</sup>, Amin Momin<sup>a</sup>, Mohit Kumar Jolly<sup>d</sup>, Muge Celiktaş<sup>a</sup>, Jaime Rodriguez-Canales<sup>c</sup>, Hui Liu<sup>c</sup>, Carmen Behrens<sup>c</sup>, Ignacio I. Wistuba<sup>c</sup>, Eshel Ben-Jacob<sup>d,1</sup>, Herbert Levine<sup>d,2</sup>, Jeffrey J. Molldrem<sup>b</sup>, Samir M. Hanash<sup>a</sup>, and Edwin J. Ostrin<sup>e,2</sup>



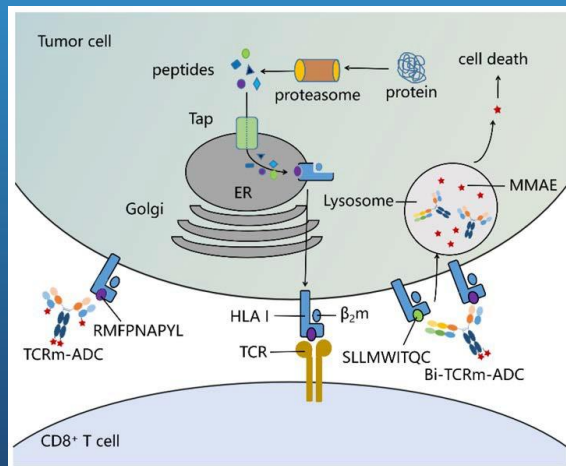
*Int. J. Mol. Sci.* 2019, 20, 3912

Newly diagnosed NSCLC patients have IgG Ab to proteins whose peptides are being presented by HLA on surface of lung cancer cell lines

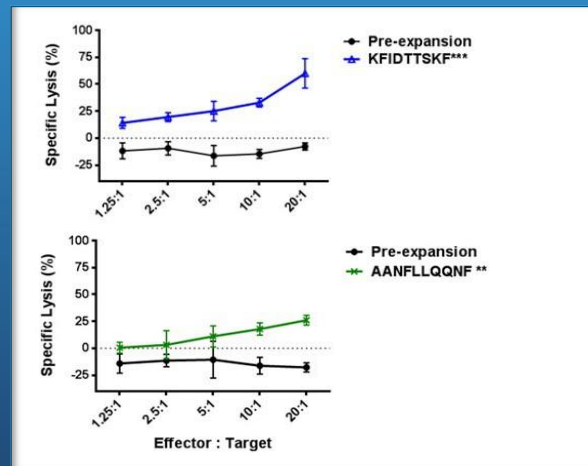
# Newly Diagnosed NSCLC Patients have a Coordinated B and T cell Response to Shared Cancer Antigens

**Immunoproteasome deficiency is a feature of non-small cell lung cancer with a mesenchymal phenotype and is associated with a poor outcome**

Satyendra C. Tripathi<sup>a</sup>, Haley L. Peters<sup>b</sup>, Ayumu Taguchi<sup>c</sup>, Hiroyuki Katayama<sup>a</sup>, Hong Wang<sup>a</sup>, Amin Momin<sup>a</sup>, Mohit Kumar Jolly<sup>d</sup>, Muge Celiktas<sup>a</sup>, Jaime Rodriguez-Canales<sup>c</sup>, Hui Liu<sup>c</sup>, Carmen Behrens<sup>c</sup>, Ignacio I. Wistuba<sup>c</sup>, Eshel Ben-Jacob<sup>d,1</sup>, Herbert Levine<sup>d,2</sup>, Jeffrey J. Mollrem<sup>b</sup>, Samir M. Hanash<sup>a</sup>, and Edwin J. Ostrin<sup>e,2</sup>



*Int. J. Mol. Sci.* 2019, 20, 3912



Patients have T cells that can be activated by peptide (IVS) to become CTL



# ***TIL Recognize Shared Melanoma Antigens***

## **Recognition of Shared Melanoma Antigens in Association With Major HLA-A Alleles by Tumor Infiltrating T Lymphocytes From 123 Patients With Melanoma**

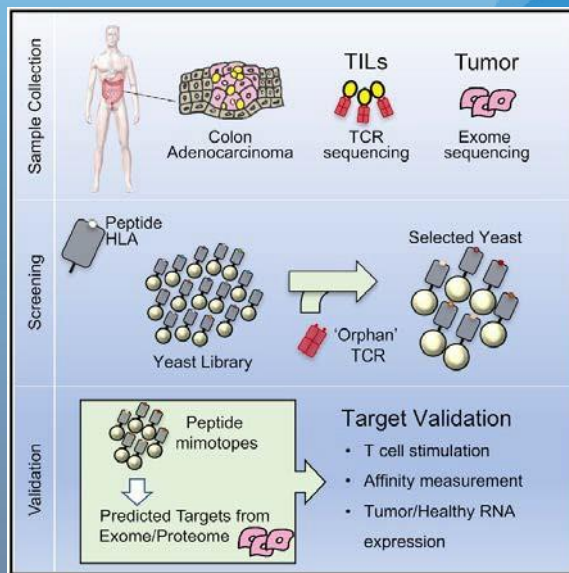
Yutaka Kawakami;Nita Dang;Xiang Wang;Janis Tupesis;Paul Robbins;Rong-Fu Wang;John Wunderlich;John Yannelli;Steven Rosenberg;

- × Screened for recognition of shared melanoma antigens
  - × tyrosinase, MART-1/melan-A, gp100, TRP1, TRP2
  - × peptides derived from MAGE-1 and MAGE-3.
- × Majority of HLA-A2 TIL recognized shared melanoma antigens
- × Recognition of gp100 by HLA-A2 restricted TIL significantly correlated with clinical response to adoptive immunotherapy with TIL in 21 HLA-A2 melanoma patients ( $p = 0.024$ ).

Journal of Immunotherapy. 23(1):17-27, JANUARY 2000

# Antigen Identification for Orphan T Cell Receptors Expressed on Tumor-Infiltrating Lymphocytes

Marvin H. Gee,<sup>1,2,11</sup> Arnold Han,<sup>3,4,11</sup> Shane M. Lofgren,<sup>3,5</sup> John F. Beausang,<sup>6</sup> Juan L. Mendoza,<sup>2</sup> Michael E. Birnbaum,<sup>1,2</sup> Michael T. Bethune,<sup>7</sup> Suzanne Fischer,<sup>2</sup> Xinbo Yang,<sup>2</sup> Raquel Gomez-Eerland,<sup>8</sup> David B. Bingham,<sup>5</sup> Leah V. Sibener,<sup>1,2</sup> Ricardo A. Fernandes,<sup>2</sup> Andrew Velasco,<sup>2</sup> David Baltimore,<sup>7</sup> Ton N. Schumacher,<sup>8</sup> Purvesh Khatri,<sup>3,5</sup> Stephen R. Quake,<sup>6,9</sup> Mark M. Davis,<sup>3,4,10</sup> and K. Christopher Garcia<sup>2,10,12,\*</sup>



- 3 of 4 TCRs characterized recognized non-mutated peptide that is overexpressed in colon cancer and other cancers
- Identification of a shared non-mutated tumor antigen between two patients (same TCR  $\alpha$ )
  - U2AF2 overexpressed in many cancers incl. COAD, NSCLC, Breast and lymphoma

Gee et al., 2018, Cell 172, 1–15

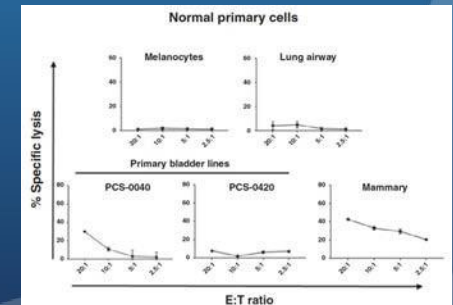
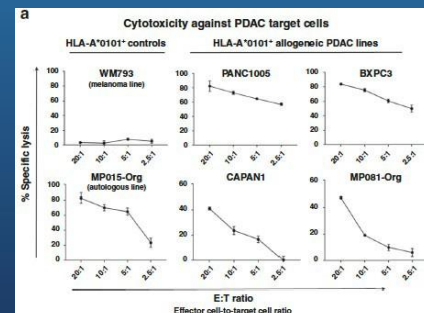
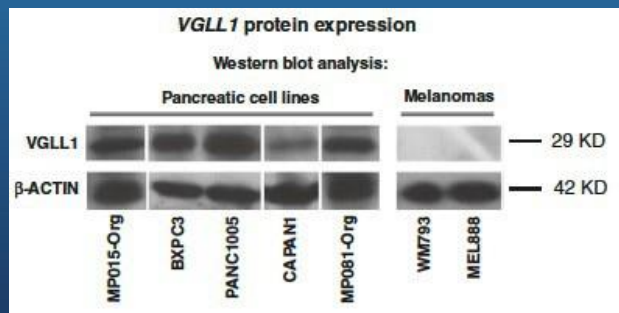
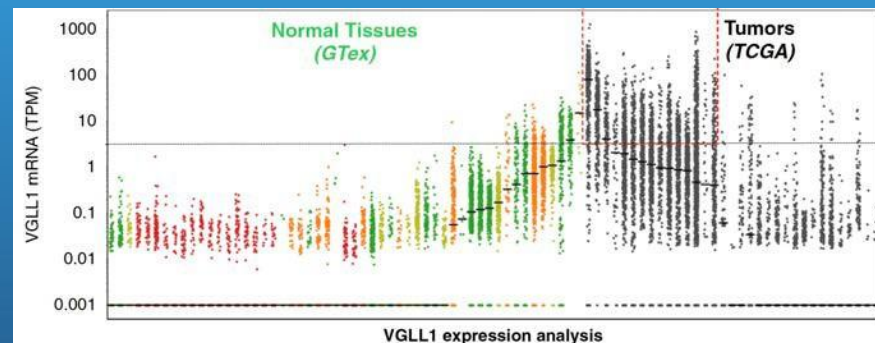
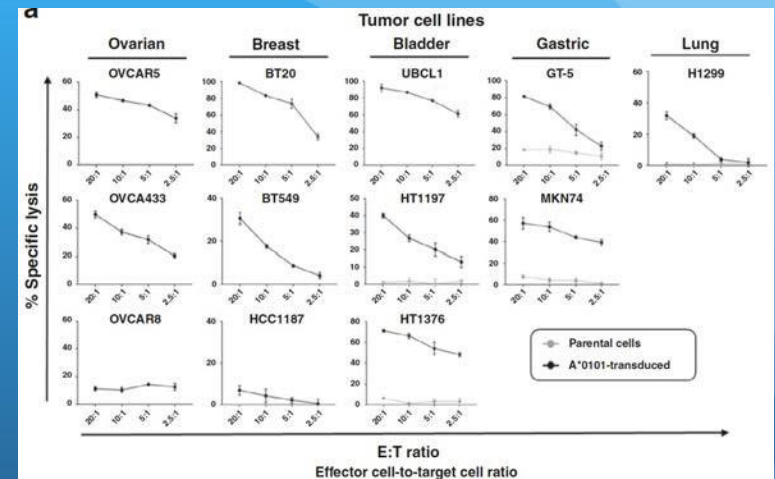
# CTL recognize VGLL1 & Kill Pancreatic, Breast, Ovarian, Bladder, Gastric & Lung Cancer / But not Normal Cells

Vestigial-like 1 is a shared targetable cancer-placenta antigen expressed by pancreatic and basal-like breast cancers

Sherille D. Bradley<sup>1</sup>, Amjad H. Talukder<sup>1</sup>, Ivy Lai<sup>1</sup>, Rebecca Davis<sup>1</sup>, Hector Alvarez<sup>2</sup>, Herve Tiriach<sup>3</sup>, Mingyong Zhang<sup>1</sup>, Yulun Chiu<sup>1</sup>, Brenda Melendez<sup>1</sup>, Kyle R. Jackson<sup>1</sup>, Arjun Katailhi<sup>1</sup>, Heather M. Sonnemann<sup>1</sup>, Fenge Li<sup>1</sup>, Yaan Kang<sup>4</sup>, Na Qiao<sup>5</sup>, Bi-Fang Pan<sup>6</sup>, Philip L. Lorenzi<sup>7</sup>, Mark Hurd<sup>8</sup>, Elizabeth A. Mittendorf<sup>4</sup>, Christine B. Peterson<sup>9</sup>, Milind Javle<sup>10</sup>, Christopher Bristow<sup>11</sup>, Michael Kim<sup>4</sup>, David A. Tuveson<sup>3</sup>, David Hawke<sup>6</sup>, Scott Kopetz<sup>10</sup>, Robert A. Wolff<sup>10</sup>, Patrick Hwu<sup>1</sup>, Anirban Maitra<sup>12</sup>, Jason Roszik<sup>1</sup>, Cassian Yee<sup>1,13</sup> & Gregory Lizée<sup>1,13</sup>

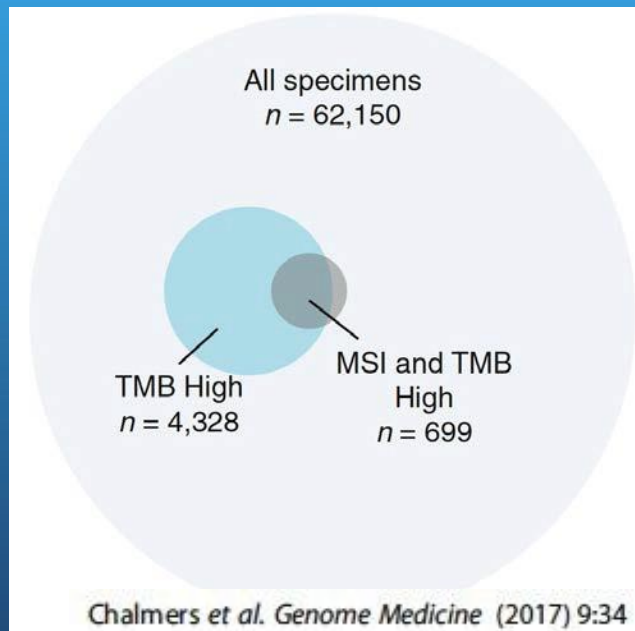


21 October 2020





# It is Good News that the Immune System can Recognize Non-Mutated Shared Ags



## Source of Antigens for Low TMB

- × Overexpressed self
- × Cancer Testis Ags
- × Viral antigens

ON MEDICINE

## The Search for Cancer Treatment Beyond Mutant-Hunting



Photo illustration by Cristiana Couceiro. Cells: National Cancer Institute, via Wikipedia.

# Shared Non-Mutated Cancer Antigens

## *Human Cancer Biology*

### **The Prioritization of Cancer Antigens: A National Cancer Institute Pilot Project for the Acceleration of Translational Research**

Martin A. Cheever,<sup>1</sup> James P. Allison,<sup>2</sup> Andrea S. Ferris,<sup>3</sup> Olivera J. Finn,<sup>4</sup> Benjamin M. Hastings,<sup>3</sup> Toby T. Hecht,<sup>5</sup> Ira Mellman,<sup>7</sup> Sheila A. Prindiville,<sup>6</sup> Jaye L. Viner,<sup>6</sup> Louis M. Weiner,<sup>8</sup> and Lynn M. Matrisian<sup>6</sup>

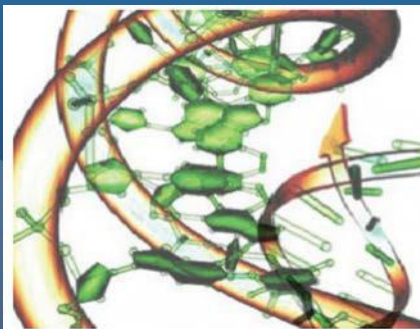
*Clin Cancer Res* 2009;15:5323-5337



## **THE CANCER GENOME ATLAS**

National Cancer Institute

National Human Genome Research Institute



- ID genes in cancer that are upregulated, amplified, mutated compared to normal tissue
- Associations with survival

# How to Tailor Therapy to a Specific Patient?

# Novel technologies and emerging biomarkers for personalized cancer immunotherapy


Journal for ImmunoTherapy of Cancer

Yuan et al. *Journal for ImmunoTherapy of Cancer* (2016) 4:3  
DOI 10.1186/s40425-016-0107-3

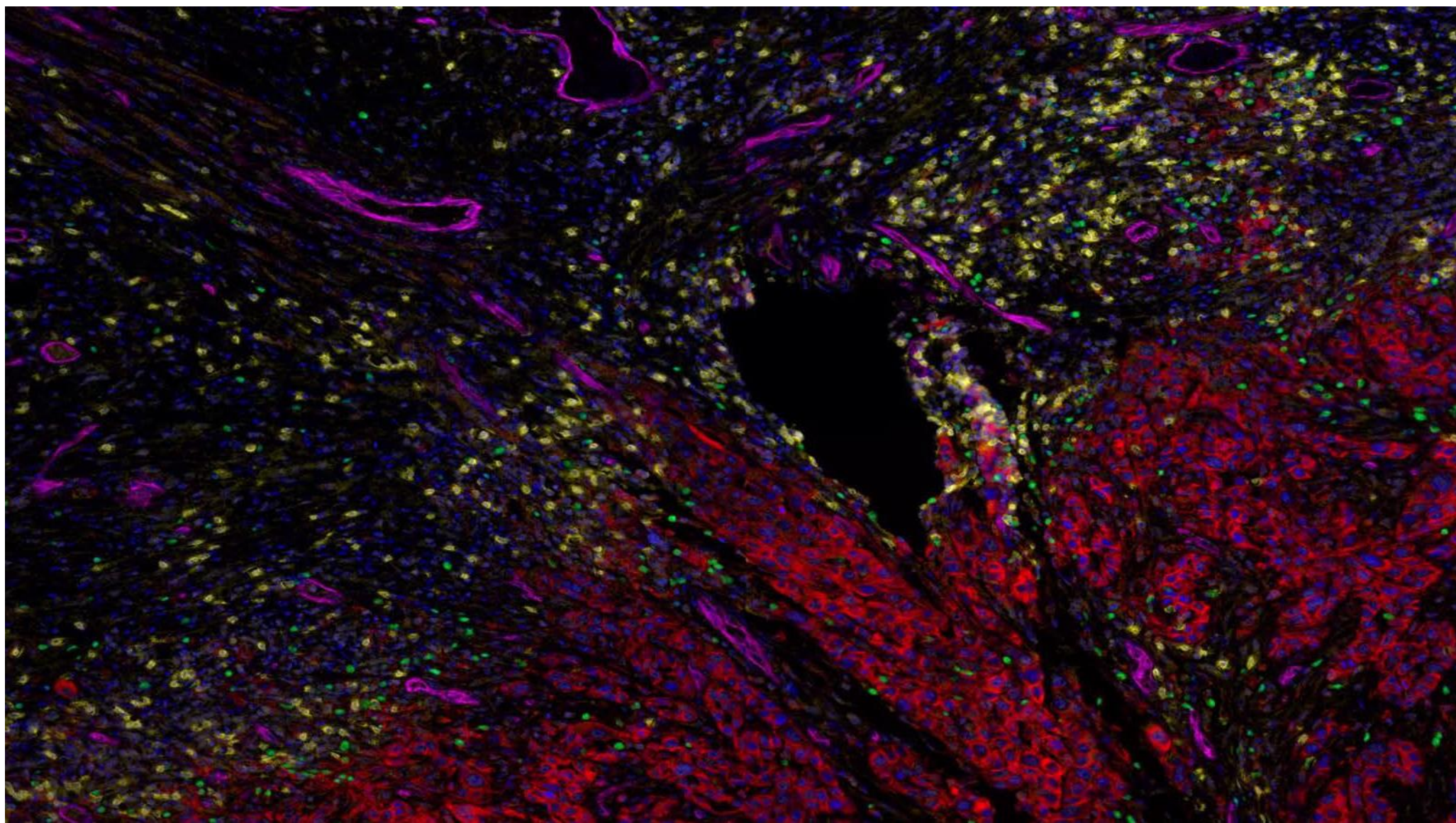
Jianda Yuan<sup>1\*</sup>, Priti S. Hegde<sup>2</sup>, Raphael Clynes<sup>3</sup>, Periklis G. Foukas<sup>4,5</sup>, Alexandre Harari<sup>4</sup>, Thomas O. Kleen<sup>6</sup>, Pia Kvistborg<sup>7</sup>, Cristina MacCalli<sup>8</sup>, Holden T. Maecker<sup>9</sup>, David B. Page<sup>10</sup>, Harlan Robins<sup>11</sup>, Wenru Song<sup>12</sup>, Edward C. Stack<sup>13</sup>, Ena Wang<sup>14</sup>, Theresa L. Whiteside<sup>15</sup>, Yingdong Zhao<sup>16</sup>, Heinz Zwiernina<sup>17</sup>, Lisa H. Butterfield<sup>18</sup> and Bernard A. Fox<sup>10\*</sup>

SITC Immune Biomarkers Task Force

## Novel immune monitoring assays for biomarker discovery and personalized cancer immunotherapy

Monitoring strategy	Immunologically-unresponsive tumor	Immunologically-responsive tumor
<b>Whole exome sequencing</b>	Low mutational burden	High mutational burden
<b>Gene signature/patterns</b>	↓ activation signature	↑ activation signature
<b>Epigenetic modification</b>	↑ Treg/CD3 ratio ↓ CD3 cells	↓ Treg/CD3 ratio ↑ CD3 cells
<b>Protein microarray</b>	Poor general antibody response	Robust general antibody response
<b>B/ T-cell receptor repertoire</b>	Low CD3 count Low clonality	High CD3 count High clonality
<b>Flow/Mass cytometry</b>	↓ effector cells ↓ Teff/Treg ratio	↑ effector cells ↑ Teff/Treg ratio
<b>Multicolor IHC</b>	↓ effector cells, ↑ suppressor cells low PD-L1 on tumor and tumor infiltrating immune cells	↑ effector cells ↓ suppressor cells high PD-L1 on tumor and tumor infiltrating immune cell
<b>Therapeutic strategy</b>	Vaccination, ablation, radiotherapy, chemotherapy, oncolytic therapy, adaptive cellular therapy first	Immune checkpoint blockade therapies and other immunotherapies first
<b>Legend</b>		







JCI insight

RESEARCH ARTICLE

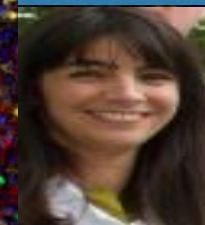
## Multiparametric immune profiling in HPV<sup>-</sup> oral squamous cell cancer

Zipei Feng,<sup>1,2</sup> Daniel Bethmann,<sup>1,3</sup> Matthias Kappler,<sup>4</sup> Carmen Ballesteros-Merino,<sup>1</sup>  
Alexander Eckert,<sup>4</sup> R. Bryan Bell,<sup>1,5</sup> Allen Cheng,<sup>5</sup> Tuan Bui,<sup>5</sup> Rom Leidner,<sup>1,5</sup> Walter J. Urba,<sup>1</sup>  
Kent Johnson,<sup>6</sup> Clifford Hoyt,<sup>6</sup> Carlo B. Bifulco,<sup>1,7</sup> Juergen Bukur,<sup>8</sup> Claudia Wickenhauser,<sup>3</sup>  
Barbara Seliger,<sup>8</sup> and Bernard A. Fox<sup>1,9</sup>

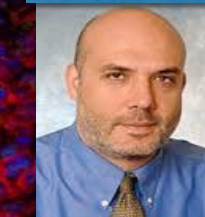
*JCI Insight 2 (14) 2017*



Zipei Feng



Carmen  
Ballesteros-  
Merino



Carlo Bifulco



EARLE A. CHILES  
RESEARCH INSTITUTE

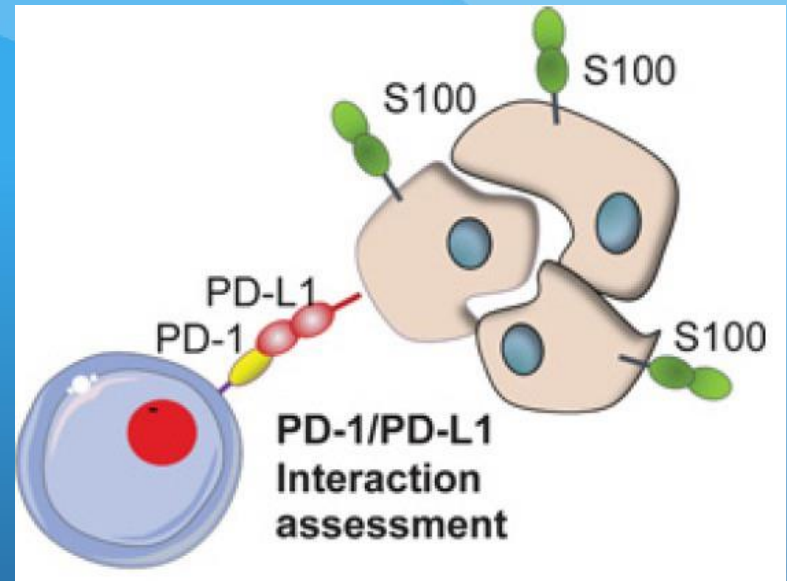
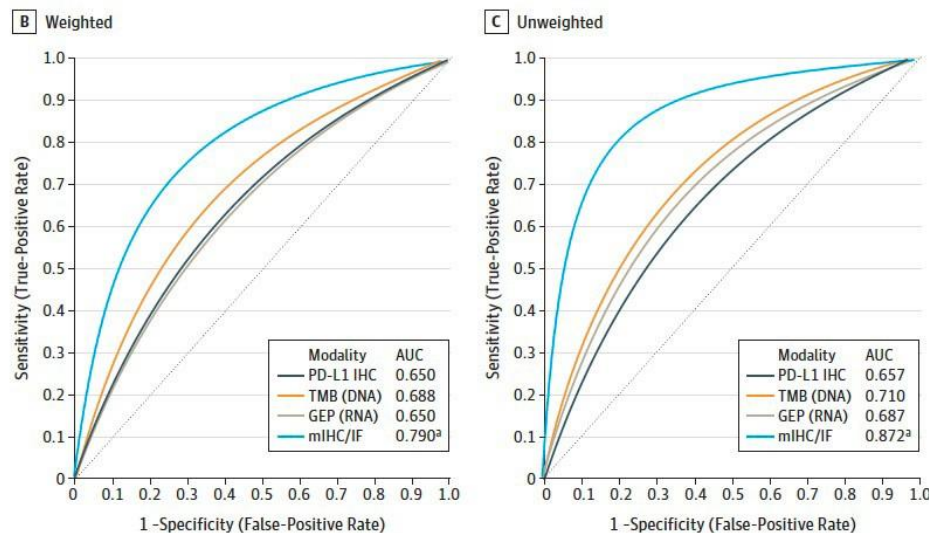


# Multiplex IHC

JAMA Oncology | Original Investigation

## Comparison of Biomarker Modalities for Predicting Response to PD-1/PD-L1 Checkpoint Blockade A Systematic Review and Meta-analysis

Steve Lu; Julie E. Stein, MD; David L. Rimm, MD, PhD; Daphne W. Wang, MS; J. Michael Bell;  
Douglas B. Johnson, MD; Jeffrey A. Sosman, MD; Kurt A. Schalper, MD, PhD; Robert A. Anders, MD, PhD;  
Hao Wang, PhD; Clifford Hoyt, MS; Drew M. Pardoll, MD, PhD; Ludmila Danilova, PhD; Janis M. Taube, MD



**Future:** These assays will be standardized/approved & enrich for patients likely to respond to CPI & ID pts who need clinical trials

# Single cell sequencing & Digital Spatial Profiling (DSP)

## Single Cell Seq:

Provides assessment of the status of all the cells in a tumor

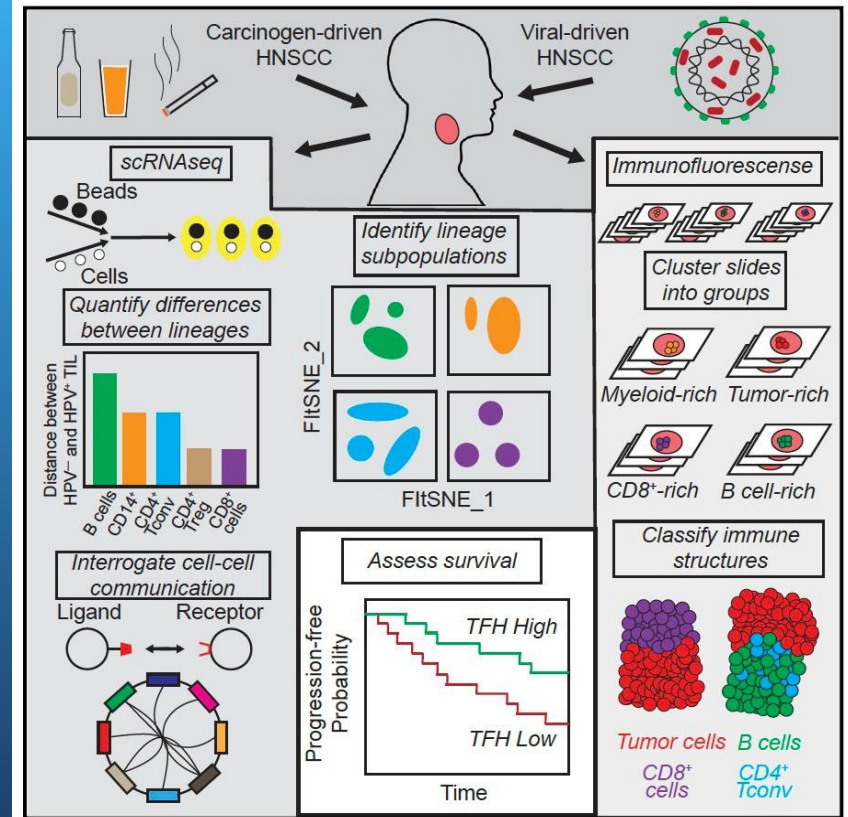
## DSP:

Allows and assessment of the cells that are interacting with each other in a specific spot

Immunity 52, 183–199, January 14, 2020

## Immune Landscape of Viral- and Carcinogen-Driven Head and Neck Cancer

Anthony R. Cillo,<sup>1,2</sup> Cornelius H.L. Kürten,<sup>2,3,4</sup> Tracy Tabib,<sup>5</sup> Zengbiao Qi,<sup>5</sup> Sayali Onkar,<sup>1,2</sup> Ting Wang,<sup>6</sup> Angen Liu,<sup>7</sup> Umamaheswar Duvvuri,<sup>3</sup> Seungwon Kim,<sup>3</sup> Ryan J. Soose,<sup>3</sup> Steffi Oesterreich,<sup>8,9</sup> Wei Chen,<sup>6</sup> Robert Lafyatis,<sup>5</sup> Tullia C. Bruno,<sup>1,2,\*</sup> Robert L. Ferris,<sup>1,2,3,\*</sup> and Dario A.A. Vignali<sup>1,2,10,\*</sup>





# Observation:

- × Patients with Melanoma, Pancreatic, Breast, Ovarian, Bladder, Gastric, Colon & Lung Cancer have immunity to non-mutated cancer epitopes
- × Once “re-activated” with Ag and cytokine - T cells can kill cancer cells

# Observation:

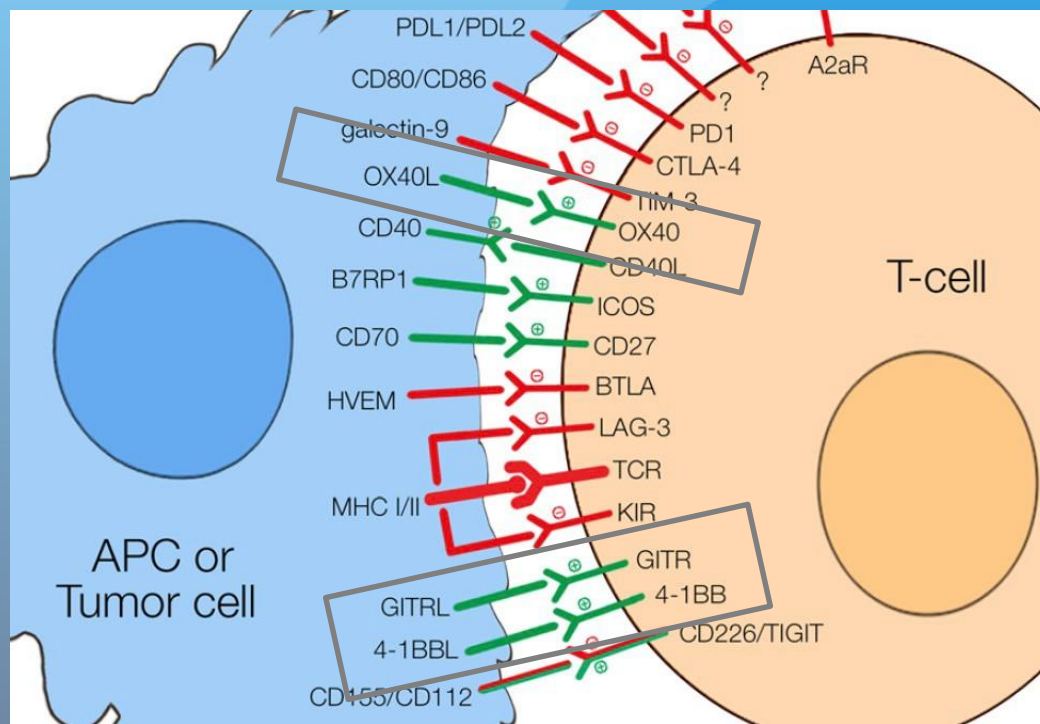
- × Many patients with Melanoma, Pancreatic, Breast, Ovarian, Bladder, Gastric, Colon & Lung Cancer have immunity to non-mutated cancer epitopes
- × Once “re-activated” with Ag and cytokine - T cells can kill cancer cells

# Challenge:

- Identify approaches that re-activate and expand tumor-reactive T cells

# Costimulation Augments T cell Function

## OX40, GITR and 4-1BB



Receptor expression is upregulated following T cell activation

Triggering co-stimulates T cells

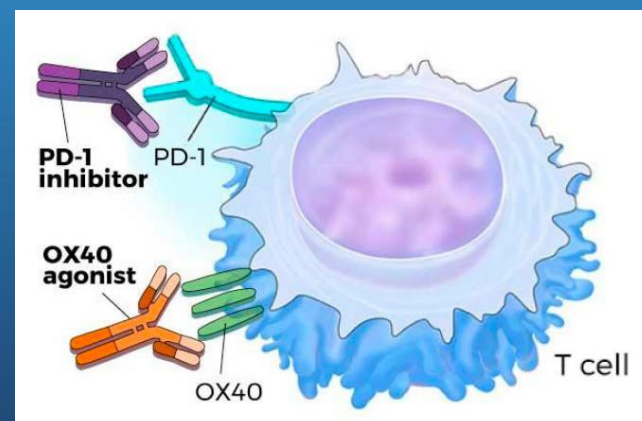
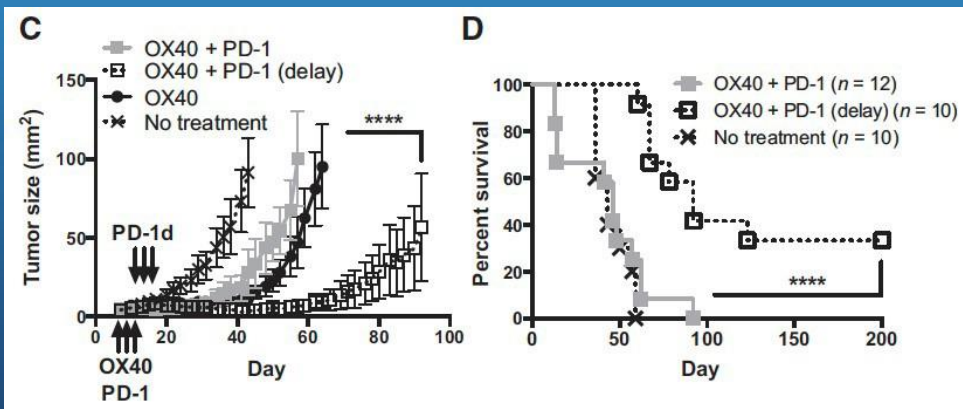
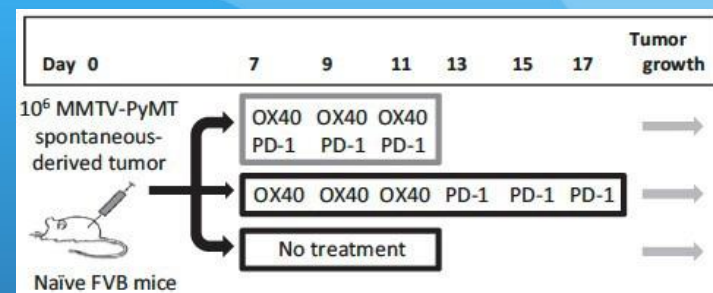
Can augment Anti-Cancer Activity

Marin-Acevedo et al. *Journal of Hematology & Oncology* (2018) 11:39

# Timing and Sequence Critical for Immunotherapy Combination

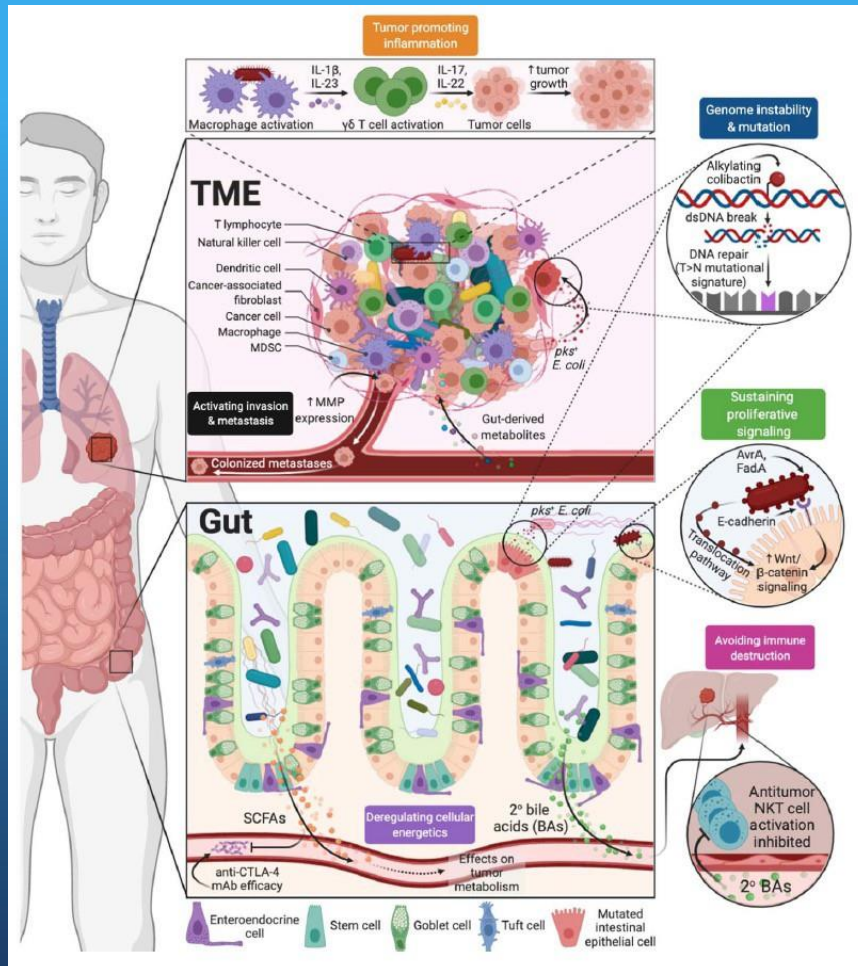
## Timing of PD-1 Blockade Is Critical to Effective Combination Immunotherapy with Anti-OX40

David J. Messenheimer<sup>1,2</sup>, Shawn M. Jensen<sup>1</sup>, Michael E. Afentoulis<sup>1</sup>, Keith W. Wegmann<sup>1</sup>, Zipei Feng<sup>1,3</sup>, David J. Friedman<sup>1</sup>, Michael J. Gough<sup>1</sup>, Walter J. Urbani<sup>1</sup>, and Bernard A. Fox<sup>1,2,3,4</sup>





# Intersection of microbial mechanisms with cancer hallmarks



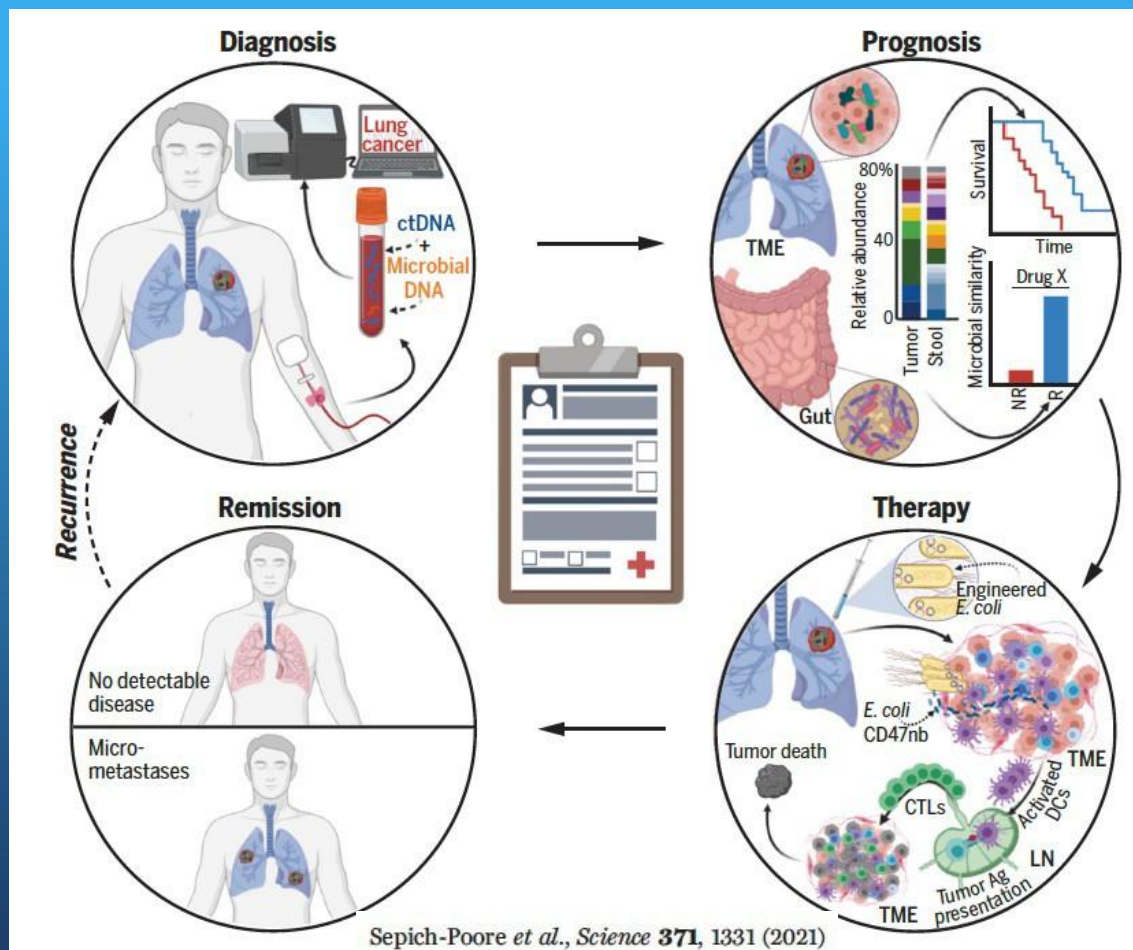
## The microbiome and human cancer

Gregory D. Sepich-Poore<sup>1</sup>, Laurence Zitvogel<sup>2,3,4,5</sup>, Ravid Straussman<sup>6</sup>, Jeff Hasty<sup>1,7,8</sup>, Jennifer A. Wargo<sup>9,10</sup>, Rob Knight<sup>1,11,12\*</sup>

Sepich-Poore *et al.*, *Science* **371**, 1331 (2021)

Microbiota-derived metabolites, genotoxins, and antigens influence host antitumor immunity, inflammation, energetics, cellular signaling, and metastasis

# Opportunities for Microbes to Affects Cancer Care



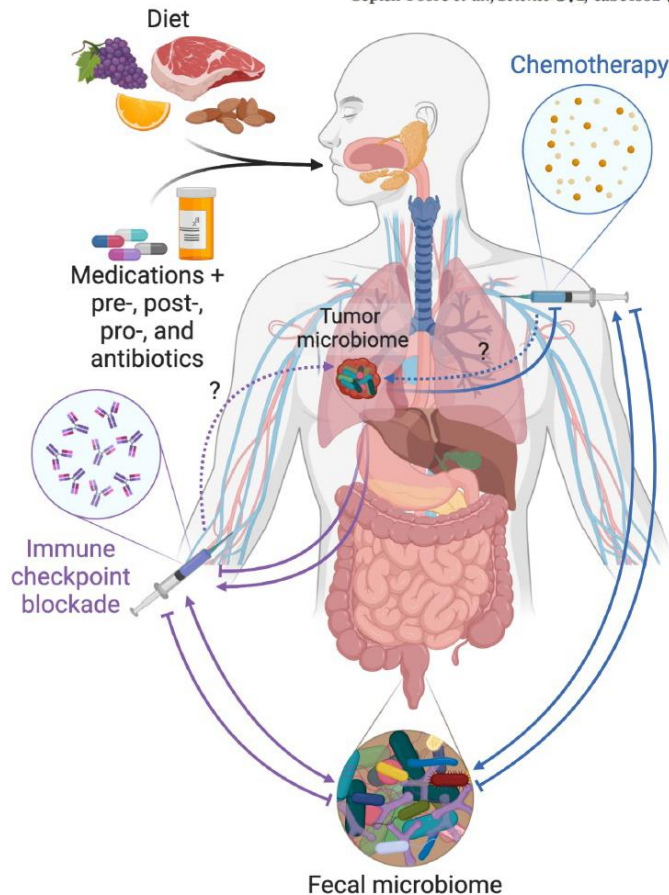
**Future:**  
Understanding the  
microbiome will  
impact:

- Diagnosis
- Prognosis
- Therapy

# The microbiome and human cancer

Gregory D. Sepich-Poore<sup>1</sup>, Laurence Zitvogel<sup>2,3,4,5</sup>, Ravid Straussman<sup>6</sup>, Jeff Hasty<sup>1,7,8</sup>, Jennifer A. Wargo<sup>9,10</sup>, Rob Knight<sup>1,11,12\*</sup>

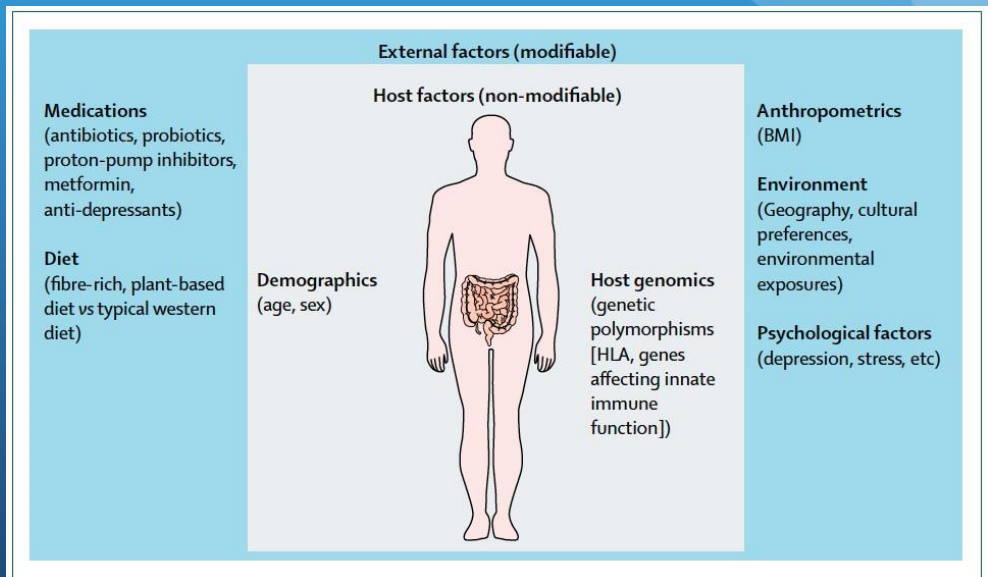
Sepich-Poore et al., *Science* **371**, eabc4552 (2021)



## Modulating the microbiome to improve therapeutic response in cancer

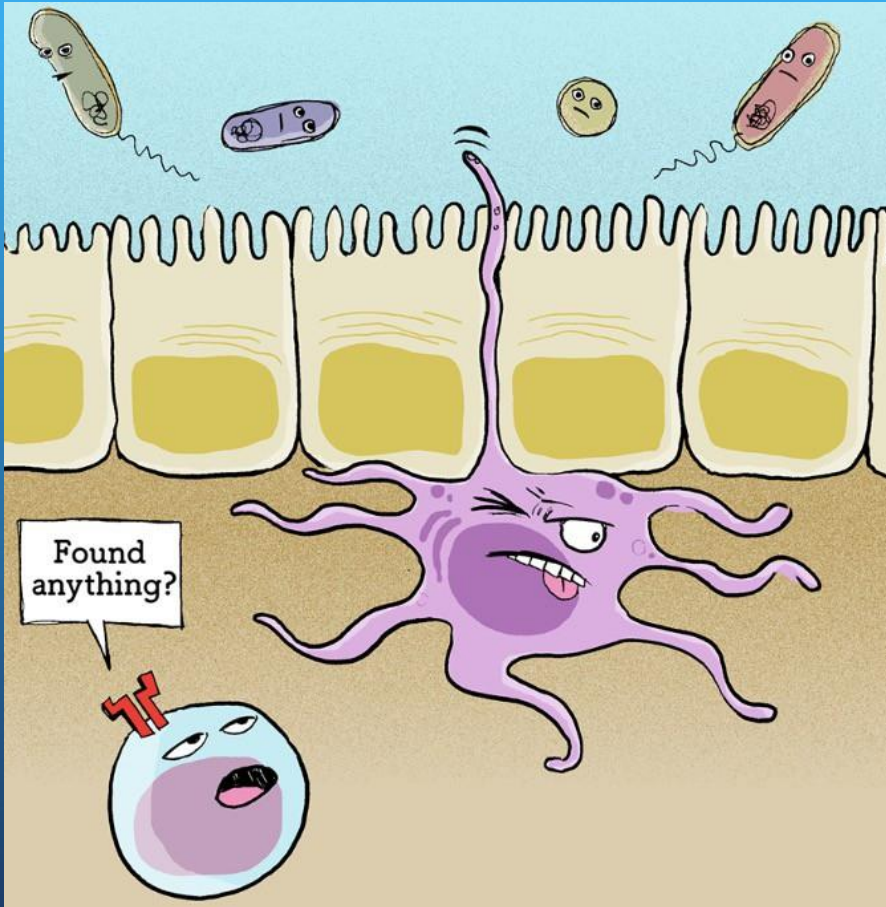
Jennifer L McQuade, Carrie R Daniel, Beth A Helmink, Jennifer A Wargo

*Lancet Oncol* 2019; 20: e77-91





# Microbiome Effects on Immune Responsiveness



## Clinical Trials.

- Fecal/microbe transplants
- Probiotics
- Diets/prebiotics
- Phage
- Antibiotics / other

**Future:** Lots of questions?  
When to treat and how to assess impact?



# Questions to consider in clinical trial design for microbiome modulation in cancer

## Before treatment

### Patients

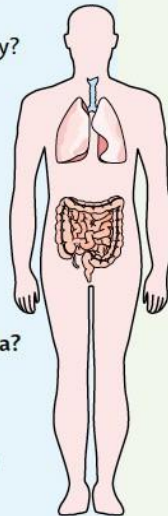
- Which patient population to treat: treatment naive or refractory?
- Should the microbiome be profiled to stratify or select patients?

### Pre-conditioning regimen

- Do we need to pre-treat the gut with antibiotics to facilitate engraftment?

### How should we optimally modulate the gut microbiota?

- FMT?
- Diet?
- Designer consortia?
- Phage, antibiotics, or other?



## During therapy

### What therapy should we combine with modulation of the gut microbiome?

- Immune checkpoint blockade anti-PD-1)?
- Other forms of immunotherapy?
- Other therapy?

### How do we optimally monitor patients during therapy?

- Microbiome analyses to assess engraftment or function?
- Immune profiling?
- Peripheral blood?
- Tumour?

### How can we facilitate stable engraftment?

- Should we recommend dietary changes?
- Any medications to avoid?

## Assessing effects

### What are appropriate primary endpoints for such studies?

- Safety and tolerability?
- Engraftment?
- Others?

### What are appropriate secondary endpoints?

- Response?
  - Radiographic (RECIST or irRC)?
  - Rate of complete responses?
  - Pathological response (on biopsy or after neoadjuvant therapy)?
- Toxicity?
- Novel markers (ctDNA, immunophenotyping)?

## Long-term effects

### Durability of engraftment

- How durable is engraftment?
- What microbes or functional phenotypes in gut microbiota are associated with responses? Can these microorganisms be used to design consortia?

### Overall responses

- What is the effect on overall and disease-specific survival?

### Toxicity

- Can we uncouple toxicity and response to immunotherapy?

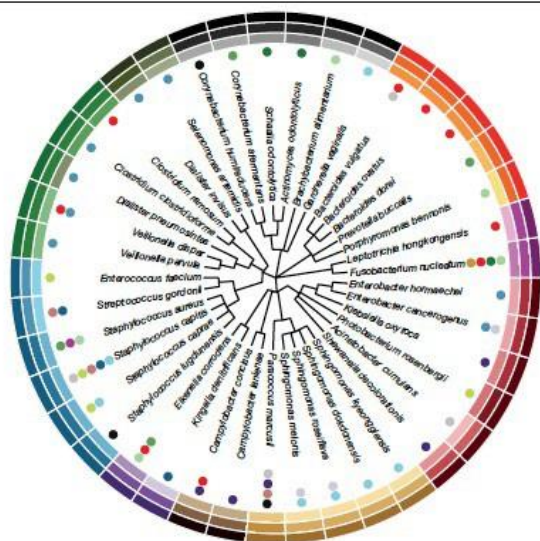
### Other transplanted traits with FMT?

- Obesity?
- Depression?
- Any potentially favourable traits?

Lancet Oncol 2019; 20: e77-91

# Identification of bacteria-derived HLA-bound peptides in melanoma

Nature. 2021 Apr;592(7852):138-143.

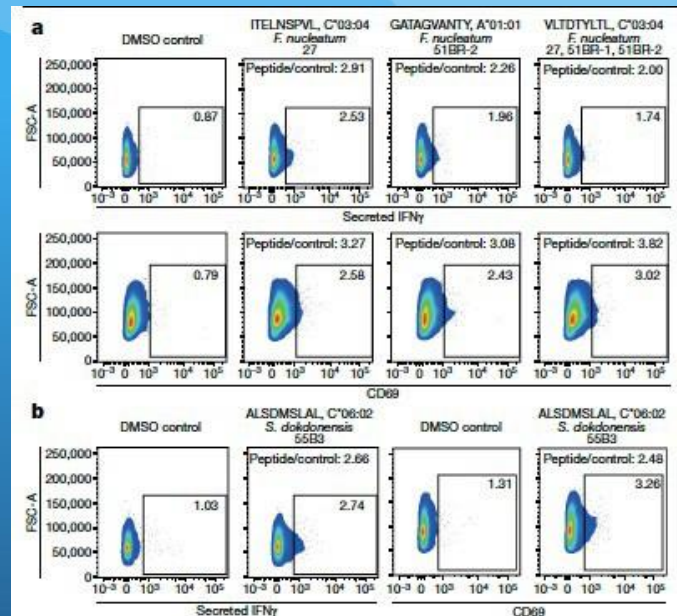


Class	Order	Genus
Bacilli	Bacillales	Staphylococcus
	Lactobacillales	Streptococcus
Bacteroidia	Bacteroidales	Bacteroides
		Prevotella
		Porphyromonas
Epsilonproteobacteria	Campylobacterales	Campylobacter
Clostridia	Clostridiales	Clostridium
		Veillonella
		Dialister
Betaproteobacteria	Neisseriales	Eikenella
		Kingella
Fusobacteria	Fusobacteriales	Fusobacterium
		Leptotrichia
Gammaaproteobacteria	Vibrionales	Photobacterium
	Enterobacteriales	Klebsiella
	Alteromonadales	Shewanella
	Pseudomonadales	Enterobacter
		Acinetobacter
Actinobacteria	Actinomycetales	Actinomyces
	Micrococcales	Schaalia
	Bifidobacteriales	Corynebacterium
		Brachybacterium
		Gardnerella
Alphaproteobacteria	Rhodobacterales	Paracoccus
	Sphingomonadales	Sphingomonas
Negativicutes	Veillonellales	Dialister
	Selenomonadales	Selenomonas

Patients and metastases									
19	42	51	55	70	86	92	112	152	
19T	42F	51A	55A3	70.1	803	92B3	112	152A2	
42F5	51B1	55A7	55B3	8032					

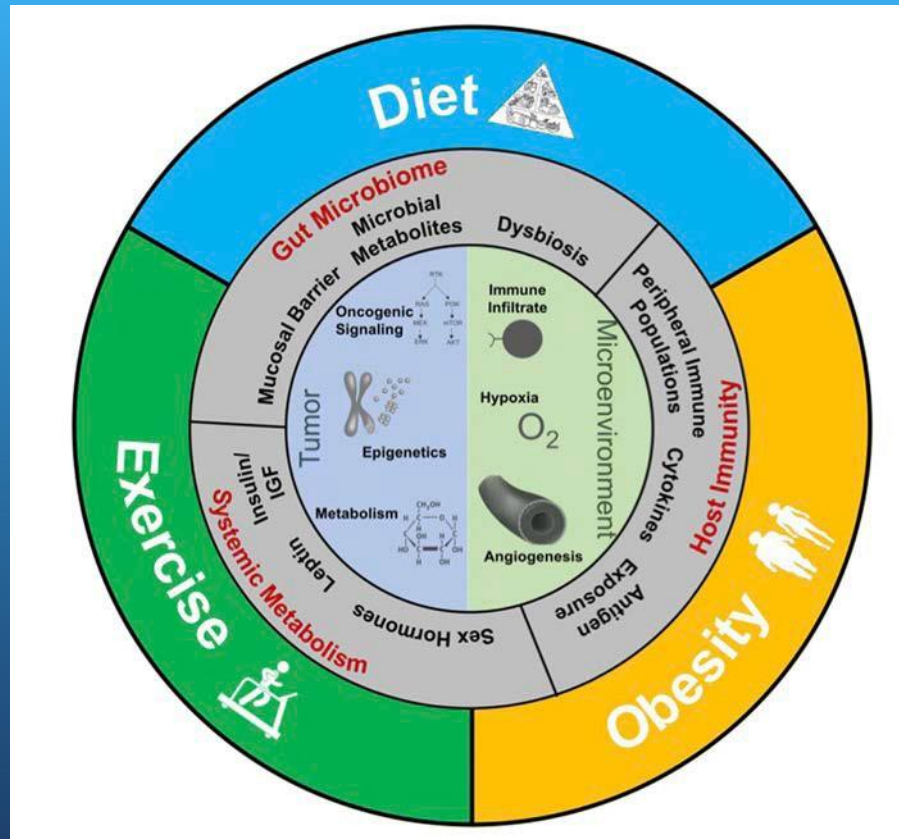
**Fig. 1 | Identification of intratumoral bacteria in melanoma.** Schematic phylogenetic tree of the bacterial composition of 17 melanoma metastases that originated from 9 patients. The analysis is based on rRNA 16S gene sequencing. The different colours and shades in the circles indicate the different

classifications of bacteria at the genus (inner circle), order (middle circle) and class (outer circle) level. Each patient is colour-coded (as in the index), and different metastases from the same patient are depicted in different shades of the same colour.



**Fig. 4 | TIL reactivity towards bacteria-derived antigens.** IFN- $\gamma$ -secreting 51A3 and 55A3 TILs were detected after 6 h of coculture with B cells loaded with a bacterial peptide or dimethyl sulfoxide (DMSO) control, using flow cytometry. TILs were also tested for the presence of the CD69 reactivity marker. The image presents 4 representative immunogenic peptides (3 out of 7 immunogenic peptides of patient 51 and 1 out of 1 peptides of patient 55) that showed at least a 2-fold change between the peptide and DMSO control. The percentage of positive IFN- $\gamma$ -secreting or CD69-expressing TILs is an average of three independent experiments (Extended Data Fig. 12, Supplementary Figs. 10, 11). a, Patient 51. b, Patient 55.

# Not Just Diet's Impact on Microbiome -



[Curr Oncol Rep. 2019 Jul 1; 21\(8\): 72.](#)





## Body Weight Affects Cancer Risk

- × Being overweight or obese is clearly linked to an overall increased risk of cancer.
- × 8% of all cancers in the United States-
- × 7% of all cancer deaths.

### Clearly linked with an increased risk of:

- Breast (in women past menopause)
- Colon and rectum
- Endometrium (lining of the uterus)
- Esophagus
- Kidney
- Pancreas

### May raise the risk of:

- Gallbladder
- Liver
- Non-Hodgkin lymphoma
- Multiple myeloma
- Cervix
- Ovary
- Aggressive prostate cancer

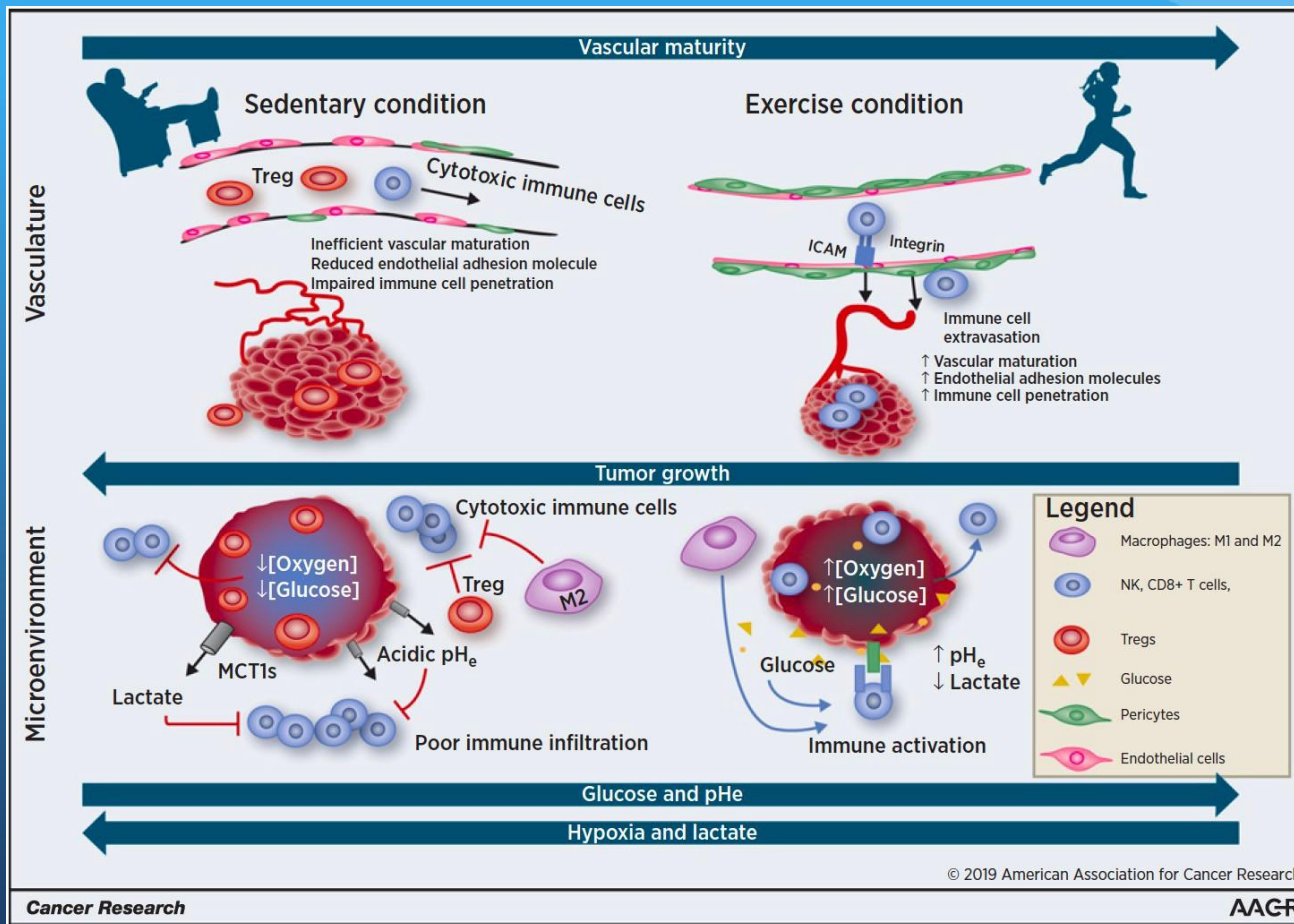
<https://www.cancer.org/cancer/cancer-causes/diet-physical-activity/body-weight-and-cancer-risk/effects.html>





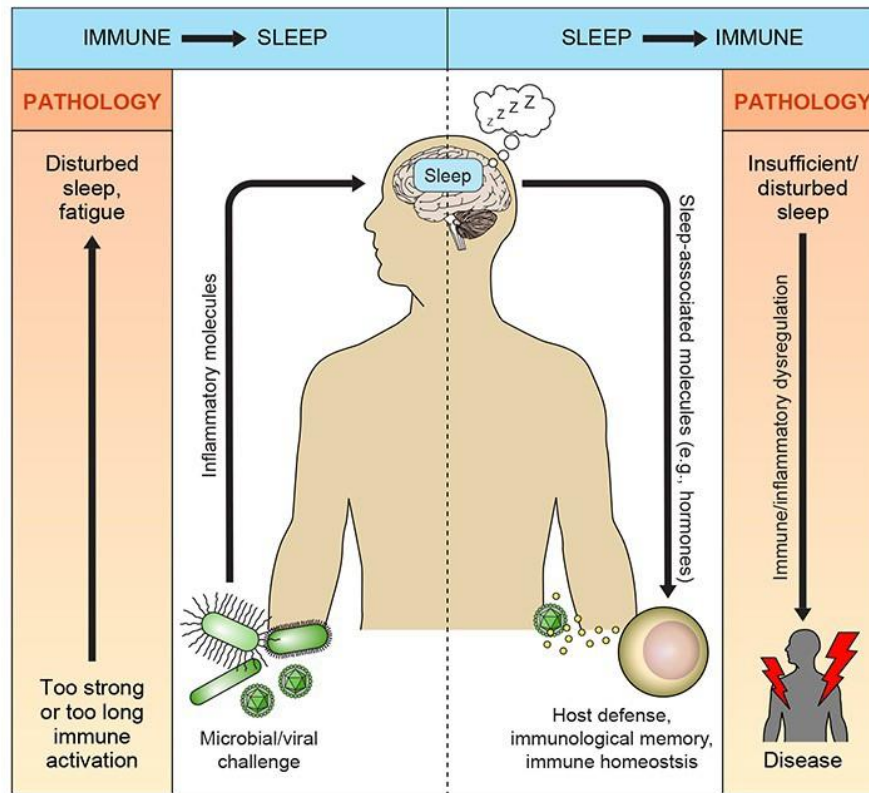
**Future:**  
Wearables are  
coming to clinical  
trials near you!  
Here is WHY!

# Impact of Exercise



**Future:**  
We will  
objectively  
assess activity  
of patients on  
clinical trials

# Sleep-Immune Crosstalk in Health and Disease



**Future:**  
Tracking sleep will provide non-invasive strategy to assess “immune response” of patients on clinical trials



## Is Insomnia a Risk Factor for Decreased Influenza Vaccine Response?

Daniel J. Taylor, Kimberly Kelly, Marian L. Kohut & Kai-Sheng Song

*Behavioral Sleep Medicine*, 00:1–18, 2016

Results indicate insomnia may be a risk factor for lowered immunity to the influenza virus

## Sleep and Antibody Response to Hepatitis B Vaccination

Aric A. Prather, PhD<sup>1,2</sup>; Martica Hall, PhD<sup>3</sup>; Jacqueline M. Fury, BS<sup>1</sup>; Diana C. Ross, MSN, RN<sup>1</sup>; Matthew F. Muldoon, MD, MPH<sup>4</sup>; Sheldon Cohen, PhD<sup>5</sup>; Anna L. Marsland, PhD, RN<sup>1</sup>

*SLEEP* 2012;35(8):1063-1069.

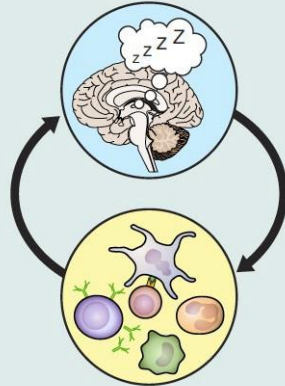
Sleep duration was associated with decreased likelihood of clinical protection, which remained significant after adjustment for age, sex, and BMI (OR, 3.53; 95% CI, 1.22-10.27,  $P = 0.02$ )

# Sleep-Immune Crosstalk in Health and Disease

## Experimental studies

### Sleep changes induced by:

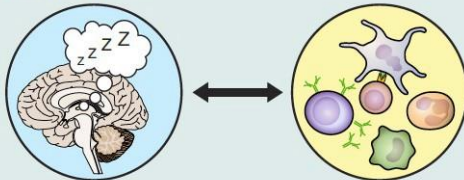
- cytokines (II B)
- prostaglandins (II B)
- LPS/infection (II C)



### Acute (1 night of total or partial SD) and subchronic (several nights of total or partial SD) effects of sleep/SD on:

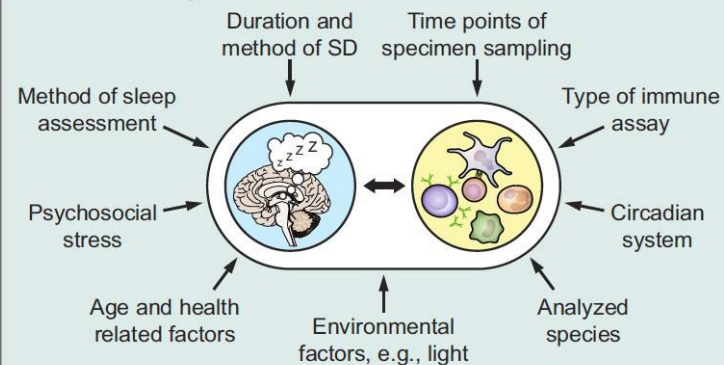
- single immune parameters (III A)
- vaccination response (III B)
- infection outcome and risk (III C)

## Field studies

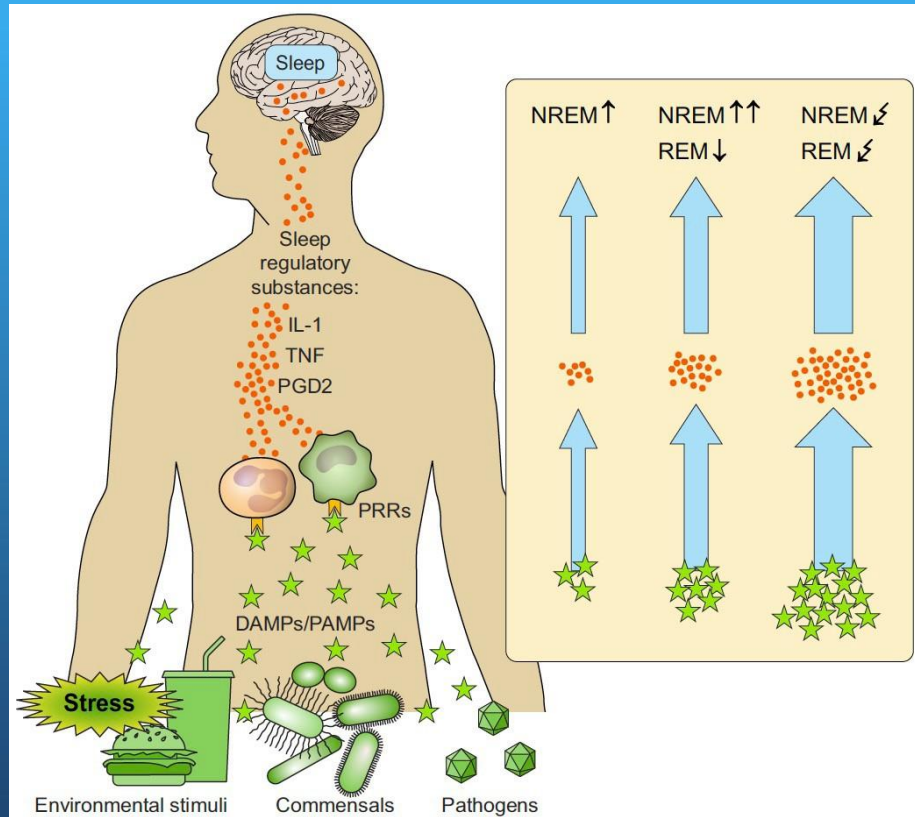


- Sleep changes during chronic immune activation (II D)
- Associations of habitual sleep with vaccination response (III B) and infection risk (III C)
- Immune measures associated with habitual sleep duration (IV A) and chronic sleep disturbances (IV B)

## Influencing factors



# Sleep-Immune Crosstalk in Health and Disease





## Summary:

54

- The role

# Earle A. Chiles Research Institute Robert W. Franz Cancer Center Providence Cancer Institute

