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## Powerful synergistic effects of a Stand agonist and L-2 superkine in eliciting NK and T cell Natalie K. Wolf<sup>1</sup>, Cristina Blaj<sup>1</sup>, Lora Picton<sup>2,3</sup>, Gail Synder<sup>1</sup>, Liw Zhang<sup>1</sup>, Christopher J. Nicolai<sup>1</sup>, Christopher Garcia<sup>2,3</sup> and David Raulet<sup>1</sup> <sup>1</sup>Department of Molecular and Cell Biology, University of California Berkeley, <sup>2</sup>Department of Molecular and Structural Biology, Stanford University, <sup>3</sup>Howard Hughes Medical

### Abstract

Most current cancer immunotherapies are based on mobilizing CD8 T cell responses. However, many types of tumors evade CD8 T cell recognition by displaying few or no antigens, or losing expression of MHC I. These considerations underlie the need for complementary therapies that mobilize other antitumor effector cells, such as NK cells, which preferentially kill MHC I-deficient cells. Cyclic dinucleotides (CDNs) activate the cGAS-STING pathway of the innate immune system and are candidates as immunotherapy agents. Intratumoral CDN injections induce type I IFNs and other mediators that amplify the CD8 T cell response and induce tumor regression (1). CDN therapy also induces long-term tumor regressions in some MHC Ideficient tumor models, mediated primarily by NK cells (2).

To extend the efficacy of CDN therapy, we have combined the IL-2 superkine, H9, or half-life extended H9, with CDNs to target and activate NK cells and T cells in the tumor microenvironment and prevent or delay the onset of NK cell desensitization (3,4). In these studies, we utilized B16-F10 and MC38 tumor cells lacking B2m to examine effects of the combination therapy on MHC I-deficient tumor growth as well as to examine the activation of NK cells by flow cytometry and cytotoxicity assays. We also utilized B16-F10 WT and the spontaneous tumor model, MCA, to assess the effect of the combination therapy on MHC I+ tumors

Here we show that H9 synergized with CDN therapy to mobilize much more powerful antitumor responses against MHC I-deficient tumors than CDN alone. The responses were mediated by NK cells and in some cases CD4 T cells, and were accompanied by increased recruitment to and sustained activation of NK cells in the tumor. This combination therapy regimen activated NK cells systemically, as shown by antitumor effects distant from the site of CDN injection and enhanced cytolytic activity of splenic NK cells against tumor cell targets ex vivo. Finally, the same combination therapy regimen synergistically mobilized powerful CD8 T cell responses in the case of MHC I+ tumor cells, suggesting the generality of the approach. The approach was effective against primary sarcomas, as well, especially when combined with checkpoint therapy, leading to tumor regressions and long-term survival of many mice with MCA-induced sarcoma.

Overall, our work demonstrates the impact of a novel combination therapy in mobilizing powerful NK and T cellmediated antitumor activity, providing important justification for evaluating this approach for treating cancers that are refractory to available treatment options.



CDNs have therapeutic effects with CD8-T cell resistant MHC I-deficient tumors, generated by knocking out B2m. C1498-B2m-/-, (B) MC38-B2m-/- and (C) B16-F10-B2m-/- tumors were established in B6 mice to a size of 50 mm<sup>3</sup> before CDNs were injected intratumorally. Tumors were eliminated (left) or delayed (right) depending on the model

## Engineered IL-2 signals more effectively to NK cells



H9 "superkine" has been engineered to bind IL-2R $\beta$  with high affinity and stimulates NK cells better than WT IL-2 (3). H9 also restores NK-mediated antitumor responses against MHC I-deficient tumors in vivo, in which NK cells become desensitized (4). We employed H9 fused to mouse serum albumin (H9-MSA) to extend the in vivo half life of the cytokine.



Tumors were established with 4 x 10<sup>6</sup> cells implanted s.c. in C57BL/6J mice and grown to approximately 50 mm<sup>3</sup> (day 0). Tumors were injected once i.t. with 50 ug of CDN or PBS. Some mice were also injected i.p. with 10 ug H9-MSA or PBS on day 0 and repeated every three days until euthanization or 1 week after complete tumor clearance. Tumor growth curves and survival of mice with B16-F10-B2m-/- (A) and MC38-B2m-/- (B) tumors. CDN/H9-MSA combination therapy led to dramatic tumor rejection in both hard-to-treat models

Berze Protein

MC38 B/2038-B2m-/-

Institution and <sup>4</sup>Aduro Biotech







![](_page_0_Figure_26.jpeg)

![](_page_0_Figure_28.jpeg)

B16-F10-B2m-/- tumors were established and treated as described above. On d2 (A) and especially on d5 (B) after treatment initiation, flow cytometry analysis of tumors revealed increased GzmB MFI, %GzmB+ NK cells and %CD69+ NK cells in mice treated with CDN/H9-MSA compared to monotherapy. (C) Spleen cells from tumor bearing mice 72 hours after treatment as above with CDN/H9-MSA showed enhanced cytotoxicity of B16-F10-B2m-/- target cells. (D) Cytotoxicity by splenocytes was mediated by NK cells, as shown by depleting mice of NK cells prior to treatment.

25 50 100 200

Spleen cell:Target Ratio

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