

The background of the slide is a histological image showing a dense population of cells, likely lymphocytes, stained with hematoxylin and eosin (H&E). The cells are small with dark purple nuclei and light pink cytoplasm/extracellular matrix. The overall texture is granular and somewhat disorganized, typical of a tissue section.

***Interleukin-1 role in human Th-17
cell responses, dendritic cell
activation, and epithelial cell
transformation***

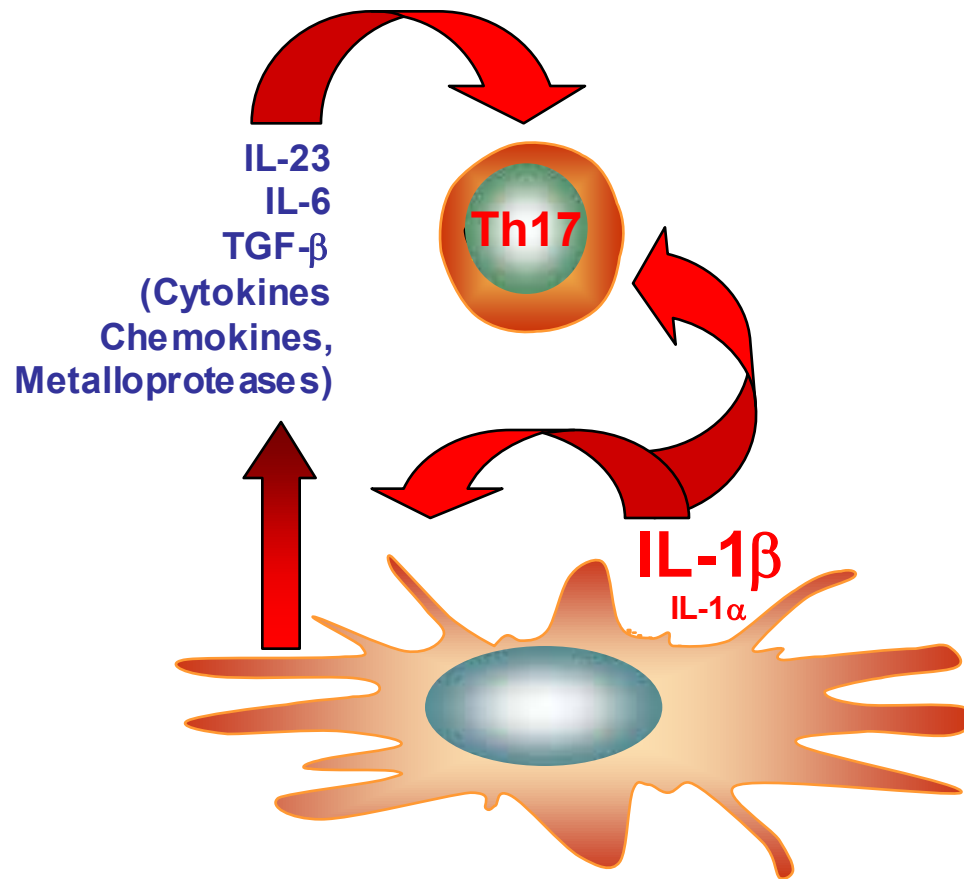
Giorgio Trinchieri

Cancer and Inflammation Program

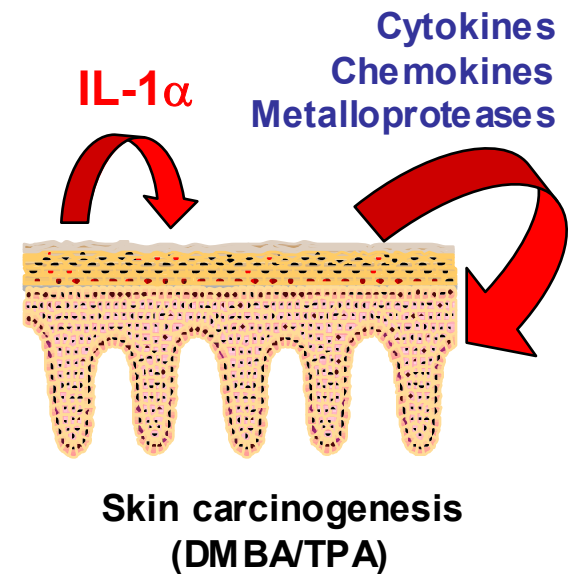
CCR, NCI

Frederick, MD, U.S.A.

BACK TO THE FUTURE: several new important biological roles of the Interleukin-1 cytokine family emerge from studies of inflammation-dependent carcinogenesis



Human Dendritic Cells
Activation by β -Glucan



Activation of human Dendritic Cells by ITAM-signaling receptors

IL-1 β



Human
Monocyte-derived
Dendritic Cells



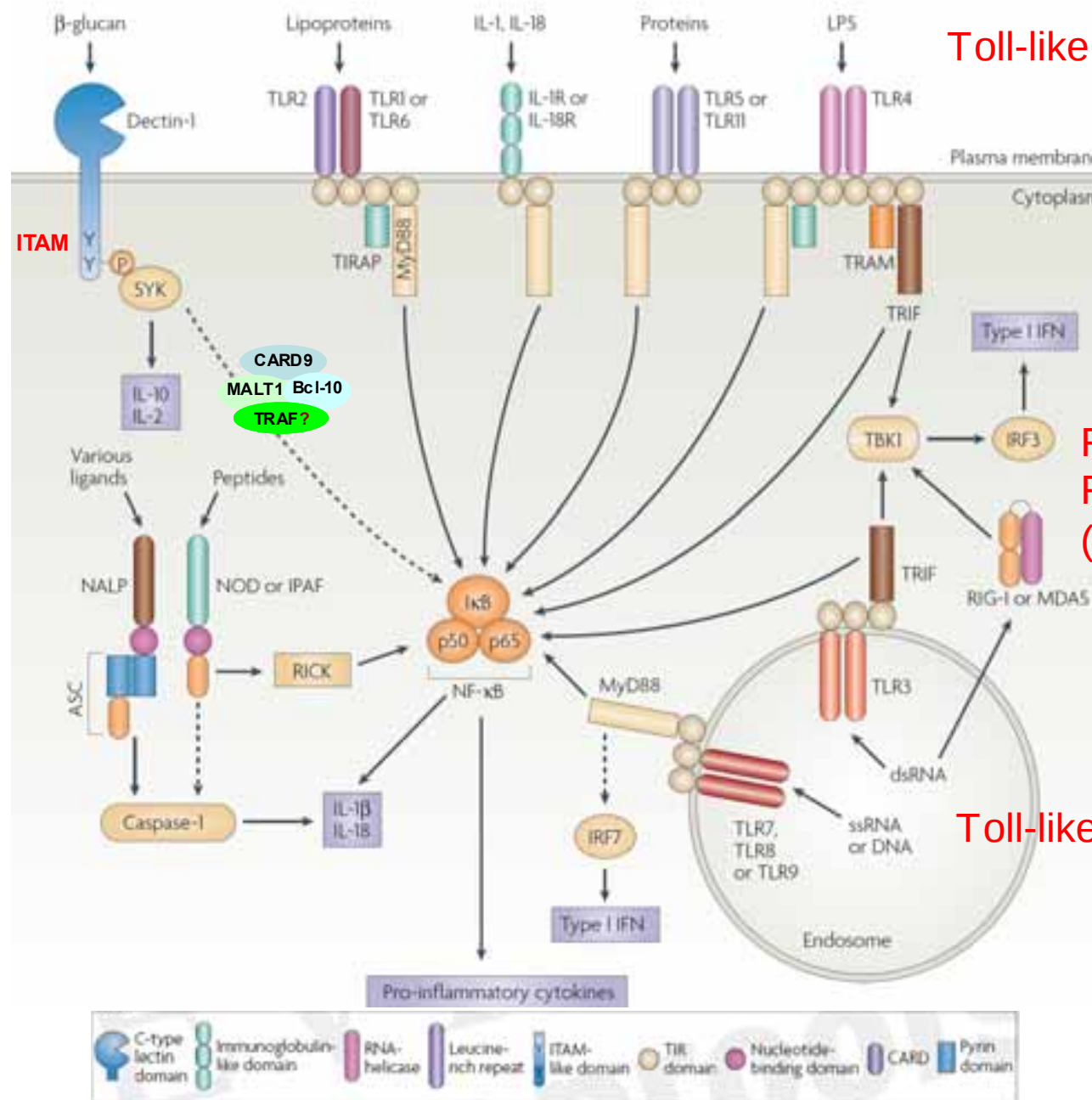
Pattern Recognition Receptors (PRR)

C-type lectins,
Scavenger r.

Toll-like receptors

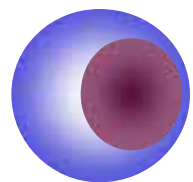
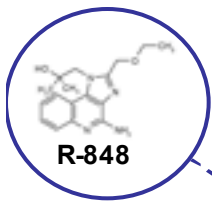
NOD-like
Receptors:

NALPs NODs

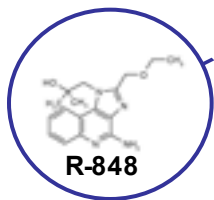


RIG-I-like
Receptors
(helicases)

Toll-like receptors

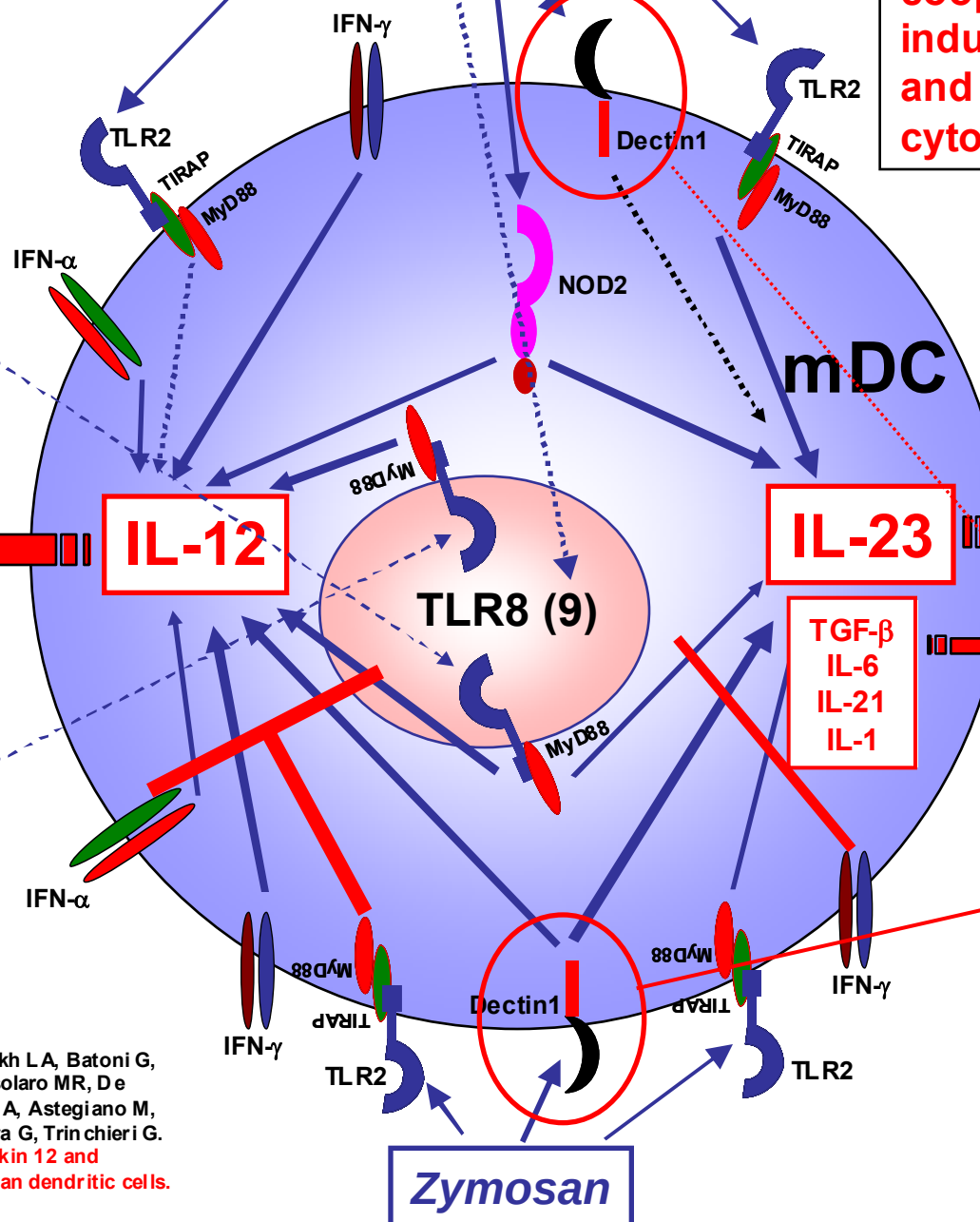


Th1

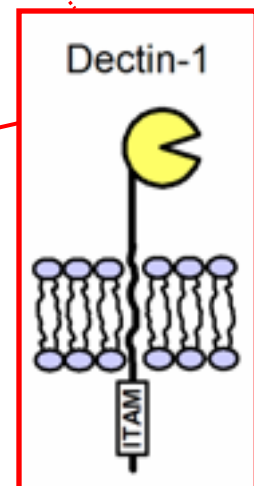


Gerosa F, Baldani-Guerra B, Lyakh LA, Batoni G, Esin S, Winkler-Pickett RT, Consolaro MR, De Marchi M, Giachino D, Robbiano A, Astegiano M, Sambataro A, Kastelein RA, Carra G, Trinchieri G. Differential regulation of interleukin 12 and interleukin 23 production in human dendritic cells. J Exp Med. 2008;205: 1447-61.

M. tuberculosis

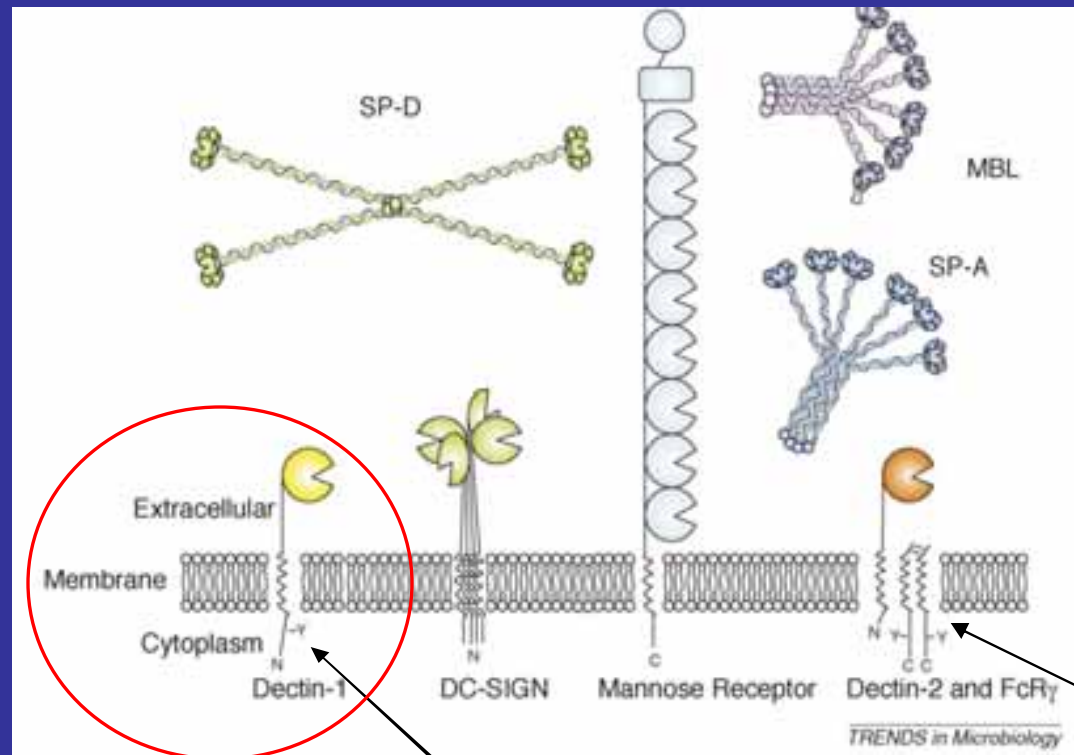
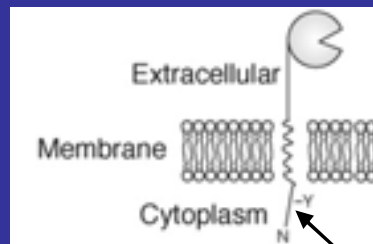


Signals from innate receptors and cytokine receptors cooperate and antagonize in inducing pro-inflammatory and immunoregulatory cytokines.



Mouse CD8 α + DC
Human BDCA3+ DC

DNGR-1



Emi-ITAM

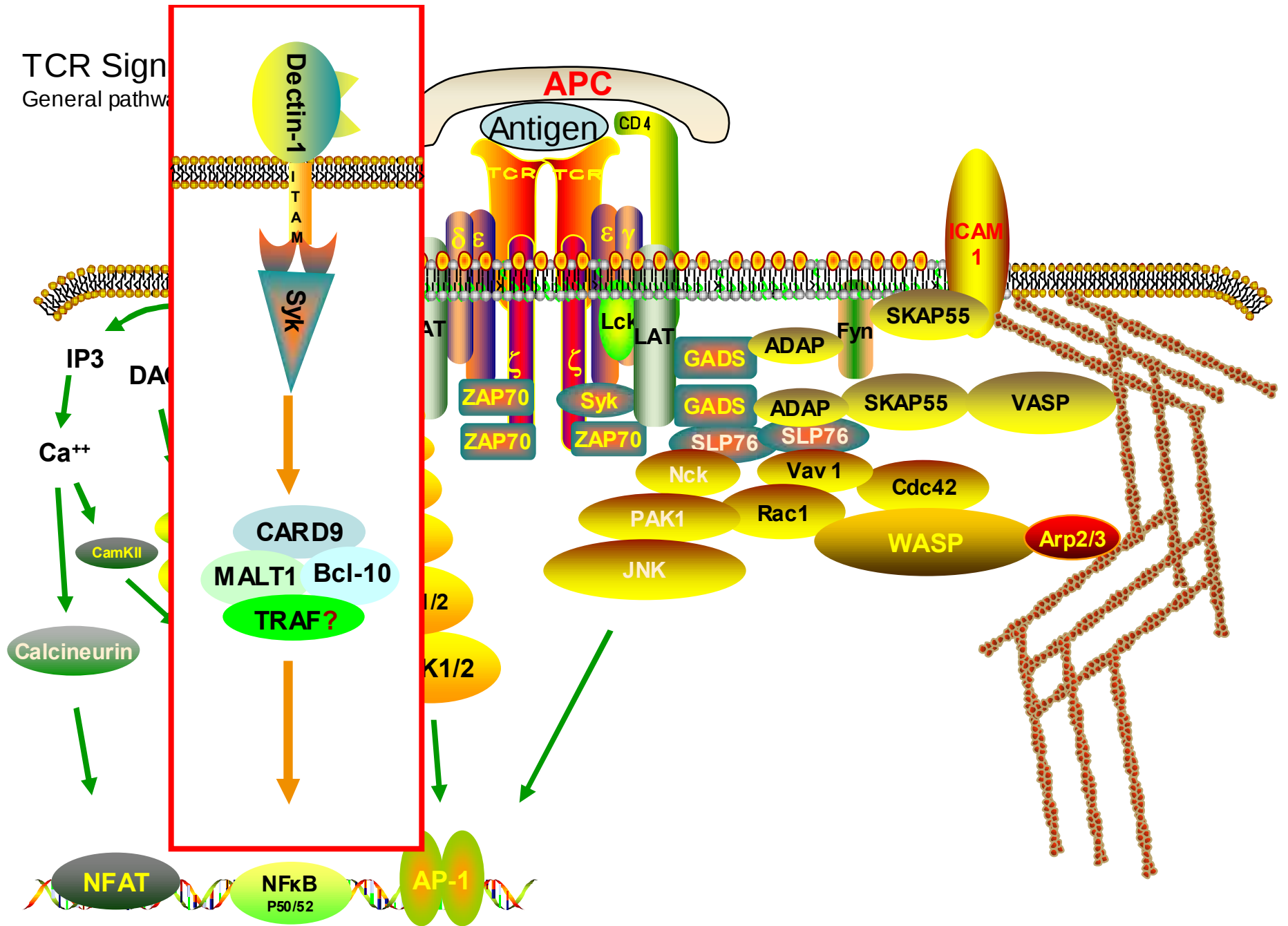
Emi-ITAM

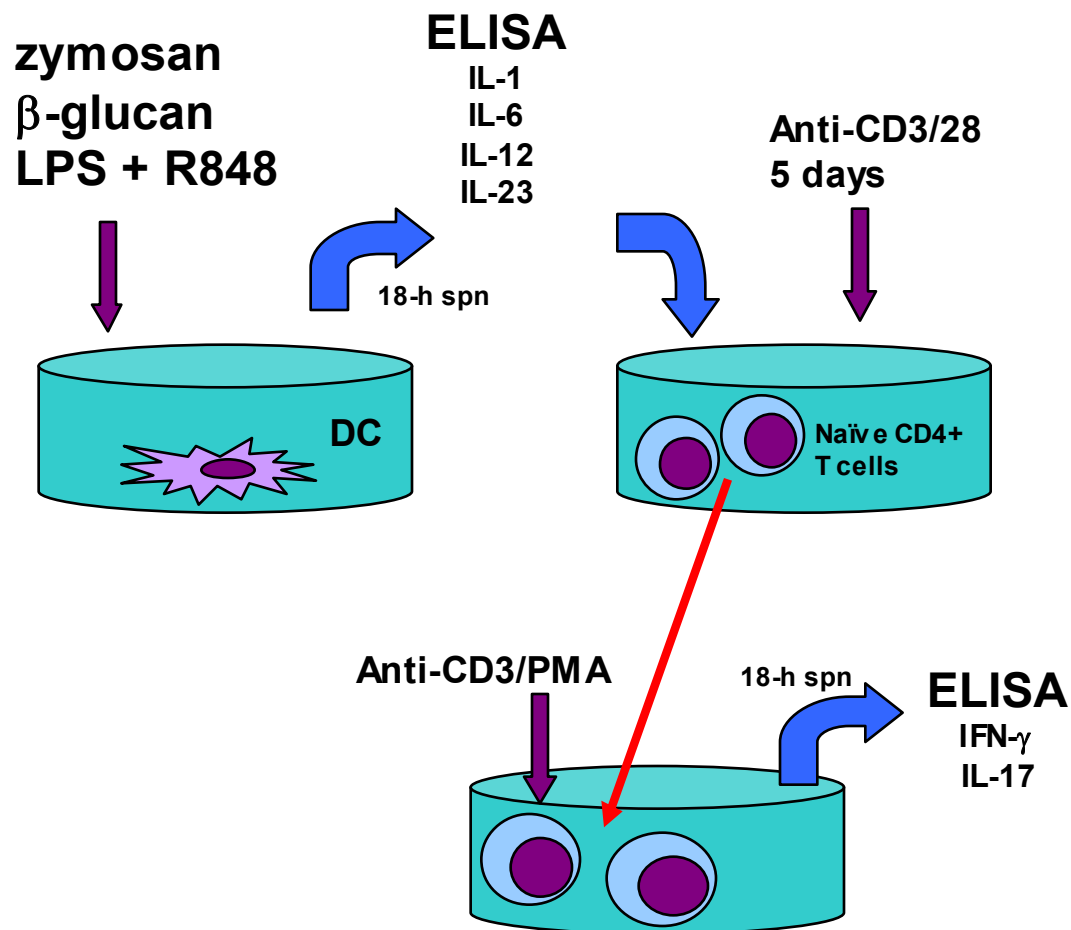
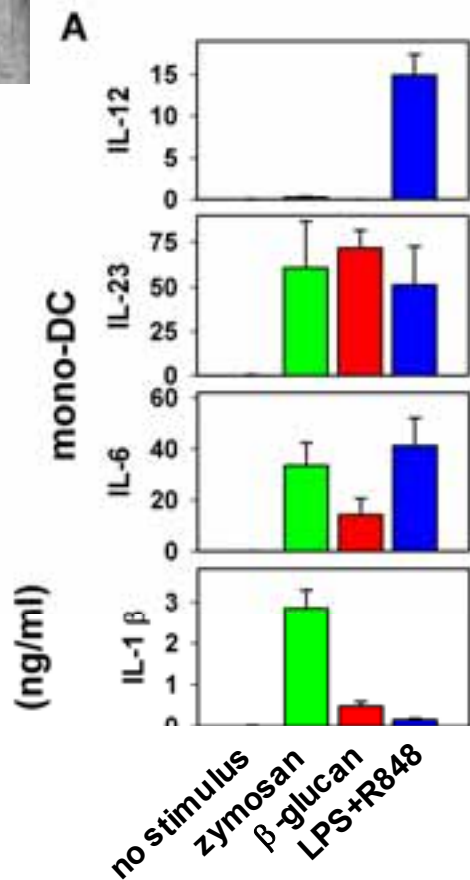
ITAM

Table 1. Fungal PAMPs and their receptors

PAMP	PRR(s)
β -1,2 mannosides	Galectin-3
β -glucan	Dectin-1, SP-D, lactosylceramide
Chitin	Unknown
Phospholipomannan	TLR2
Glucuronoxylomannan	CD14, TLR4
Mannan	TLR4; SP-A; SP-D, MR, DC-SIGN, Dectin-2, MBL
Galactomannan	PTX3 (pentraxin-3)
DNA	TLR9

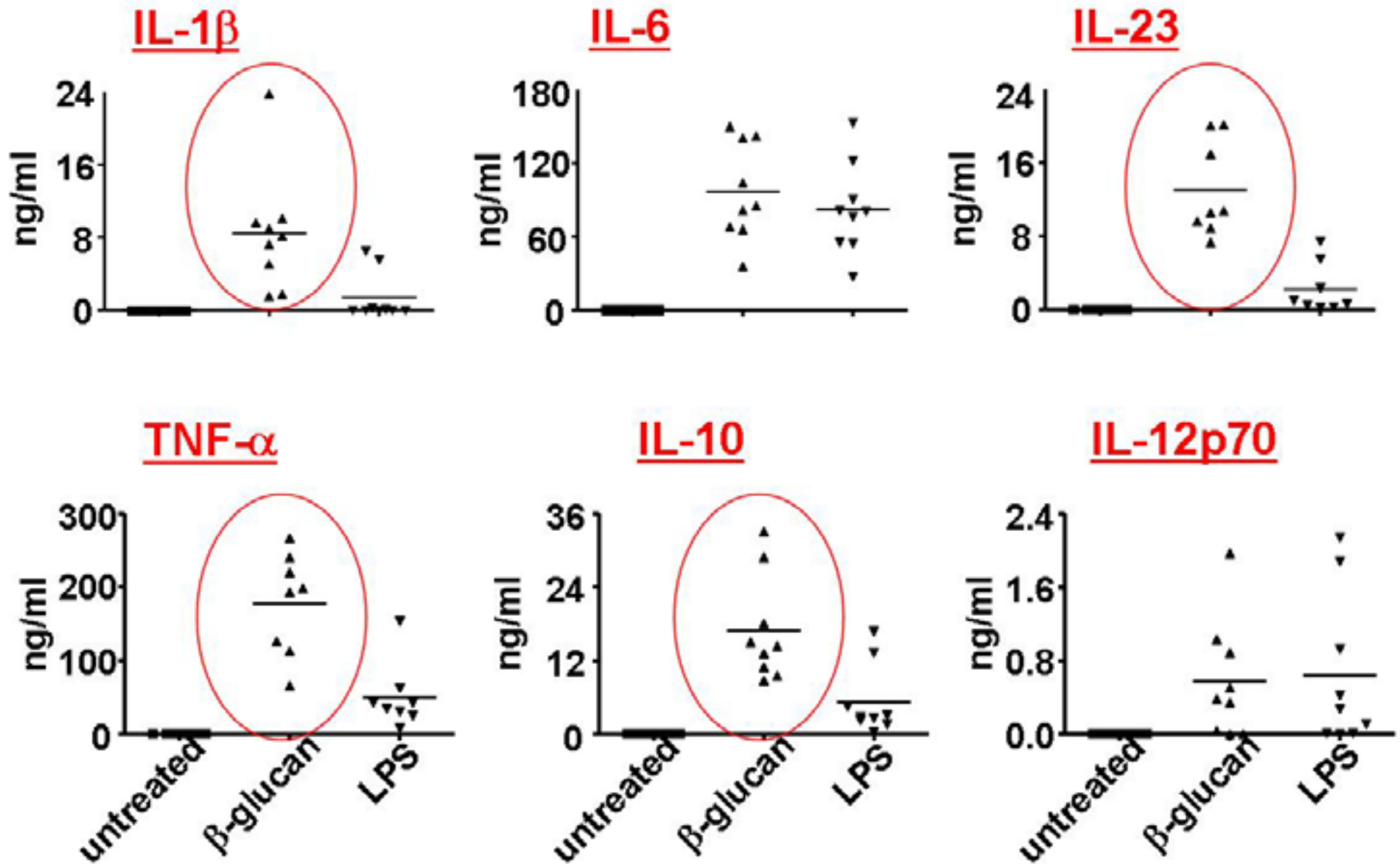
TCR Signaling General pathway





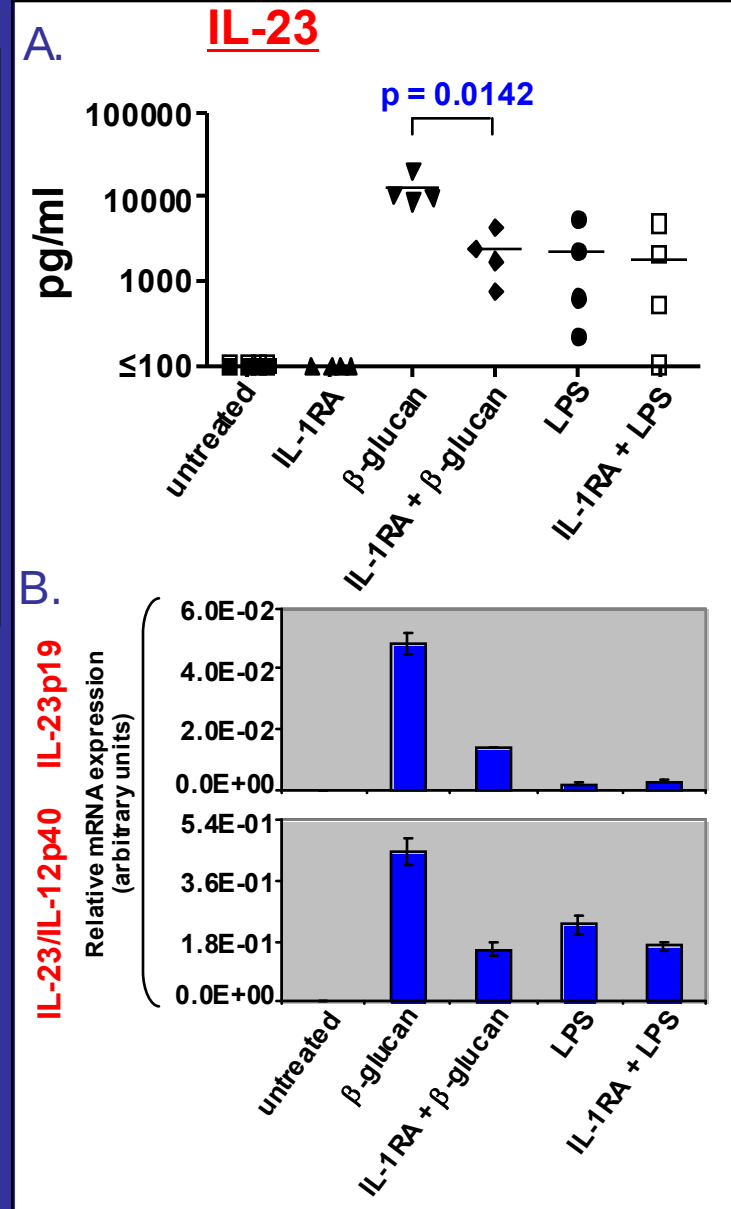
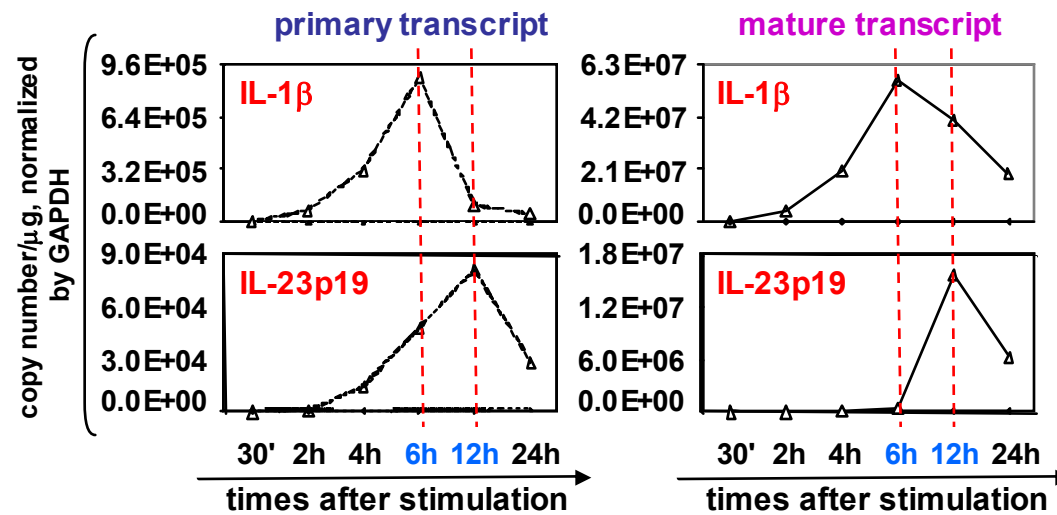


Cytokine profile induced by β -glucan and LPS in human monocyte-derived DCs





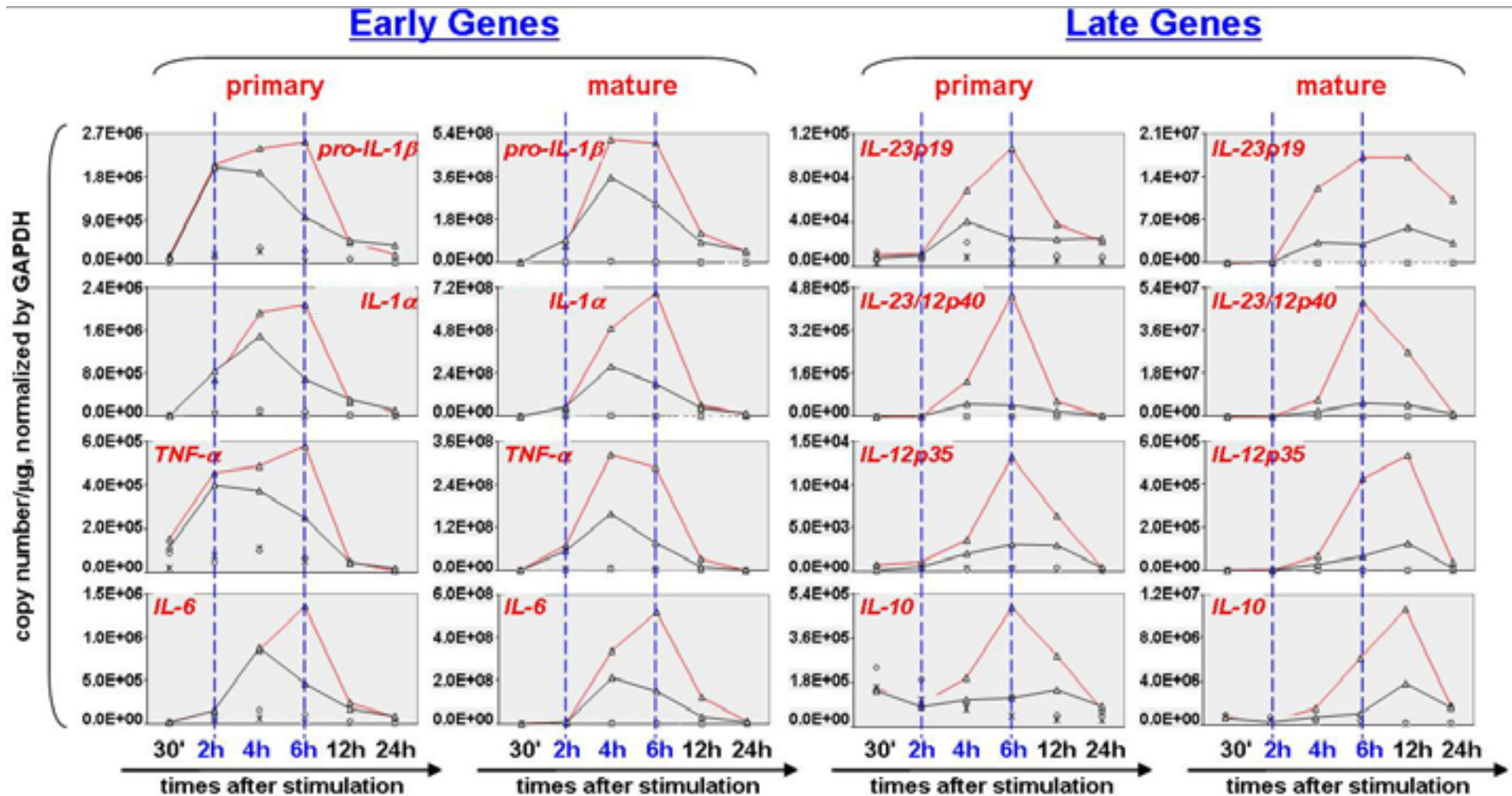
Role of endogenous IL-1 in IL-23 production in human monocyte-derived DCs activated by β -glucan



✓ In the presence of the IL-1RA the ability of β -glucan treatment to induce IL-23 protein production (A) and IL-23 p19 and IL-12 p40 mRNA expression (B) in human mono-DCs is decreased.



Role of endogenous IL-1 in the regulation of the late gene expression induced by β -glucan in human mono-DCs

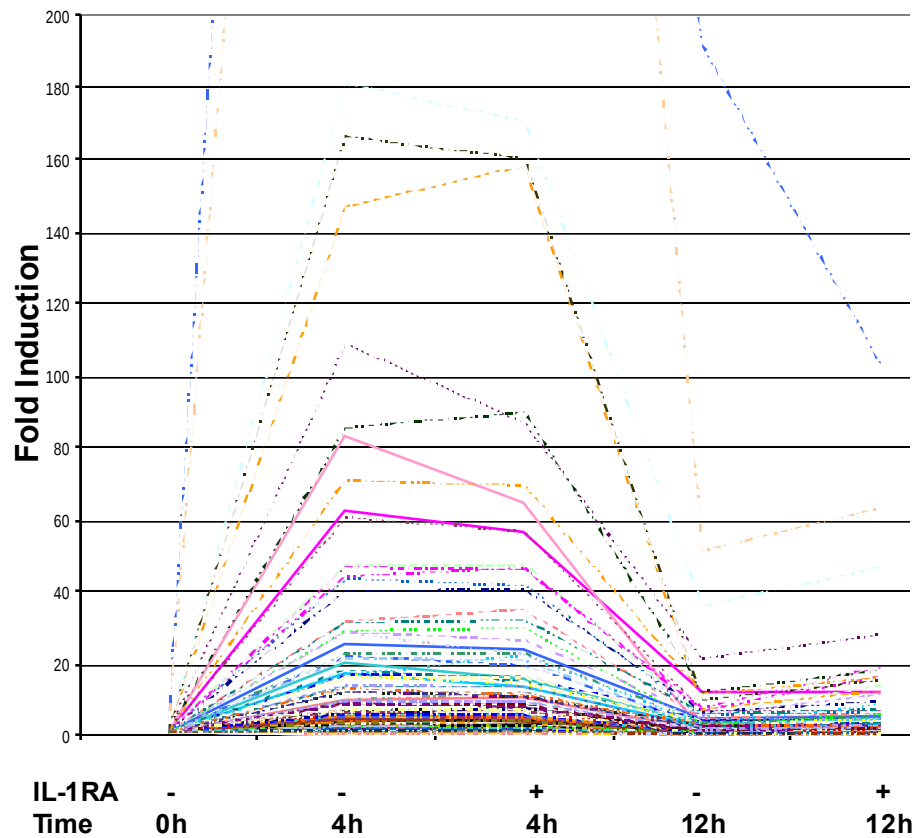


···x··· untreated $\blacktriangle$ β -glucan
 -◆- IL-1RA 2.5 $\triangle$ IL-1RA 2.5 + β -glucan

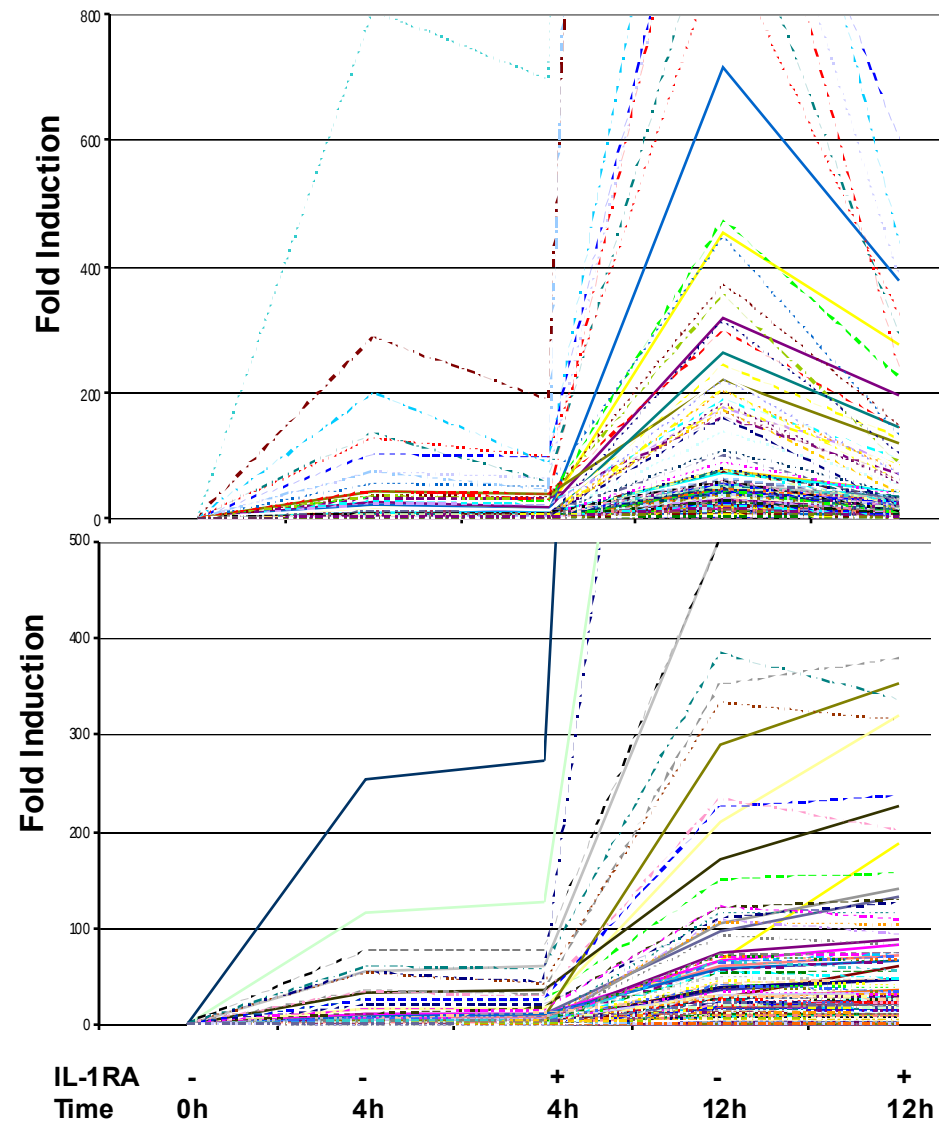


Microarray analysis of the transcripts induced by b-glucan in human mono-DCs in the presence or not of IL-1RA

“Early genes”

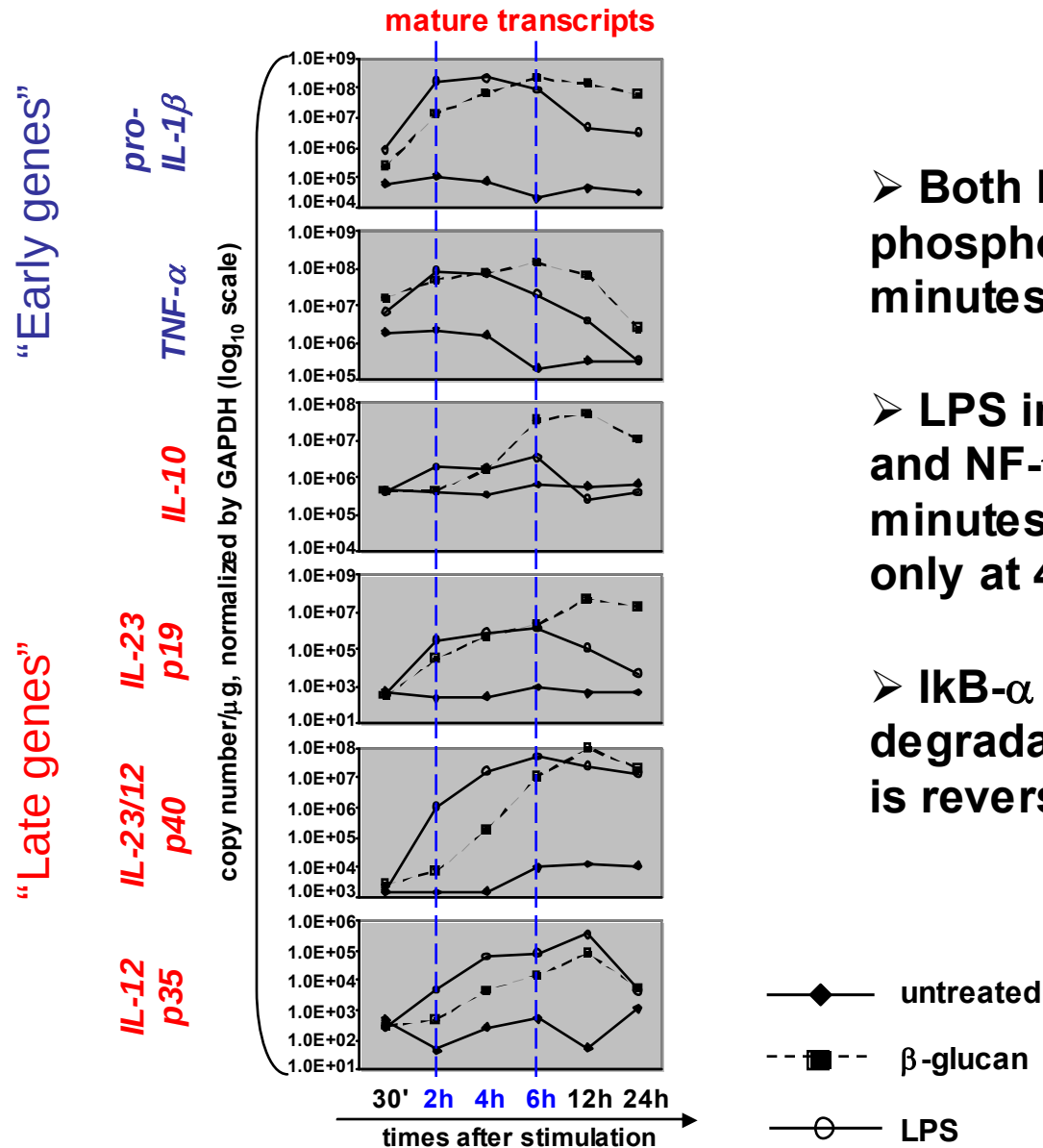


“Late genes”





LPS induces “late genes” expression with a faster kinetic than β -glucan in human mono-DCs



➤ Both LPS and β -Glucan induce phosphorylation of p38 and ERK1/2 at 30 minutes or earlier.

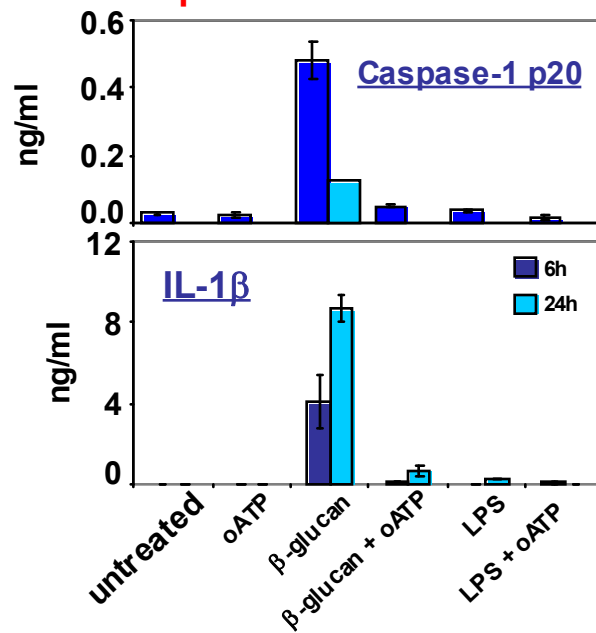
➤ LPS induces phosphorylation of I κ B- α and NF- κ B activation already at 30 minutes whereas β -Glucan induces them only at 4-6 hours.

➤ I κ B- α phosphorylation and degradation by β -glucan but not by LPS is reversed by Interleukin-1RA.

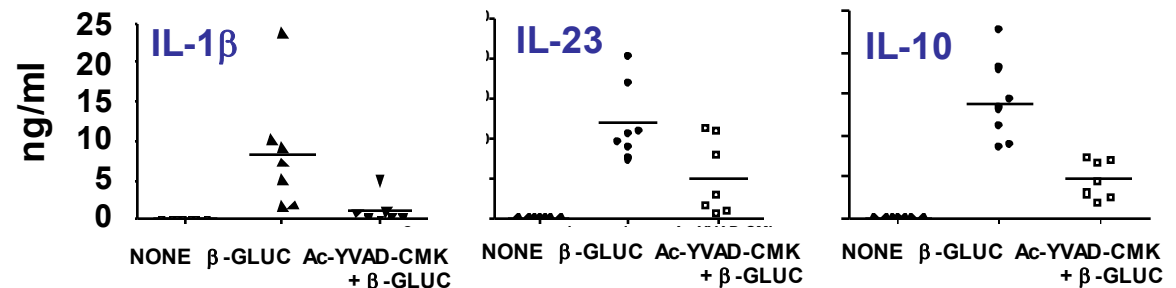


The expression of “Late genes” in human DCs stimulated by β -Glucan is dependent on IL-1 β and, to a lesser extent, IL-1 α

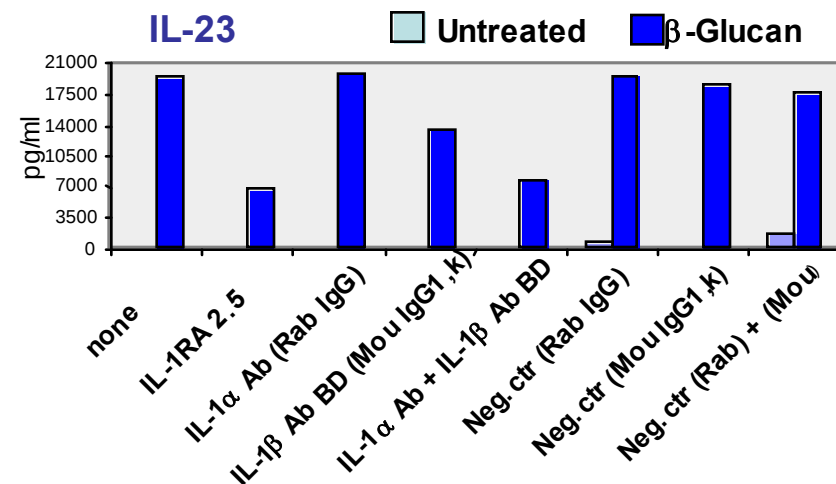
β -glucan but not LPS activates Caspase-1 in mono-DC



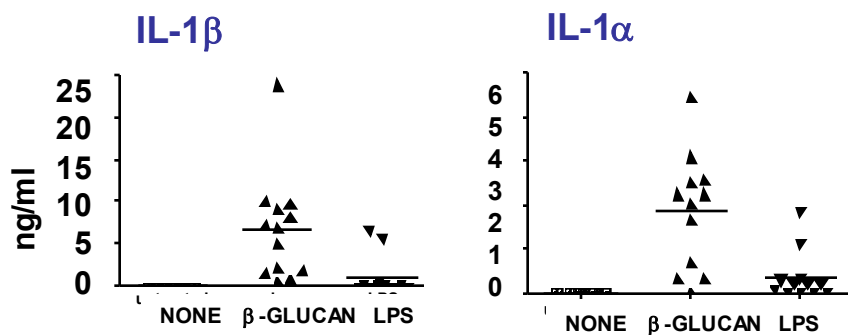
Caspase-1 inhibitors decrease IL-1 β and also IL-23 and IL-10 secretion by β -glucan-activated mono-DC

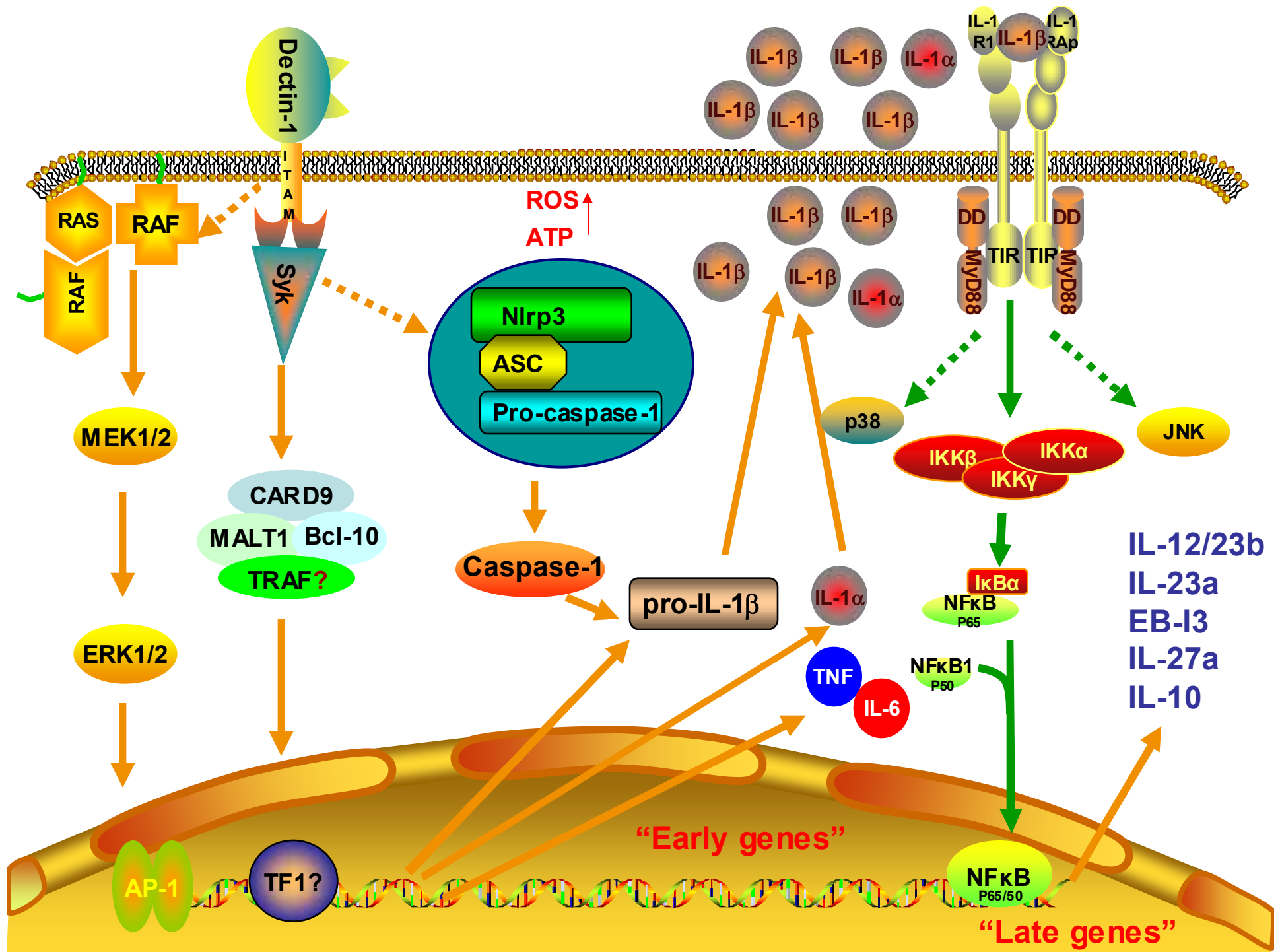


Anti-IL-1 β is more effective than anti-IL-1 α to inhibit IL-23 production by β -glucan-activated mono-DC



Production of IL-1 β and IL-1 α by β -glucan-stimulated mono-DC

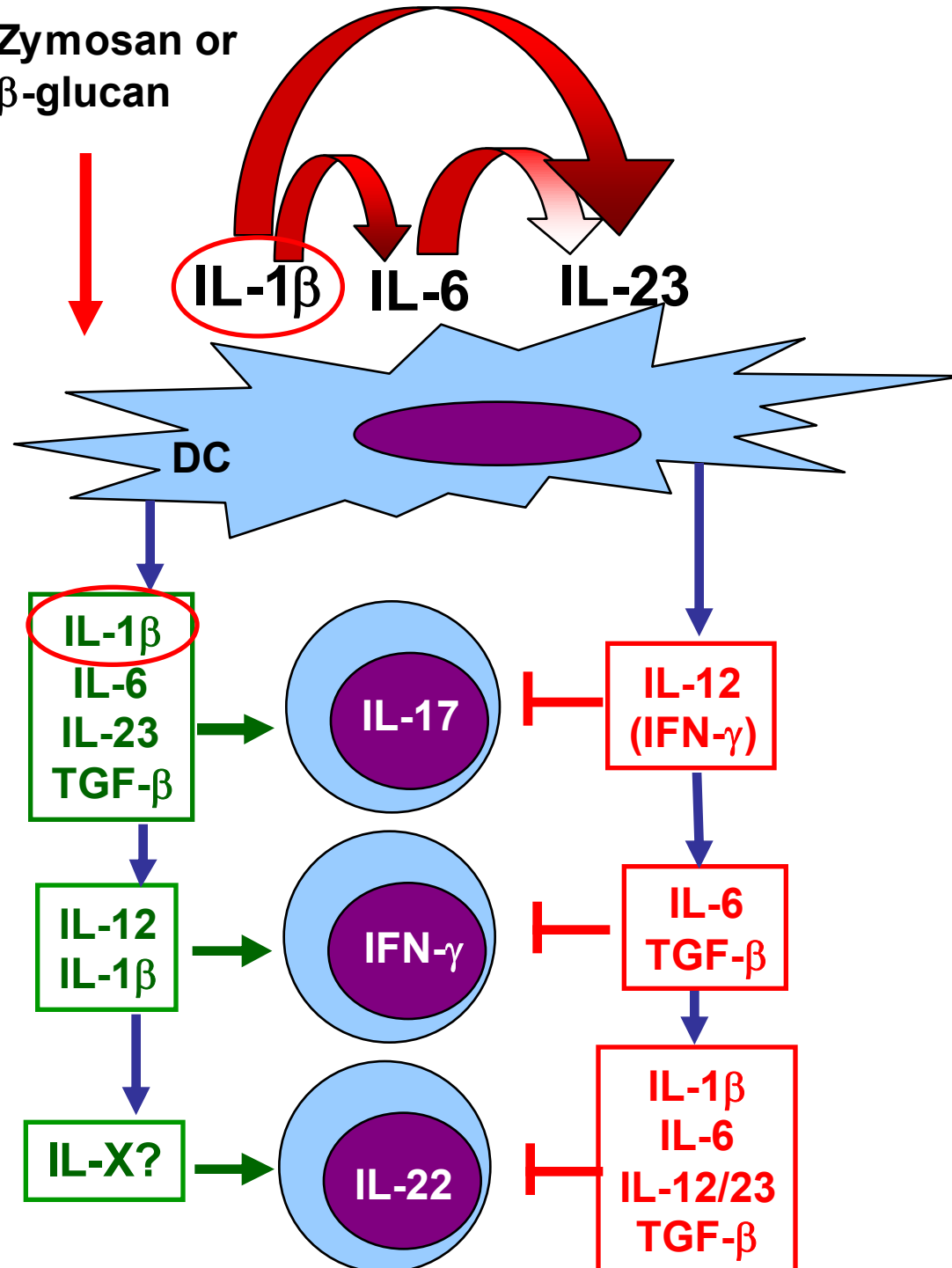




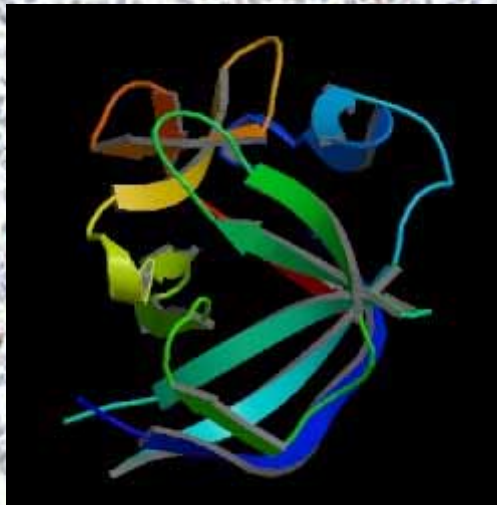
Zymosan or
 β -glucan

IL-1 β has a central role in the Th17 response by being involved in an autocrine way to increase production of the Th-17 inducing cytokine IL-6 and IL-23 from DC and by directly acting on the CD4⁺ T cells in the induction of IL-17 production.

→ Induction
┐ Inhibition



Role of IL-1 α in skin carcinogenesis





MyD88 is required for DMBA-induced Mouse Skin Tumorigenesis

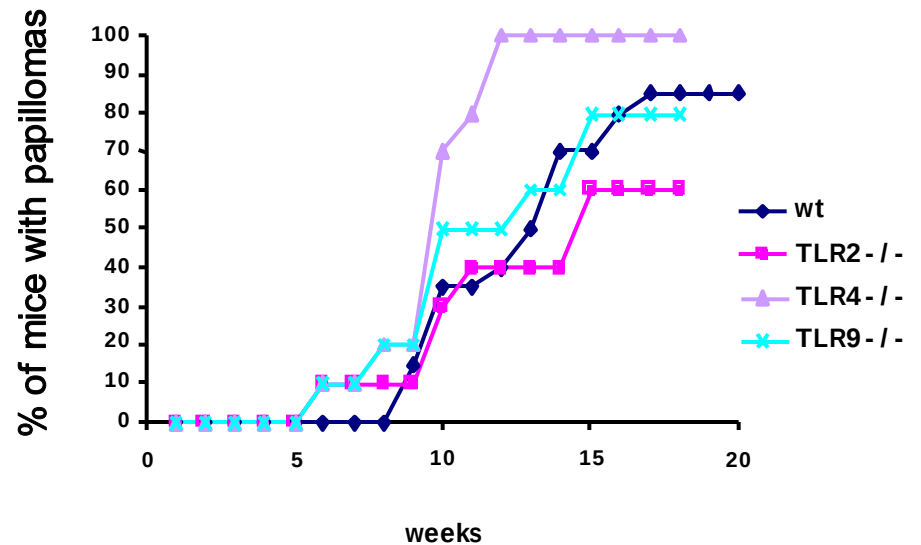
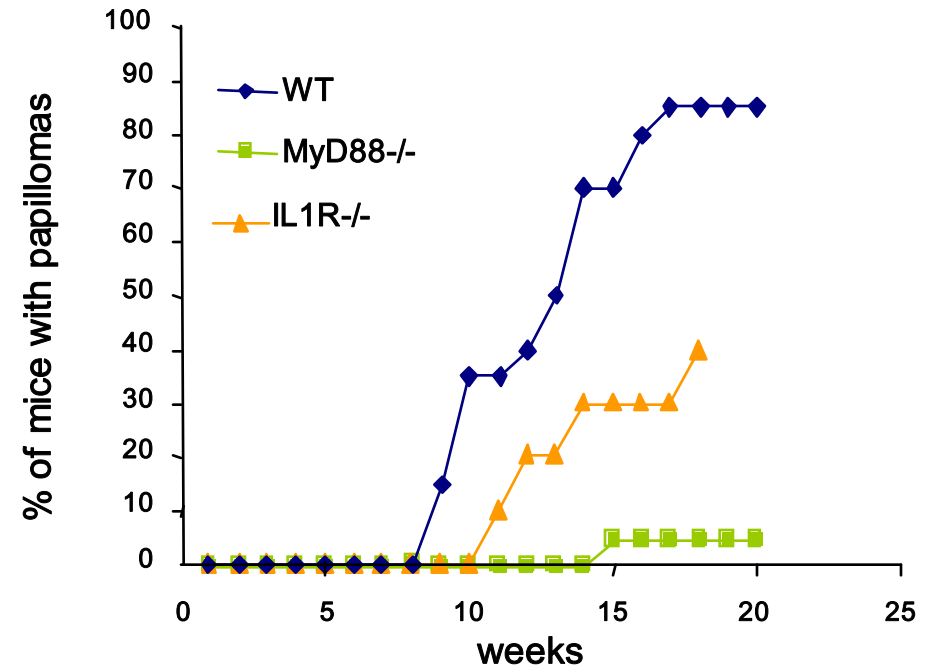
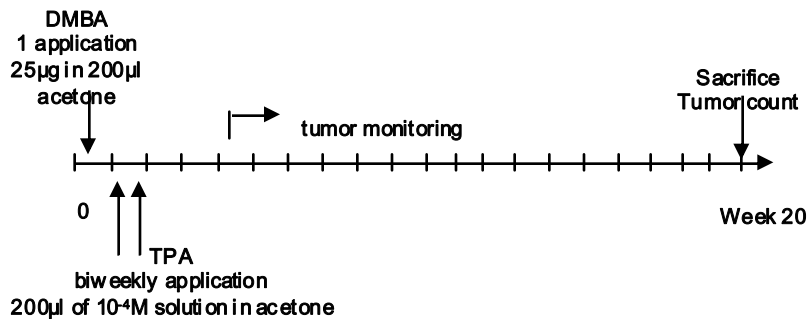
(19) **United States**

(12) **Patent Application Publication** (10) Pub. No.: US 2006/0141490 A1
Jun. 29, 2006

(54) **MYD88 AS A THERAPEUTIC TARGET FOR CANCER**

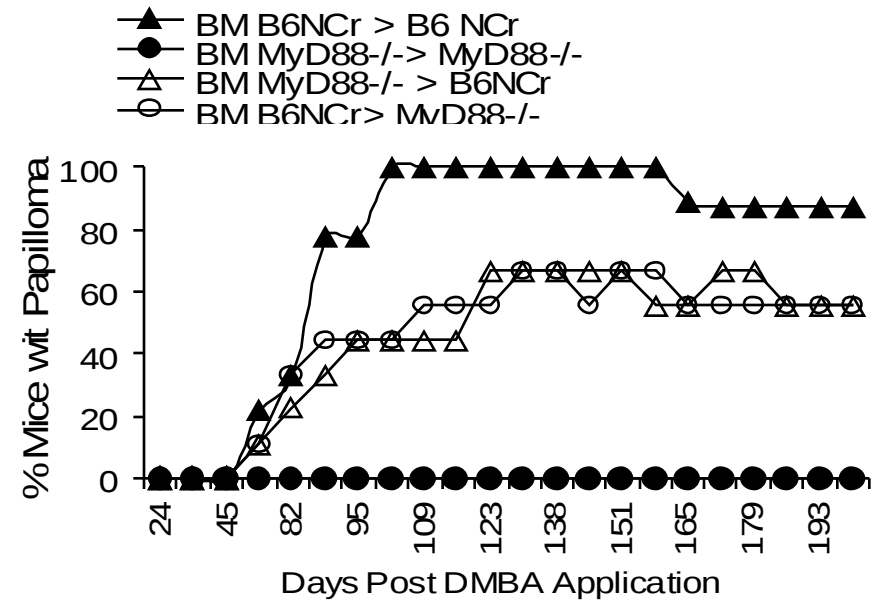
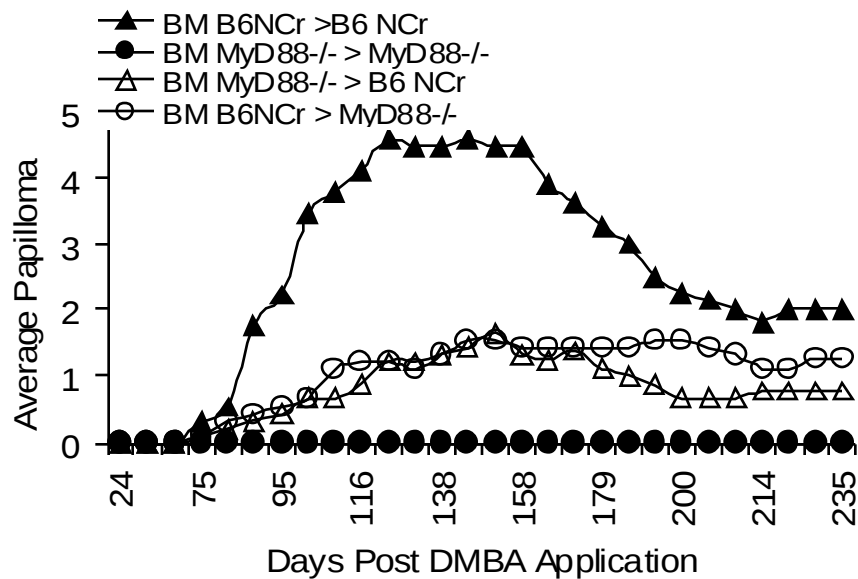
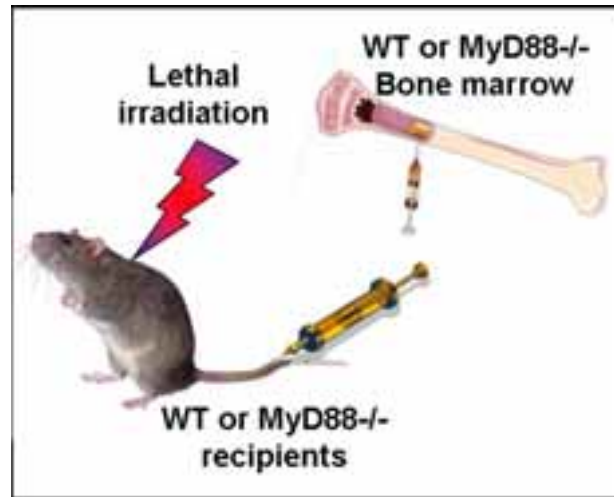
(76) Inventors: **Isabelle Coste-Invernizzi**, Chazay d'Azergues (FR); **Toufic Renno**, Civrieux d'Azergues (FR)

Correspondence Address:
SCHERING-PLOUGH CORPORATION



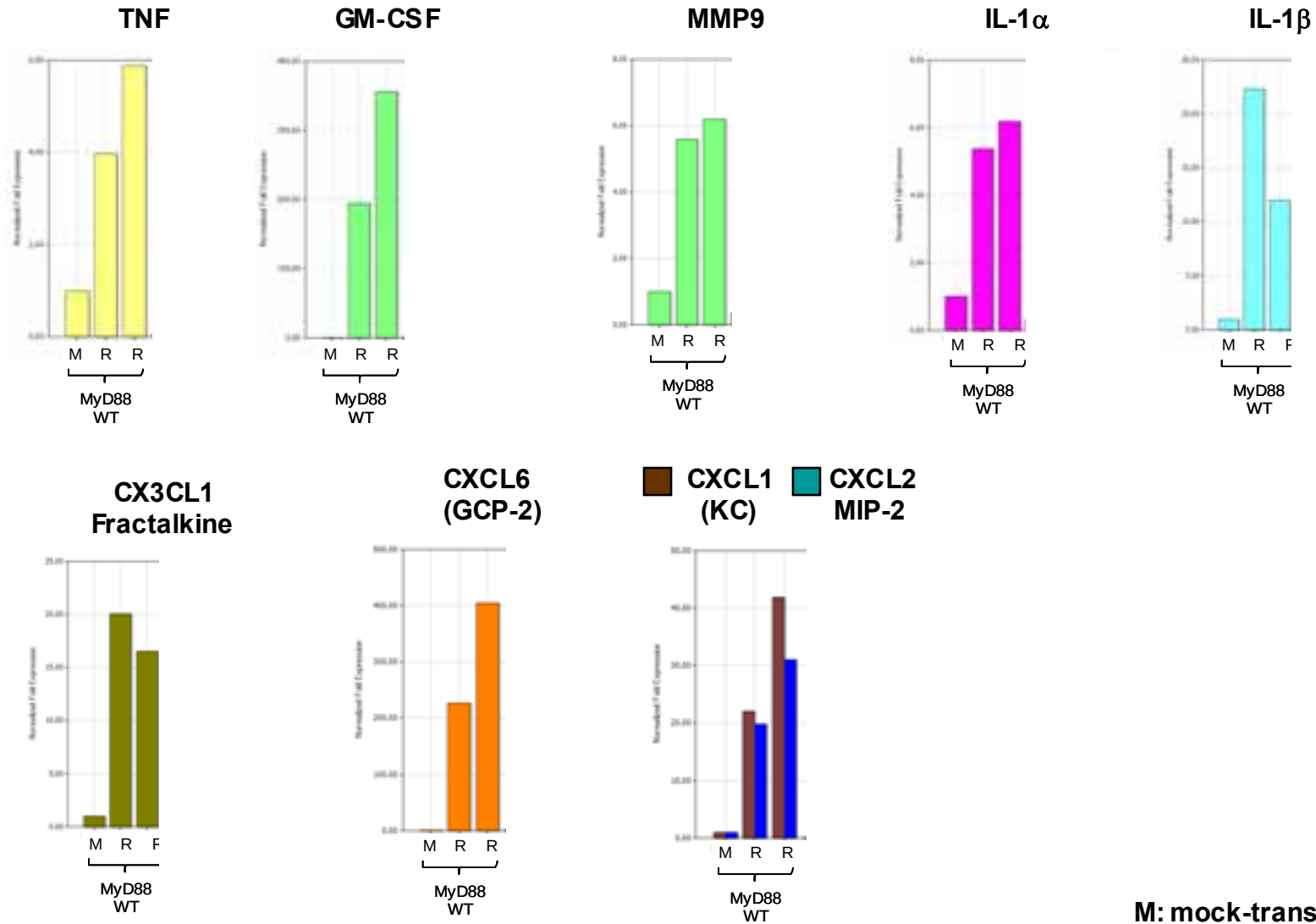


Bone marrow chimera experiments indicate that optimal skin carcinogenesis requires MyD88 expression both in hematopoietic cells and in recipient cells



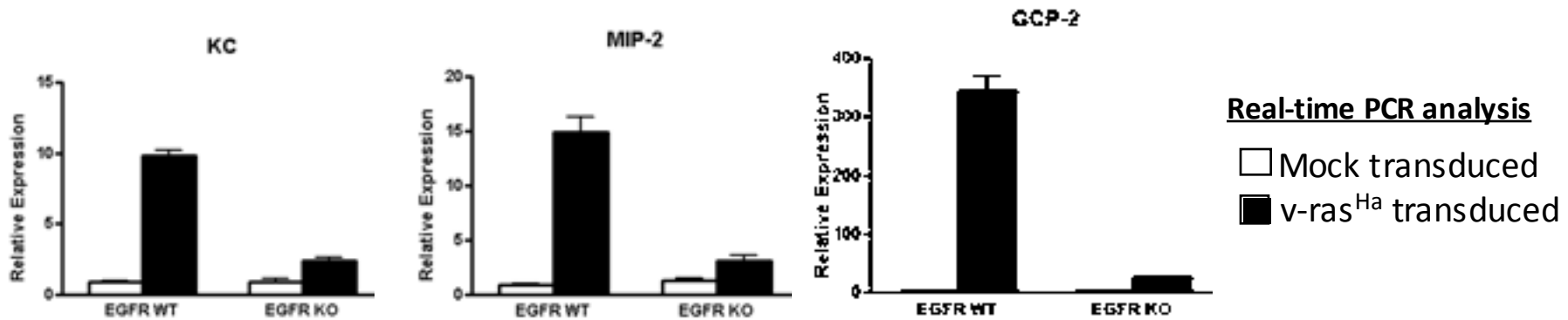


v-ras^{Ha} transformed keratinocytes from wild type mice produce CXCR2 ligands and other cytokines

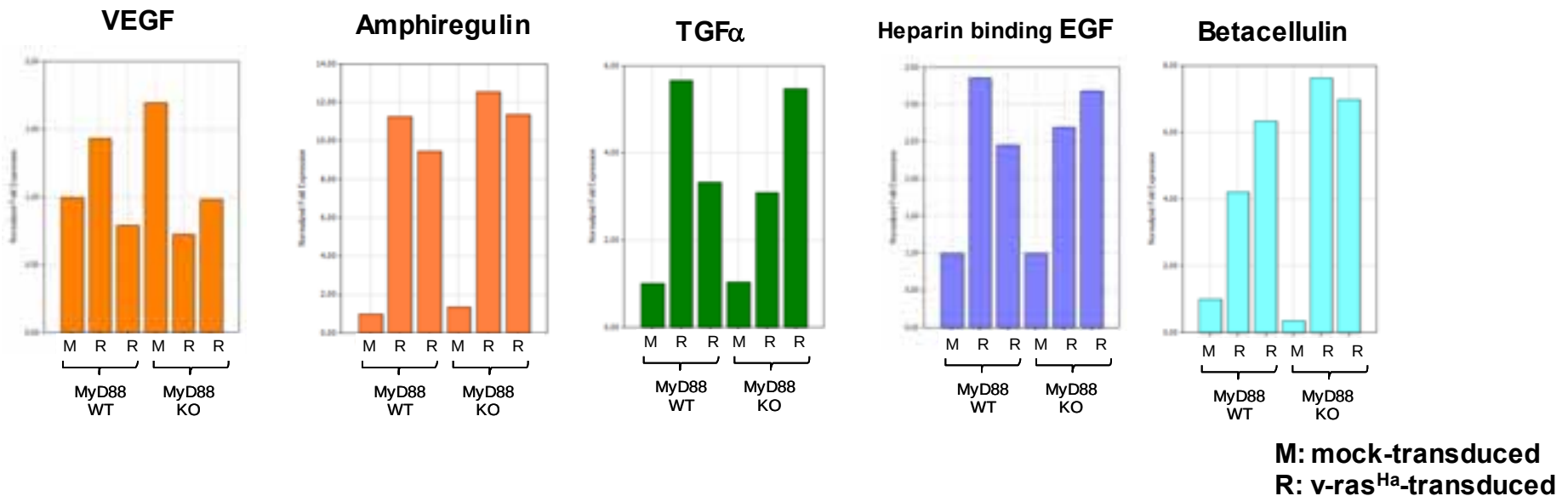




Upregulation of CXCR2 ligands by oncogenic v-ras^{Ha} in primary keratinocytes requires EGFR expression

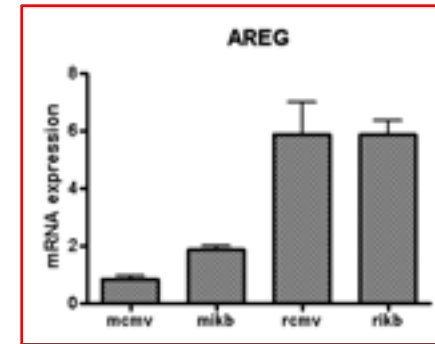
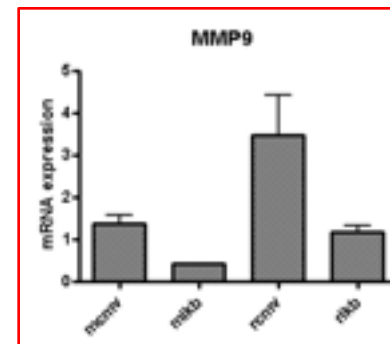
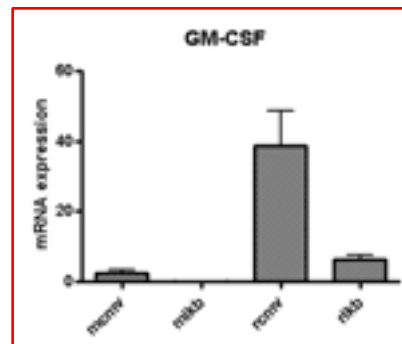
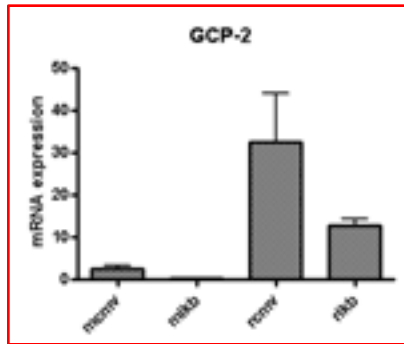


v-ras^{Ha} transformed keratinocytes from MyD88^{-/-} mice are able to produce EGFR ligands



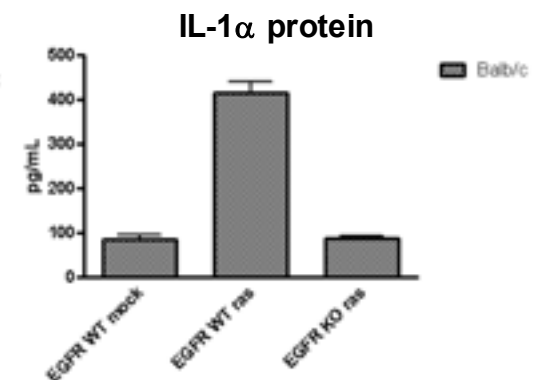
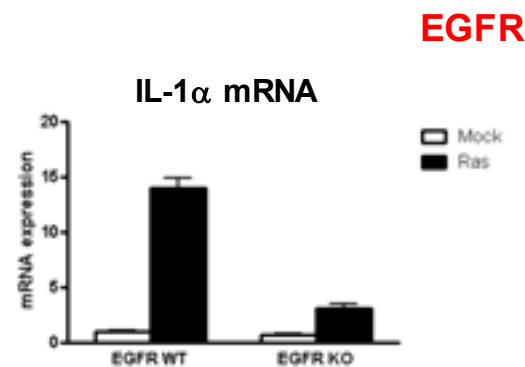
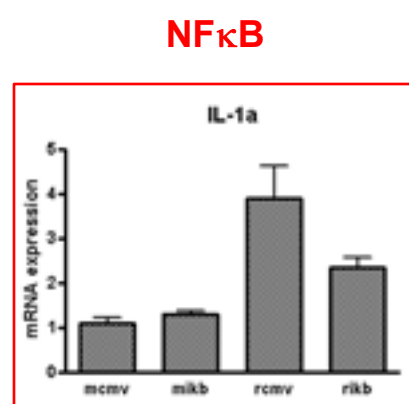
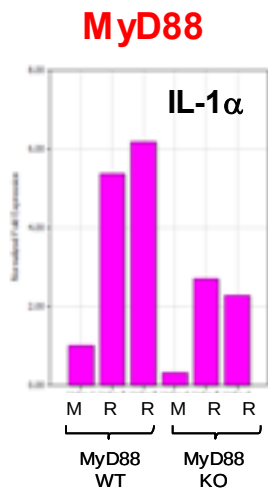


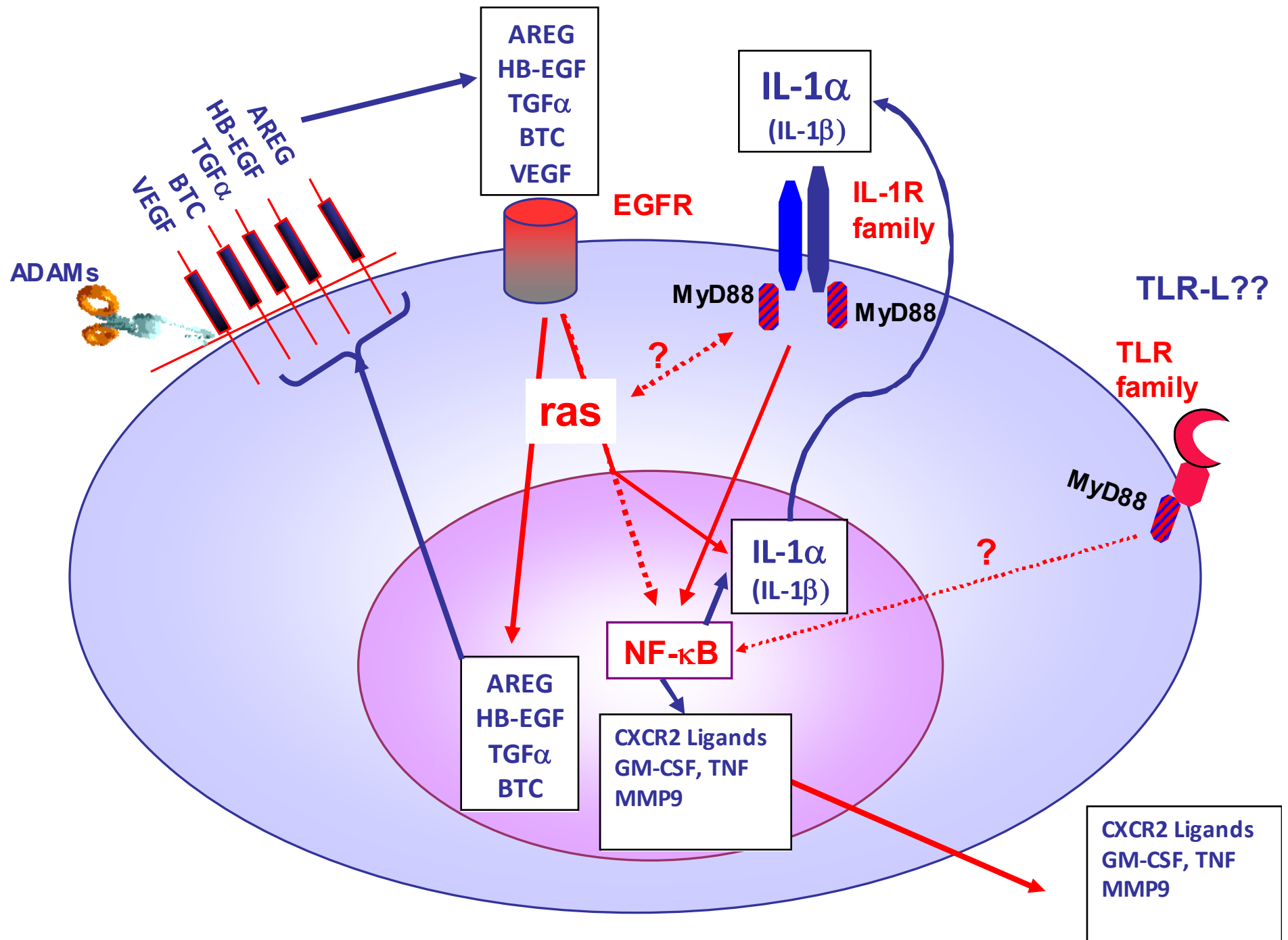
Ras target genes affected by MyD88 deficiency are NF- κ B regulated

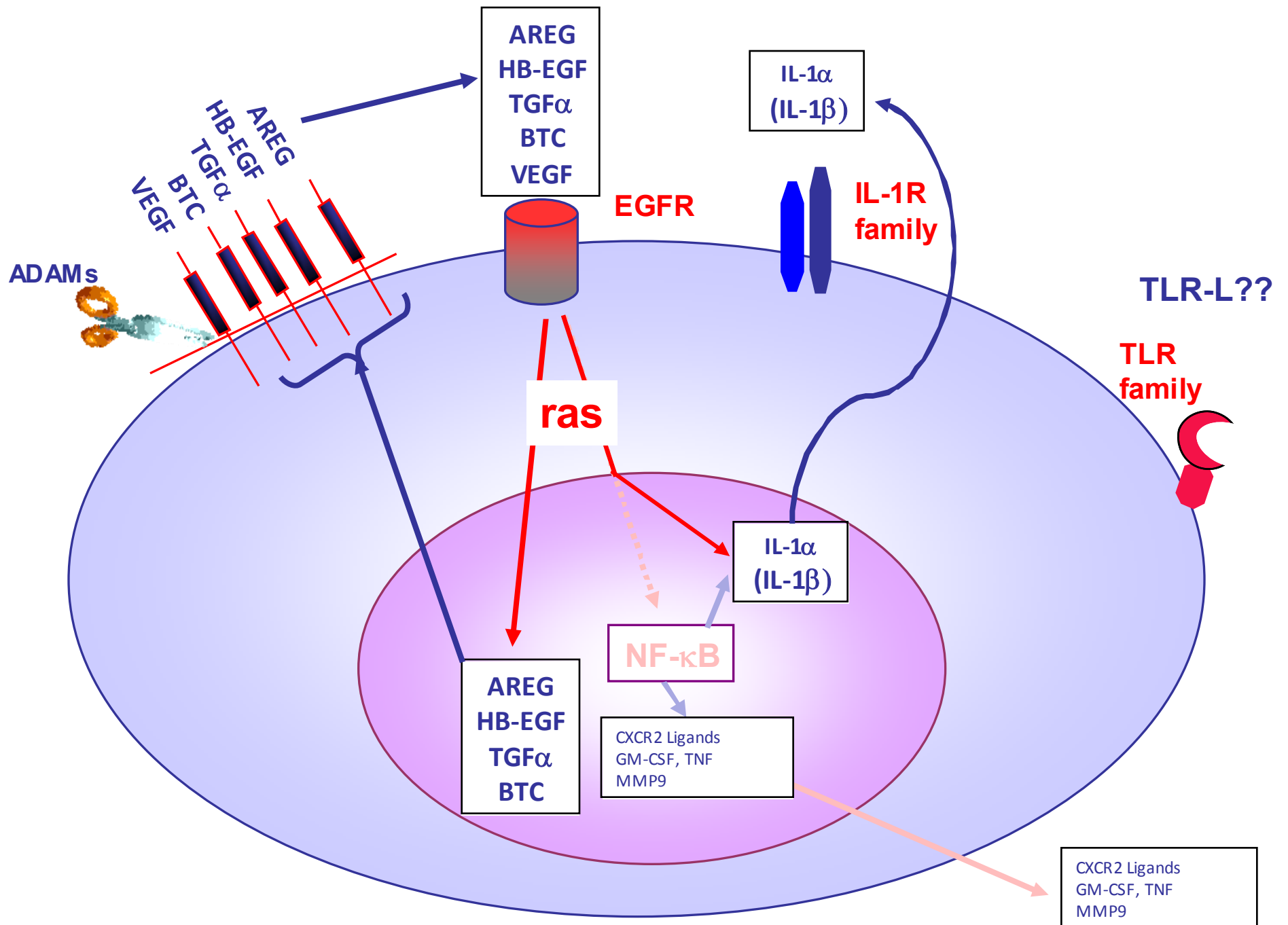


mcmv: mock, control Ad; **mikb**: mock, I κ B α SR Ad
rcmv: v-ras, control Ad; **rikb**: v-ras, I κ B α SR Ad

Ras induction of IL-1 α is EGFR-dependent but only partially MyD88 and NF κ B dependent

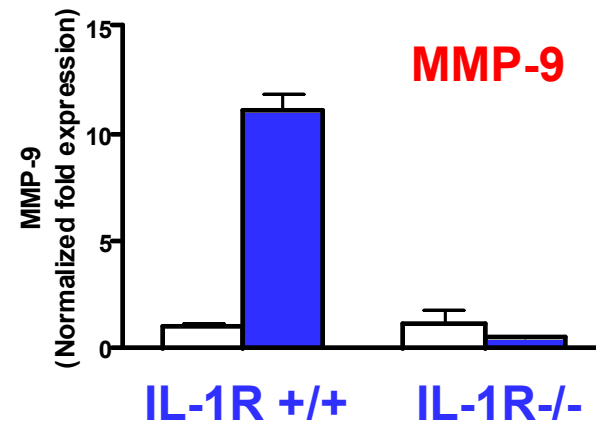
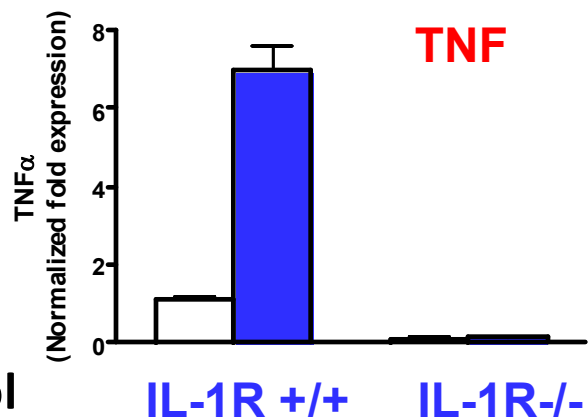
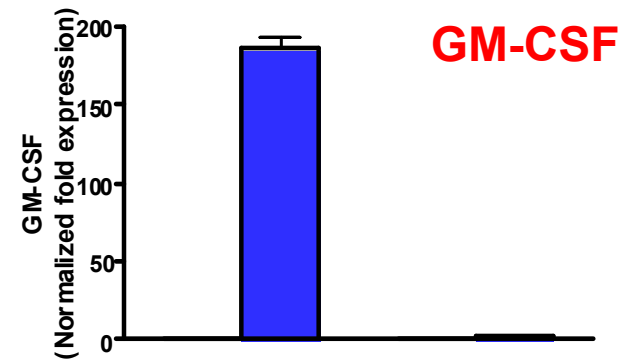
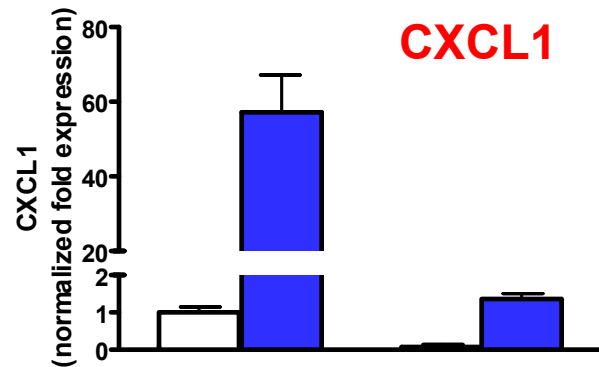








$v\text{-ras}^{\text{Ha}}$ transformed keratinocytes from IL-1R^{-/-} mice are defective in production of CXCR2 ligands

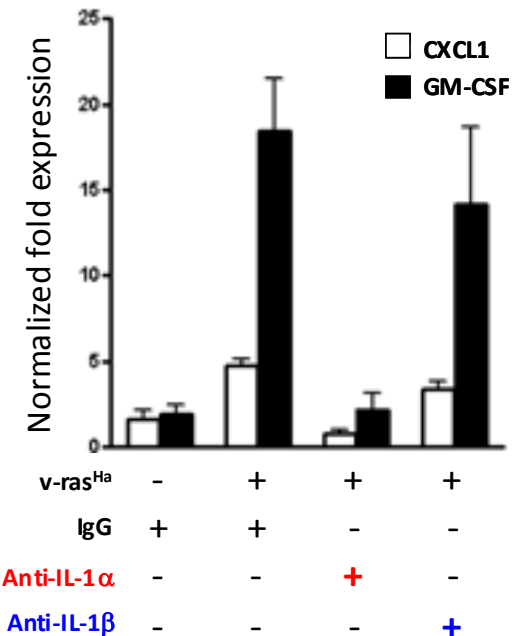
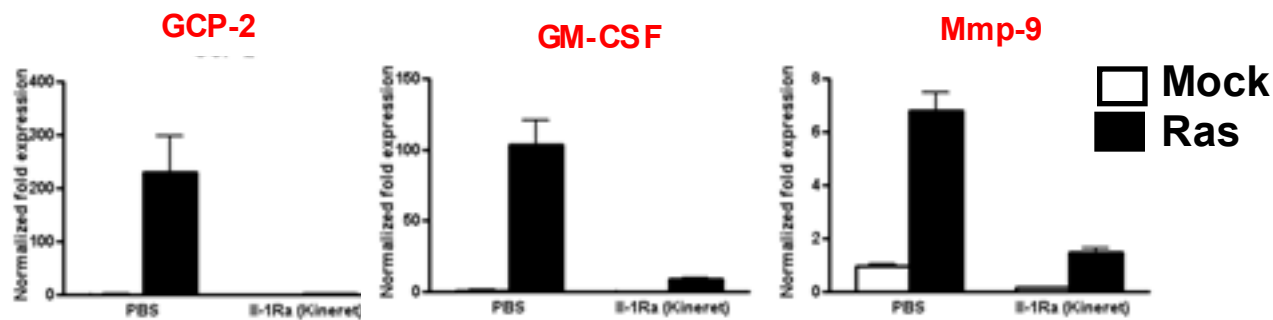
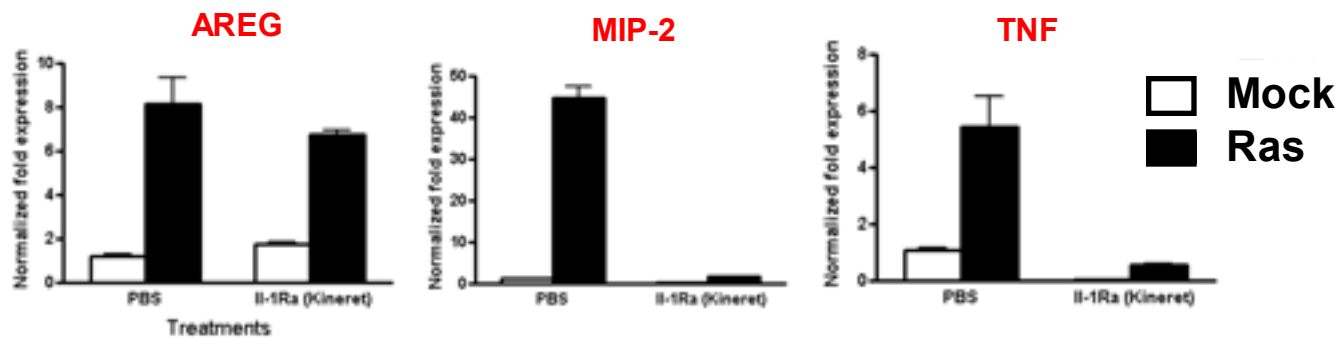
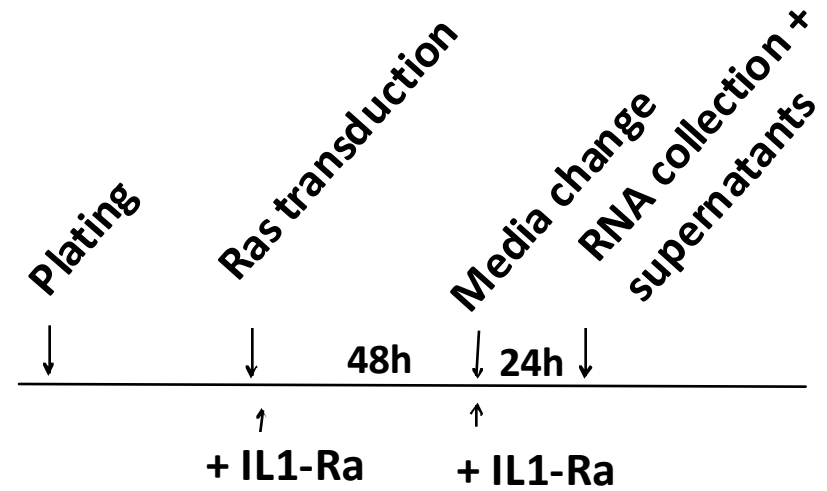


 Control

 $v\text{-ras}^{\text{Ha}}$



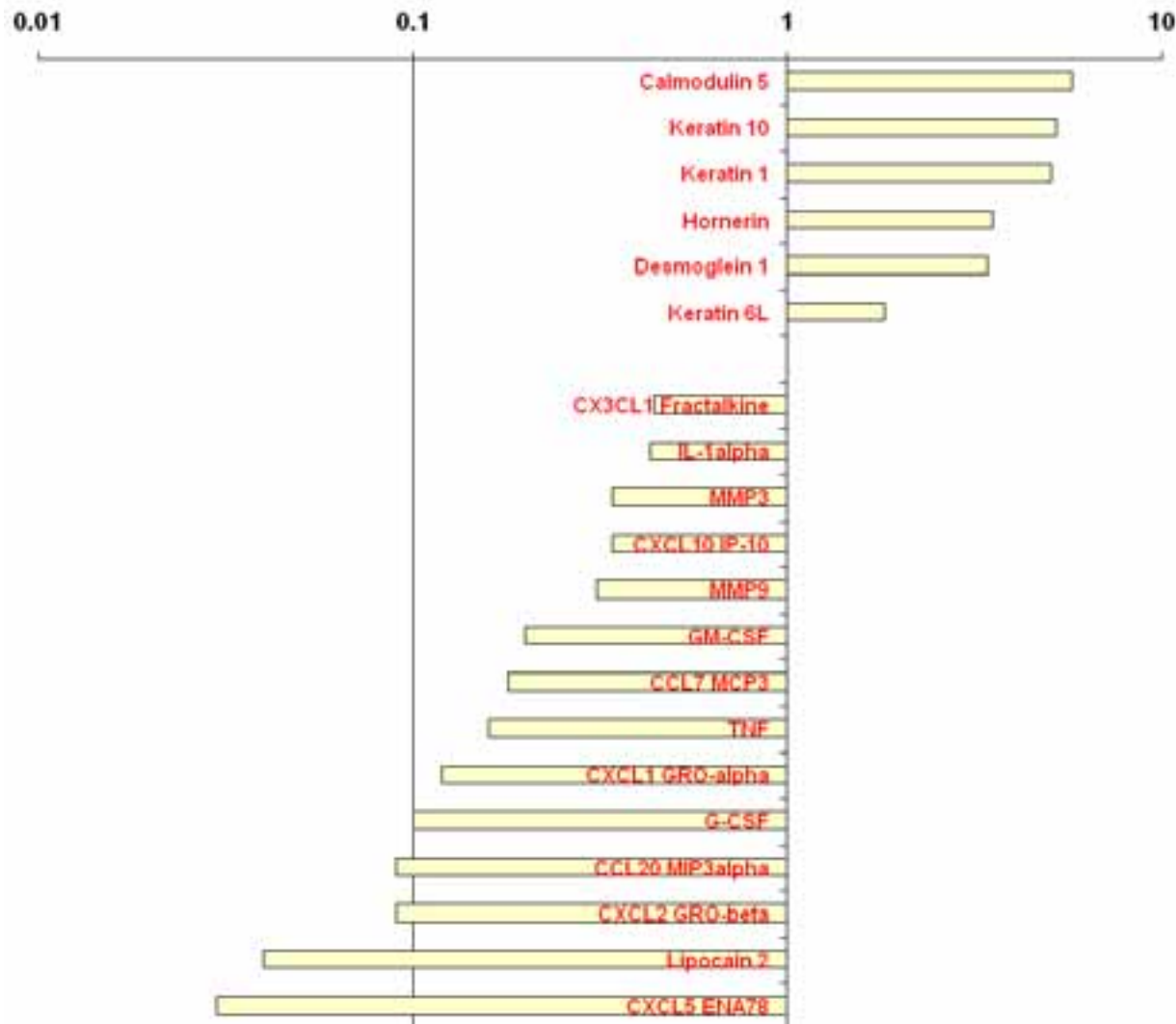
IL-1RA inhibits the production of pro-inflammatory chemokines and cytokines by v-ras^{Ha} transformed keratinocytes



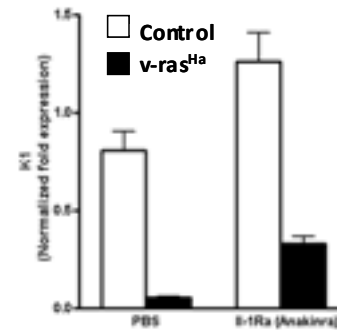


Gene expression (microarray analysis) in v-ras^{Ha} transduced mouse keratinocytes treated with IL-1RA

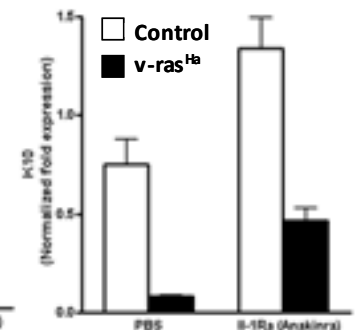
Fold gene expression IL-1RA-treated vs untreated



K1



K10



Ca⁺⁺ (mM) 0.05 0.12

Anakinra - + - + - +
v-ras^{Ha} - - - - + +

K1

Actin

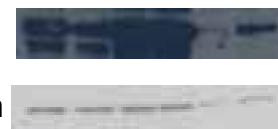


Ca⁺⁺ (mM) 0.05 0.12

Anakinra - + - + - +
v-ras^{Ha} - - - - + +

K10

Actin

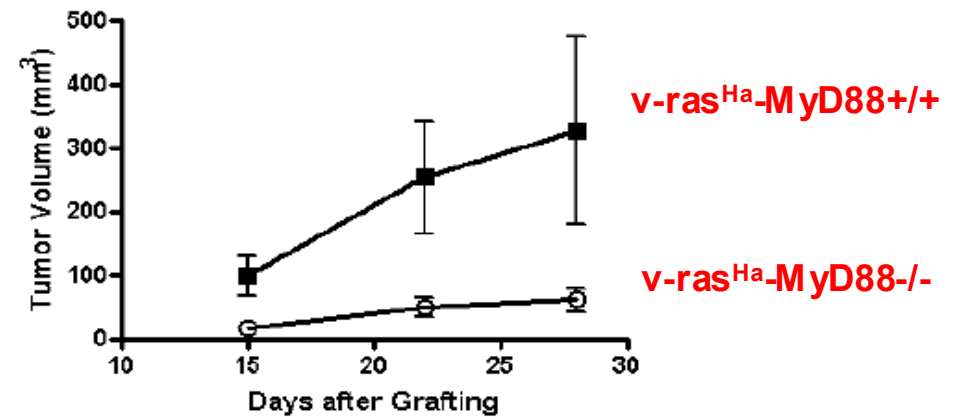


(high)



Nude mouse grafting of v-ras^{Ha} transduced keratinocytes

v-ras^{Ha} transduced primary
keratinocytes (4×10^6)
C57/Bl6
or
MyD88ko
+
primary dermal fibroblasts
(SENCAR) (6×10^6)



MyD88^{+/+}

MyD88^{-/-}

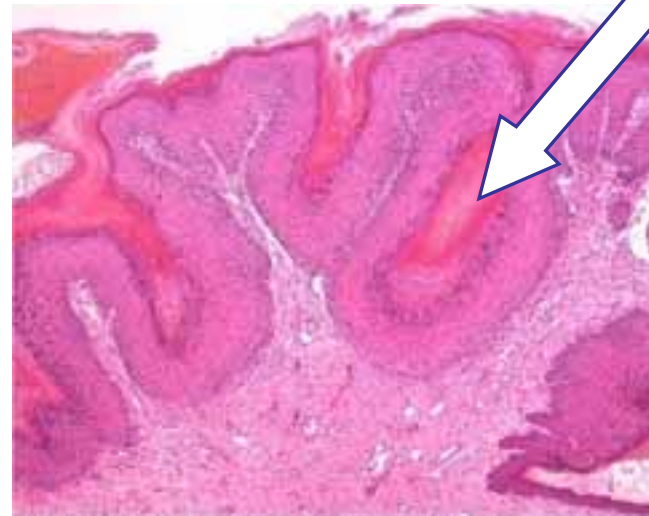


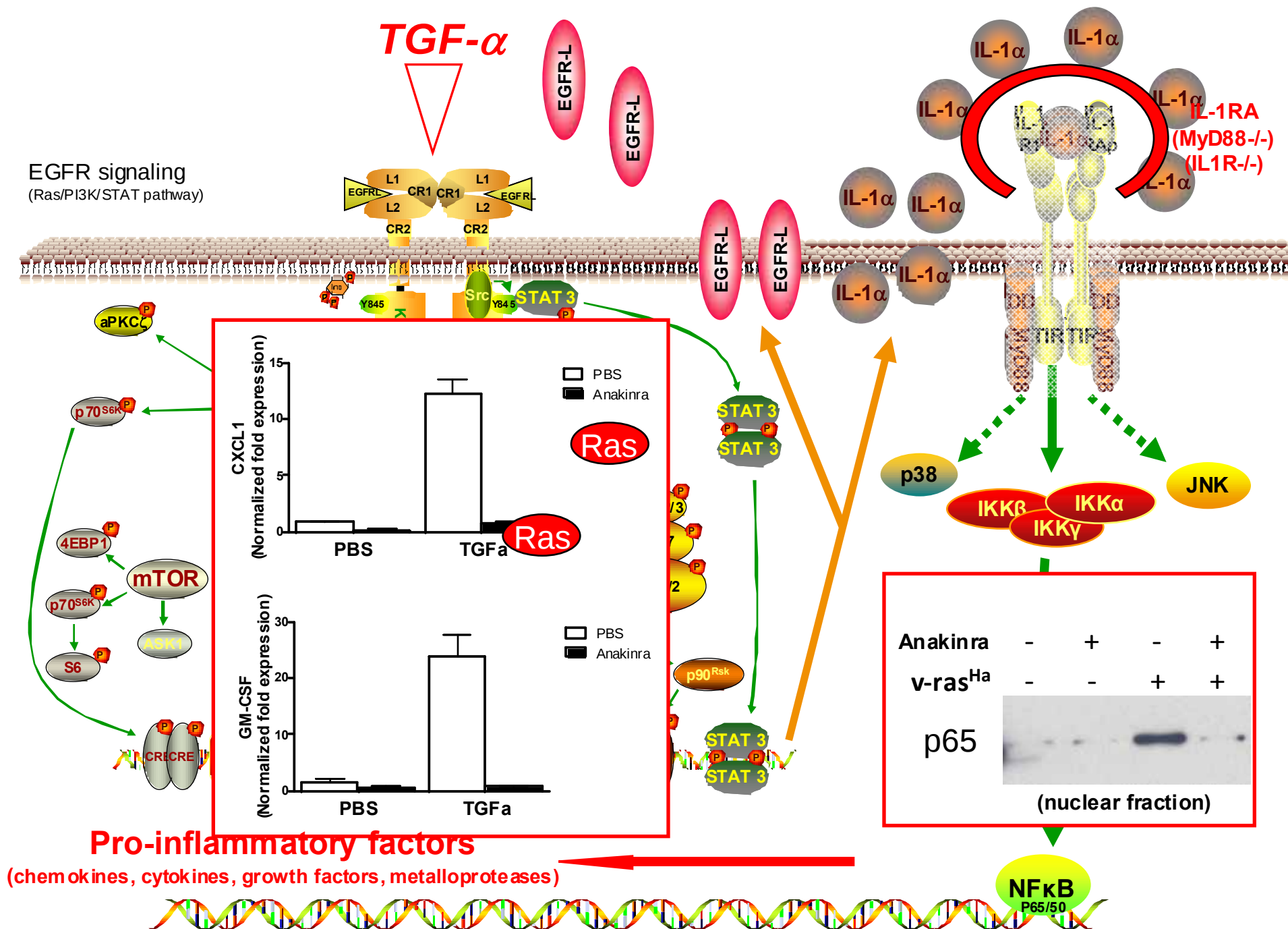
Nude mouse grafting of v-ras^{Ha} transduced keratinocytes

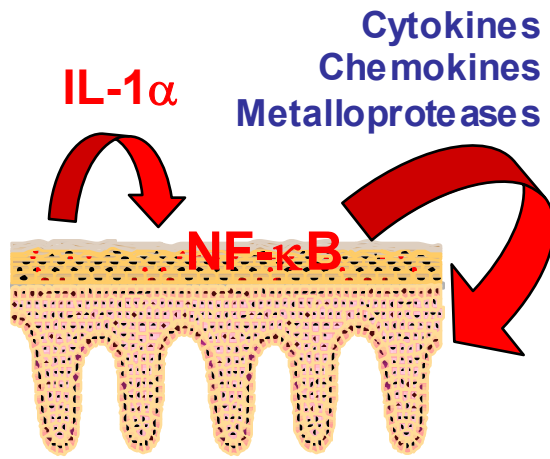
WT



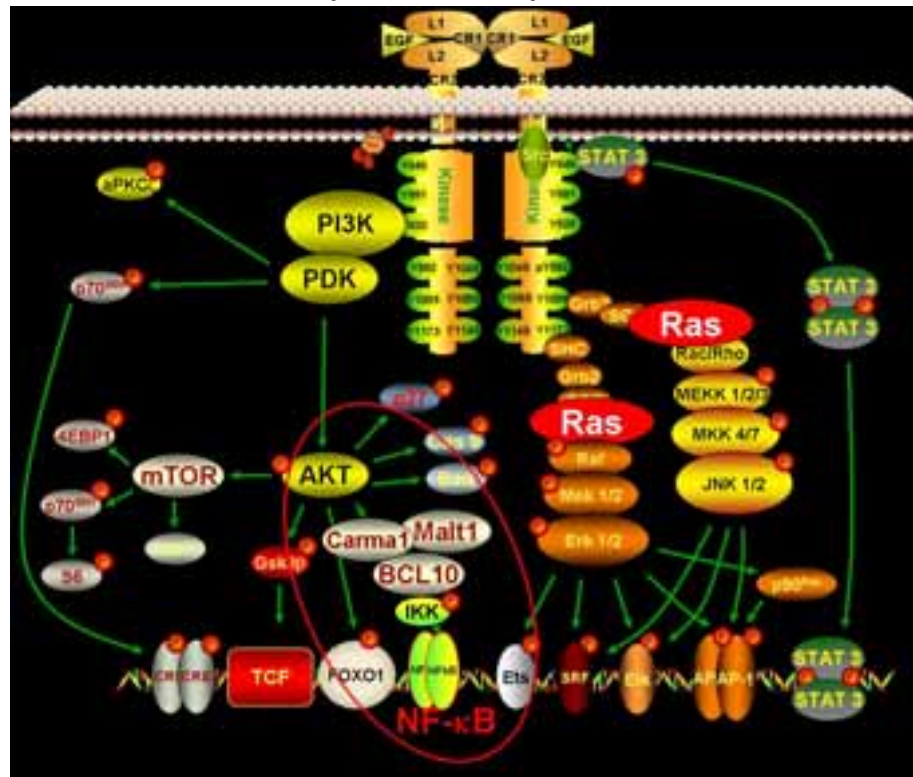
MyD88 KO



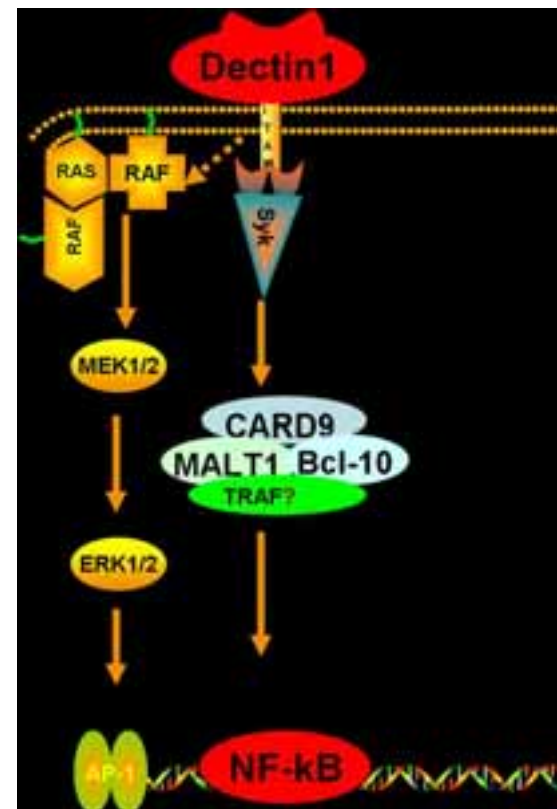
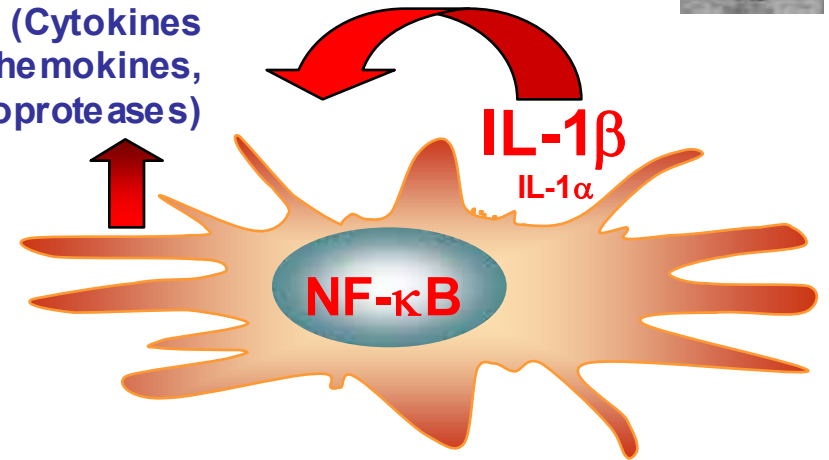




Skin carcinogenesis
(DMBA/TPA)



IL-23
IL-6
TGF- β
(Cytokines
Chemokines,
Metalloproteases)



Human
Dendritic
Cells
Activation by
 β -Glucan



Uzma Hasan
Vladimir Vlach

Isabelle Coste
Toufic Renno

LIR, Schering-Plough
Research Institute
Dardilly, France



Franca Gerosa **Verona University**
Giuseppe Carra **Italy**



Rosalba Salcedo
Zsofia Gyulai
Yava Jones
Marco Cardone
Lyudmila Lyakh

Robin Winkler Pickett
Loretta Smith
Ren Meng Dai
Anna T Mason

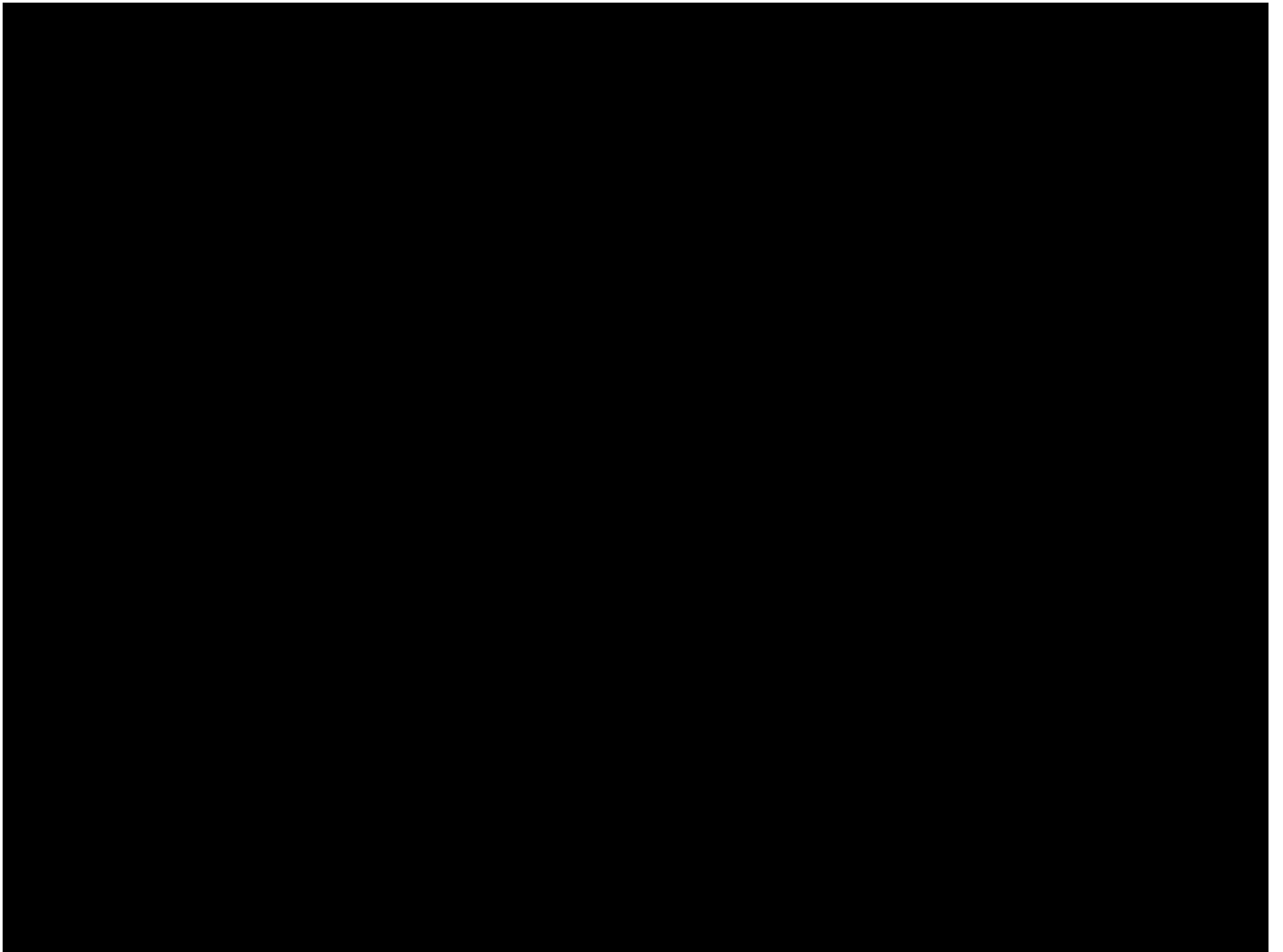
Cancer and Inflammation
Program,
CCR, NCI, Frederick, MD

Christophe Cataisson
Stuart Yuspa

Franco Marincola
Ena Wang

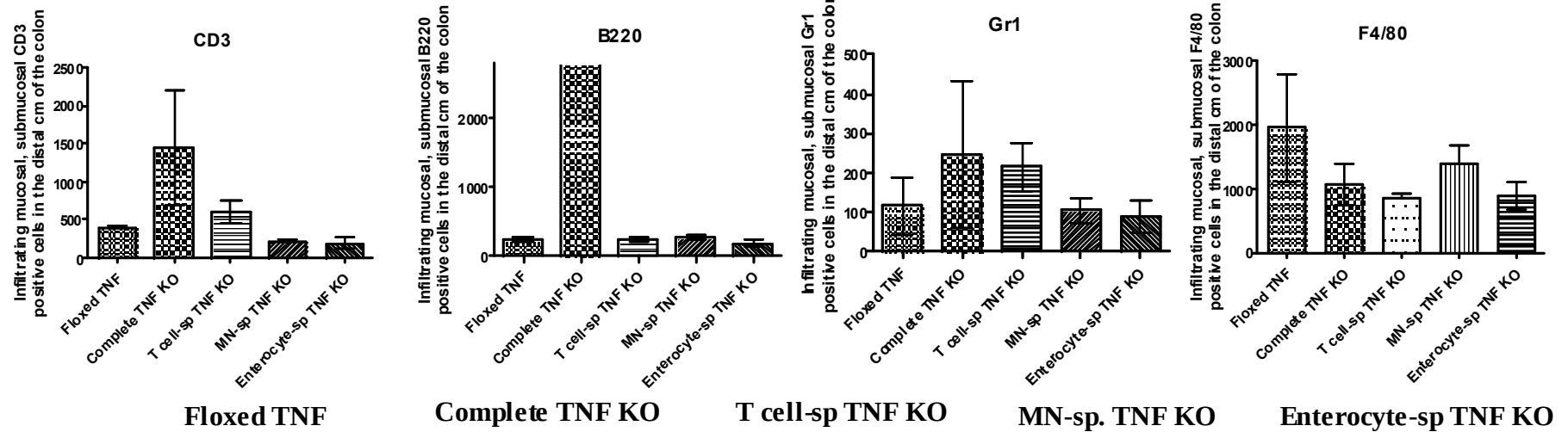
NIH, Bethesda, MD







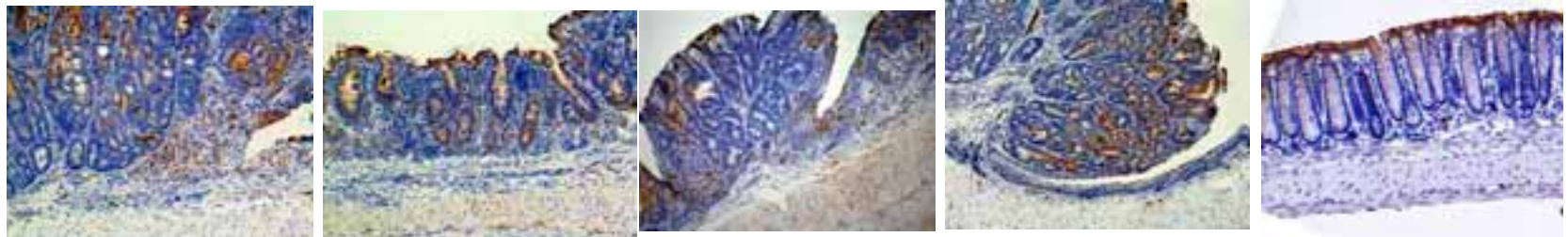
TNF -/- mice have more lymphocytic infiltration and more pronounced inflammation following 4 cycles of DSS



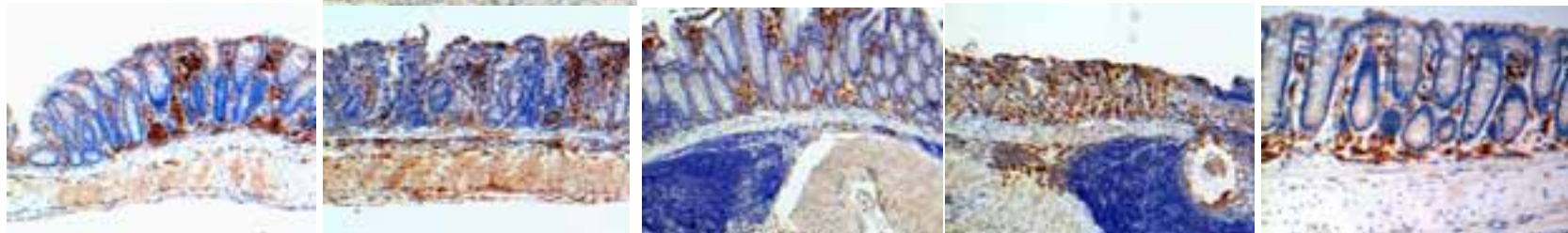
CD3/B220



Gr1

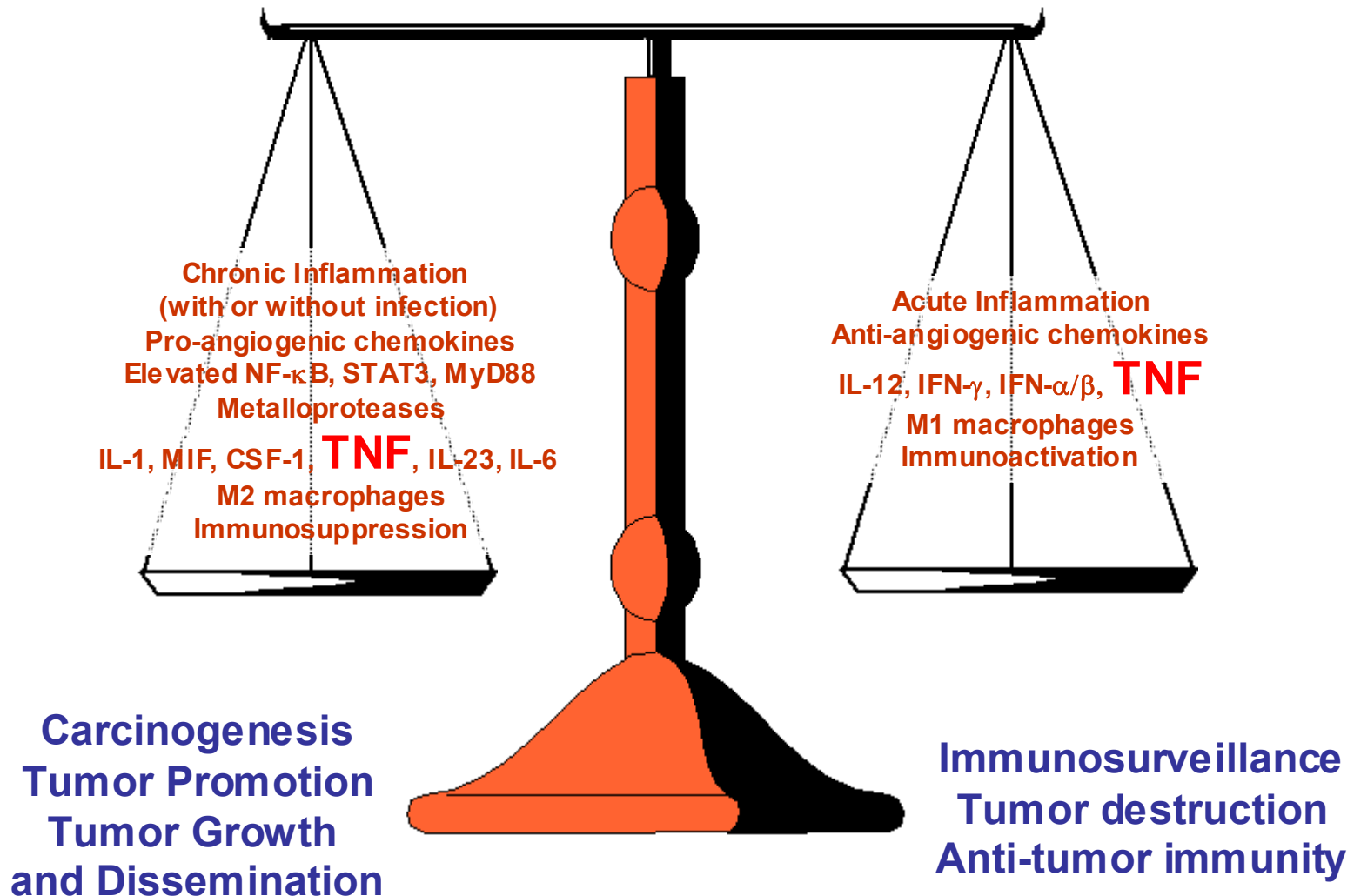


F4/80



Cancer and Inflammation

Tumor Necrosis Factor can be an anti-tumor or a tumor promoting factor depending on the producer cell type and likely the location and time of production



Targeting inflammation for preventing cancer initiation and progression



William B. Coley (1862-1936)

**Coley's mixed toxins
(mixed bacterial vaccine, MBV):**

Heat-killed
Streptococcus pyogenes
and
Bacillus prodigiosus
(*Serratia marcescens*)

W. B. Coley: "The treatment of malignant tumors by repeated inoculation of erysipelas: With a report of ten original cases". *A. J. Med. Sci.* 105:487, 1893

W. B. Coley: "The therapeutical value of the mixed toxins of streptococcus of erysipelas and *Bacillus prodigiosus* in the treatment of inoperable malignant tumors, with a report of 160 cases".
A. J. Med. Sci. 112:251, 1896

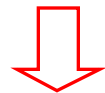


Infection



Cancer

**Innate (Natural) Resistance
(Inflammation)**

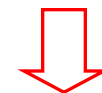


**Natural resistance to tumors
Non-specific inflammatory anti-tumor effects**

**Pro-inflammatory cancer therapy
(Coley Toxin, BCG, TLR ligands)**



Adaptive Immunity



**Tumor-specific Immunity
Immuno-surveillance**

Antigen-specific immunotherapy





Claudius Galenus of Pergamon
ca 130- ca 200

The term cancer was originally applied by Galenus to certain tumors of the breast in which superficial veins appeared much swollen and radiated somewhat like the claws of a crab. Later the name was extended to include all malignant and infiltrating growths.



Although the role of inflammation in favoring carcinogenesis has generated much interest in the last 10-15 years, the Greek physician Claudius Galenus already observed almost 2 thousand years ago some similarity among cancer and inflammation.

In 1863 Rudolf Virchow noted leucocytes in neoplastic tissues and made a connection between inflammation and cancer. He suggested that the "lymphoreticular infiltrate" reflected the origin of cancer at sites of chronic inflammation.

The recent upsurge of studies linking cancer and inflammation were inspired by the observation made two decades ago by Harold Dvorak, M.D., of Harvard University, who observed that inflammation and cancer share some basic developmental mechanisms (angiogenesis) and cells (lymphocytes, macrophages, and mast cells), and that tumors act like "wounds that do not heal."



Rudolph Virchow
1821-1902



Dr. Harold Dvorak
Harvard University



Role of Toll-like receptors (TLRs) in cancer

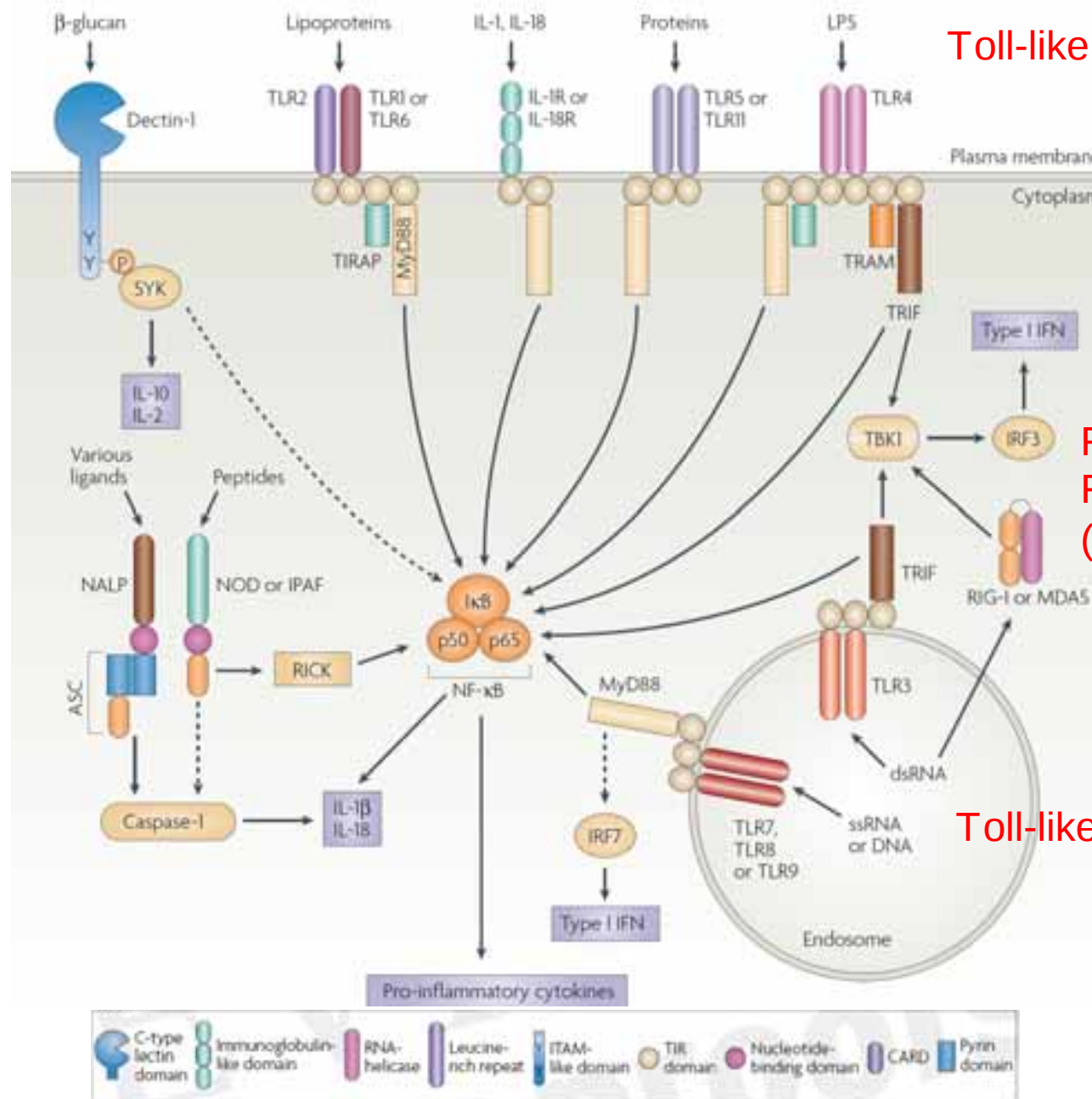
- Toll-like receptors were originally studied prevalently in hematopoietic cells (primarily dendritic cells, phagocytes, and B lymphocytes) but at least some members of this receptor family are widely expressed on other cell types including tumor cells.
- Although they have been described to recognize products of foreign organisms (pathogenic or not) they also participate in the regulation of inflammation by recognizing endogenous ligands (“alarmins”) that are present in inflamed tissues.
- The cellular response to TLR ligands is not only production of pro-inflammatory mediators but they are also involved in control of tissue homeostasis and regulate cellular differentiation, proliferation, and apoptosis. The balance between MyD88 and TRIF signaling and the production of type I IFN determine proliferation versus apoptosis in tissue and tumor cells and activation versus survival in dendritic cells.
- Epidemiologic/genetic evidence indicates a role of TLRs and signaling molecules (TLR1,2,6,10,3,4, MyD88, IRAK4) in the frequency and progression of human cancer.

Pattern Recognition Receptors (PRR)

C-type lectins,
Scavenger r.

Toll-like receptors

NOD-like
Receptors:
↓
NALPs NODs



RIG-I-like
Receptors
(helicases)

Toll-like receptors

Role of MyD88 in inflammation-dependent carcinogenesis: A tale of three Interleukins-1

