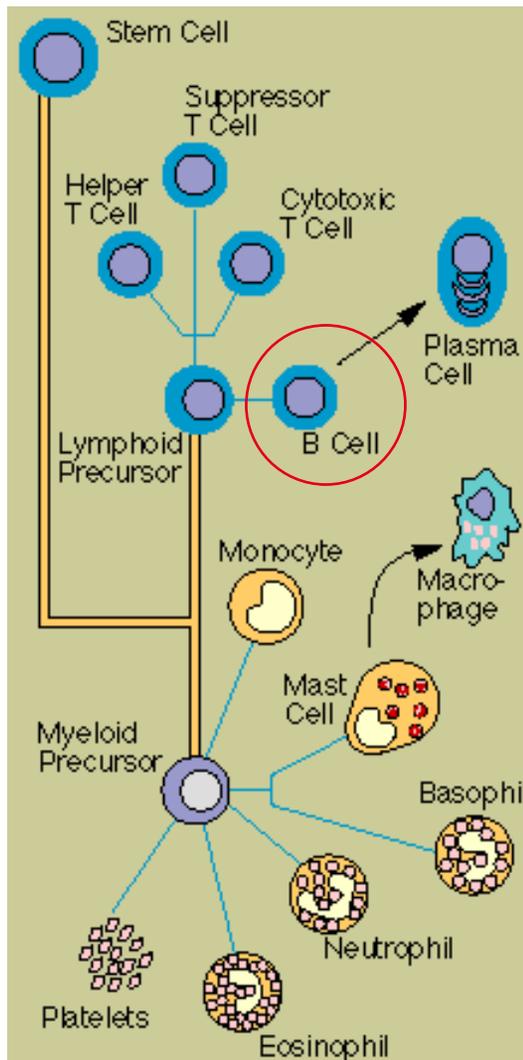


B Lymphocyte (B cell)



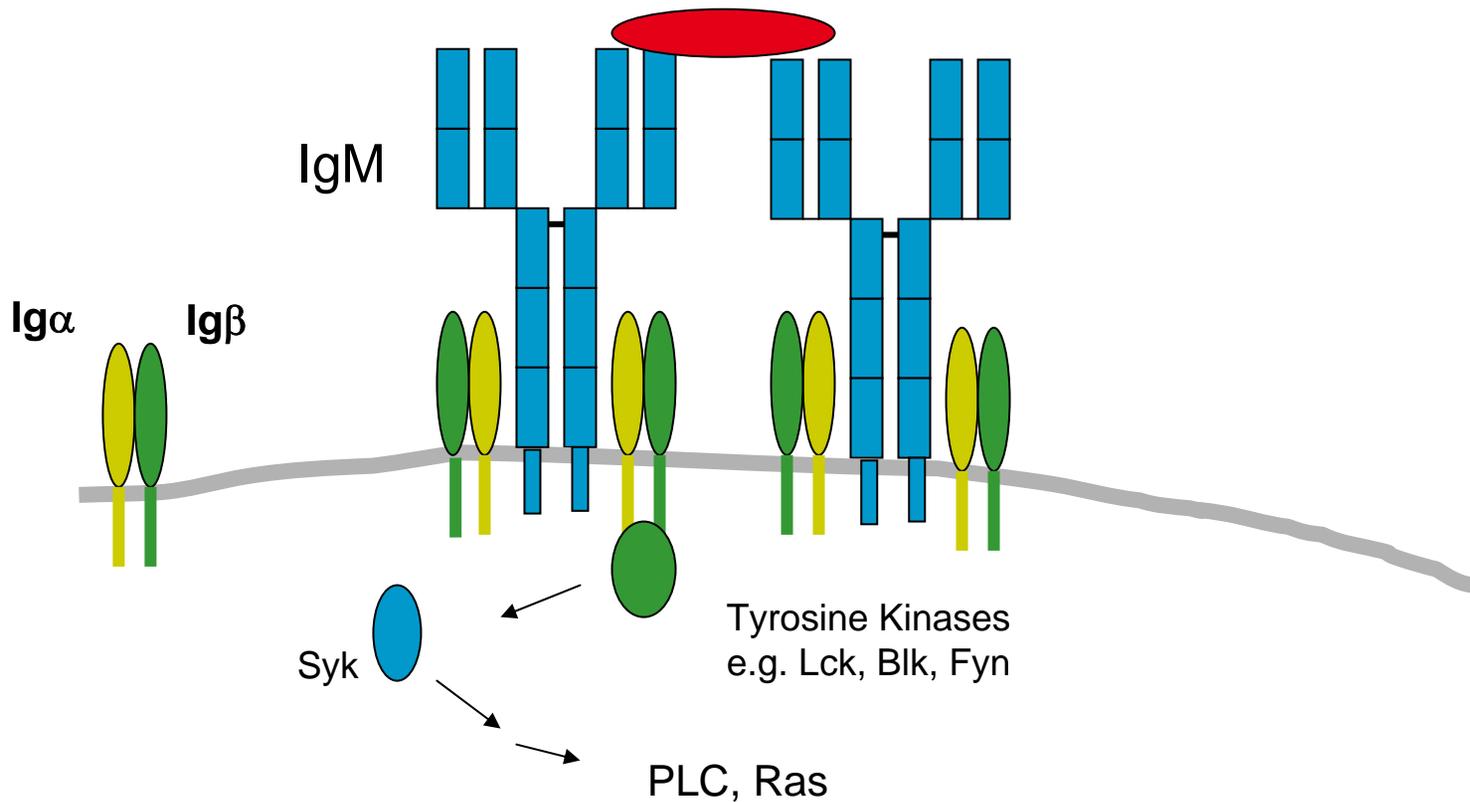
- **B cells:**

- Develop from stem cells in the bone marrow and differentiate into antibody-producing plasma cells in the blood
- Are capable of making a vast number of antibody specificities from a limited number of genes
- Antibodies are both secreted to fight disease and get displayed on the B cell surface for antigen-specific signaling and activation

B cell receptor



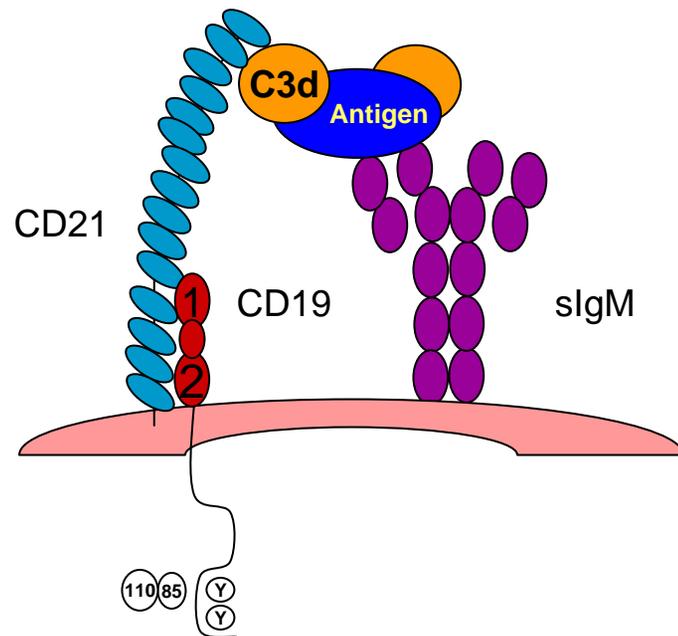
Antigen can cross-link the BCR and induce a signaling cascade



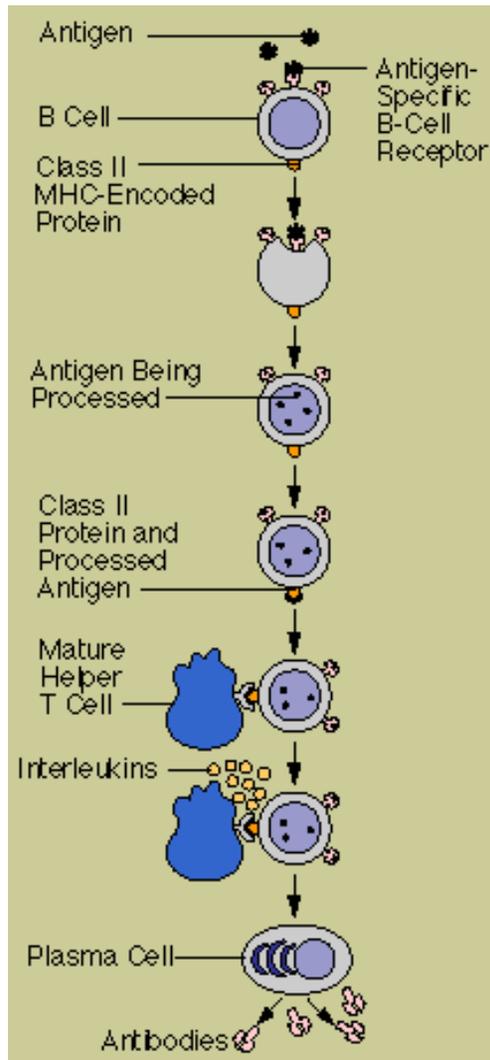
B cell co-receptor



- Lowers the threshold for cell activation in combination with antigen-specific triggering



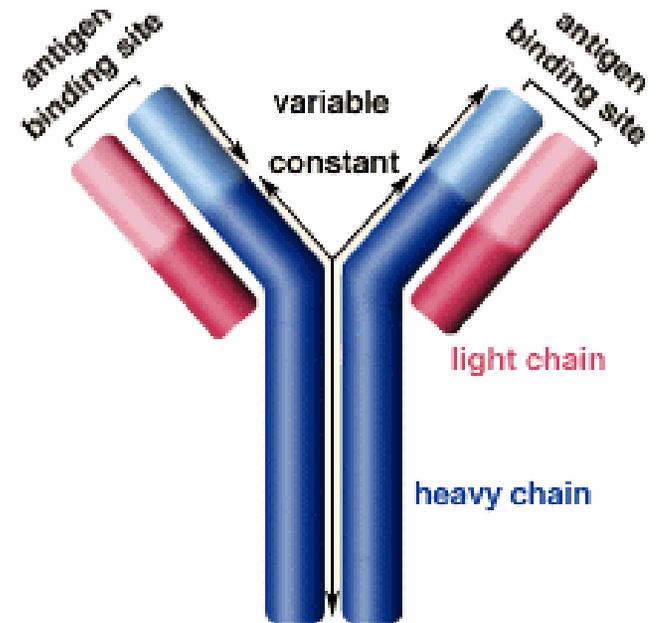
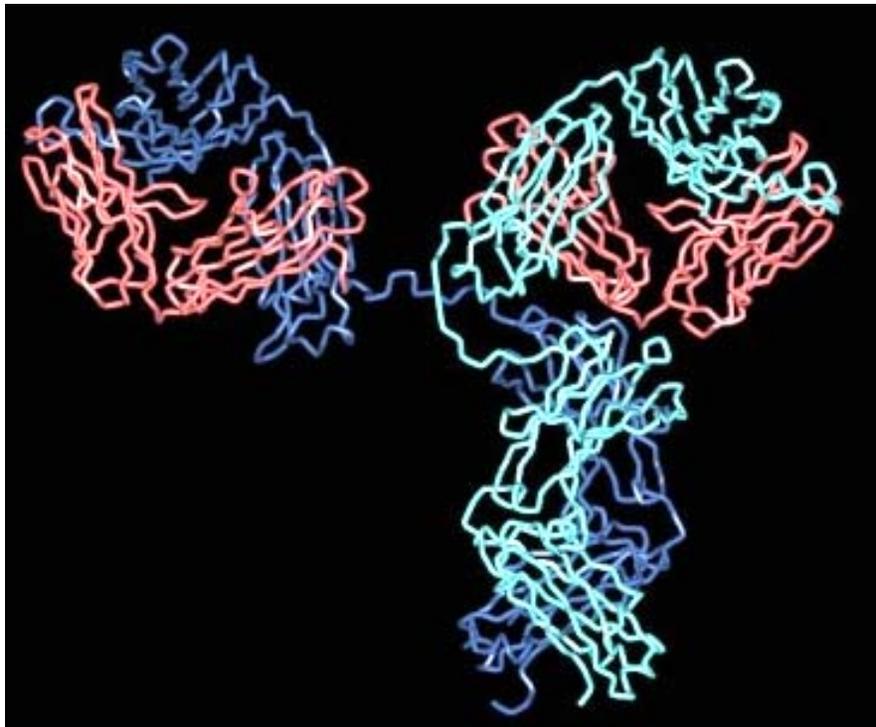
B cell activation



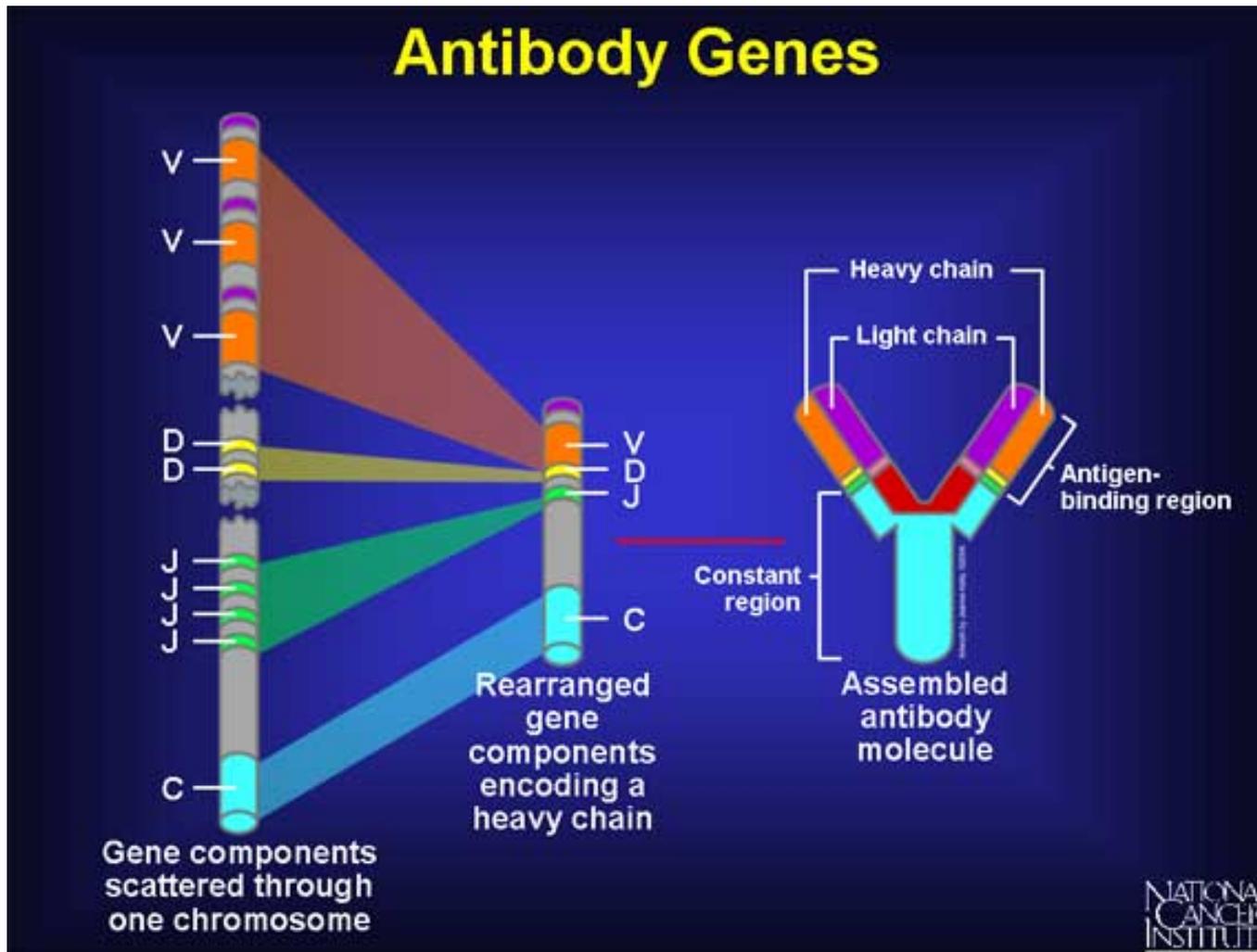
- **B cells:**

- Interact with other immune cells such as helper T cells that provide essential cytokines
- Soluble antigens bind to the B cell receptor (BCR) complex and are taken into the cell where they are degraded into peptides
- Peptide:MHC class II complexes bind to TCR on helper T cells and stimulate cytokines essential for differentiation into antibody-producing plasma cells

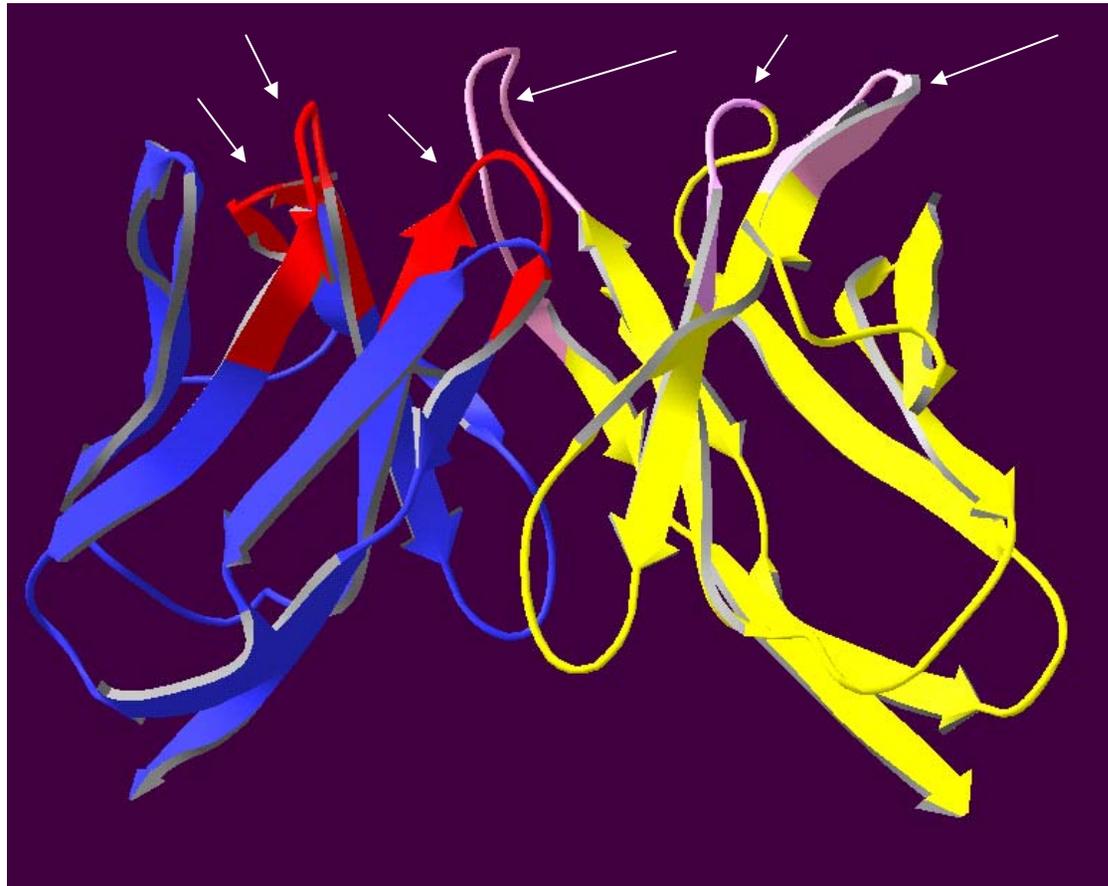
Antibody Structure



From Genes to Antibody



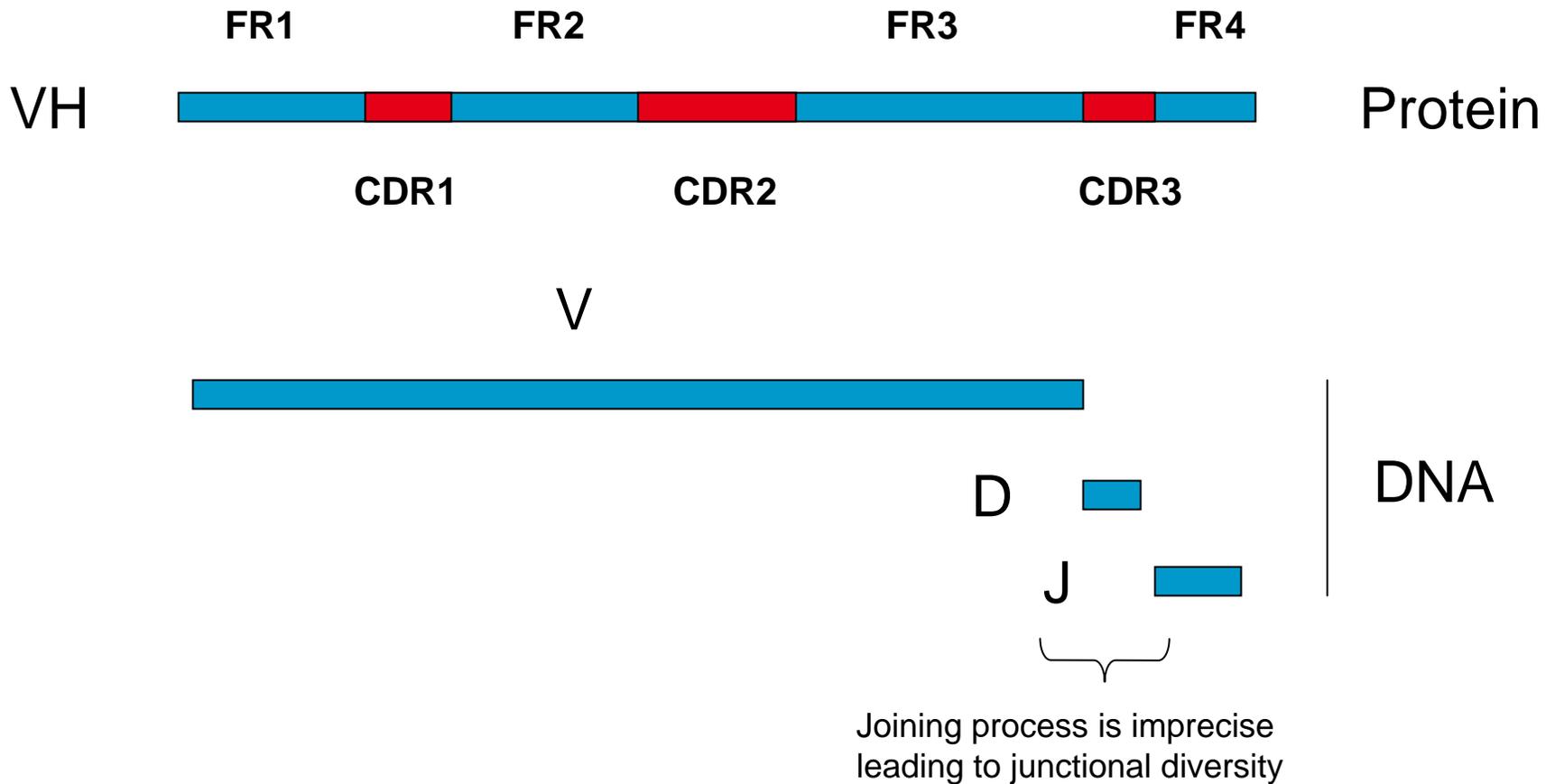
Antibody V region structure



CDRs

Antigen-binding
regions

Variable regions - H chain



Generators of antibody diversity



- **Recombination of different V + D + J regions**
 - Genomic repertoire partly responsible for diversity
- **Imprecision of recombination events**
 - Varied cross-over events result in different D segment lengths
 - Non-template encoded N-nucleotide additions (TdT enzyme)
- **Somatic hypermutation**
 - Occurs when B cells respond to antigen stimulation
 - Mutations in and around CDRs

Antibody gene organization



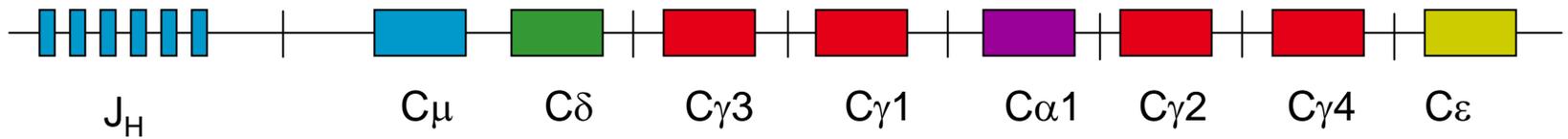
V_H (1-51)



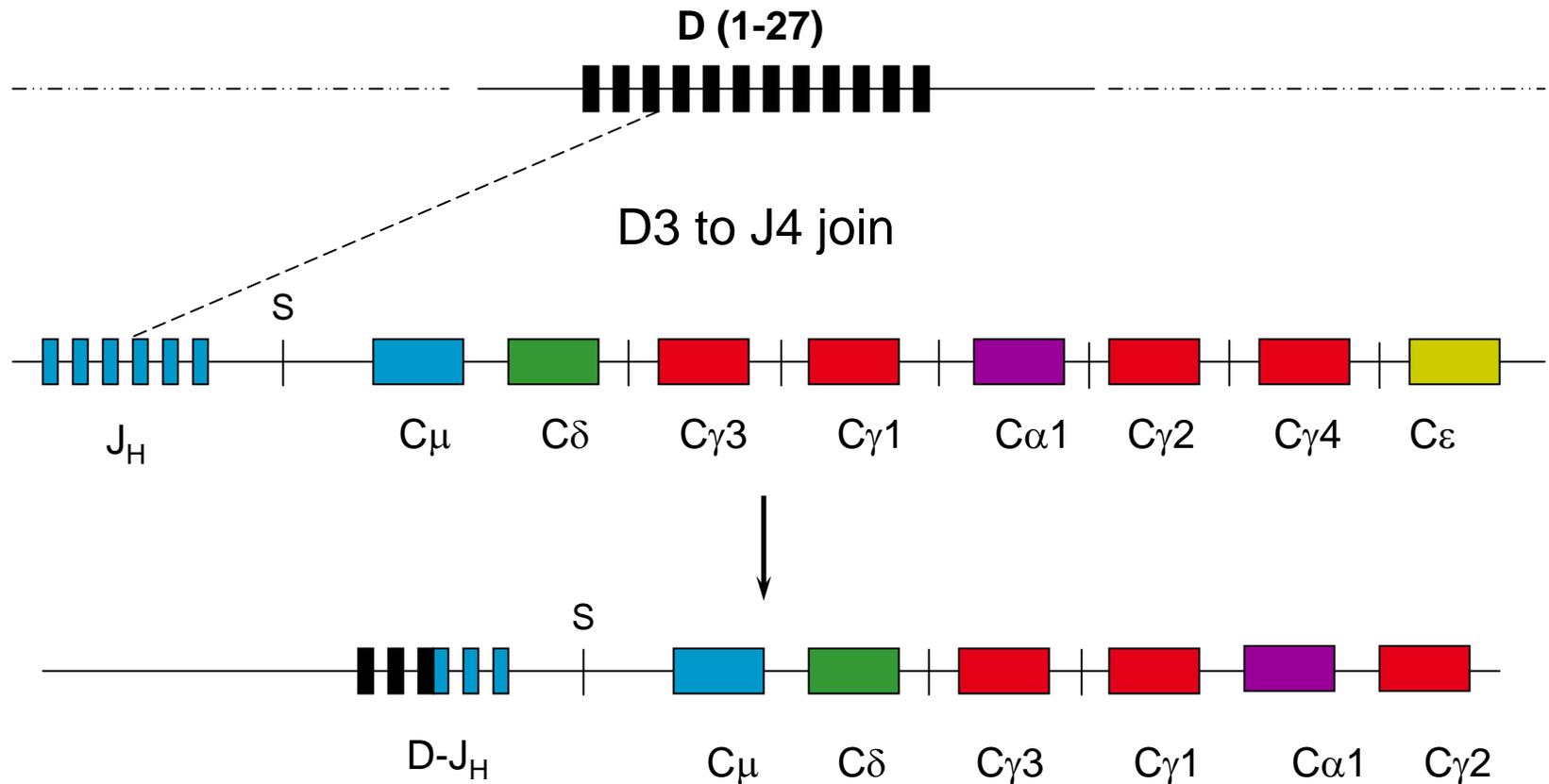
D (1-27)



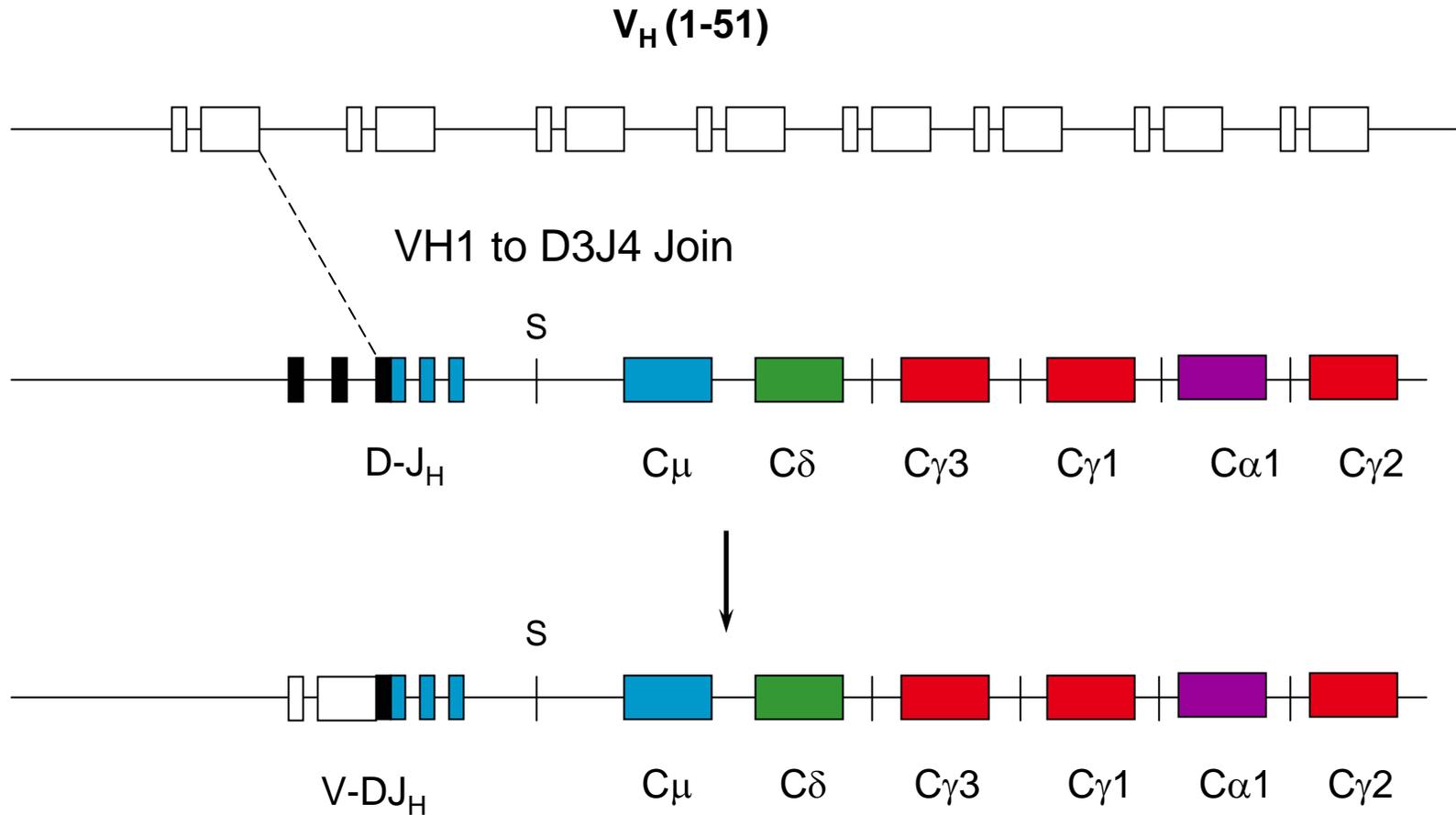
S



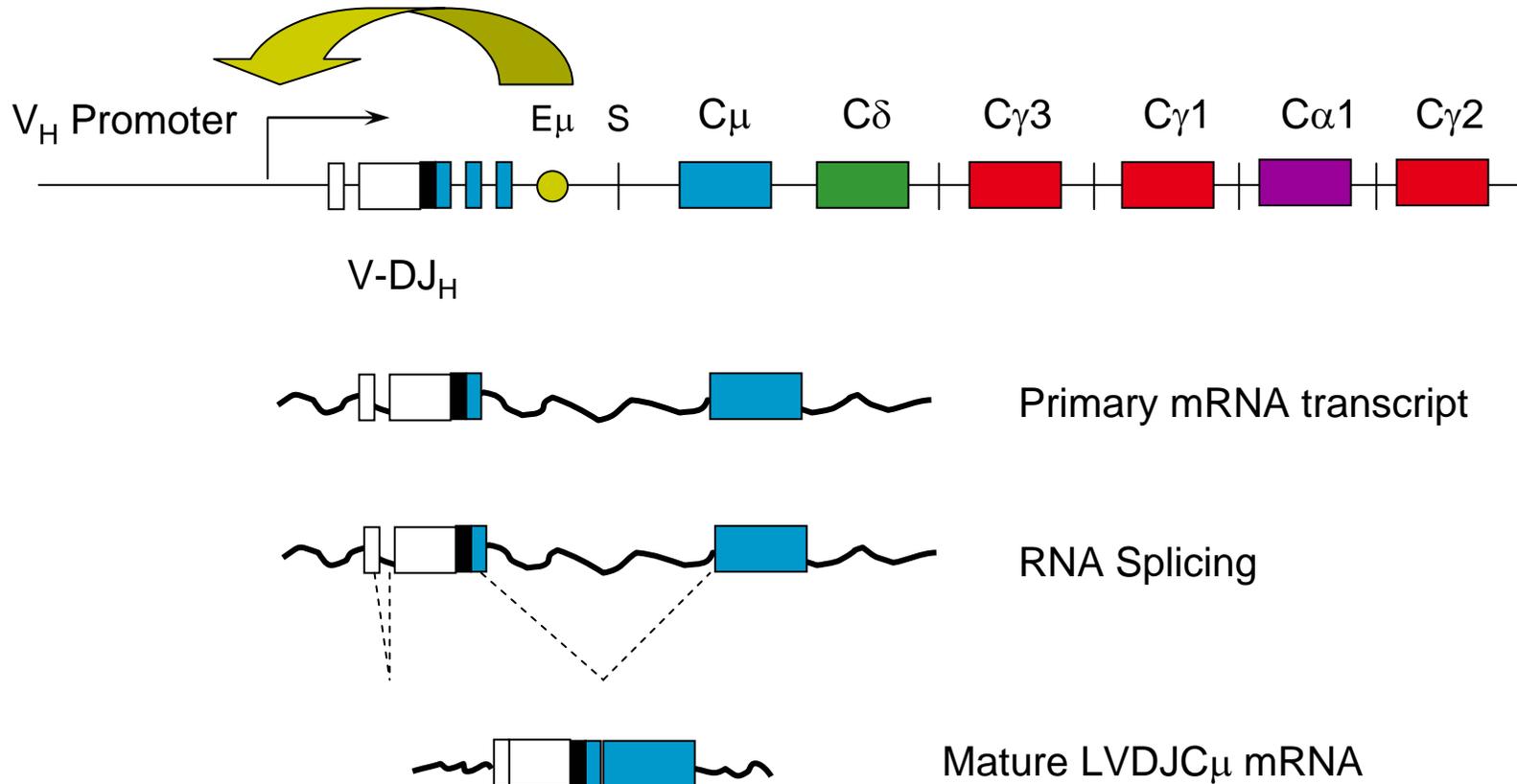
Antibody H chain gene rearrangement in **early pro-B cells**



Antibody H chain gene rearrangement in **late pro-B cells**



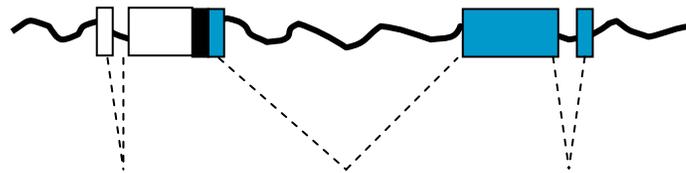
Productive rearrangement results in gene activation and expression



Alternative splicing creates “membrane” form of μ chain as part of BCR

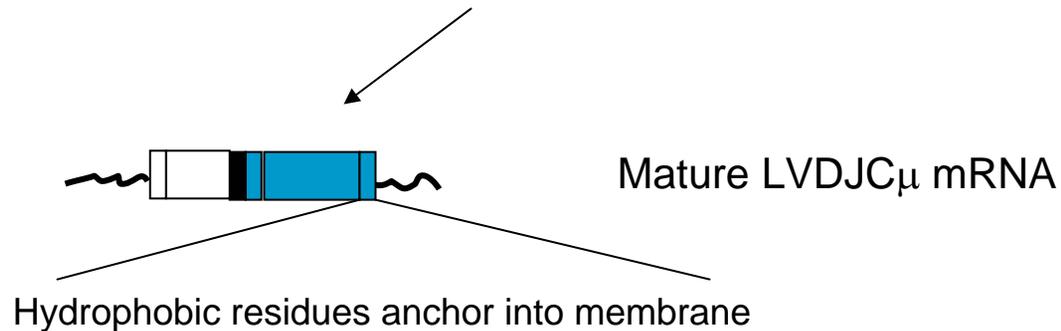


Primary mRNA transcript



Alternative RNA Splicing

Addition of “membrane” exon



Mature LVDJC μ mRNA

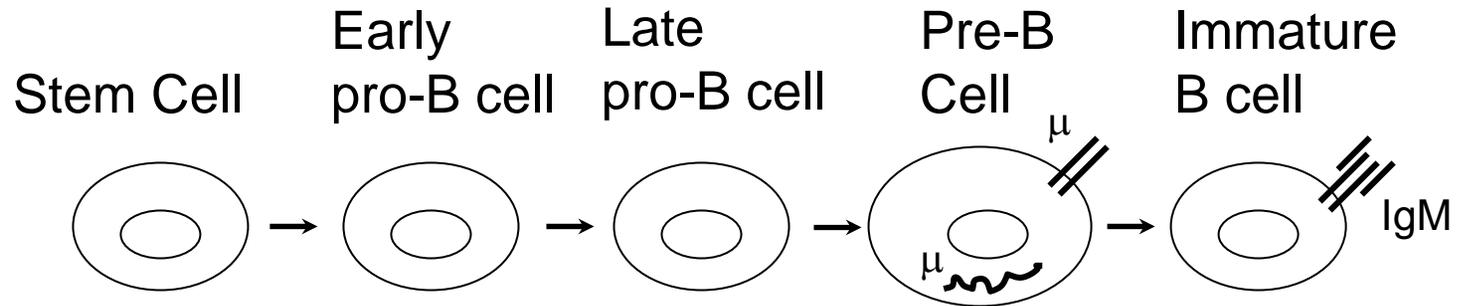
Hydrophobic residues anchor into membrane

Steps in B cell development



- **Antigen-independent steps**
 - Occurs in bone marrow
 - H and L chain rearrangement
 - Selection of B cells with “productive” gene rearrangement
 - Self reactive clones are deleted
- **Antigen-dependent steps**
 - Occurs in periphery, e.g. lymph nodes
 - Expression of IgM on surface (Ag-specific BCR)
 - Additional rearrangement for isotype switch (e.g. IgG)
 - Somatic mutation drives additional diversity

Antigen-independent (early) Development



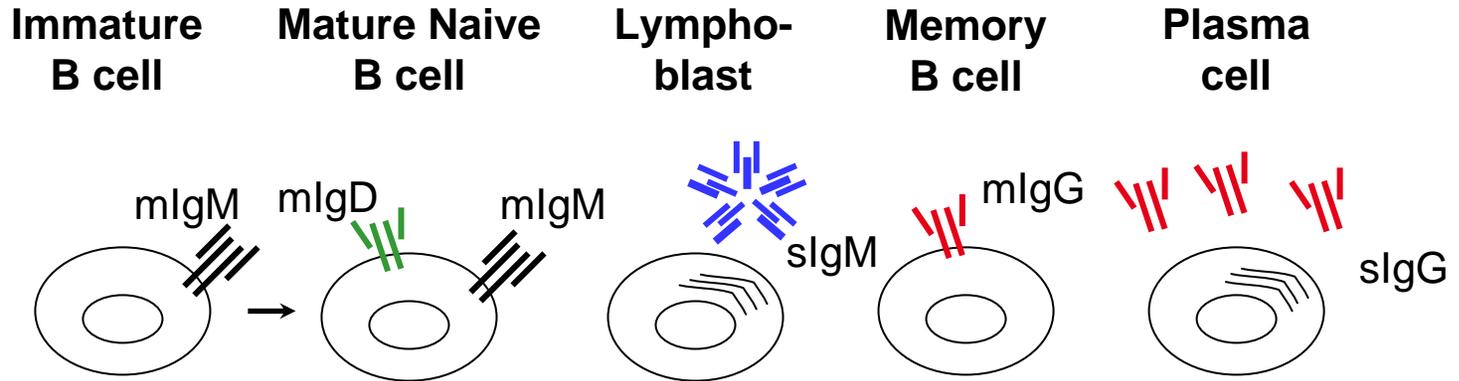
H chain	Germline	D-J Rearranged	V-DJ Rearranged	V-DJ Rearranged	V-DJ Rearranged
L chain	Germline	Germline	Germline	V-J Rearranging	V-J Rearranged
Signals	VLA4 / ICAM	Stem cell factor / Kit	IL-7 / IL7-R	IL-7 / IL7-R Other CAM	
Enzymes Transcription factors		Rag-1 and 2 TnT	Rag-1 and 2 TnT	Rag-1 and 2 NFκB	NFκB

Antigen-independent development – additional points



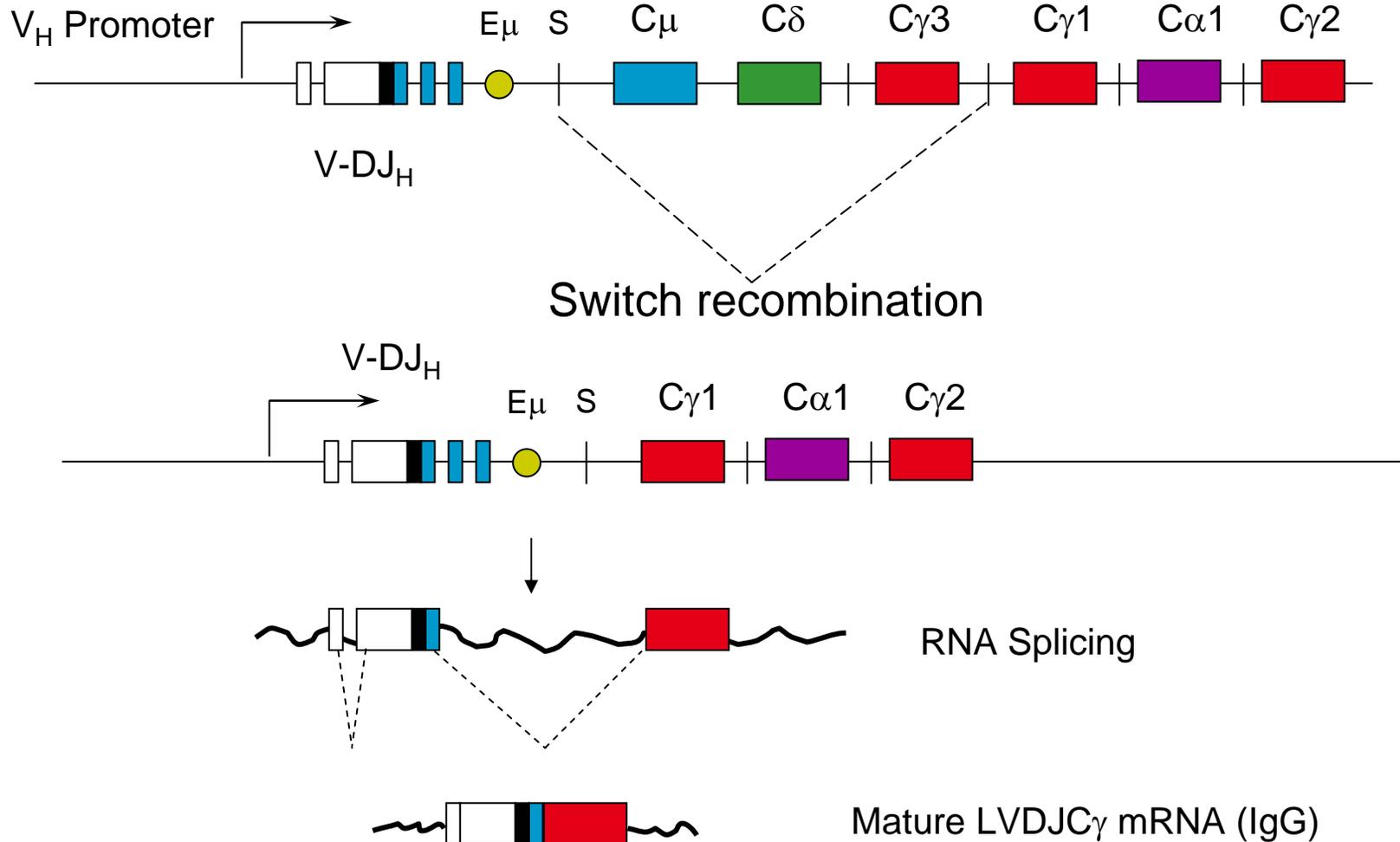
- Productive rearrangement of μ chain
 - blocks further rearrangement of H chain gene
 - Induces rearrangement of L chain genes until a successful rearrangement occurs – then further blocked
 - “Allelic Exclusion”
- Pre-B cells with expressed μ chain expand
 - Multiple chances to use a given H chain with multiple L chains

Antigen-dependent (late-stage) Development



H chain	μ chain in membrane form	μ and δ chains in membrane form via alternative splicing	Alternative splicing of μ to secreted form	Somatic hyper mutation Isotype switch to $C\gamma$, $C\alpha$, $C\epsilon$	Isotype switching and alternative splicing to secreted Ab
L chain	V-J Rearranged	V-J Rearranged	V-J Rearranged	V-J Rearranged Somatic hyper mutation	

Isotype switching occurs by recombination and gene deletion



Consequences of isotype switching



- Change in binding valency
 - Pentameric IgM → Dimeric Ig
 - Compensated by affinity maturation
- Change in antibody properties
 - Circulating half-life $IgG > IgM > IgE$
 - Effector functions - ADCC, complement fixation
 - Biodistribution
 - IgG blood
 - IgA mucosal surfaces
 - IgE mast cells

Summary



- Antibody diversity created in concert with B cell development and activation by antigen
- Involves diversity of binding
 - Affinity of Ag:Ab interaction
 - Specificity
- Also diversity of function
- Nature has been performing Antibody Engineering
- Knowledge gained inspired genetic engineering of Abs

Antibody Engineering

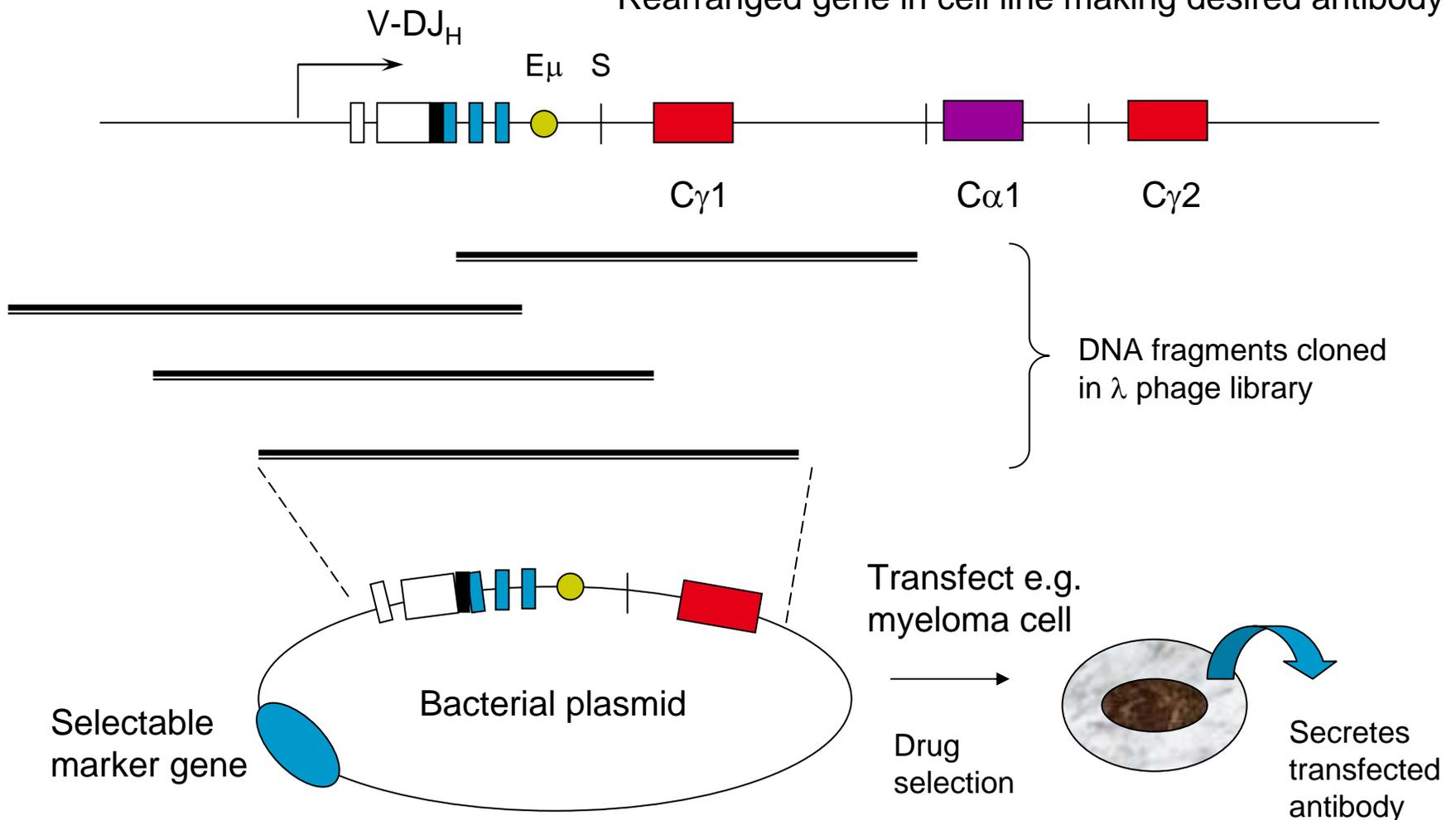


- **First step - development of monoclonal antibodies**
 - Fusion of antibody-producing B cell with myeloma
 - Results in immortalized monospecific Ab-producing cell line
- **Second step - ability to clone and re-express Abs**
 - Initially done with cloned, rearranged genes from hybridomas
 - Parallel work with isolated Fab fragments in bacteria
- **Third step - re-engineering for desired properties**
 - Reducing immunogenicity of mouse antibodies
 - Tailoring size and half-life for specific need
 - Adding or removing functions
- **Engineered diversity – phage display approach**

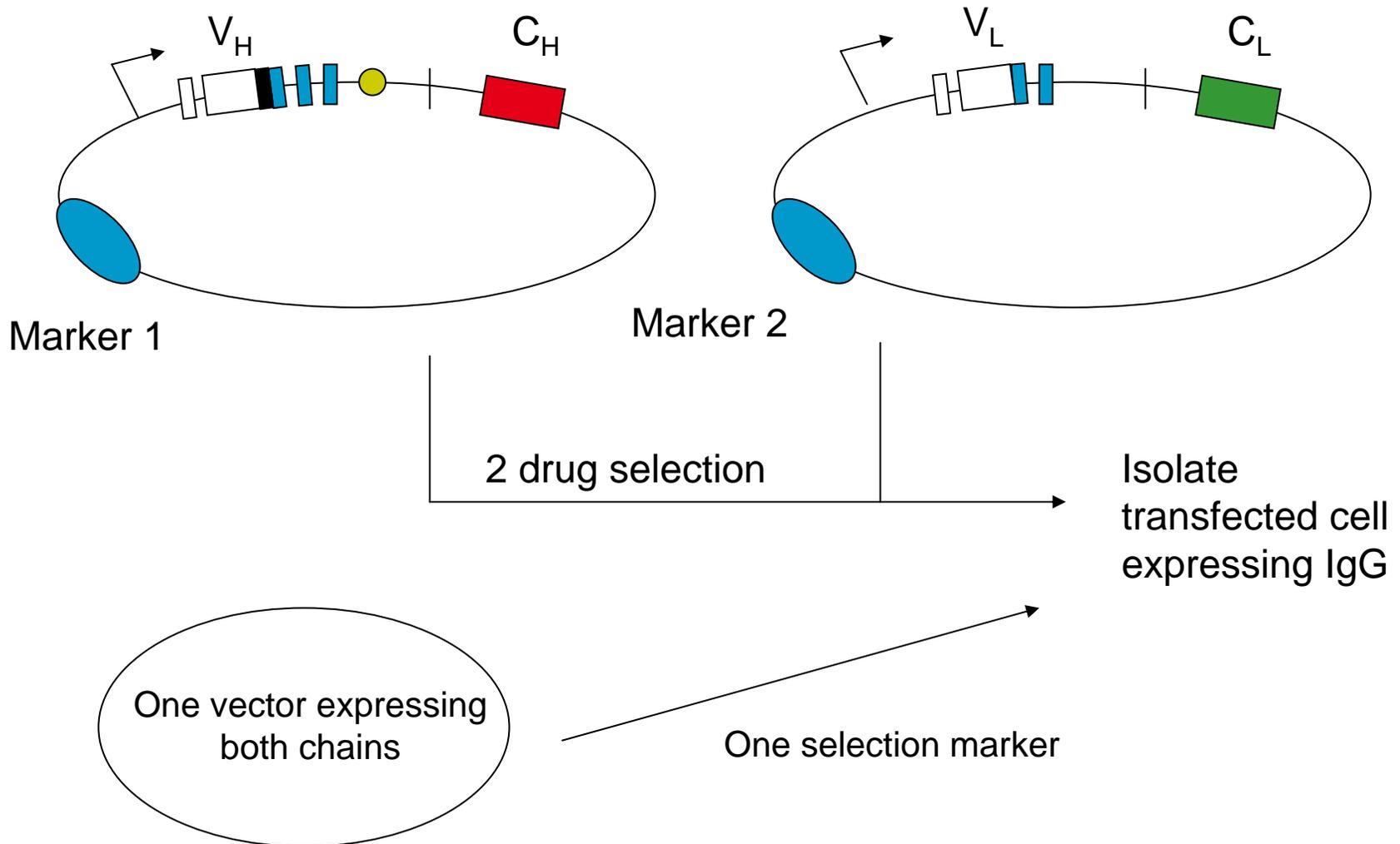
Cloning and re-expressing antibodies



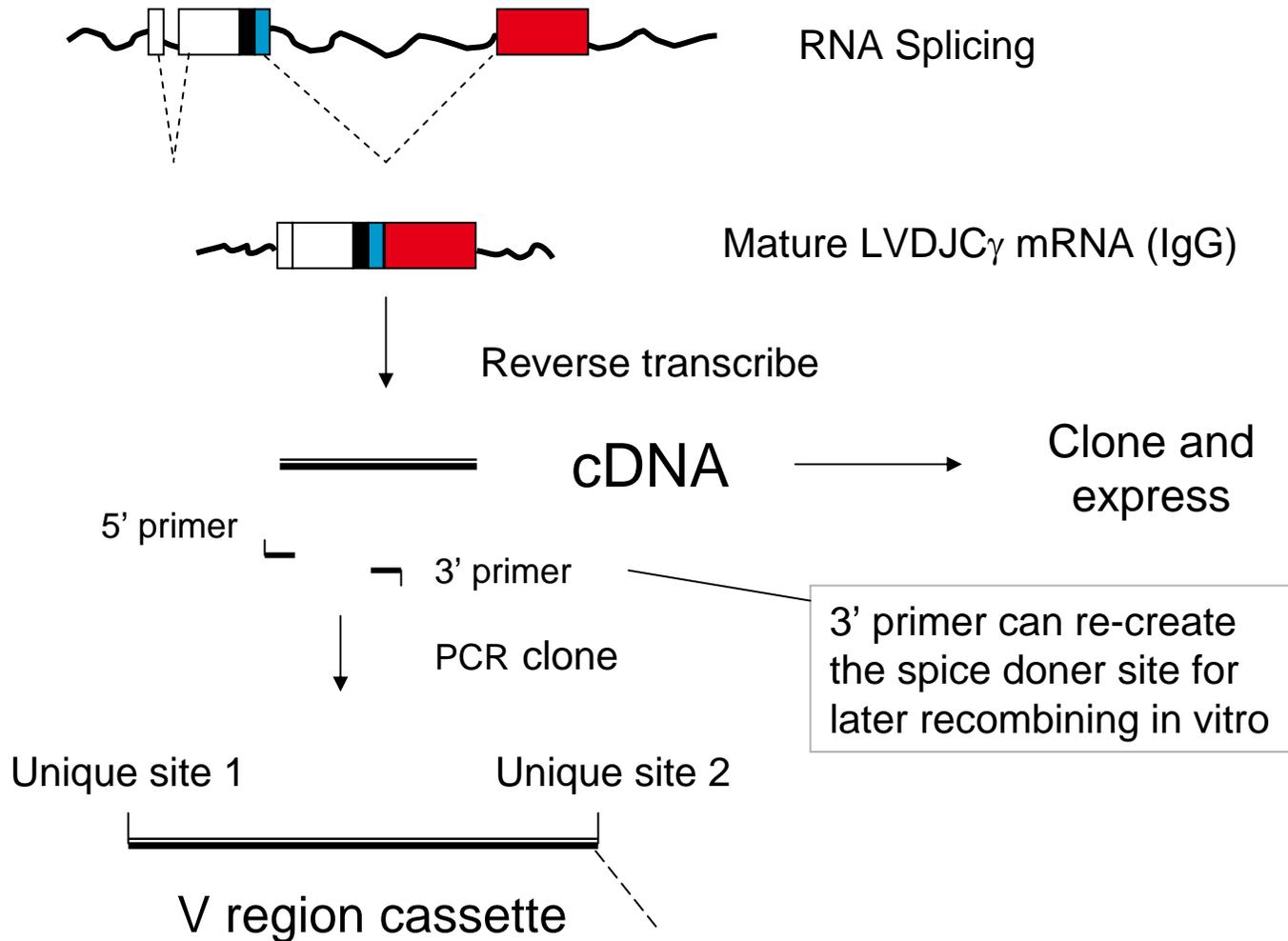
Rearranged gene in cell line making desired antibody



Cloning and re-expressing antibodies



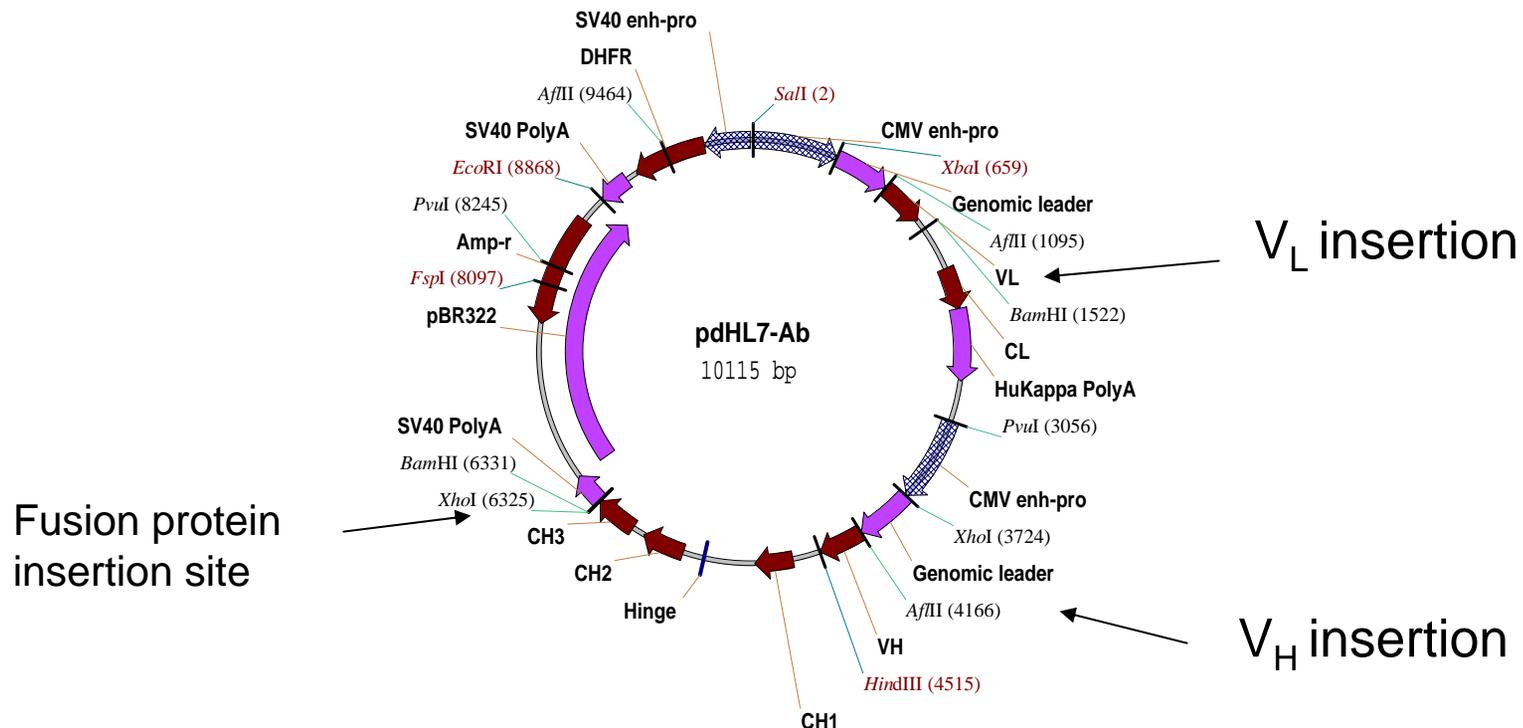
More efficient cloning



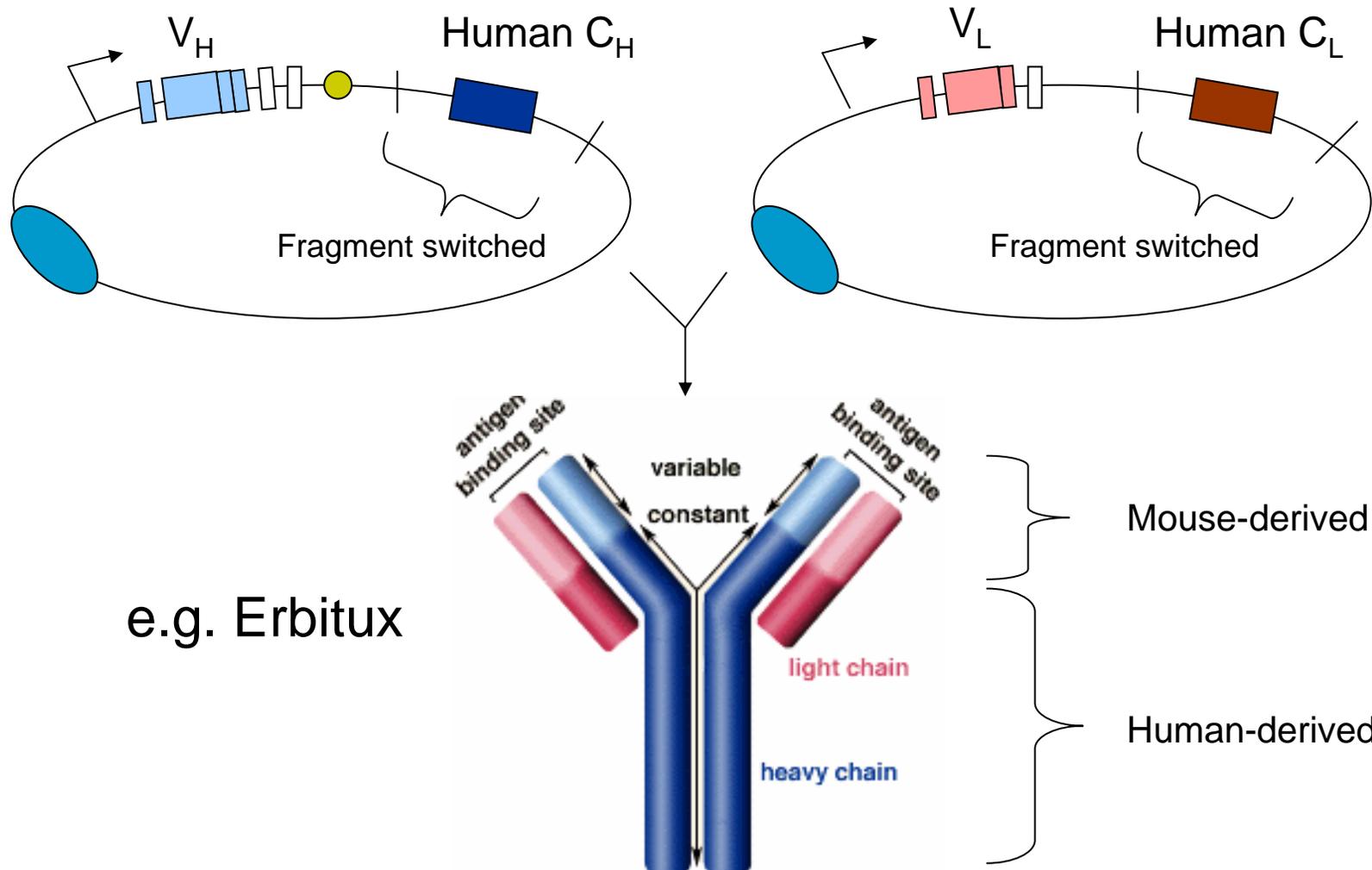
Single vector for high level Ab expression in lymphoid or non-lymphoid cells



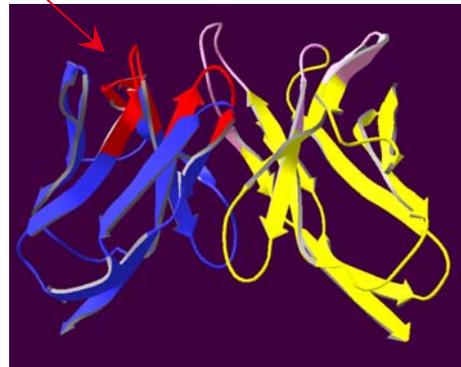
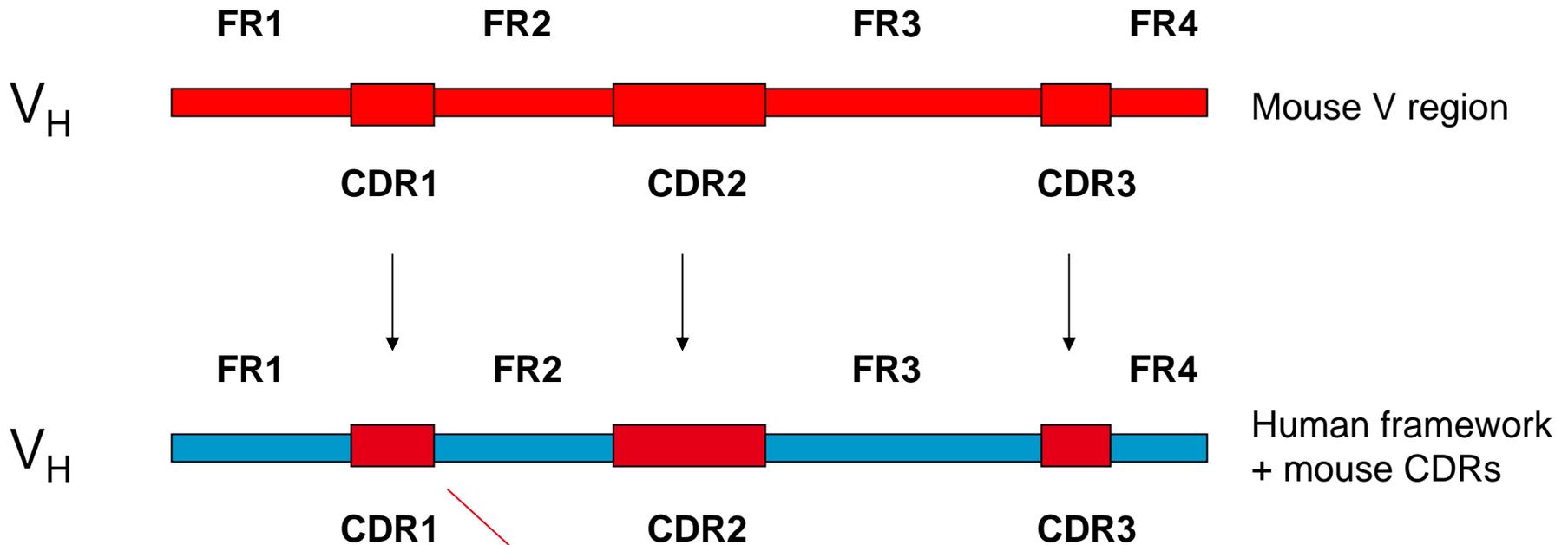
- V_H and V_L cassettes are inserted into pre-designed sites of modular expression vector
- Modular design allows for easy isotype or fusion protein switching
- Promoter/enhancers optimized for multiple cell types
- High producers obtained by gene amplification or forced selection of optimal integration



Chimeric Mouse-human antibodies



V Region “humanization”



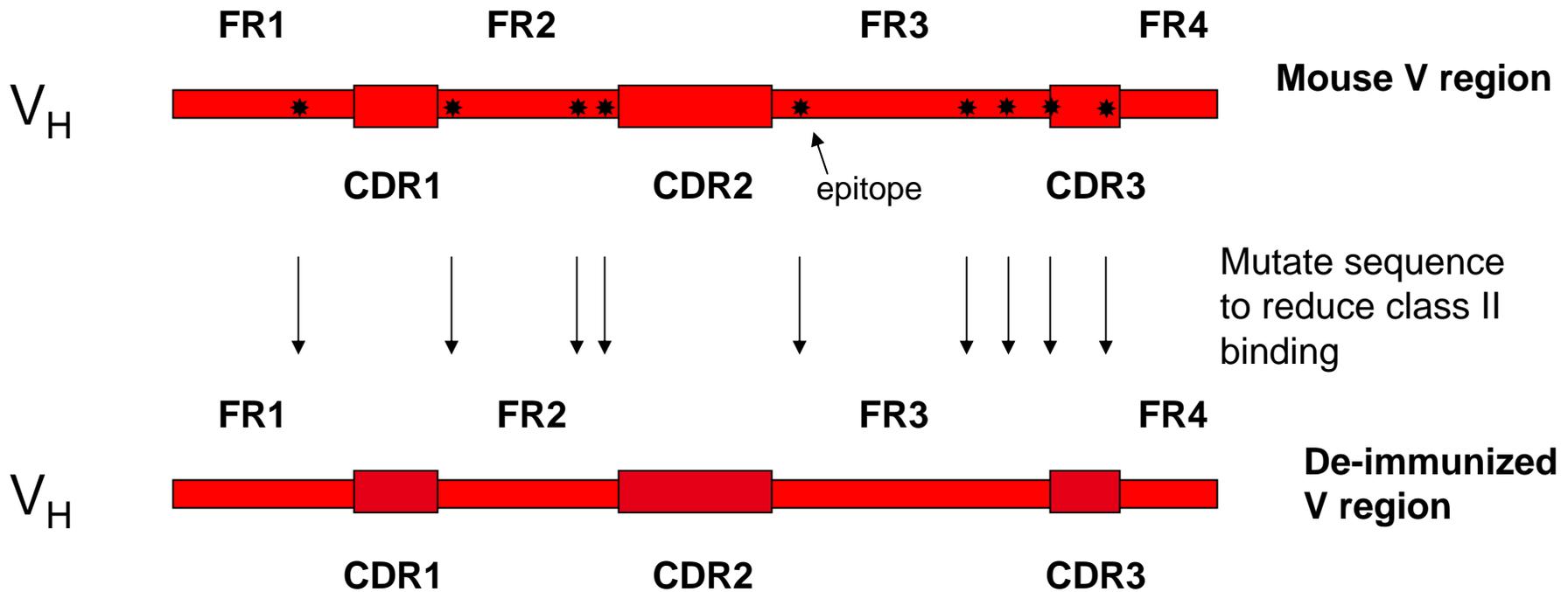
This approach has worked but often leads to mis-folding of CDRs requiring additional FR back-mutations

Is “humanization” what we want

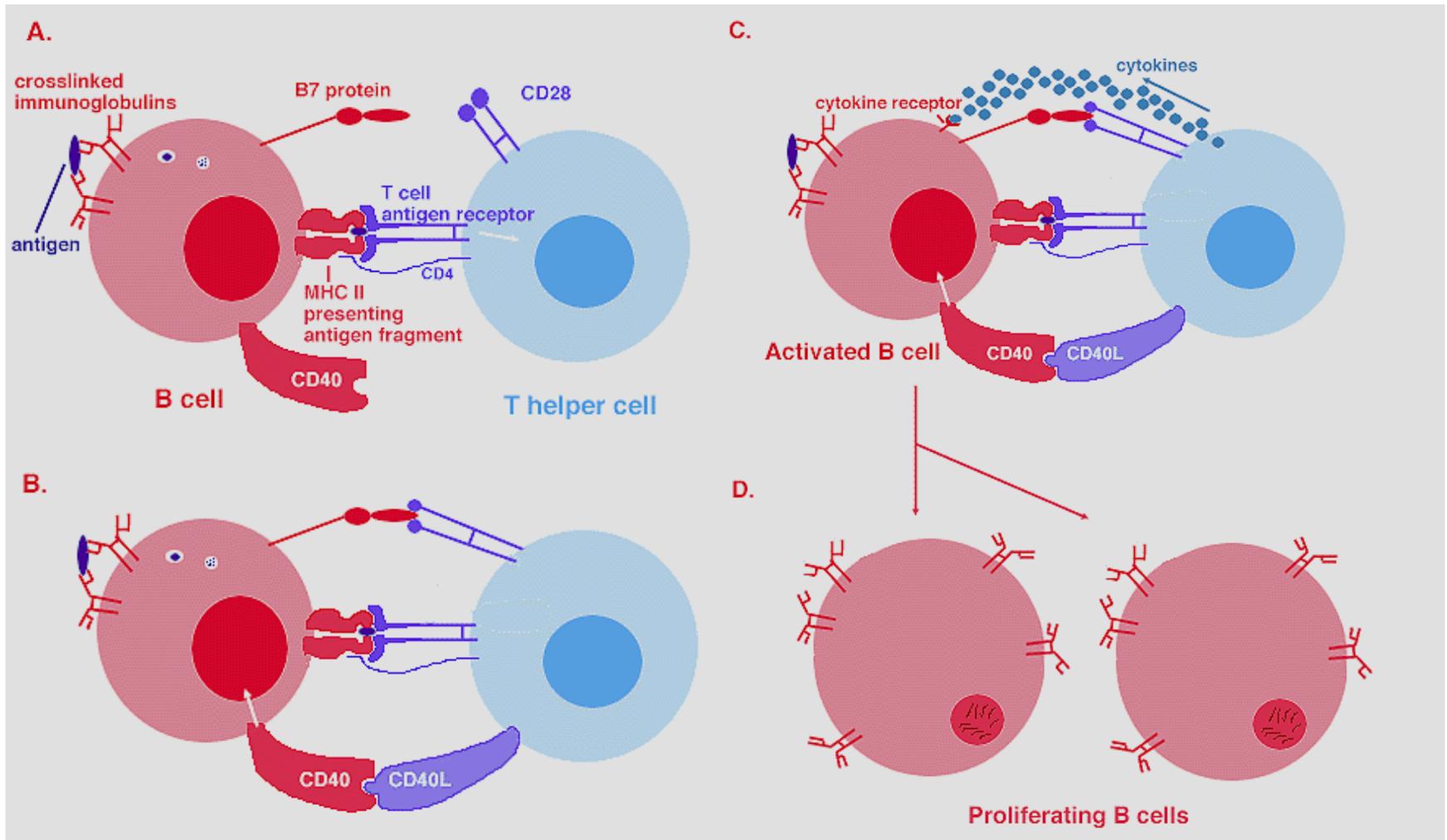


- No – we want the protein to be non-immunogenic
- Humanized and even human V regions can contain T cell epitopes that differ from the germline (via somatic mutation or recombination events)
- What is “self” for a V region sequence donor can be “non-self” for the general population

V Region “de-immunization”



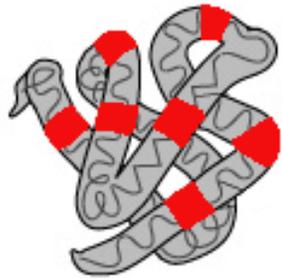
Antibody production requires MHC Class II antigen presentation



Protein Deimmunization Based Upon *in silico* Prediction Of T-cell Epitopes



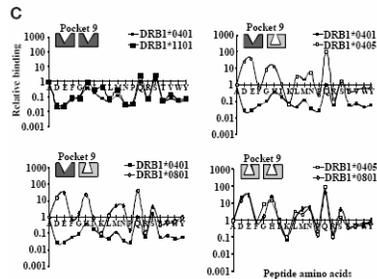
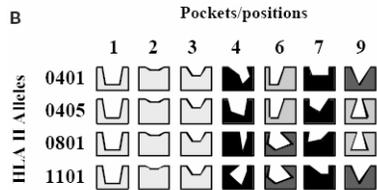
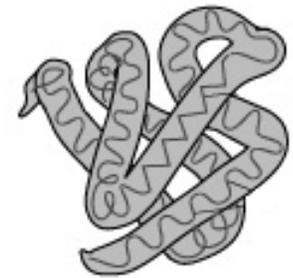
Immunogenic Protein



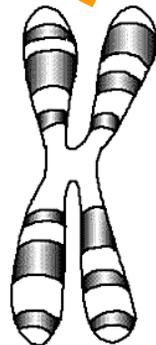
Lexitope 1.0
 By Gordon Webster (gwebster@emdlexigen.com)
 (c) 2004, EMD Lexigen Research Center



Deimmunized Protein



HLA Binding Matrices



Germlines

BLAST Alignments

```

        . . . . . 20 . . . . . 30 . . . . . 40 . . . . . 50 . . . . .
        -X A Q I T G R P E T L L A L C T A L M C L C T L Y F L W K C M C V S D P D E
        ----- C I G T L L M L I C T F Y F I A R C G V T D K K E
        ----- L M L C T A C M F L C M L Y F I A R C G V T D K R I
        A D L L G D C R P E T L L G I C T L L M L I C T F Y F I W K C M C V T D K E E
        S Q A Q I T G R P E T L L A L C T A L M C L C T L Y F L W K C M C V S D P D E
        - D L L G D C R P E T L L G I C T L L M L I C T F Y F L W K C M C V T D K D E
        C M S T Y V P G G E S I F L W V G T A C M F L C M L Y F I A R C G V S D Q R I
        - X A Q I T G R P E T L L A L C T A L M C L C T L Y F L W K C M C V S D P D E
        - M P A P E C E A I L L L C T A C M F L C M L Y F I A R C G V E T D S R I
        --- M P E P C S E A I L L M L C T A C M F L C M L Y F I A R C G E T D S R I
    
```

BLAST Alignments

	A	C	D	E	F	G	H
A	4	0	-2	-1	-2	0	-2
C	0	9	-3	-4	-2	-3	-3
D	-2	-3	6	2	-3	-1	-1
E	-1	-4	2	5	-3	-2	0
F	-2	-2	-3	-3	6	-3	
G	0	-3	-1	-2	-3		
H	-2	-3	-1	0			

BLOSUM 62

Substitution Matrices

Artificial Ab libraries



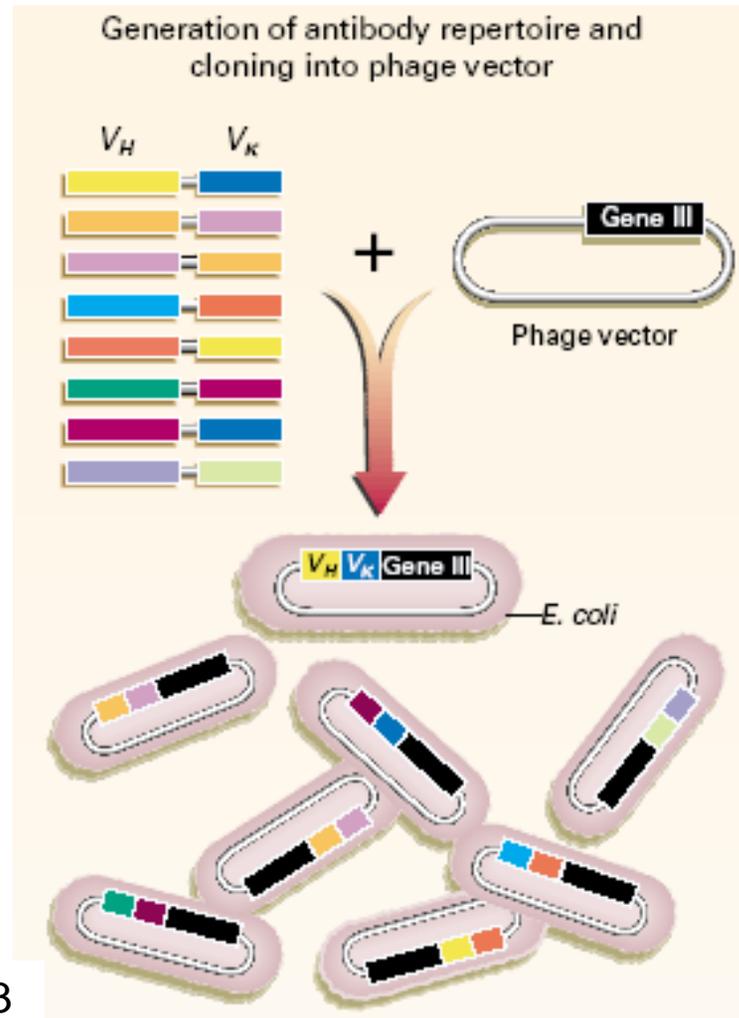
- **Phage-display methods (pros)**
 - Allow for rapid screening of huge numbers (high diversity)
 - Are not biased against “self” binders due to human-mouse homology
 - Can be used for affinity maturation
 - Can be used to isolate human antibody V region binders
- **Phage-display methods (cons)**
 - Lack of in vivo editing for cross-reactivity
 - May still contain immunogenic epitopes
 - Costly to commercialize

Phage library construction



Different sources of V gene input:

- cDNA from immune or naïve individual
- germline V regions



Therapeutic Antibody Approaches



- Whole Antibodies (chimeric, human, de-immunized, etc.)
 - Antagonists
 - Anti-EGFR, CTLA4
 - Agonists
 - Anti-CD40
 - Anti-virals
 - Neutralizing RSV
 - Depleting immune cells
 - Anti-CD20, CD19, CD22, CD3, CD4

Therapeutic Antibody Approaches



- Antibodies as carriers
 - Radio-immune therapy (Zevalin, Bexxar)
 - Immunotoxins
 - Antibody-drug conjugates
 - Antibody-cytokine fusions
- Bi-specific antibodies
 - Dual specificity (same cell)
 - Cross-linking cells (e.g. anti-T cell / anti-tumor cell)