

Immune monitoring of cancer immunotherapies:

Next generation tools to identify baseline immunocompetence and predictive biomarkers

Sacha Gnjatic, PhD

*Associate Professor, Department of Medicine
Division of Hematology and Medical Oncology
Tisch Cancer Institute and Immunology Institute
Icahn School of Medicine at Mount Sinai, New York NY
Associate Director of the Human Immune Monitoring Center*



**Mount
Sinai**

**Precision
Immunolog
y Institute**

*The Tisch
Cancer
Institute*

Disclosure information: Sacha Gnjjatic, PhD

The following relationships exist:

Immune Design: Research Support

Janssen R&D: Research Support

Agenus: Research Support

Genentech: Research Support

Pfizer: Research Support

Third Rock Ventures/Neon Therapeutics: Consulting Fees (e.g., advisory boards)

B4CC: Consulting Fees (e.g., advisory boards)

OncoMed: Consulting Fees (e.g., advisory boards)

Merck: Consulting Fees (e.g., advisory boards)

Aims of immune monitoring

To find better ways to predict patients who may benefit from immunotherapies, and to design new approaches for those who don't

Just measuring tumor growth and survival in immunotherapy clinical trials leaves too many questions unanswered

Multidisciplinary approach to find molecular, genetic, microbial, or cellular signatures that are useful to select patients for the most appropriate treatment

Explore markers at the tumor site and in the periphery

Learn from immune monitoring of untreated and treated tumors, and their antigenic profile for mechanisms and biomarker discovery

Need: High-dimensional immune monitoring and analysis tools

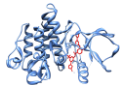
Challenge to find biomarkers for immunotherapy

Small Molecule Targeted Therapies

Tumor cells



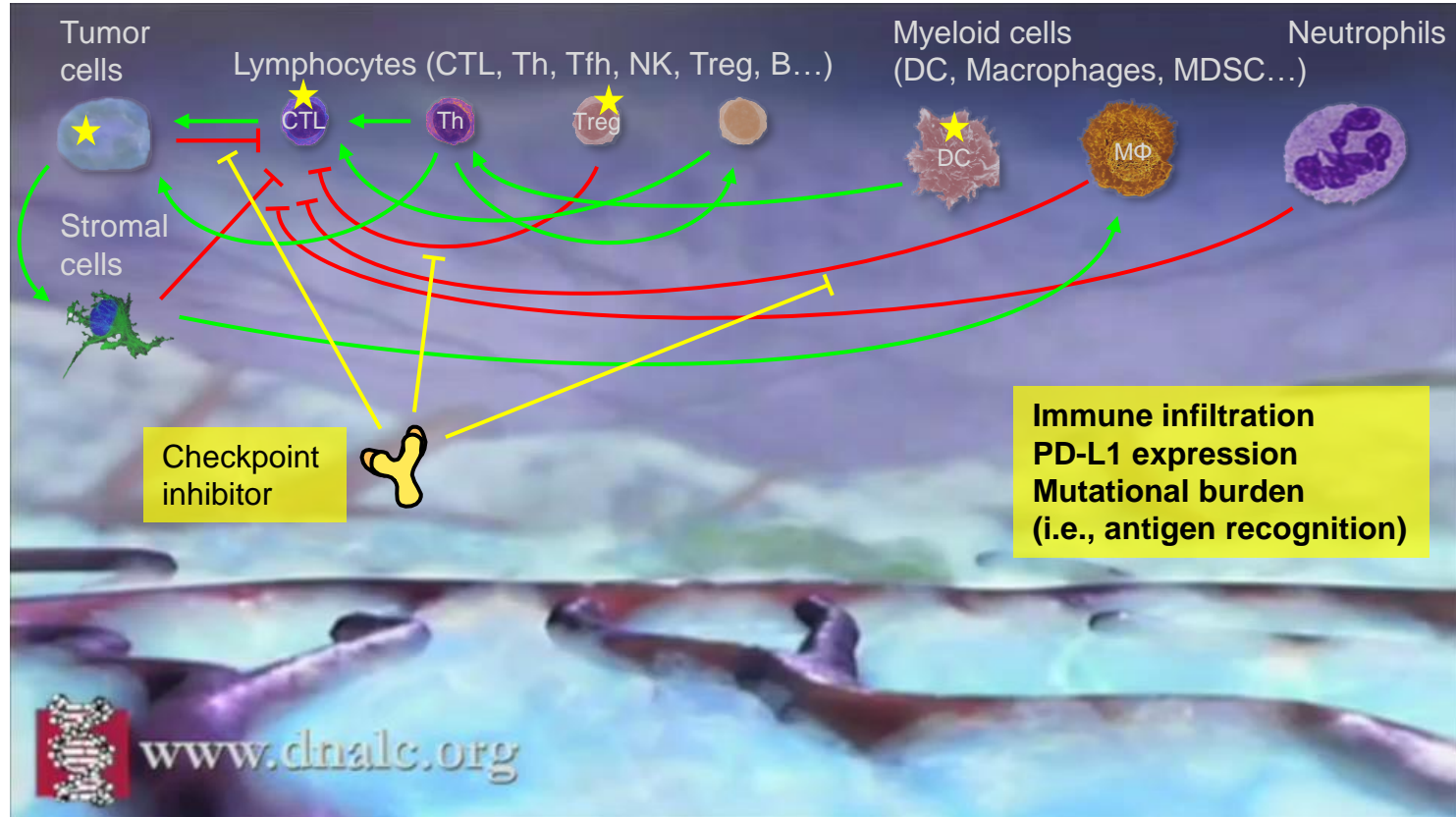
Mutated Kinase...



Kinase Inhibitor



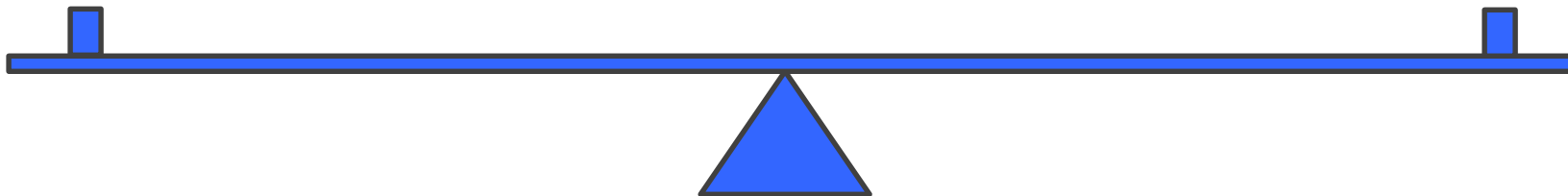
Immunotherapies



The ideal immune monitoring program

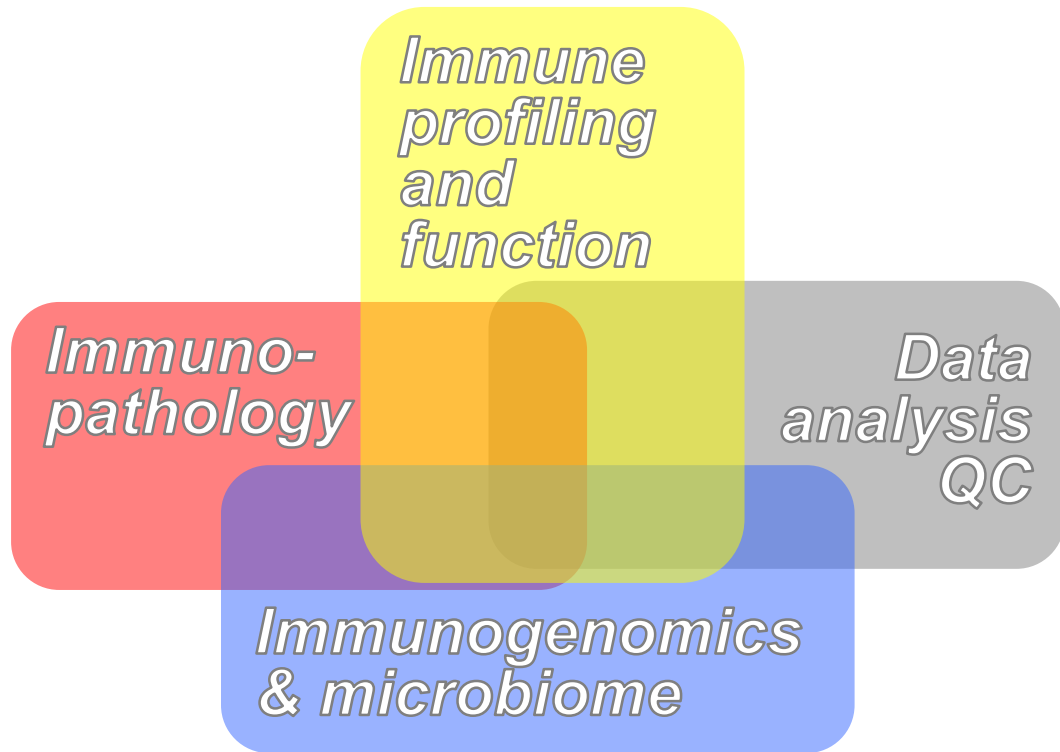
Develop innovative assays to monitor disease-relevant immune signatures and discover new mechanisms, biomarkers and immune targets

Improve assay standardization and minimize experimental variability to maximize data quality and reproducibility



Balancing innovation and standardization.

A multidisciplinary high-dimensional approach



All using analytically validated assays and procedures

In situ (tumor site)

Periphery

Stool

Microbiome
sequencing

P	A
T	C

Biopsy tumor / normal

Surgical material

Fine needle aspirate
Ascites

Single cell
suspension

PBMC

CYTOF mass cytometry
Flow cytometry

P	A
T	C
P	A
T	C

Immune composition

Multiplex IHC/MICSSS
Immunofluorescence

P	A
T	C
P	A
T	C

Immunopathology

WES / RNAseq
TCR sequencing
Nanostring / RT-PCR
Single cell sequencing
BCR sequencing / CTC

P	A
T	C
P	A
T	C
P	A
T	C
P	A
T	C

Immunogenomics

Sorted
cells or
whole
blood

IVS / ELISPOT
ICS (flow / CYTOF)
Tetramers
Phospho-CYTOF/flow
Epitope definition & mapping
NK cell assay / cytotoxicity
Suppression / prolif. assay
CD40L guided T cell sorting
Magnetic / flow cell sorting

P	A
T	C
P	A
T	C
P	A
T	C
P	A
T	C
P	A
T	C
P	A
T	C

Immune cell function

Serum / Plasma
Urine / Ascites

ELISA (grand serology)
Seromics (antibody profiling)
Soluble analytes (multiplex)

P	A
T	C
P	A
T	C
P	A
T	C

Humoral immune profile

Culture
supernatants

Priority

P	A
T	C

Amounts
needed

Throughput

Cost

high
low

Example of budget prioritization plan for assays

CITN-09

Pembrolizumab in Merkel Cell Carcinoma.

Assay and cost rundown per patient.

Priority

#6

#2

#12

#7

#4

#5

#1

Sample reception, barcoding, storage, management, realiquoting post-assay	\$
Serum antibodies to MCPyV	
ELISA (3 time points)	\$
Serum antibody profiling for tumor specificity	
Grand Serology (3 time points)	\$\$
Seromics (subset only, 50%)	\$\$\$
Soluble protein analytes, including Flt3L	
O-Link (3 time points)	\$
Phenotyping of biopsies and of peripheral blood	
CyTOF of tissue (2 time points)	\$\$
CyTOF of blood (3 time points)	\$\$
Tissue multiplex IHC from biopsies	
MICSSS (2 time points)	\$\$

Priority

#3

#8

#9

#10

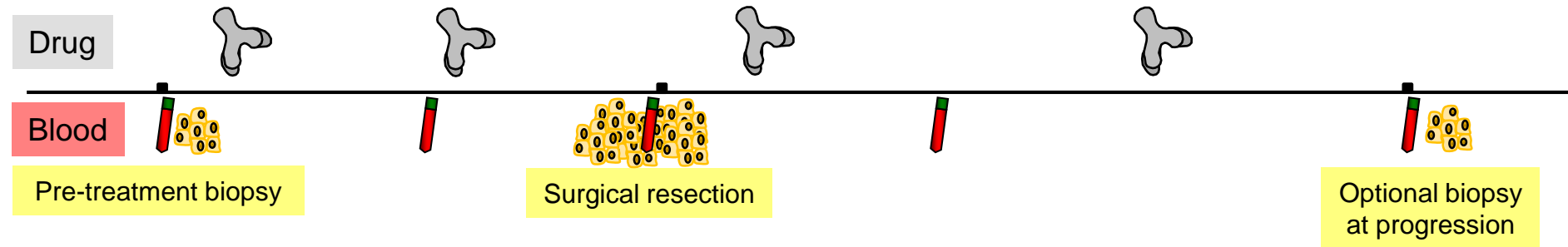
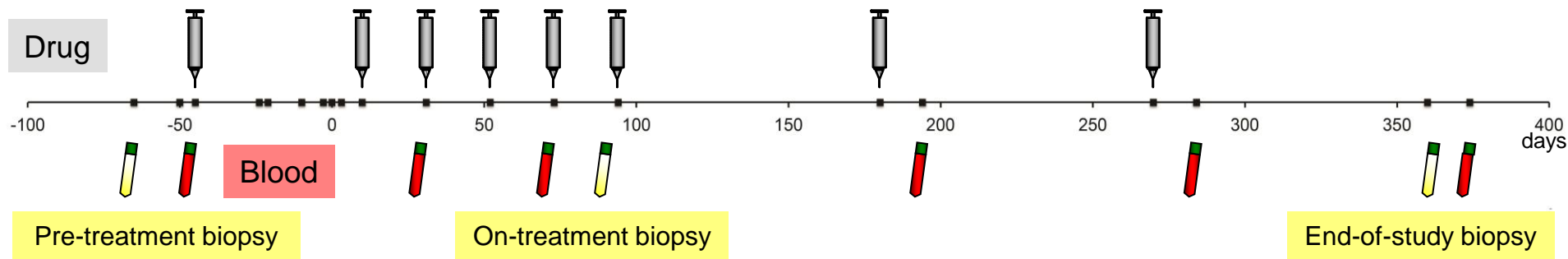
#14

#11

#13

Tumor gene expression from biopsies	
Nanostrings (2 time points)	\$
Tumor mutational profile and neoepitope prediction	
WES / RNAseq (1 time point)	\$\$
Peptides for neoantigen	\$\$
Neoantigen identification of T cell, characterization (priced at 50% of cost if planned only in subset)	
IVS + ELISPOT (2 time points)	\$\$
CD154 sort / tetramer (subset)	\$\$
T cell diversity from biopsies or peripheral blood	
TCRSeq (2 time points)	\$\$\$
Microbiome analyses	
16S sequencing (2 time points)	\$
Data analysis pricing included in assays	
Data management, storage, sharing	\$

Planning ahead for sample collection



- Blood collected before or after each treatment
- Tumor tissue biopsies or surgical material collected throughout treatment
- Clinical annotations

Areas of focus

Immune microenvironment by multiplex immunohistochemistry

Phenotyping by CYTOF mass cytometry

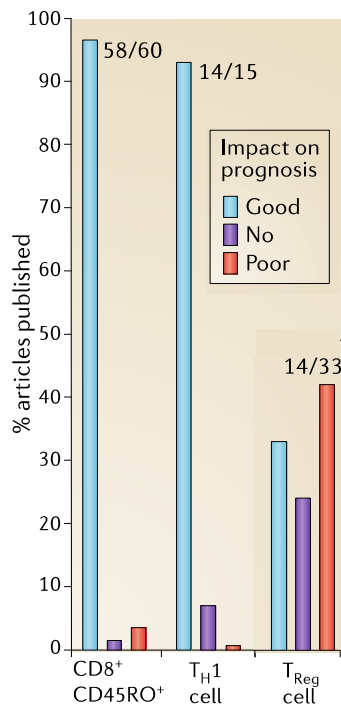
Immunosupportive role of microbiome composition

Defining antigen specificity and quality (neoantigens, seromics)

Modeling, integration of data, and automated analyses pipelines

T cell tumor infiltration as a prognostic marker in various tumors and a predictive biomarker of PD-1 response in melanoma

Meta-analysis of 124 articles (20 cancer types)

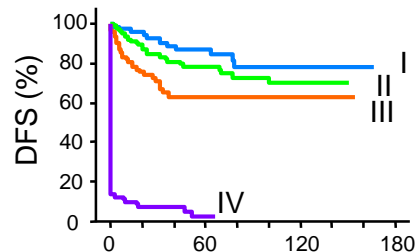


The immune contexture of the tumor influences prognosis

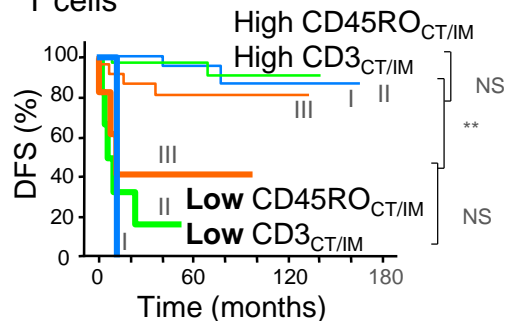
Fridman et al., *Nature Rev. Cancer*, 2012

Immunoscore (colorectal cancer)

TNM classification

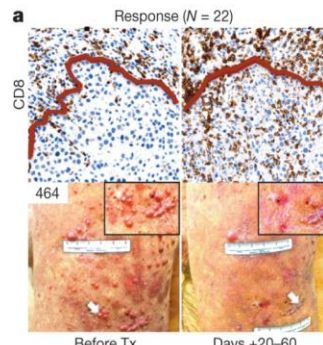


T cells

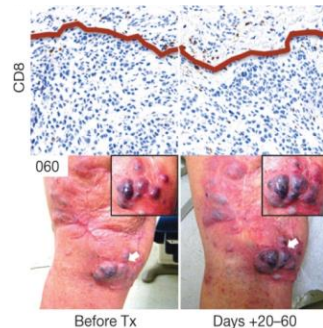


Galon et al., *Science*, 2006;
Pagès et al., *J Clin Oncol*, 2009

Responders (n=22)



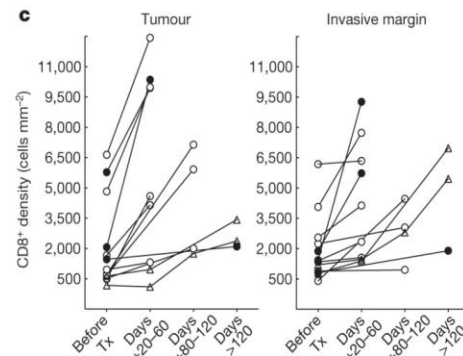
Progressors (n=24)



CD8 T cell infiltration before and during pembrolizumab in advanced melanoma.

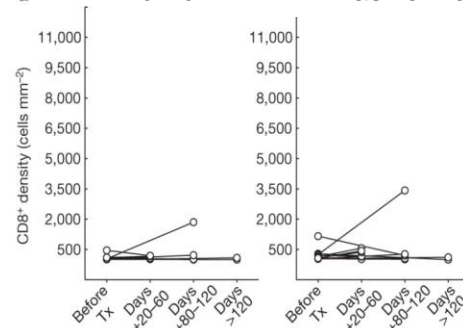
Tumor

Invasive margin



Tumor

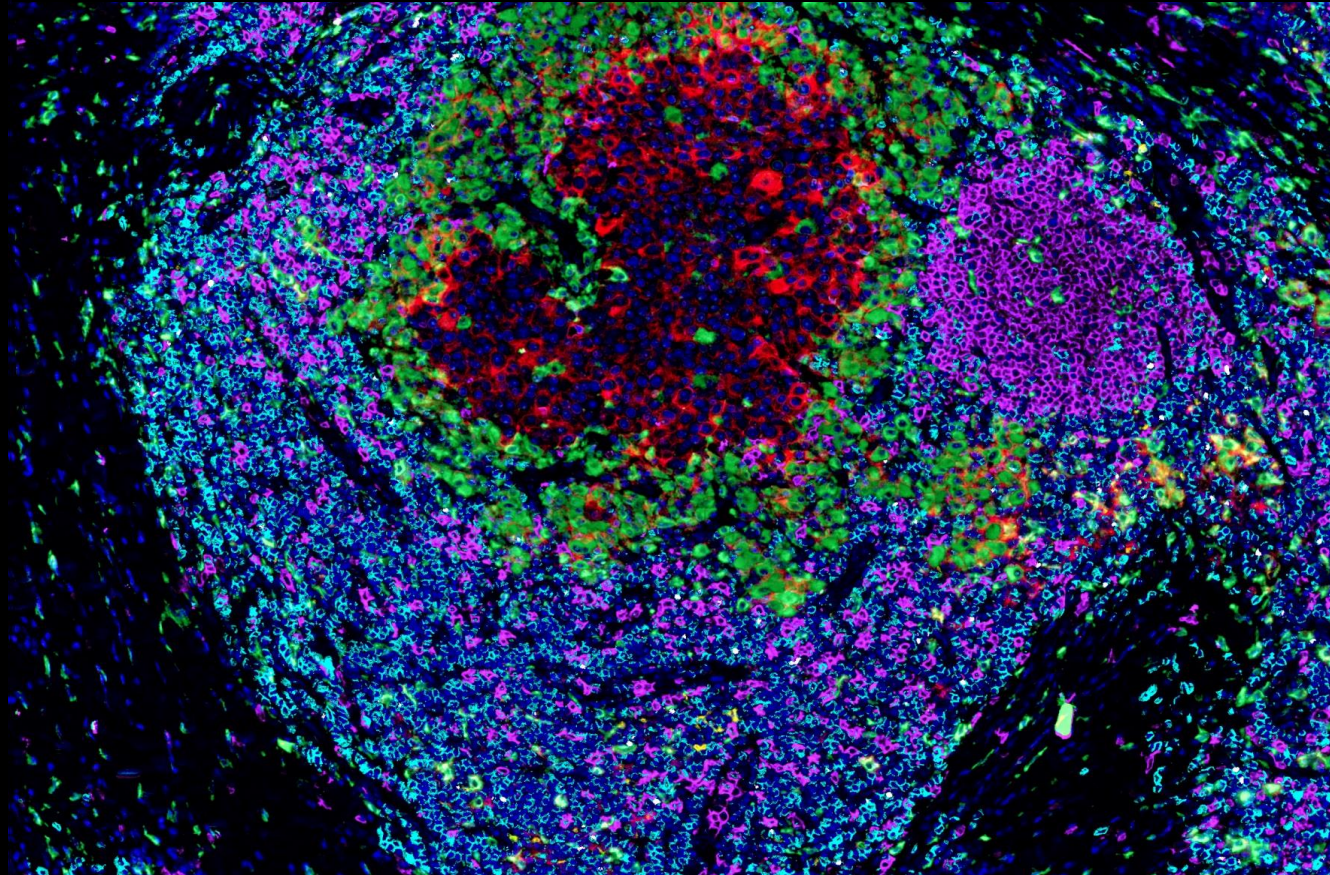
Invasive margin



PC Tumeh et al. *Nature* 515, 568-71 (2014)

Applying multiplex IHC to query effect of checkpoint blockade

Hematoxylin/PD-L1/CD68/DC-LAMP/CD20/CD3/FoxP3



Melanoma lesion biopsied after CTLA-4 treatment (1 FFPE slide)

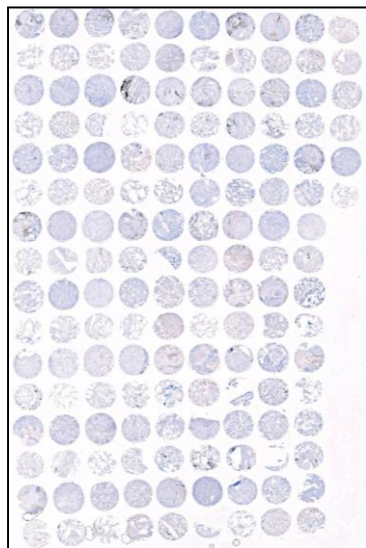
x100

Remark R, Merghoub T,
Grabe N, Litjens G,
Damotte D, Wolchok
JD, Merad M, Gnjatic S.
Science Immunology;
1:aaf6925 (2016).

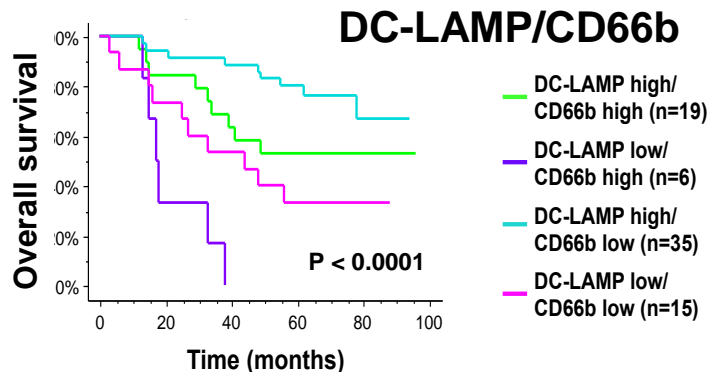
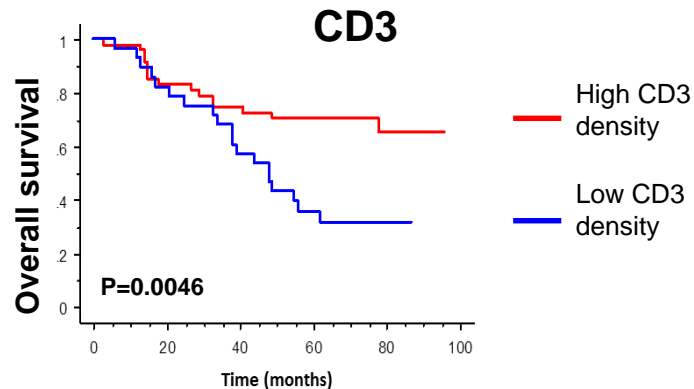
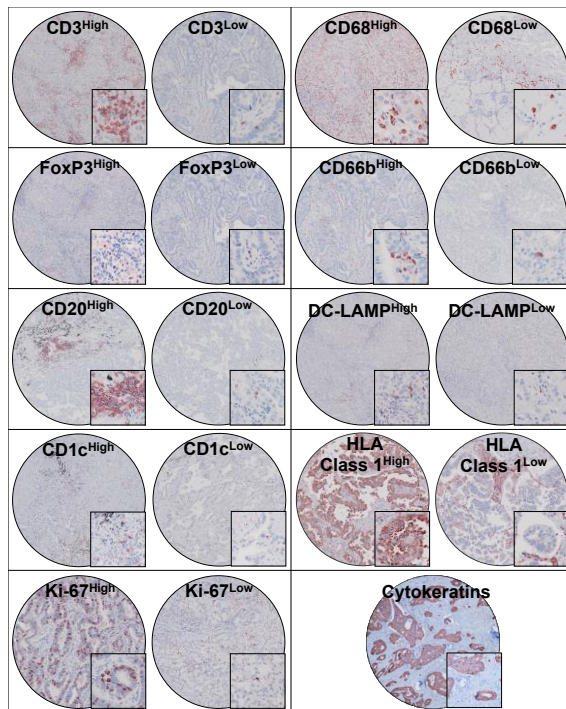
Multiplex IHC on tissue microarrays to identify prognostic biomarkers

Heterogeneity of immune markers in non-small cell lung cancer (NSCLC)

NSCLC tissue microarray



n=75 patients

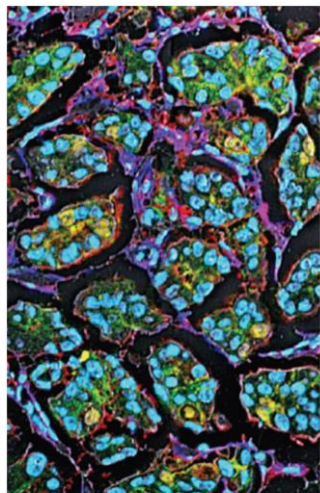
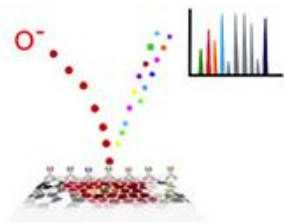


Remark R, Merghoub T, Grabe N, Litjens G, Damotte D, Wolchok JD, Merad M, Gnjatich S. *Science Immunology*; 1:aaf6925 (2016).

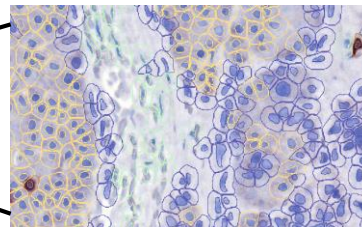
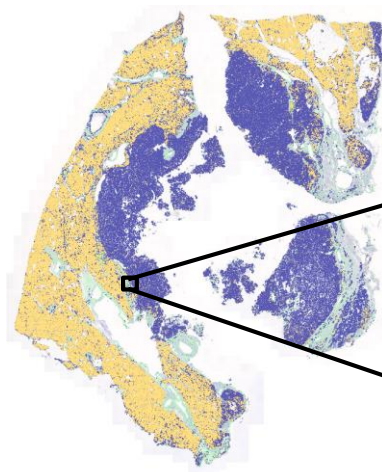
Review on NSCLC immune contexture in *Am J Respir Crit Care Med*. 2015;191:377-90

Next frontier in tissue imaging: higher multiplexing, 3D-4D analyses, and neural network image learning

More than 40 markers using metal-conjugated antibodies
And an ion-beam or CYTOF for mass cytometry analysis



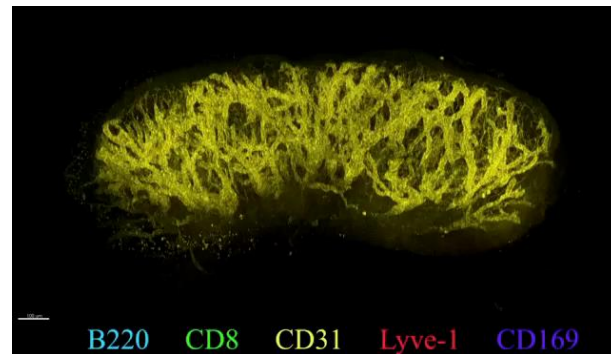
From Stanford.edu



Normal pancreas
Stroma
Neuroendocrine pancreatic tumor
CD3 lymphocytes

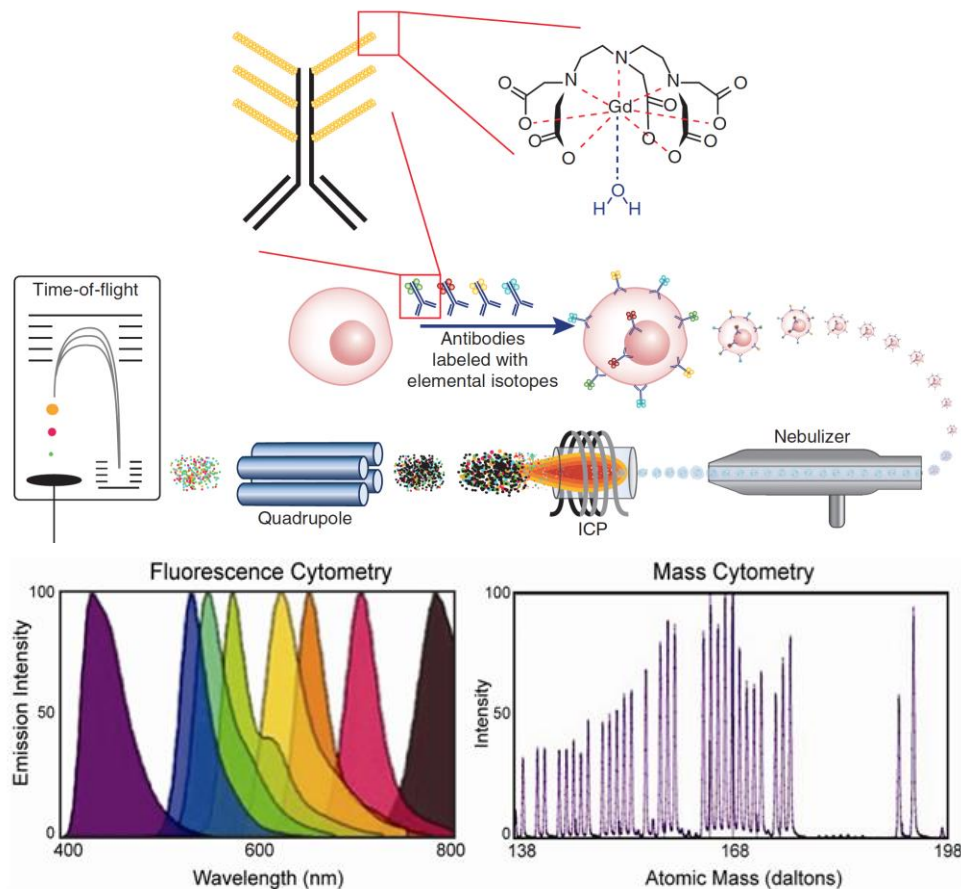
Gnjatic et al. Unpublished. With QuPath software
(<http://biorxiv.org/content/early/2017/01/12/099796>)

3D imaging of lymph node

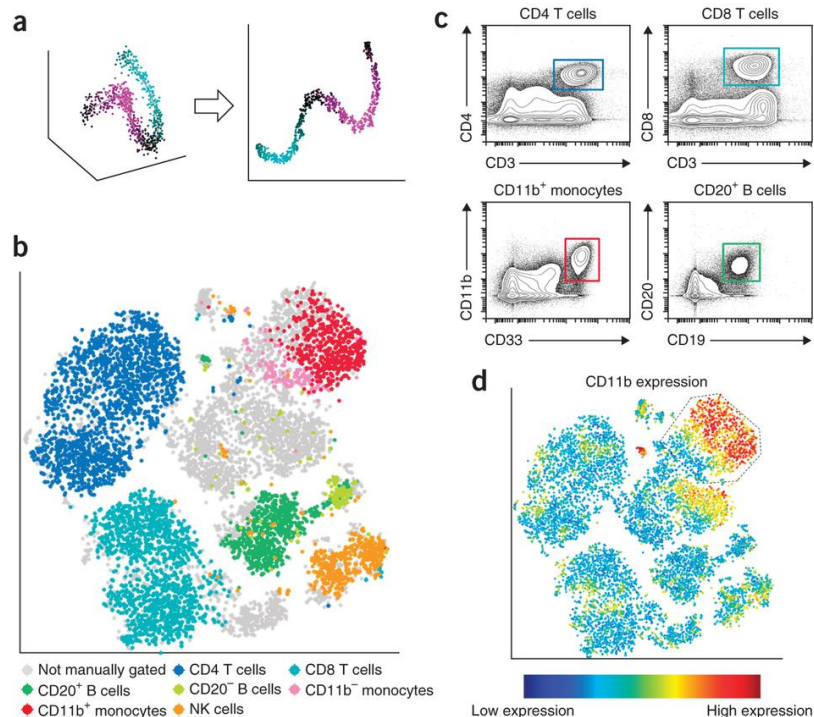


Li, Germain, Gerner. PNAS 2017; 114:E7321

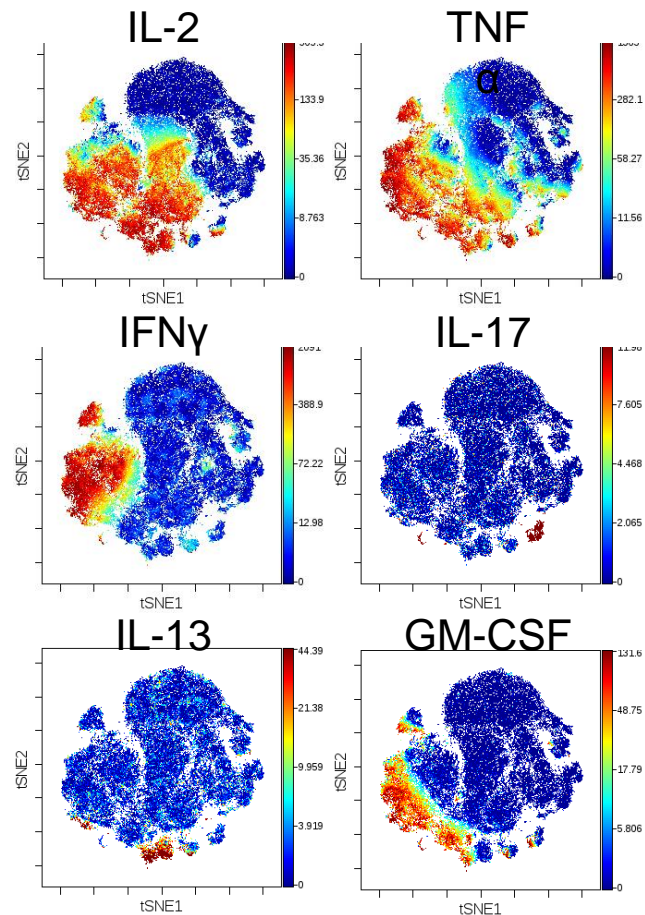
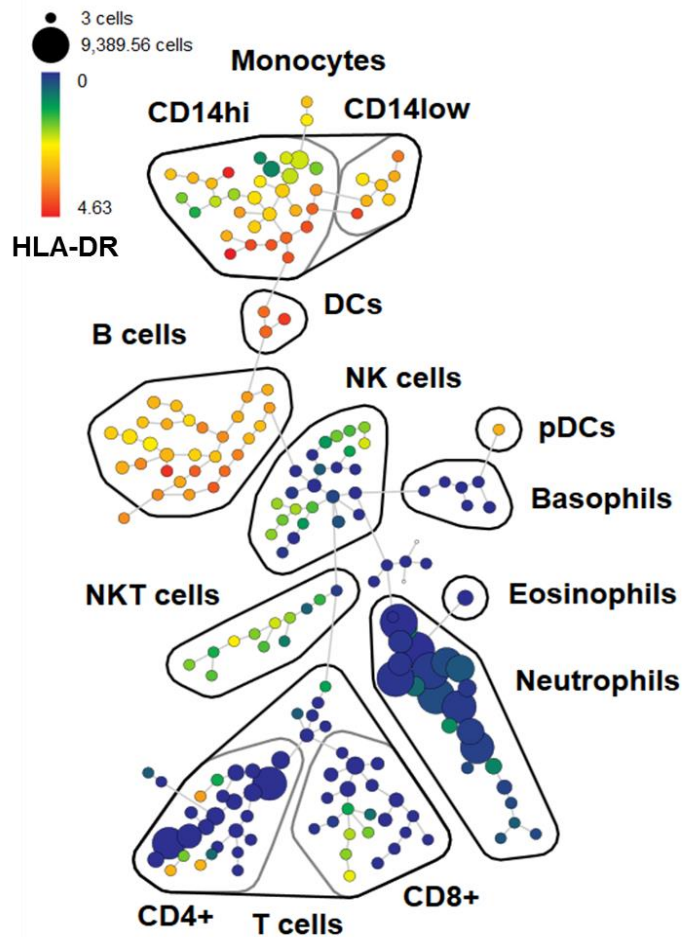
Mass cytometry (CyTOF) to explore phenotypic and functional composition in high dimensions



35 markers can be routinely analyzed using dimensionality-reduction algorithm (viSNE, etc.)

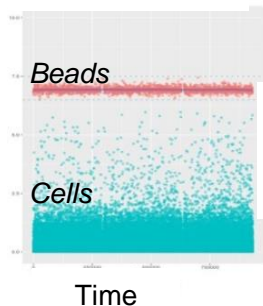


Functional applications of CyTOF mass cytometry

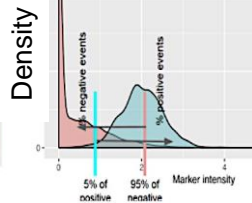


Optimization of CyTOF with beads and lyophilized panels

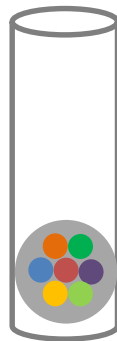
Beads for QC



Average overlap frequency (AOF) between channels



Core set of consensus markers to identify major immune cell subsets



Core Panel

CD45	CD19
CD3	CD27
CD4	CD56
CD8	CD16
CD45RA	CD14
CD38	CD123
HLADR	CD11c



Additional custom conventional liquid antibodies

Optimized panels to provide detailed characterization of specific subsets

T cell module

CD161	ICOS
CD57	OX40
TIM3	CD39
CD103	CD73
CCR7	PD-1
CXCR3	CD25
CCR6	CD127
CCR4	CD69
CXCR5	CD28
41BB	TCRgd
2B4	CD44

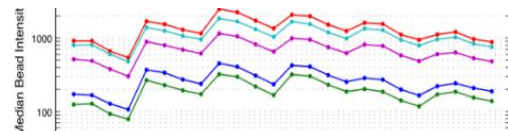
Myeloid module

CD33	CX3CR1
CD64	CD85j
CD1c	CD141
CD66b	CD11b
CD163	CD86
CD206	CD40
CD169	CD117
CD15	PDL1
CD141	PDL2
TLR2	TLR4
CCR7	SIRPa

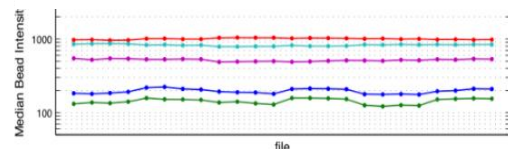
NK cell module

NKp30	CD244
NKp44	CD103
NKp46	CD69
NKG2A	CD96
NKG2C	CD94
NKG2D	Siglec7
KIR3DL1	DNAM1
KIR2DL3	CD132
CD57	CD25
CD161	TIM3
LILRB1	PD-1

Bead intensity before normalization



Bead intensity after normalization



AOF for all antibody channels



Overall staining quality score

Cross-tissue immune profiling by mass cytometry in NSCLC patients

(Yonit Lavin, Adeeb Rahman, Christian Becker, Sacha Gnjjatic, Miriam Merad, Cell 2017;169:750-765)

Resection tissue
Stage I/II adenocarcinoma

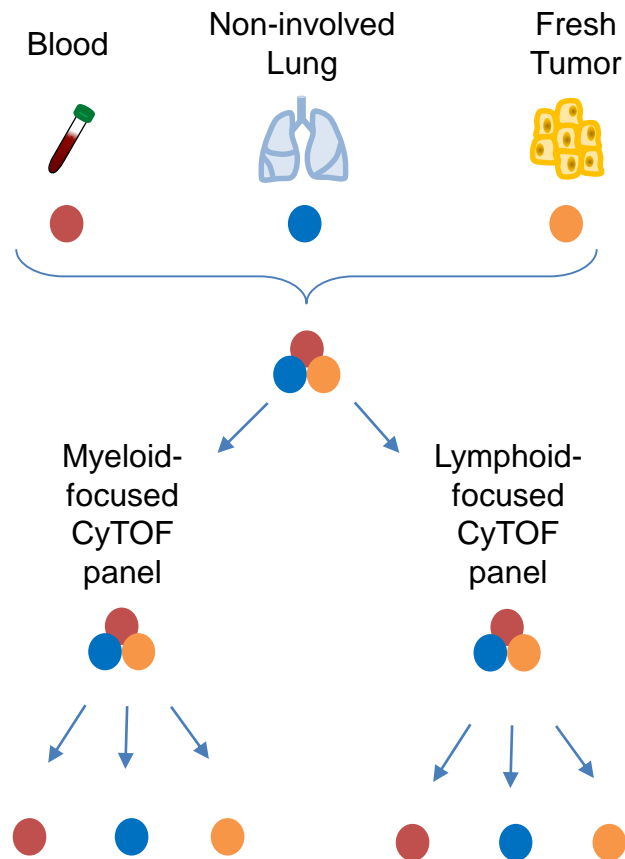
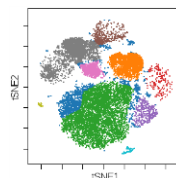
Barcode with tissue-specific CD45 isotopes

Pool

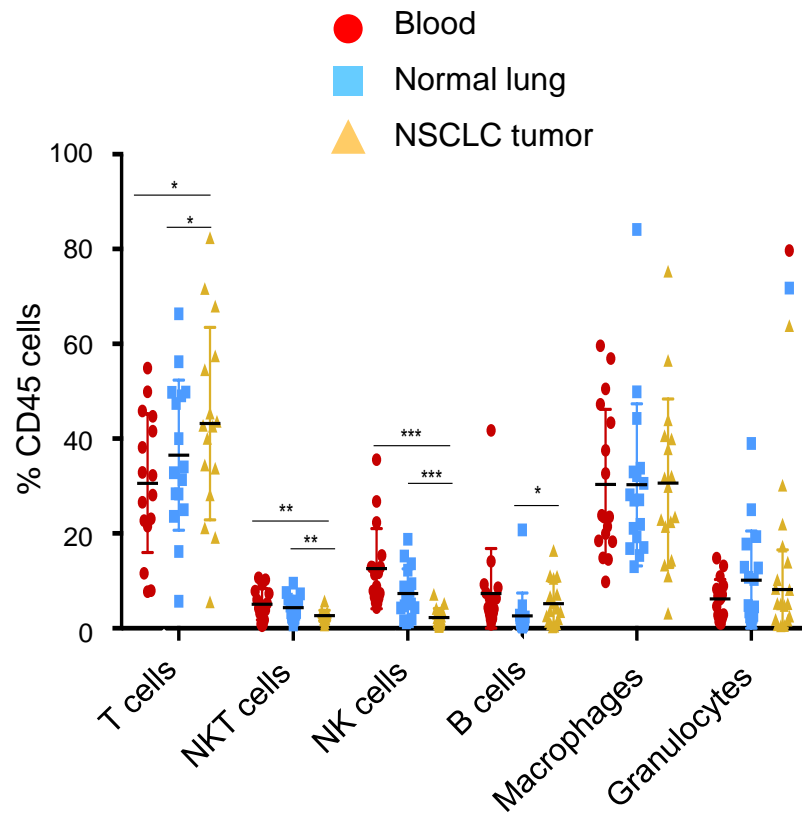
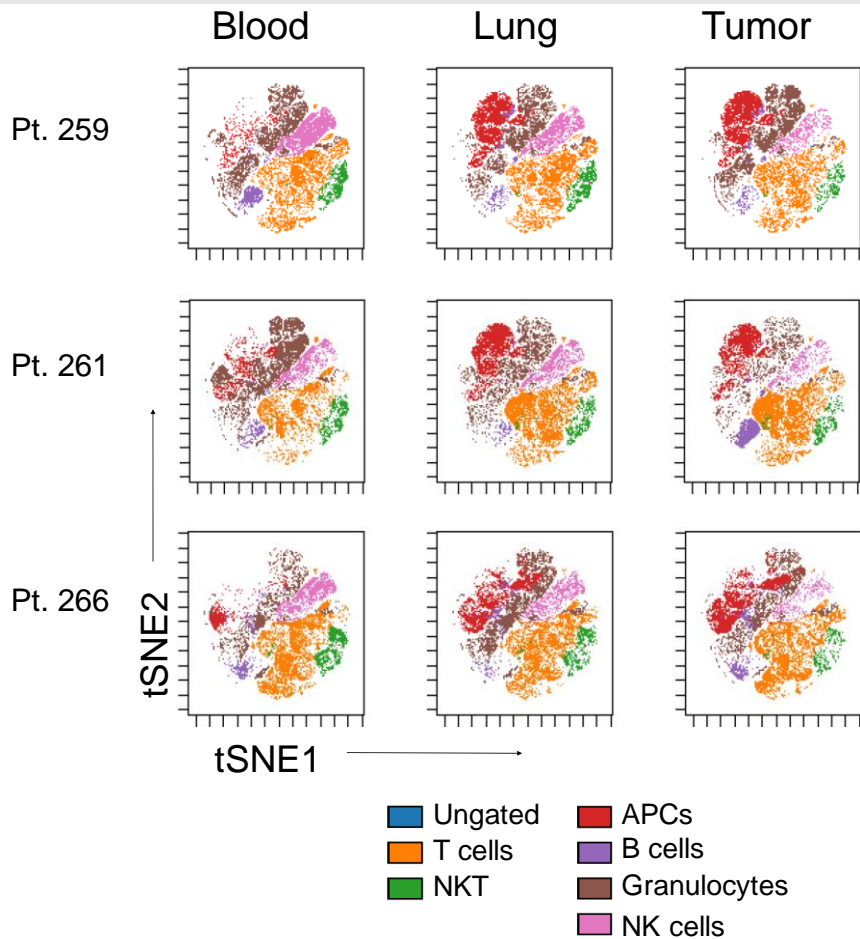
Stain

Acquire on mass cytometry

Deconvolute barcodes and analyze

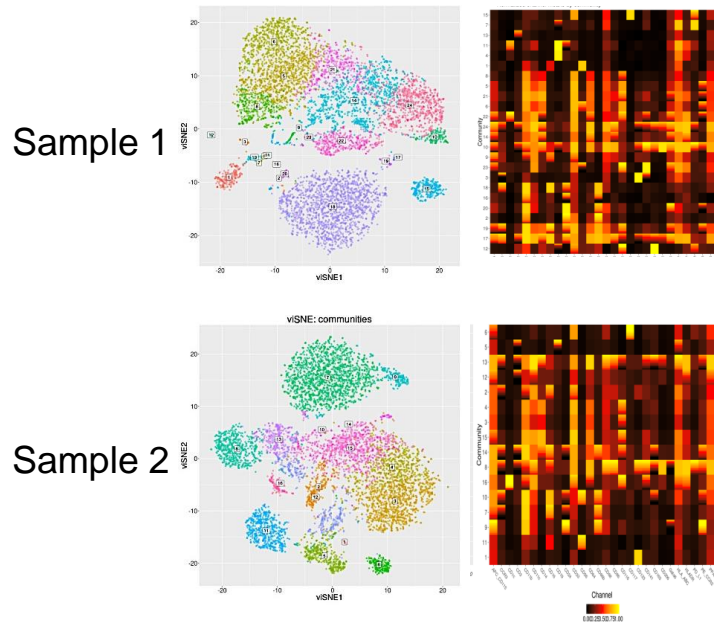


Immune composition of early non-small cell lung carcinoma (NSCLC) by mass cytometry (Lavin, Rahman, Gnjjatic, Merad, Cell 2017;169:750-765)



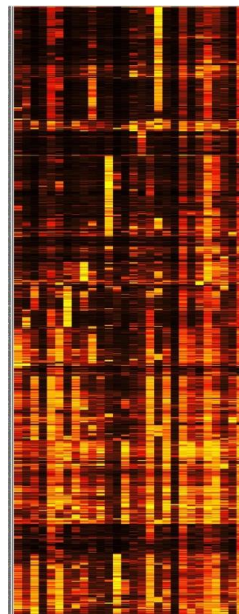
Automated analysis pipeline for CyTOF

Unbiased identification and characterization of cell populations in individual CyTOF samples

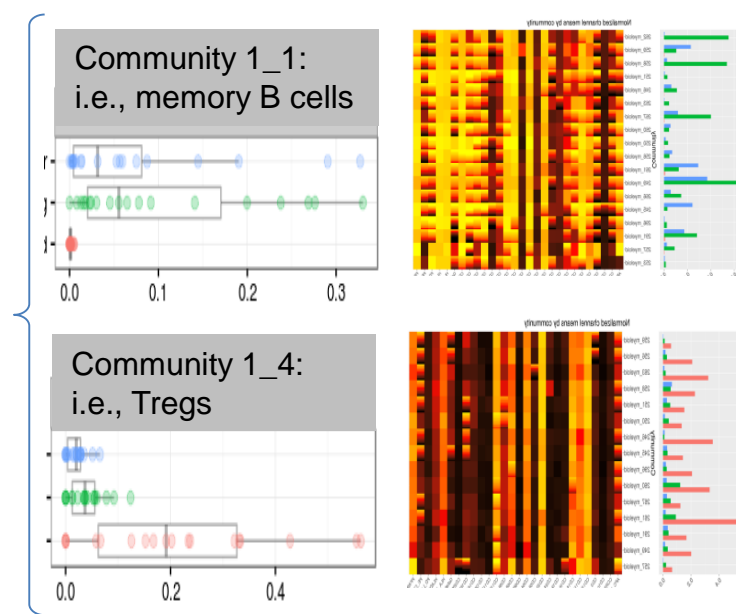


... multiple samples

Automated meta-clustering of populations across multiple samples



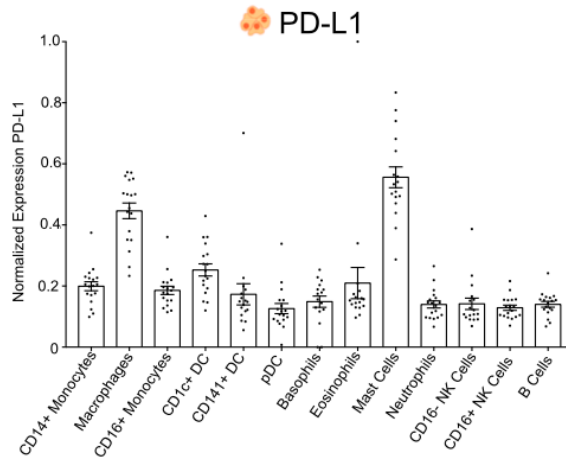
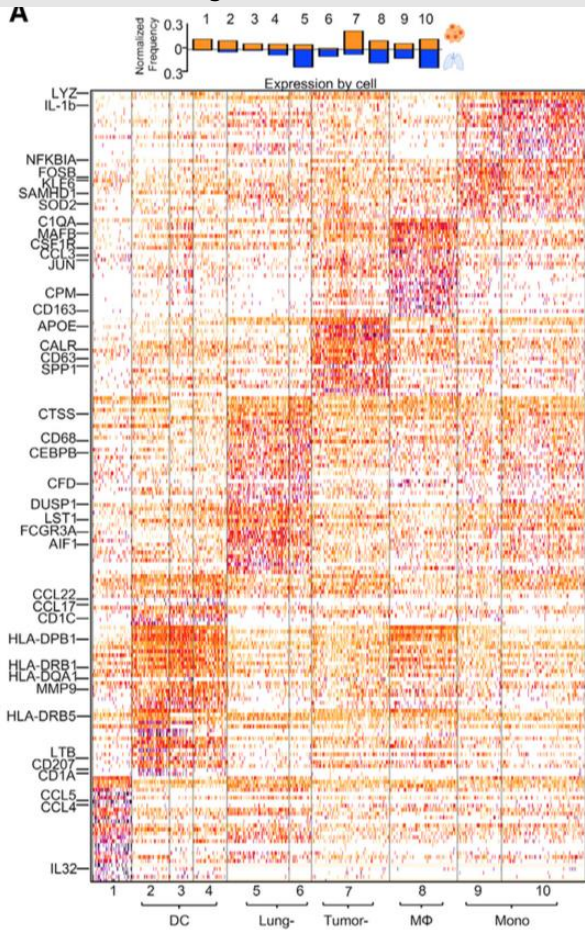
Automated analytics to identify populations and protein expression patterns that differ between treatment groups



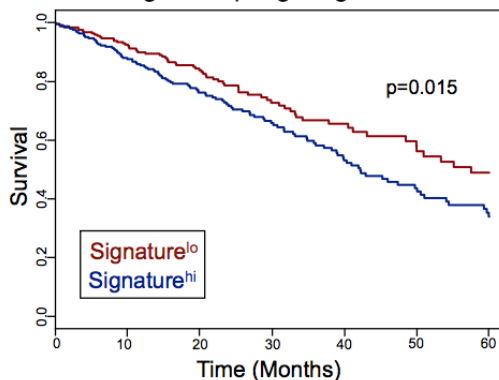
... multiple differing features across samples

Single cell analyses reveal impaired immune profiles at the tumor site vs. adjacent non-involved tissue in early NSCLC

A

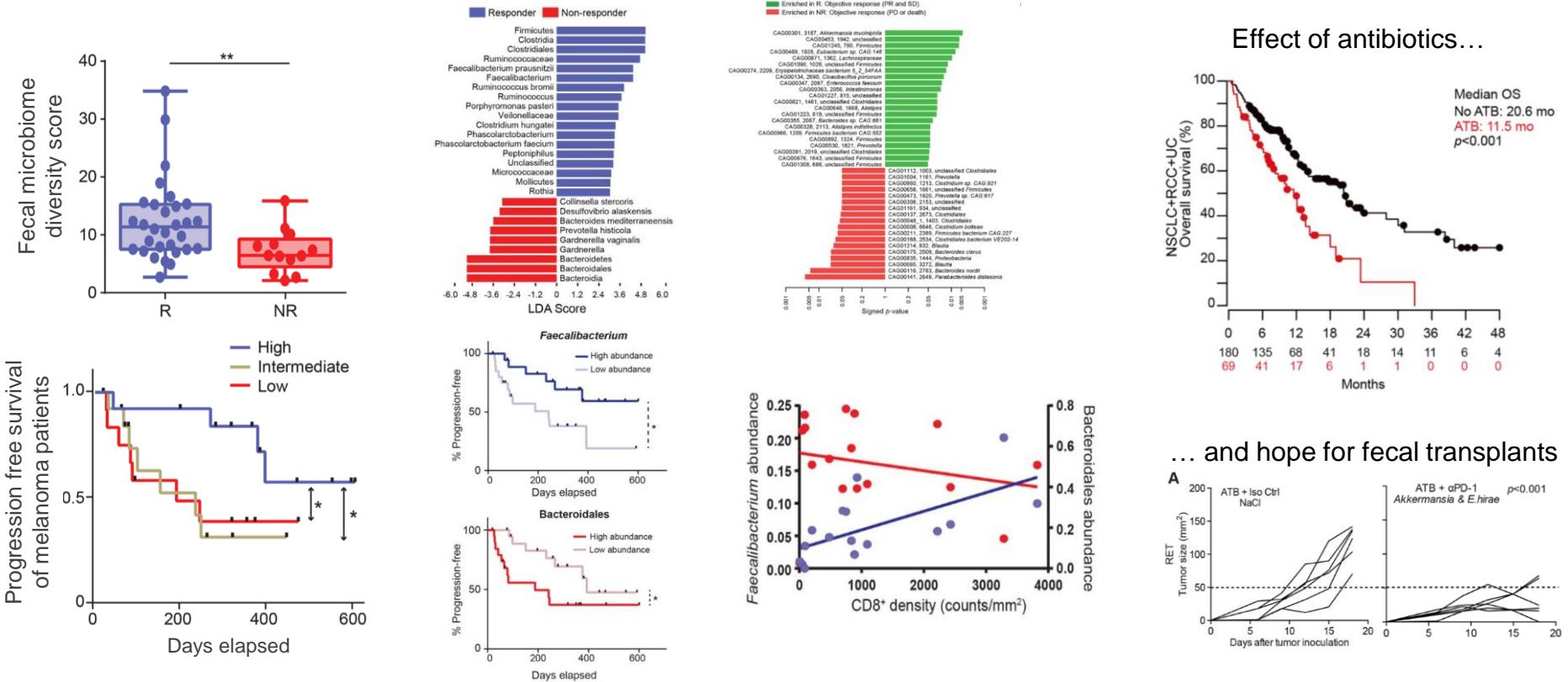


Tumor/Lung Macrophage Signature Survival



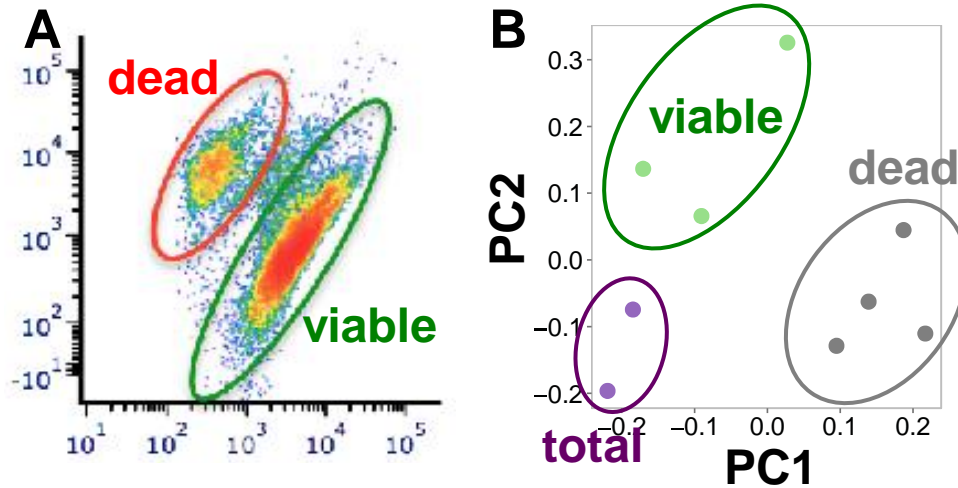
Role of microbiome as a tumor extrinsic factor contributing to immune recognition during immunotherapy (Jenq+Wargo, Kroemer+Zitvogel)

Cancer patients treated with PD-1 blockade, sequenced for gut bacteria by 16S or shotgun metagenomics



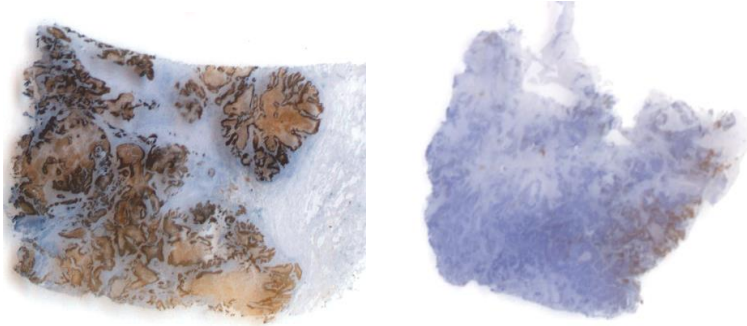
Improving microbiome analyses with shotgun metagenomics and live cell detection (Clemente, Faith)

Importance of distinguishing live from dead bacteria for QC aspects



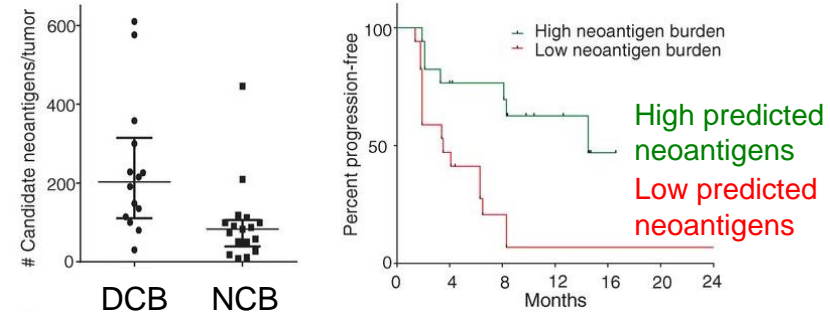
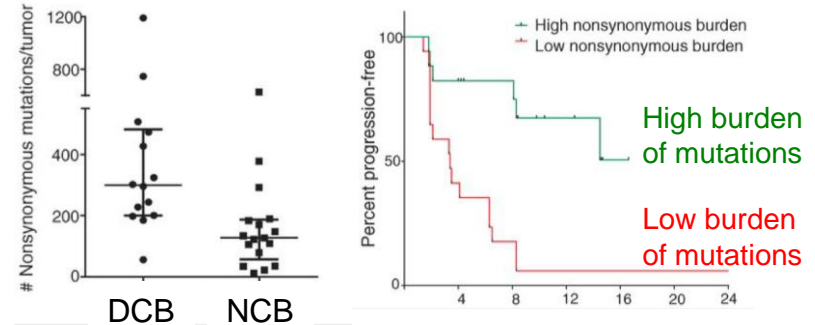
Identifying antigens recognized at the tumor site

- Whole exome sequencing and RNASeq to identify **tumor-specific mutations** that may give rise to neoepitopes, followed by high-throughput tetramer screening or cytokine production of T cells (Schumacher et al)
- Immunohistochemistry and RT-PCR to look for presence of known tumor antigens, such as **cancer/testis antigens**



- **Serological assays** to quickly screen for immunogenic target antigens

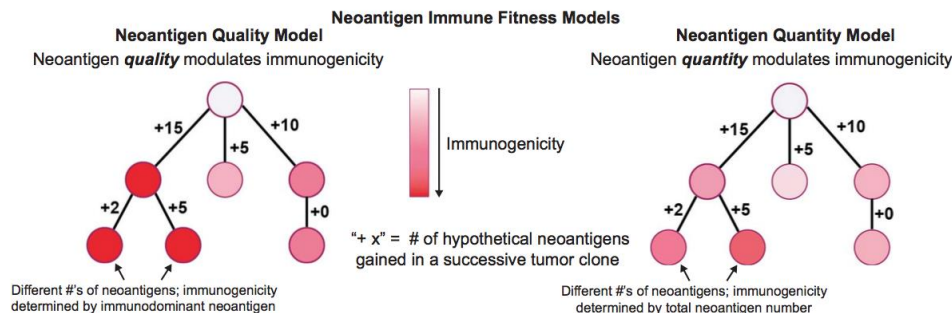
Mutational burden correlates with response to PD-1 blockade in NSCLC



DCB = durable clinical benefit
NDB = no durable clinical benefit

In silico modeling improves prognostic value of neoantigens by assessing their quality

Pancreatic cancer patients, including subset with long-term survival



Quality Score

a) Assess sequence similarity of neoantigen to pathogen

Wild type epitope: PPSAR**G**GPL

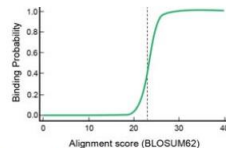
Tumor neoepitope: PPSAR**R**GPL

Human Herpes Virus (HHV)-8: PPSG**R**GPVAFRTRV

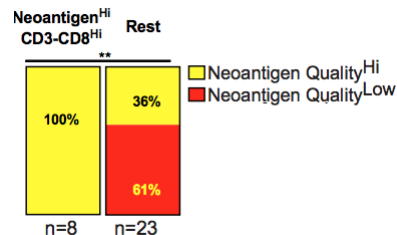
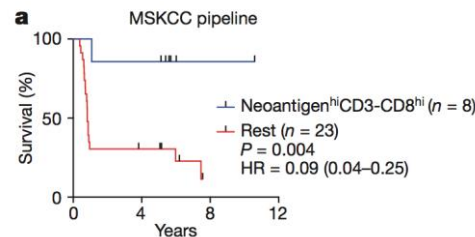
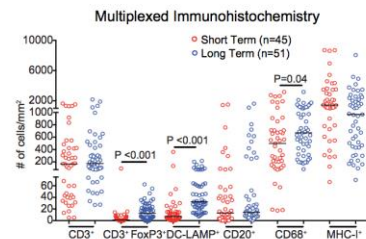
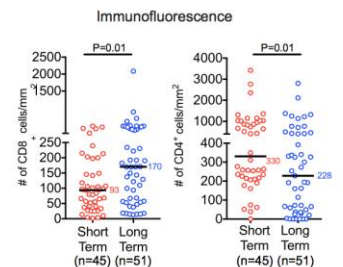
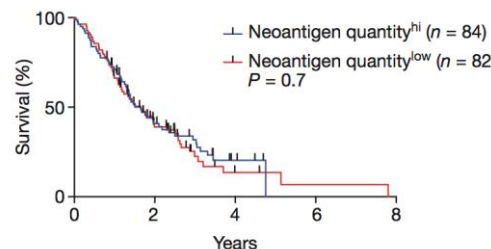
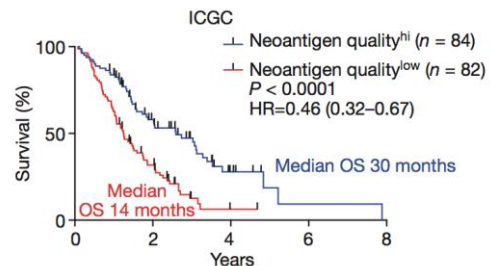
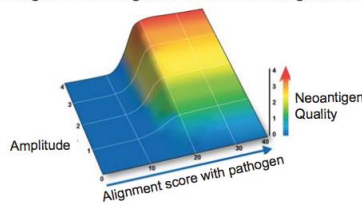
Calculate Alignment score (BLOSUM62)

b) Scale alignment score to binding probability of a TCR to the cross reacting antigen

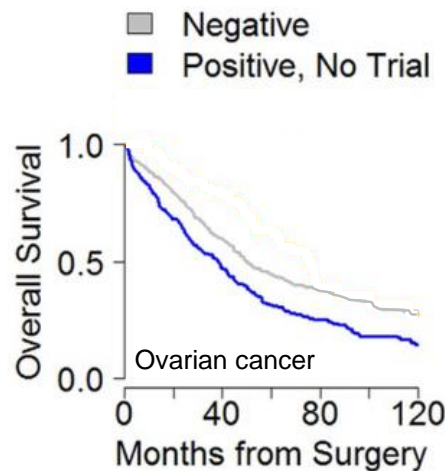
TCR binding probability is a sigmoid function of alignment score



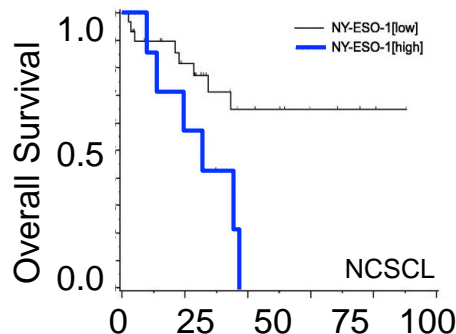
c) Neoantigen cross reactivity for a given neoantigen is a function of alignment score and amplitude (K_d^{WT}/K_d^{Mutant})



NY-ESO-1 expression is a poor prognostic factor but it may be a good predictive marker for immunotherapy



Based on IHC or RT-PCR expression of ovarian cancer patients and participation or not in immunotherapy clinical trials.



Based on RT-PCR expression of non-small cell lung adenocarcinoma cancer patients with stage II disease ($p=0.02$).

Metastatic melanoma patients with **baseline** NY-ESO-1 serum antibodies **before CTLA-4 (ipilimumab) treatment**

Status at wk 24	# patients (%)	NY-ESO-1 seronegative # (%)	NY-ESO-1 seropositive # (%)
CR	4 (2.9%)	3	1
PR	14 (10.0%)	10	4
SD	30 (21.4%)	23	7
Clinical Benefit	48 (34.3%)	36 (30.5%)	12 (54.6%)
No Clinical Benefit	92 (65.7%)	82 (69.5%)	10 (45.4%)
Total	140 (100%)	118	22

According to immune-related response criteria:

Clinical Benefit:

CR: Complete Response

PR: Partial Response

SD: Stable Disease

No Clinical Benefit:

POD: Progression of Disease (includes MR)

DOD: Dead of Disease

Fisher's exact test

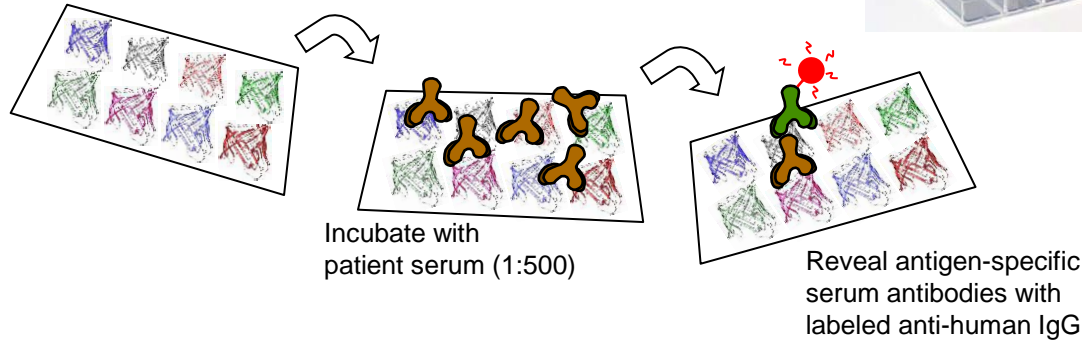
(two-tailed):

P value 0.0481

RR = 1.8 (1.1-2.9)

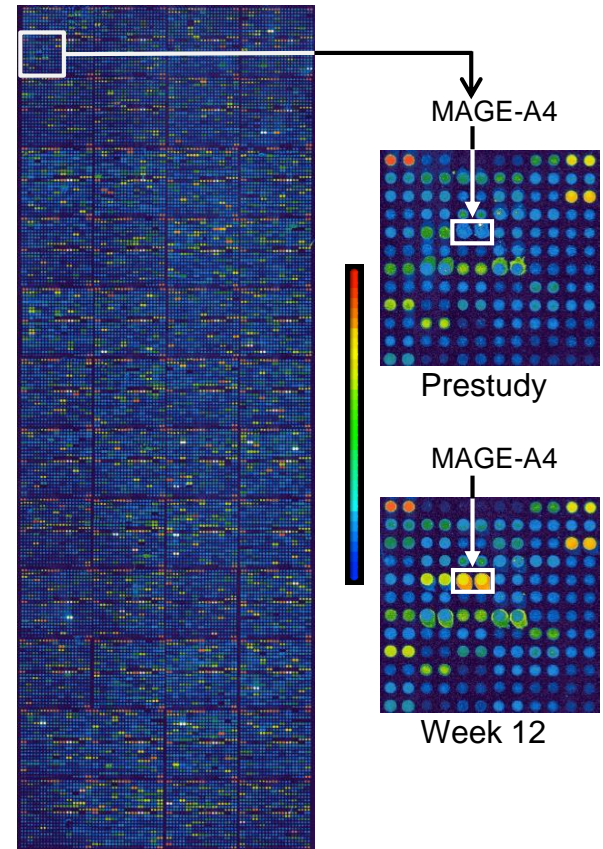
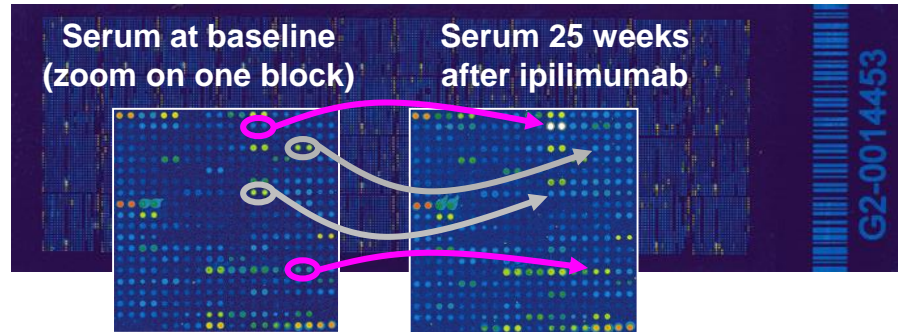
Seromics detects antigen-specific changes in autoantibody profiles during treatment (H. Wada, Osaka JP; H. Shiku, Mie JP; unpublished)

Protoarrays™ contain >9000 proteins mostly full-length baculovirus-produced GST-fusion proteins randomly selected, both known and predicted sequences



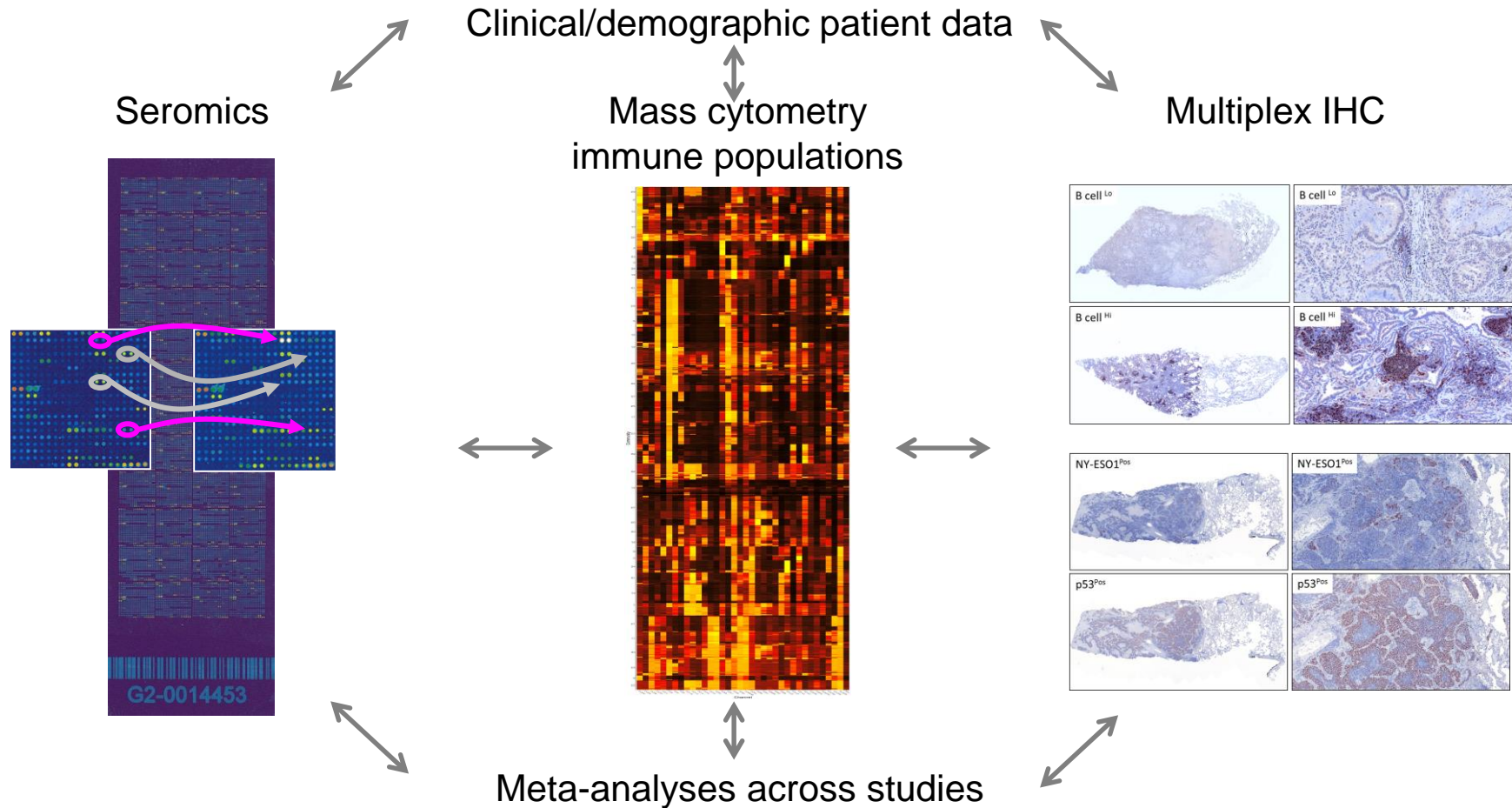
From Invitrogen.com

for biomarkers of treatment:

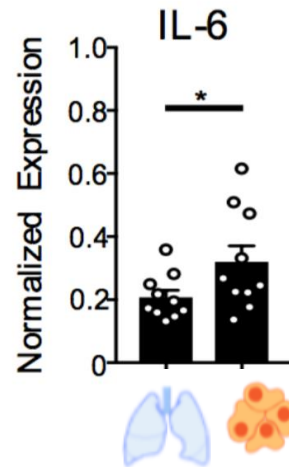
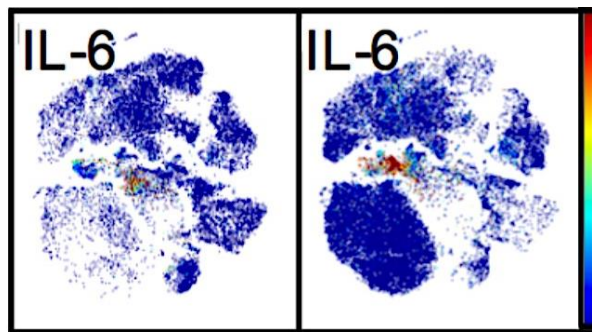
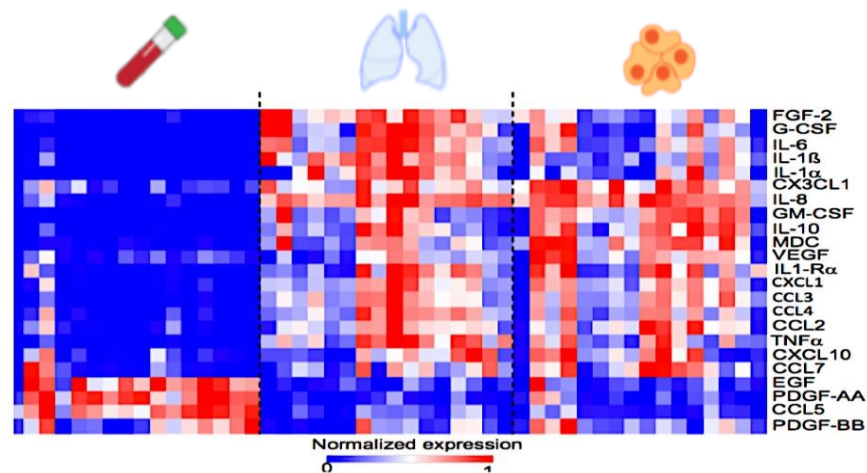


MAGEA4 protein vaccination of esophageal cancer patient

Cross-assay correlations with various data sets



Integrating datasets: Luminex, CyTOF, RNAseq



Take home message

High-dimensional immune monitoring assays are poised to explain mechanisms of novel drugs or treatment and provide complex signatures to predict outcome

It is unlikely that a single predictive biomarker will be found for immuno-oncology

Single cell data analyses and data mining are the next frontiers for discoveries in immunotherapy

Immune monitoring supports immune atlas efforts, to define baseline characteristics and mechanisms of response or resistance to various immuno-oncology drugs

Era of personalized combined biomarkers

Acknowledgments

Icahn School of Medicine at Mount Sinai, New York, NY

Romain Remark	Seunghee Kim-S
Adeeb Rahman	Christian Becker
Yeray Arteaga	Sarah Nataraj
Luis Ferran	Ilaria Laface
Yonit Lavin	Naoko Imai
El-Ad Amir	Takuro Saito
Mary Beth Beasley	Raja Flores
Patricia Kovatch	Michael Donovan
Jose Clemente	Jeremiah Faith
Jeff Hammerbacher	Ben Greenbaum
Nina Bhardwaj	Miriam Merad

MSKCC, New York, NY

Jianda Yuan	Jedd Wolchok
Achim Jungbluth	Erika Ritter
Steven Leach	Taha Merghoub

Yale University, New Haven, CT

Mario Sznol	Ruth Halaban
-------------	--------------

Roswell Park Cancer Institute, Buffalo, NY

Kunle Odunsi	Takemasa Tsuji
Junko Matsuzaki	

Ludwig Institute for Cancer Research, New York, NY

Lloyd J. Old

Support from:

Cancer Research Institute
Ludwig Institute for Cancer Research

NCI P01 CA190174

NCI U24 CA224319 (CIMAC)

ARRA RC2 NCI