



Immunotherapy/Immunotherapy combinations

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Rationale and foundation for discussion

- Important disclaimer: we are not suggesting that "vaccines don't work" and therefore combinations of vaccines plus other therapies will automatically be required
- Rather, our view is that an anti-tumor immune response is a complex and multi-stage process that can become dysregulated at several levels in the context of a growing tumor
- Overcoming each of these defects may require a distinct intervention, and therefore combination therapies may be important in order to translate immune responses into tumor regression
- Another way to look at it: with T cell-based immunotherapy, the "drug" is not necessarily the product administered (e.g. vaccine)—rather, the therapeutic entity is the properly generated tumor antigen-specific effector T cell population that has penetrated the tumor microenvironment and maintained effector function there

At what levels can a spontaneous antitumor T cell response fail?

Lymph node **Tumor microenvironment** 3. APC maturation/ 2. Antigens/ costimulation Ag processing innate immune awareness Lymphatic APC 8. Target cell apoptosis **Granzymes** nCD8 perforin eCD8 **Blood** IFN-γ **Chemokines** 4. T cell repertoire/ eCD8 eCD8 activation 6. Effector T cell 5. T cell differentiation/ trafficking 7. T cell effector function expansion/persistence (negative regulation)

Hypothetical barriers point towards strategies for intervention

Vaccines

- 1. Innate immune awareness/Ag presentation/APC maturation
 - Are there "danger" signals to ensure productive antigen display?
- 2. T cell repertoire/initial activation
 - Repertoire may be restricted or of low avidity
 - Immune suppression may carry over to DLN compartment

Adoptive Tx 3.

T cell differentiation/expansion/persistence

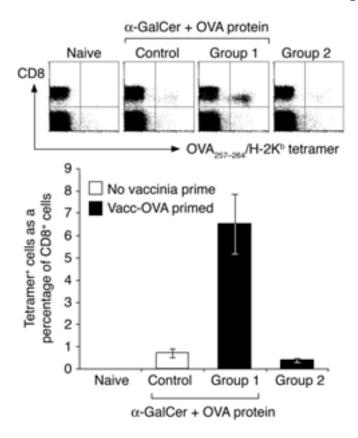
- Proper T cell phenotype might not be induced (Th1/CTL/memory)
- Magnitude or duration of T cell response may be inadequate
- 4. T cell trafficking into tumor sites
 - Lack of proper chemokine receptors on T cells, or chemokines at tumor site
 - Signals for penetrating extracellular matrix?
- 5. Executing effector function in tumor microenvironment
 - Dominant negative regulatory pathways
 - Poor maintenance of effector function (e.g. regeneration of cytotoxic granules)
- 6. Tumor cell susceptibility to recognition and killing
 - Loss of antigens, processing machinery, MHC
 - Anti-apoptotic mechanisms: tumor cells can be resistant
 - Interface with tumor cell-intrinsic biology: oncogenic pathways orchestrating resistance

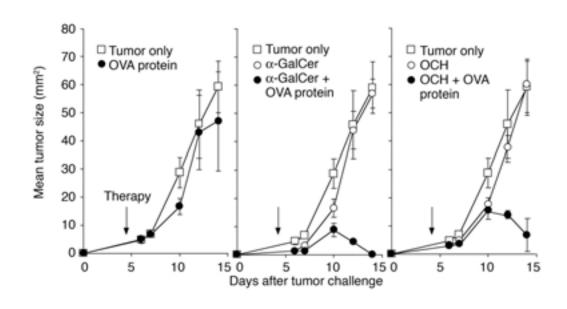
- 1. Innate immune awareness/Ag presentation/APC maturation
 - Innate immune cells and cytokines, TLR agonists, CD40 ligands, vaccination—novel Ag sources
- 2. T cell repertoire/initial activation
 - B7 and other costimulatory ligands
 - Interference with lymph node-based or systemic negative regulators (CTLA4, IDO, arginase, anergy, Tregs)
- 3. T cell differentiation/expansion/persistence
 - Differentiation cytokines (IL-12, IL-18)
 - Expansion, survival factors (IL-2, IL-7, IL-15, anti-41BB; homeostatic proliferation)
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 - Intratumoral chemokines, LIGHT
 - Pro-inflammatory treatments (XRT, TLR agonists, innate cytokines)
- 5. Executing effector function in tumor microenvironment
 - Blockade of tumor microenvironment-based negative regulators (IDO, PD-1/PD-L1, Tregs, anergy, TGF-β, IL-10, iNOS)
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 - Blockade of key anti-apoptotic molecules (Bcl2 and Spi inhibitors)
 - Inhibit oncogenic pathways that create resistant phenotype and/or resistant microenvironment (Stat3; MEK? Notch? Wnt?)

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Example 1: α **-GalCer**

Administration of protein and α -GalCer can synergistically expand CD8+ T cells





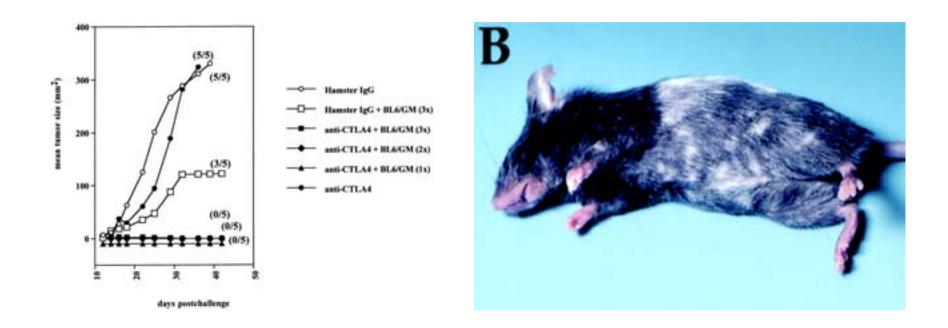
Silk, Cerundolo et al. J. Clin. Invest. 2004



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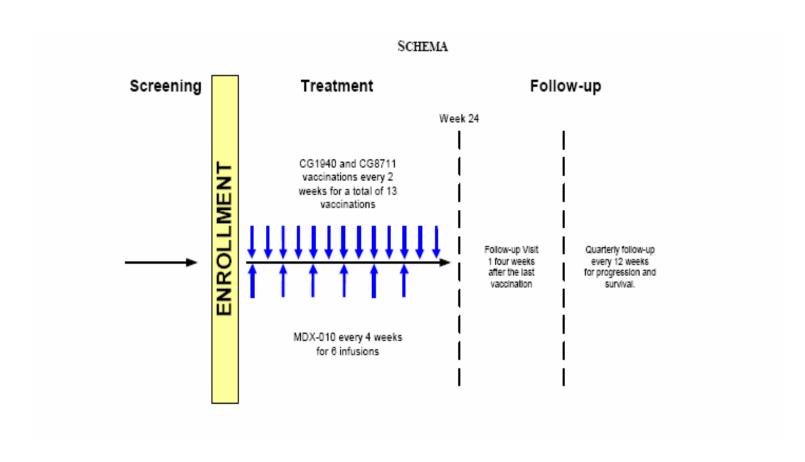
Example 2: CTLA-4

Anti-CTLA-4 mAb + GM-CSF-transduced B16 vaccine induces tumor rejection and leads to vitiligo



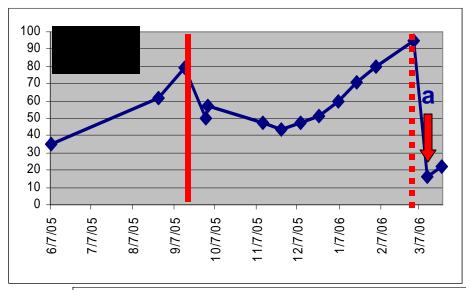
van Elsas, Allison et al. JEM 1999

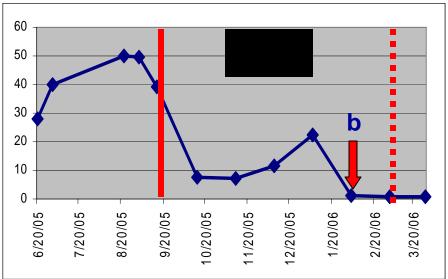
GVAX Immunotherapy (CG1940/CG8711) + Ipilimumab (MDX-010: anti-CTLA-4) for HRPC

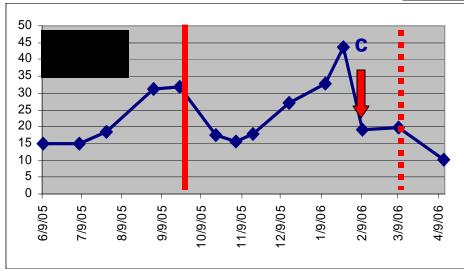


VUmc Cancer Center Amsterdam

GVAX + anti-CTLA-4 in prostate cancer: PSA curves – Dose Level 3 (3 mg/kg)







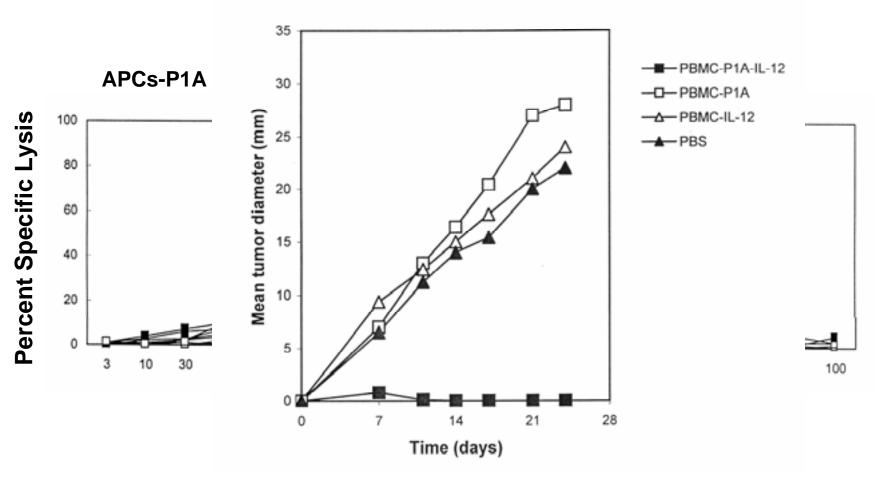
- **a**:13Mar06: SAE -Hypophysitis (7 mo)
- **b:** 03Feb06: Hypophysitis (5 mo)
- **c:** 09Feb06: SAE Hypophysitis (5 mo)

Gerritsen et al. ASCO 2006

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Example 3A: IL-12

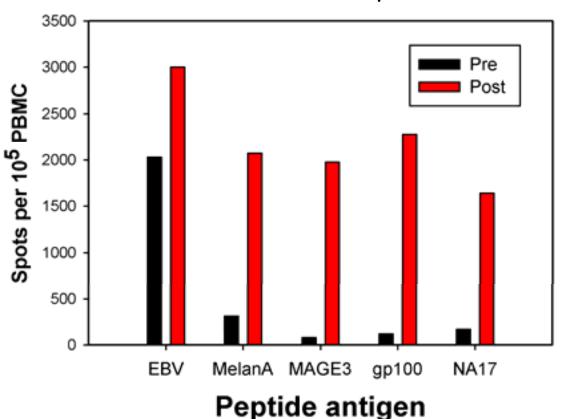
Superior induction of specific CTL responses in mice using peptide-loaded APCs + IL-12



Fallarino et al. Int. J. Cancer 1999

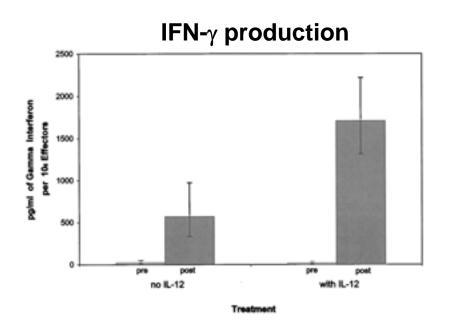
Potent T cell response against multiple antigens post-immunization of melanoma patients with peptide-pulsed PBMC + IL-12

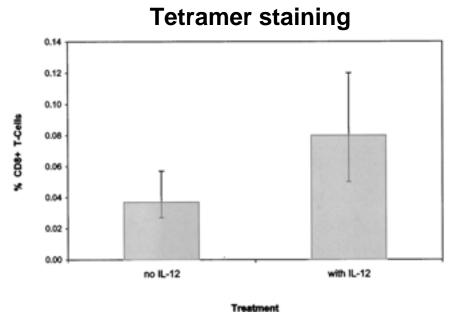




3 patients with CR post-vaccination

Superior immune responses with IL-12 + peptides in Montanide in patients with melanoma

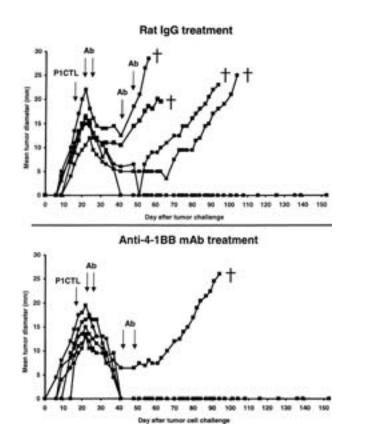


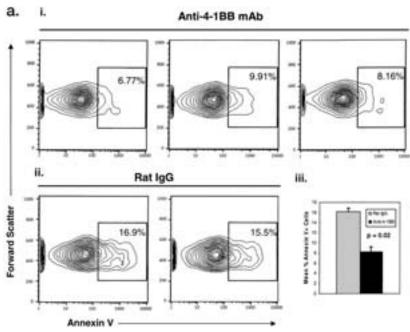


Lee, Weber et al. JCO 2001

Example 3B: Anti-4-1BB

Co-administration of anti-4-1BB mAb with adoptively transferred T cells induces superior tumor rejection and T cell survival in mice

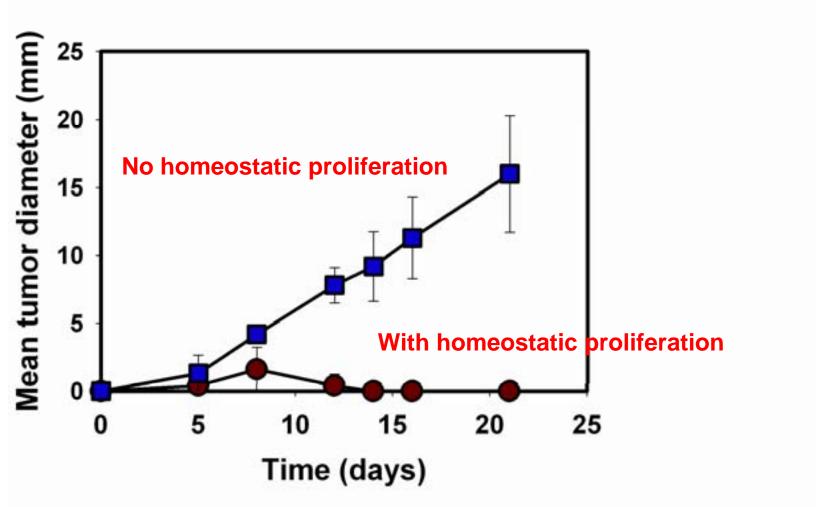




May, Liu et al. Cancer Res. 2002.

Example 3C: Homeostatic proliferation

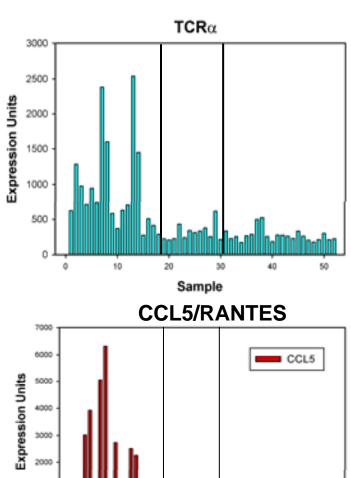
Anergic 2C T cells reject tumors after homeostatic proliferation in RAG2-/- hosts



Brown et al., J. Immunol., 2006

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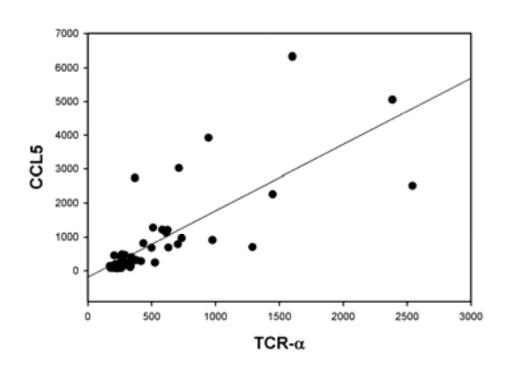
T cell transcripts in melanoma metastases are associated with expression of specific chemokine genes



Sample Number

Cell Lines

1000



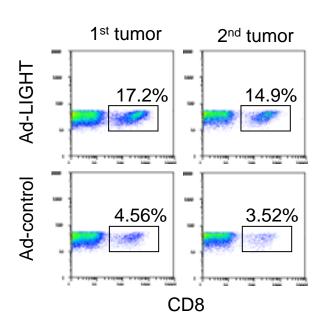
Example 4: Intratumoral LIGHT adenovirus in B16 melanoma promotes greater recruitment of CD8+ T cells in primary tumor and leads to rejection of non-injected distant metastases

Tumor rejection

4000

Subject to the state of t

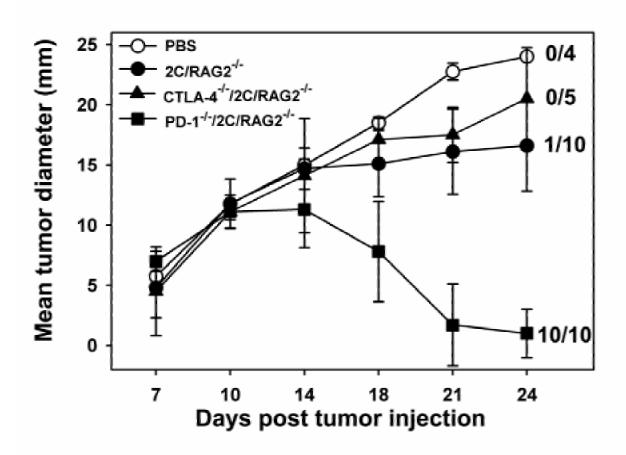
CD8+ T cell infiltrate



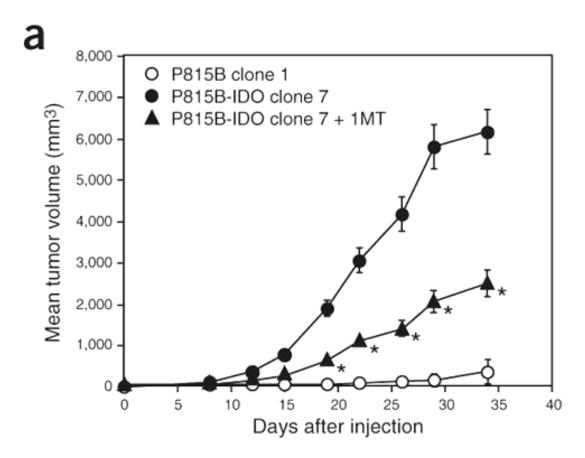
Yu et al, J. Immunol. 2007

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Example 5A: PD-1^{-/-} 2C TCR Tg T cells are superior at tumor rejection in vivo

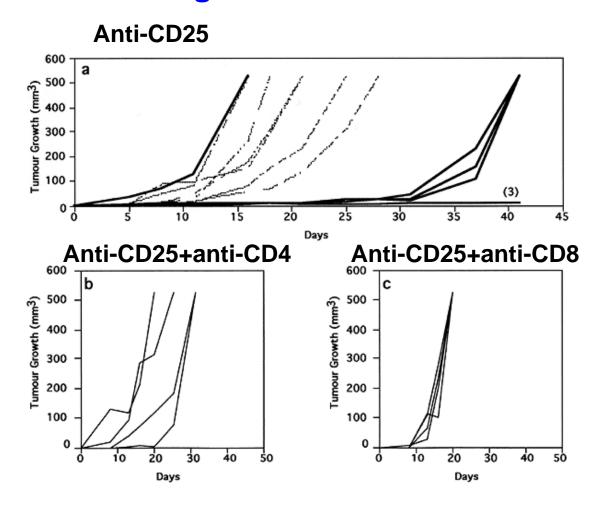


Example 5B: 1-methyltryptophan reverses immunosuppression by IDO and improves tumor control in vivo



Example 5C: CD25+ Tregs

CD25 depletion can partially control B16 melanoma growth in vivo

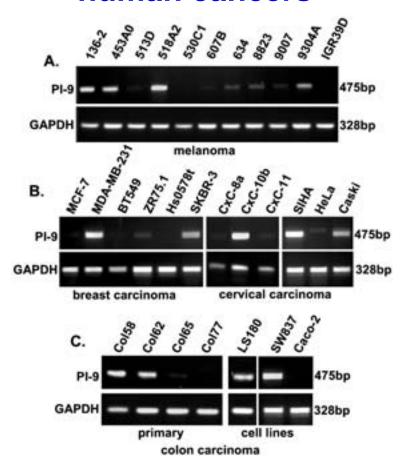


Jones, Gallimore et al. Cancer Immunity 2002

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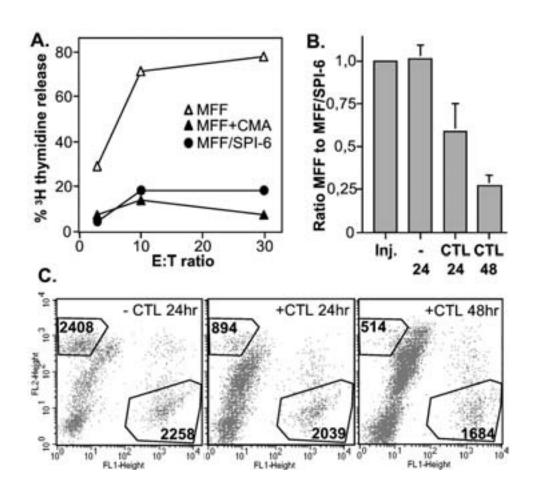
Example 6: PI-9/Spi6

Serine protease inhibitor PI-9 is frequently expressed in human cancers



Medema, J. P. et al. Proc. Natl. Acad. Sci. USA 2001

Introduction of the murine equivalent Spi6 into tumor cells decreases susceptibility to T cell-mediated lysis in vitro

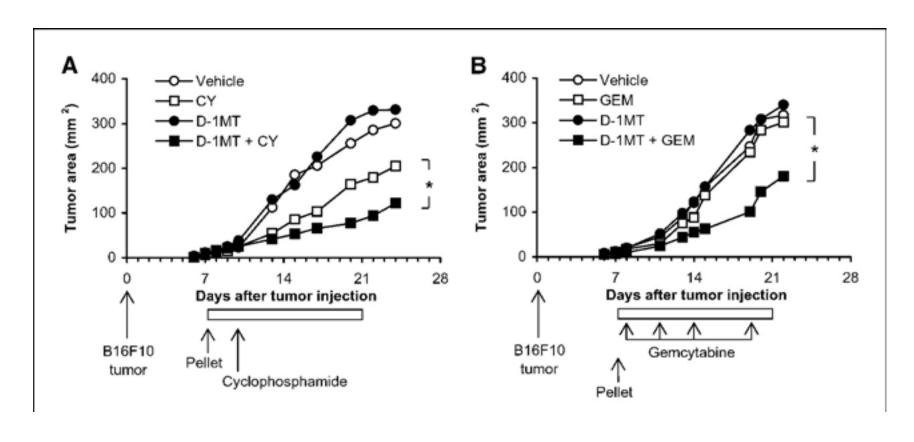


Multiple combinations:

Another layer of complexity and excitement through combined manipulation of regulatory checkpoints

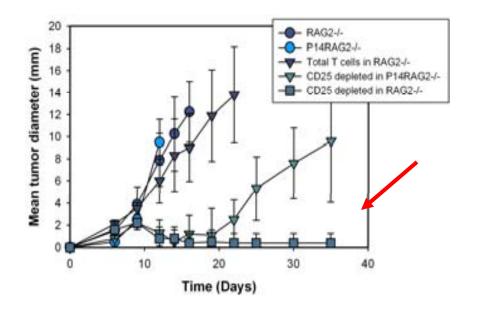
- 1-MT + lymphodepletion
- Anergy reversal + Treg-depletion
- Anti-4-1BB + anti-CTLA-4
- Anti-4-1BB + anti-PD-L1
- Anti-CTLA-4 + Treg depletion
- TLR agonist + Treg-depletion

1-MT + lymphodepleting chemotherapy: Partial control of B16 melanoma



Hou, Munn et al. Cancer Res. 2007

Treg depletion + anergy reversal CD25-depleted T cells transferred into lymphopenic hosts gives long-lived rejection of B16 melanoma and vitiligo

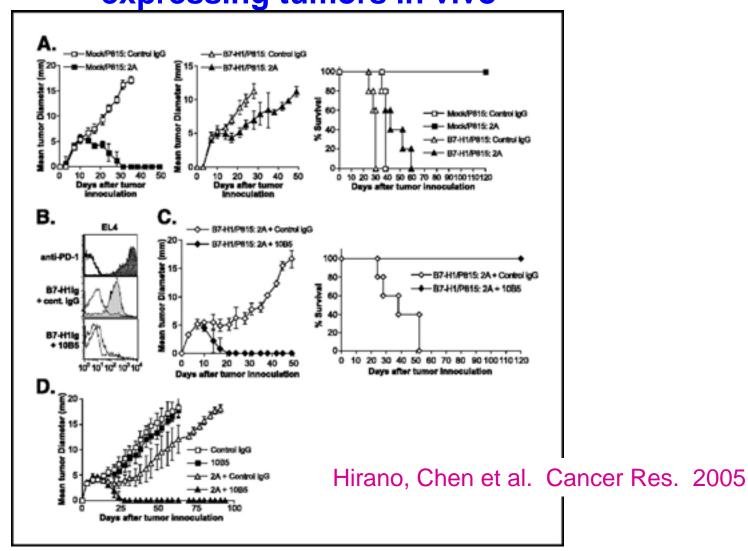




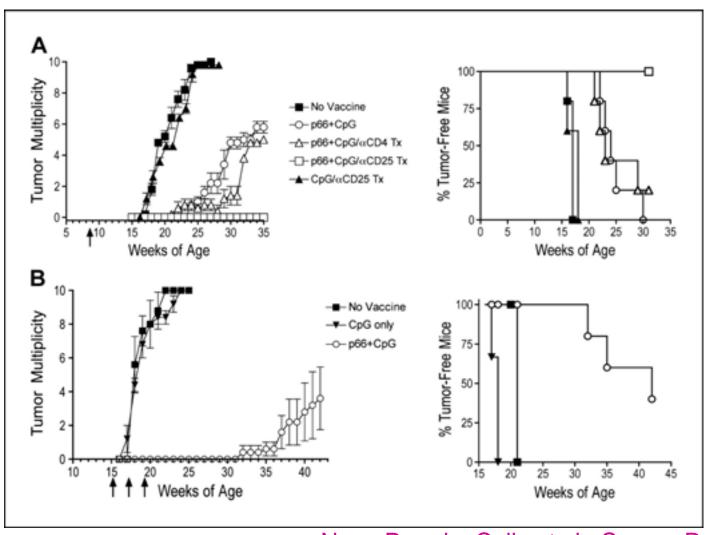
Kline et al. Submitted.

Anti-4-1BB + anti-PD-L1

Combination induces induces rejection of PD-L1expressing tumors in vivo



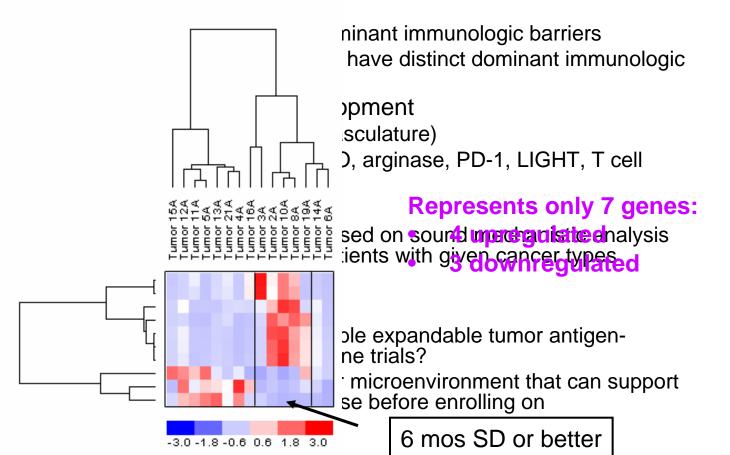
Vaccine + CpG + Treg depletion: Control of mammary tumors in Neu Tg mice



Nava-Parada, Celis et al. Cancer Res. 2007

Additional issues

- Tumor hete
 - Differen
 - Differen barriers
- Opportuniti
 - Cellular
 - Molecul signalin
- How to price
 - Many point
 of immu
 - Preclinic
- Patient selection
 - Can we specific
 - Similarly effector immuno



Conclusions

- Spontaneous anti-tumor T cell responses may fail at one of several levels
- Specific mechanisms of failure have identified new targets and strategies for intervention
- There is a strong scientific basis for combination therapies with the aim of overcoming specific barriers and immunologic checkpoints to increase the therapeutic efficacy of T cell-based immunotherapy of cancer
- Some agents are becoming available for clinical translation, but others need broad-based community support to be made available for clinical studies based on a sound rationale and preclinical data

Agents prioritized by scientific community for clinical development

Table 1. Final Rankings of Agents with High Potential for Use in Treating Cancer

Rank*	Agent	Agent Category
1	IL-15	T-Cell Growth Factor
2	Anti-Programmed Death-1	**T-Cell Checkpoint Blockade
	(PD1)and/or anti-B7-H1 (PD1	Inhibitor
	Ligand)	
3	IL-12	Vaccine Adjuvant
4	Anti-CD40 and/or CD40L	Antigen Presenting Cell
		Stimulator
5	IL-7	T-Cell Growth Factor
6	CpG	Vaccine Adjuvant
7	1-Methyl Tryptophan	Enzyme Inhibitor
8	Anti-CD137 (anti-4-1BB)	T-Cell Stimulator
9	Anti-TGF-beta	Signaling Inhibitor
10	Anti-IL-10 Receptor or Anti-	Suppression Inhibitor
	IL-10	••
11	Flt3L	Dendritic Cell Growth Factor/
		Vaccine Adjuvant
12	Anti-Glucocorticoid-Induced	T-cell Stimulator
	TNF Receptor (GITR)	
13	CCL21 Adenovirus	T-Cell Attracting Chemokine
14	Monophosphoryl Lipid A	Vaccine Adjuvant
	(MPL)	,
15	Poly I:C and/or Poly ICLC	Vaccine Adjuvant
16	Anti-OX40	T-Cell Stimulator
17	Anti-B7-H4	T-Cell Checkpoint Blockade
		Inhibitor
18	Resiguimod and/or 852A	Vaccine Adjuvant
19	LIGHT and/or LIGHT vector	T-Cell Stimulator
20	Anti-Lymphocyte Activation	T-Cell Checkpoint Blockade
	Gene-3 (LAG-3)	Inhibitor

Clinical development of anti-CTLA-4 mAb: Example of MDX-010 (Ipilumumab)

- Fully human IgG1 monoclonal antibody to human CTLA-4 created by Medarex
- Blocks binding of CTLA-4 to CD80 and CD86
- Augments immune responses in primate models
- Co-developed by Medarex and Bristol-Myers Squibb in multiple cancer indications
 - Phase III study in metastatic melanoma ongoing
 - Phase II studies in renal cell carcinoma, prostate cancer, ovarian cancer, and others

GVAX/anti-CTLA4 trial Contributors



Dept Medical Oncology

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Sinéad Lougheed

Helen Gall

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Shirley Clift

Karin Jooss

David Rhodes

Sayeh Morali



Israel Lowy

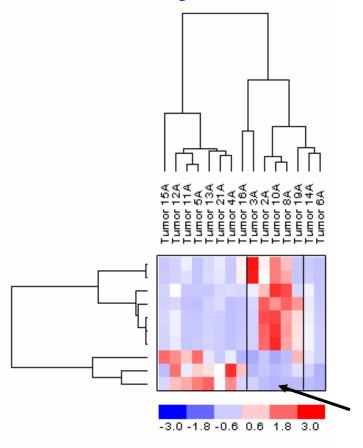
Steven Fischkoff

Elizabeth Levy





Affymetrix gene array analysis of pre-treatment biopsies from patients on melanoma vaccine sorted by clinical outcome



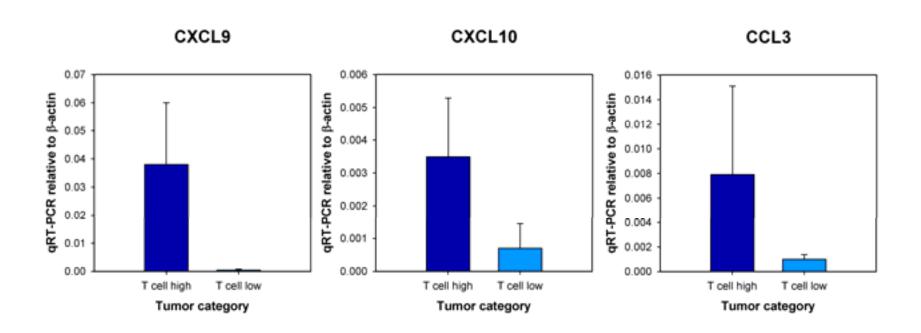
Represents only 7 genes:

- 4 upregulated
- 3 downregulated

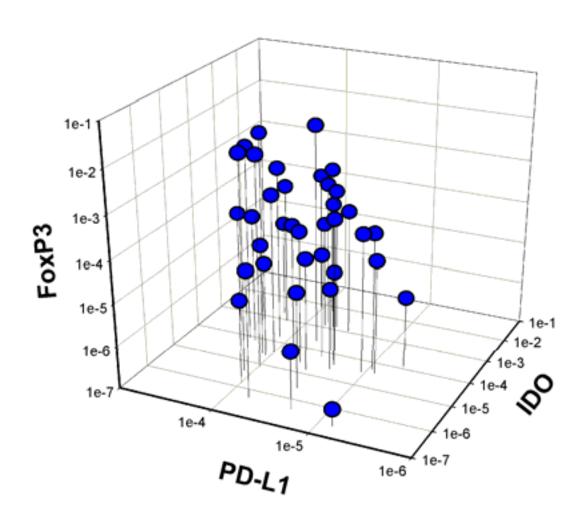
6 mos SD or better

Has implications for patient selection on vaccine trials, and understanding biology

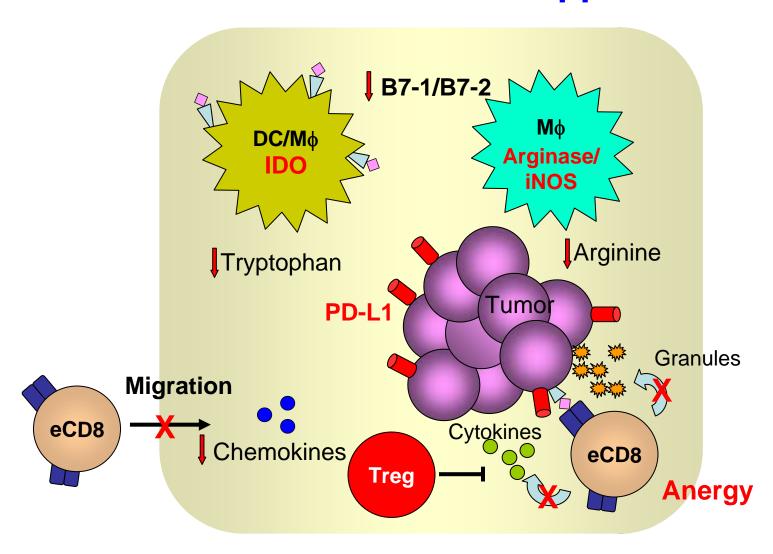
Differential chemokine expression in melanoma metastases with high versus low T cell transcripts



Co-expression of IDO, PD-L1, and FoxP3 transcripts in individual tumors



Summary of tumor microenvironment barriers: Need to promote T cell trafficking and overcome local immunosuppression



Resolution of cutaneous metastases following immunization with melanoma peptide-pulsed PBMC + rhIL-12

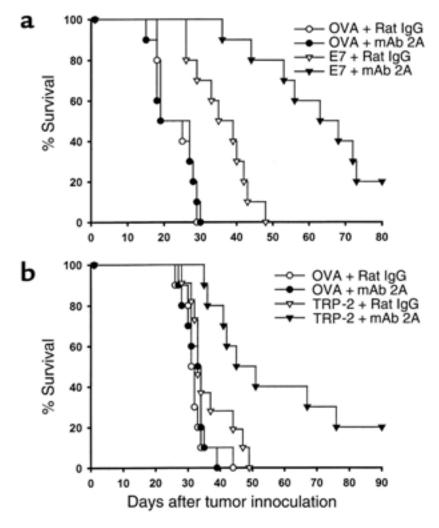
After 3 vaccines



After 9 vaccines



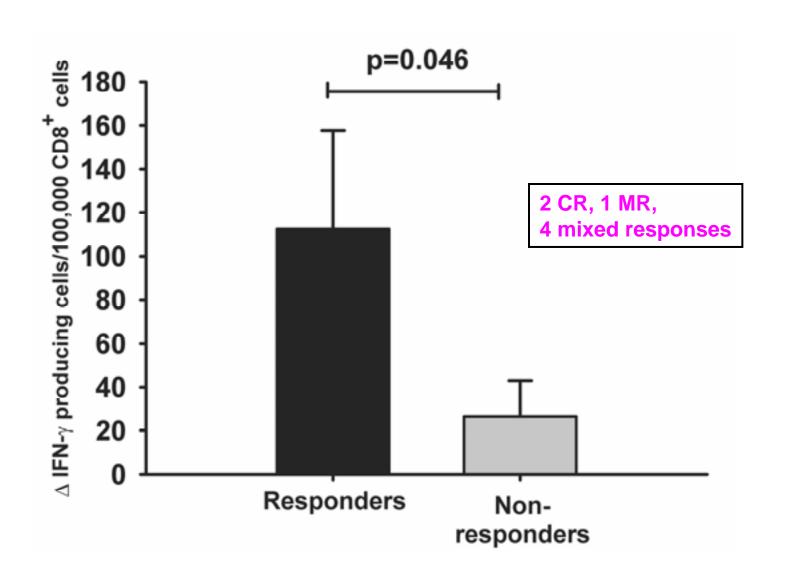
Peterson, Gajewski et al. JCO 2003.

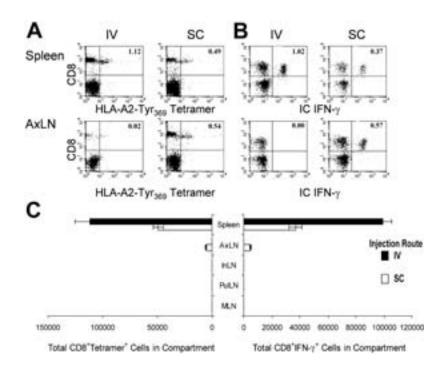


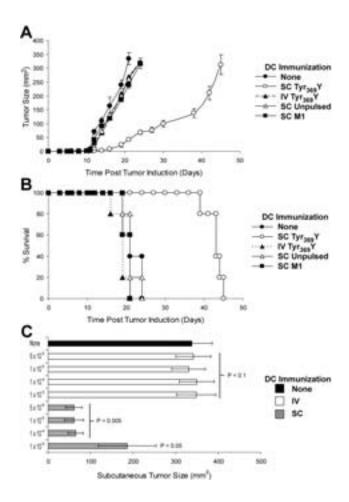
Wilcox, R. A. et al. J. Clin. Invest. 2002;109:651-659



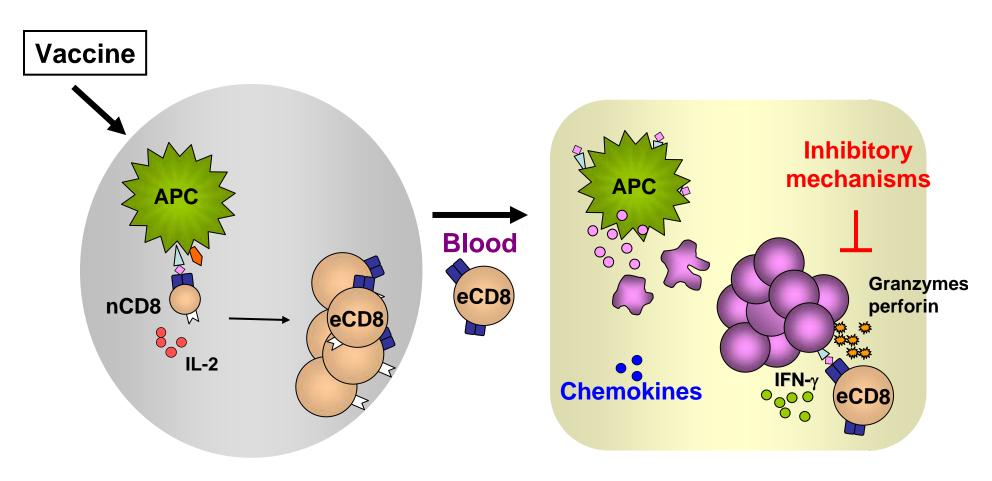
Greater increase in Melan-A-specific CD8+ T cells in clinical responders







Complexities of anti-tumor immune responses: Taking into account the effector phase



Lymph node (Priming phase)

Tumor microenvironment (Effector phase)

I. Priming phase/vaccine: considerations for combinations

- Antigen choice(s)
 - Peptides, protein, DNA, RNA, bulk tumor cells
 - Type of antigen (e.g. necessary for malignant phenotype)
 - Class I MHC, class II MHC, non-classical (glycolipids)
- Adjuvant components
 - Emulsions in oil-based formulations
 - TLR agonists (LPS/MPL + CpG)
 - Cytokine additions—differentiation promoters
 - Microbial vectors
 - Dendritic cell-oriented
- Dose, schedule, route of administration
 - Issue of tissue-specific homing of T cells

IV. Negative regulatory pathways: considerations for combinations

- Inhibitory receptors on T cells
 - CTLA-4
 - PD-1
 - KIRs
- Inhibitory cytokines
 - TGF-β
 - IL-10
- Inhibitory cell populations
 - CD4+CD25+FoxP3+ Tregs
 - Other Tregs
 - Myeloid suppressor cells
 - B cells
- Metabolic regulation
 - IDO
 - Arginase
 - Nutrient deprivation (glucose)

V. Tumor cell susceptibility: considerations for combinations

- Expression of "signal 1"
 - Antigens
 - Antigen processing machinery
 - MHC, β2M
- Overcoming anti-apoptotic mechanisms
 - Survivin
 - Bcl2-family members
 - Serine protease inhibitors
- Interface with tumor cell-intrinsic oncogenes
 - Ras/MAP kinase pathway & DC activation
 - Stat3 pathway and chemokines
 - Notch pathway and survival, immune gene expression

III. T cell trafficking: considerations for combinations

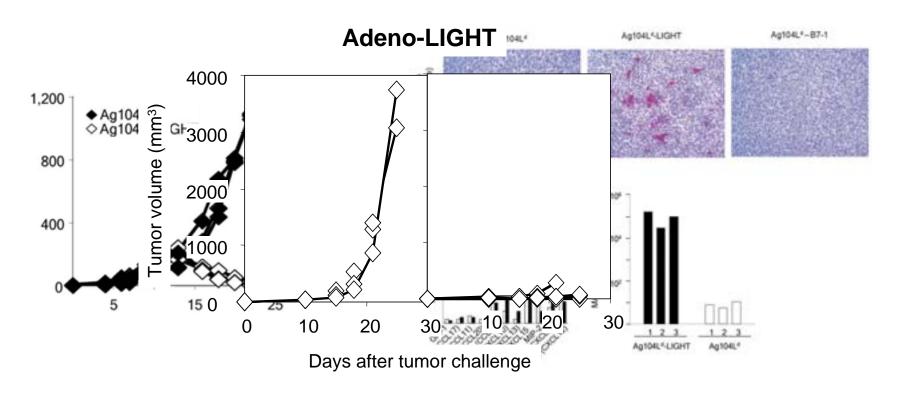
- Intratumoral chemokines
 - Mig, IP-10, MIP-1 α
 - CCL21
 - (Blockade of TARC/MDC?)
- Intratumoral LIGHT
 - Promotes secondary generation of chemokines
- Homing receptors/adhesion molecules
 - Intratumoral ICAM-1 (component of TRICOM)
 - Immunizing via optimal route (tissue specific homing)
- Angiogenesis targeting

II. T cell expansion and persistence: considerations for combinations

- Survival/homeostatic cytokines
 - IL-7
 - IL-15
 - IL-21
- Costimulatory receptors
 - B7 family members
 - 4-1BB
 - Other TNFR family members

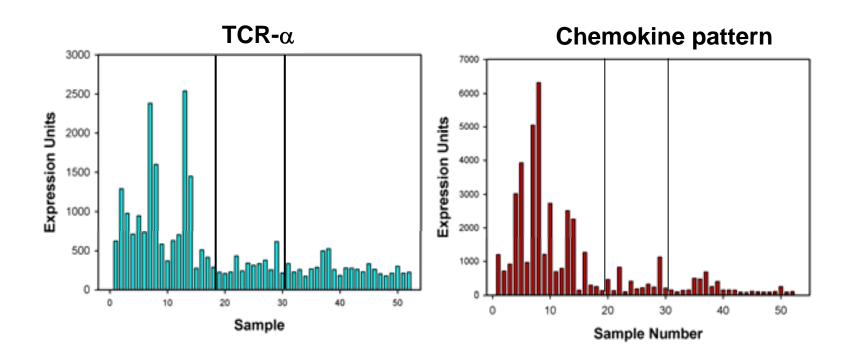
Example 4: LIGHT

Intratumoral LIGHT can induce T cell recruitment and tumor rejection in multiple tumor models



Fu et al, submitted

Only a subset of melanoma metastases appear to have the appropriate signature for T cell recruitment



Harlin, Gajewski et al. Manuscript in preparation.