



Is there a role for combining radiation therapy and immunotherapy?

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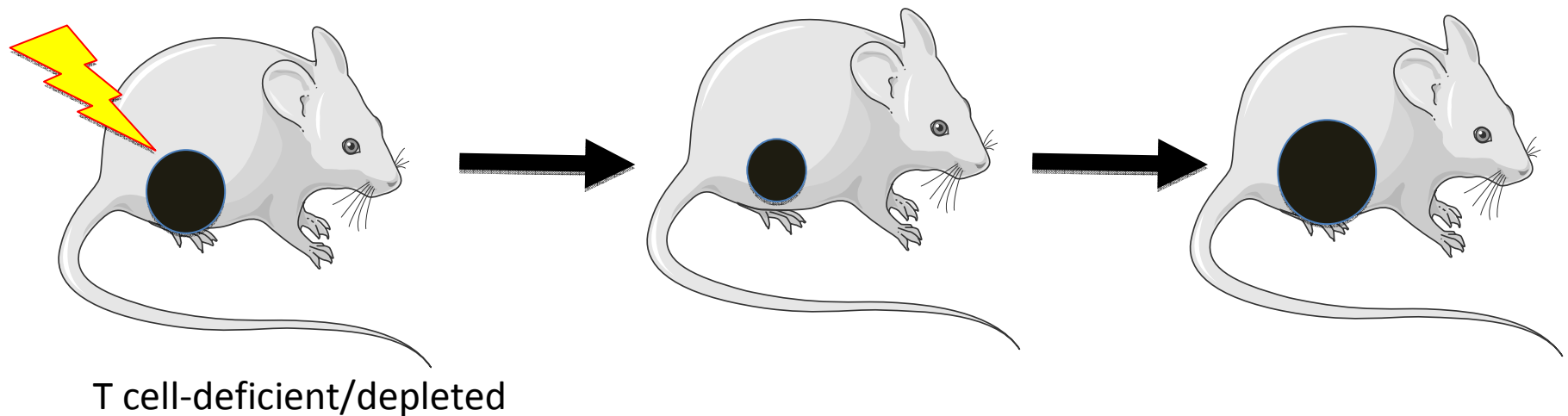
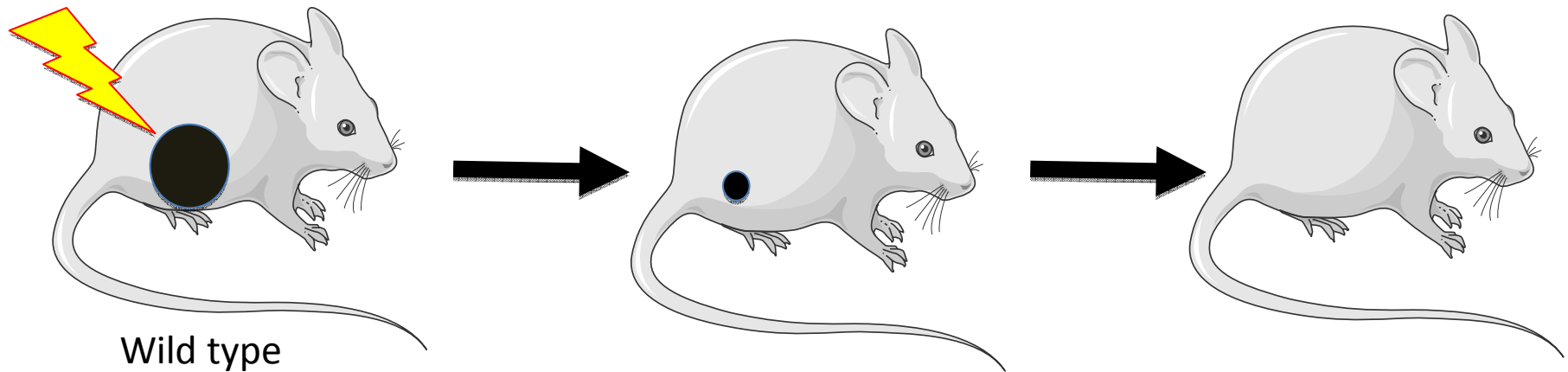


No conflicts to disclose

Outline

- Evidence for a role of the immune system in response to ionizing radiation
- Radiation-induced tumor cell death
 - Apoptosis and necrosis
 - Mitotic catastrophe and senescence
 - Autophagy
 - Immunogenic Cell Death (ICD)
- Effects of radiation on the tumor microenvironment
 - Pro-inflammatory
 - Anti-inflammatory
- Combination of radiation and immunotherapy
 - Overview
 - Checkpoint receptor inhibitor (anti-CTLA-4)

Immunocompetence of the host affects the response to local radiotherapy



T cells contribute to the response to radiotherapy

<u>FSA Tumor</u>	<u>TCD₅₀ values</u>	<u>Metastases</u>
Normal mice	30.0Gy (28.5-32.4)	1%
Immunosuppressed (6Gy)	50.8Gy (47.6-54.3)	4%
T cell deprived mice	64.5Gy (62.0-67.1)	79%

Stone et al., JNCI 63:1229, 1979

Evidence for induction of tumor-specific immune responses by radiation

IN MICE

Lugade et al., J Immunol 2005

B16-OVA model, induction of CD4 and CD8 T cells after irradiation with 15 Gy x 1 or 3 Gy x 5,

Lee et al, Blood 2009

B16-SIY model, induction of CD8 T cells after irradiation with 20 Gy x 1

Schaue et al., IJROBP 2012

B16-OVA, best induction of CD8 T cells with 7.5 Gy x 2

& MAN

Nesslinger et al., Clin Cancer Res 2007

Tumor-specific antibodies in 14% of prostate cancer patients treated with EBRT and in 25% receiving brachytherapy

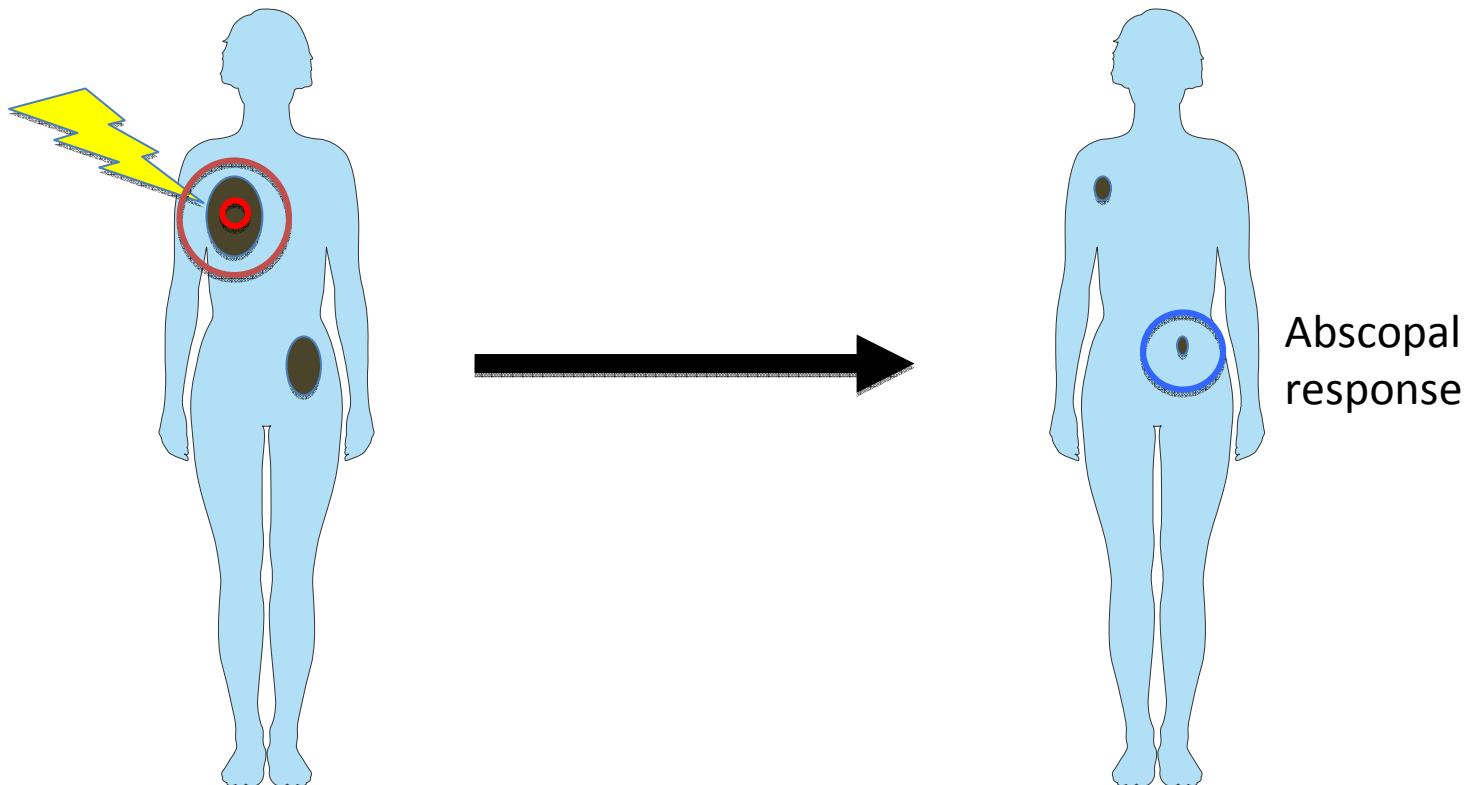
Schaue et al, Clin Cancer Res 2008

T cell responses to survivin in prostate and colorectal patients after radiotherapy

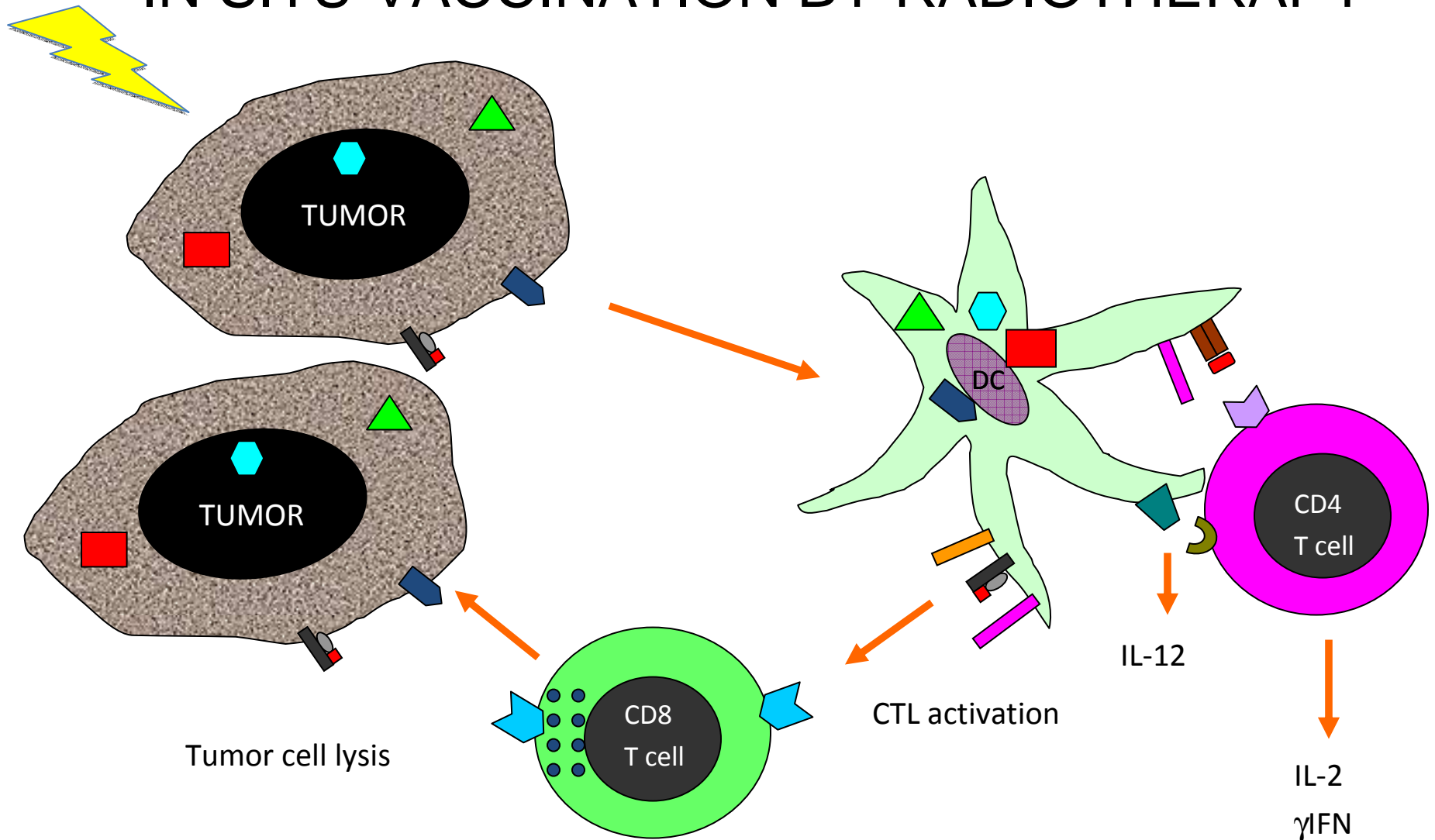
Abscopal effect

Effect of ionizing radiation on cancer
outside the radiation field

Latin *ab* (position away from) and *scopus* (mark or target)

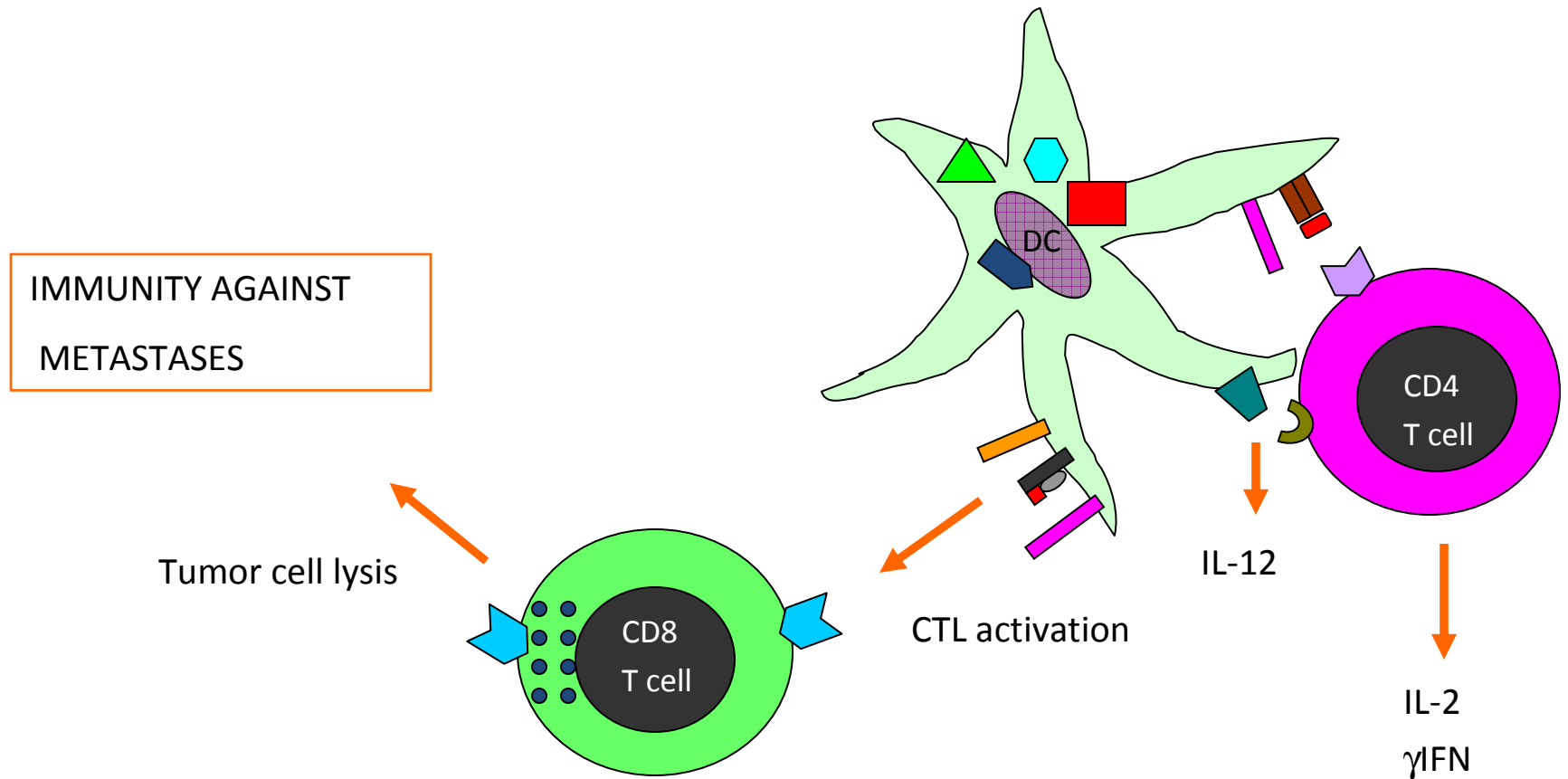


IN SITU VACCINATION BY RADIOTHERAPY



Demaria et al., IJROBP 2005
Formenti & Demaria, IJROBP 2012

IN SITU VACCINATION BY RADIOTHERAPY



Radiation-induced tumor cell death

Radiation Damage → DNA, proteins, lipids

DIRECT energy deposition

INDIRECT free radicals (H_2O radiolysis)



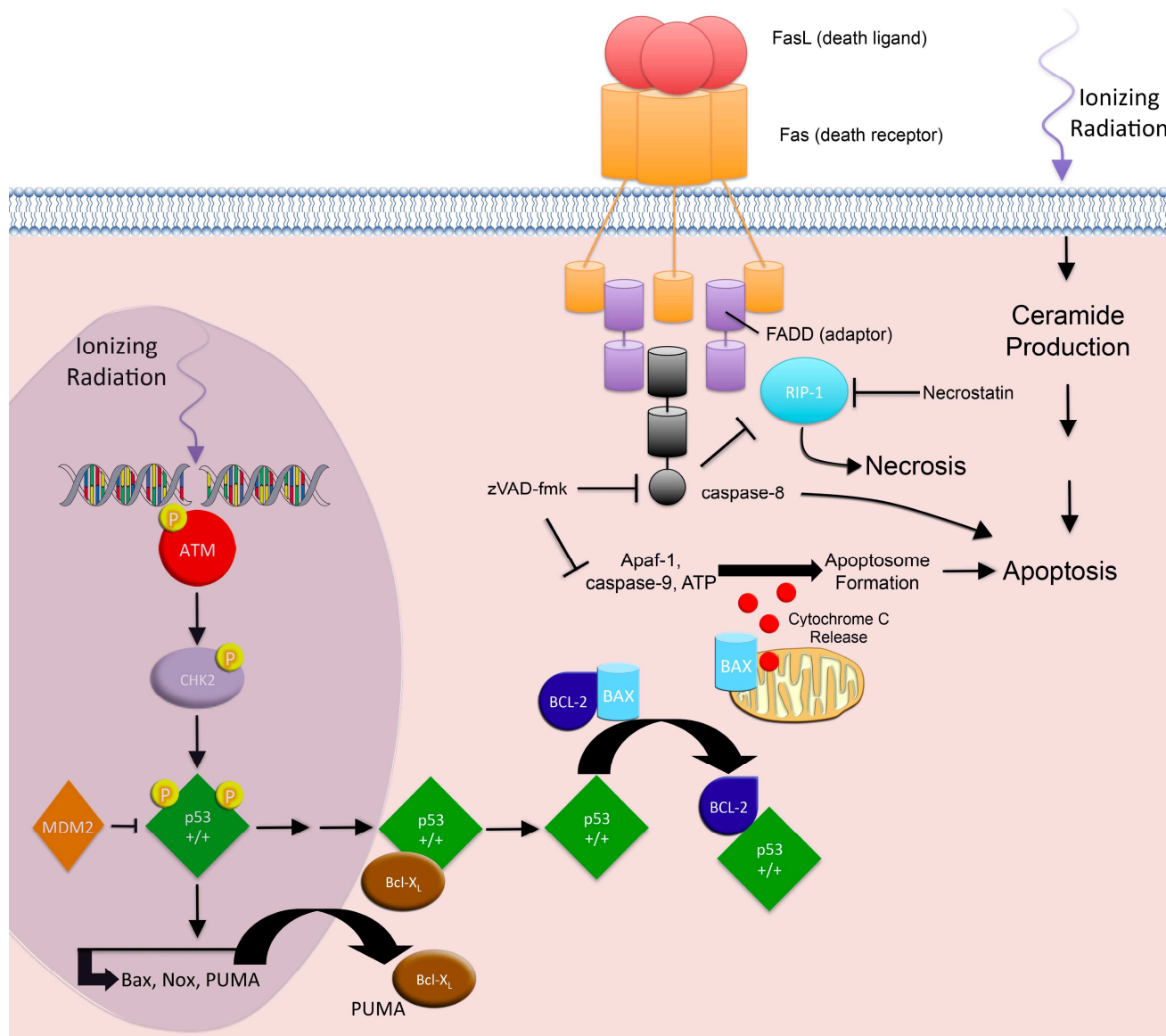
REPAIR

DEATH

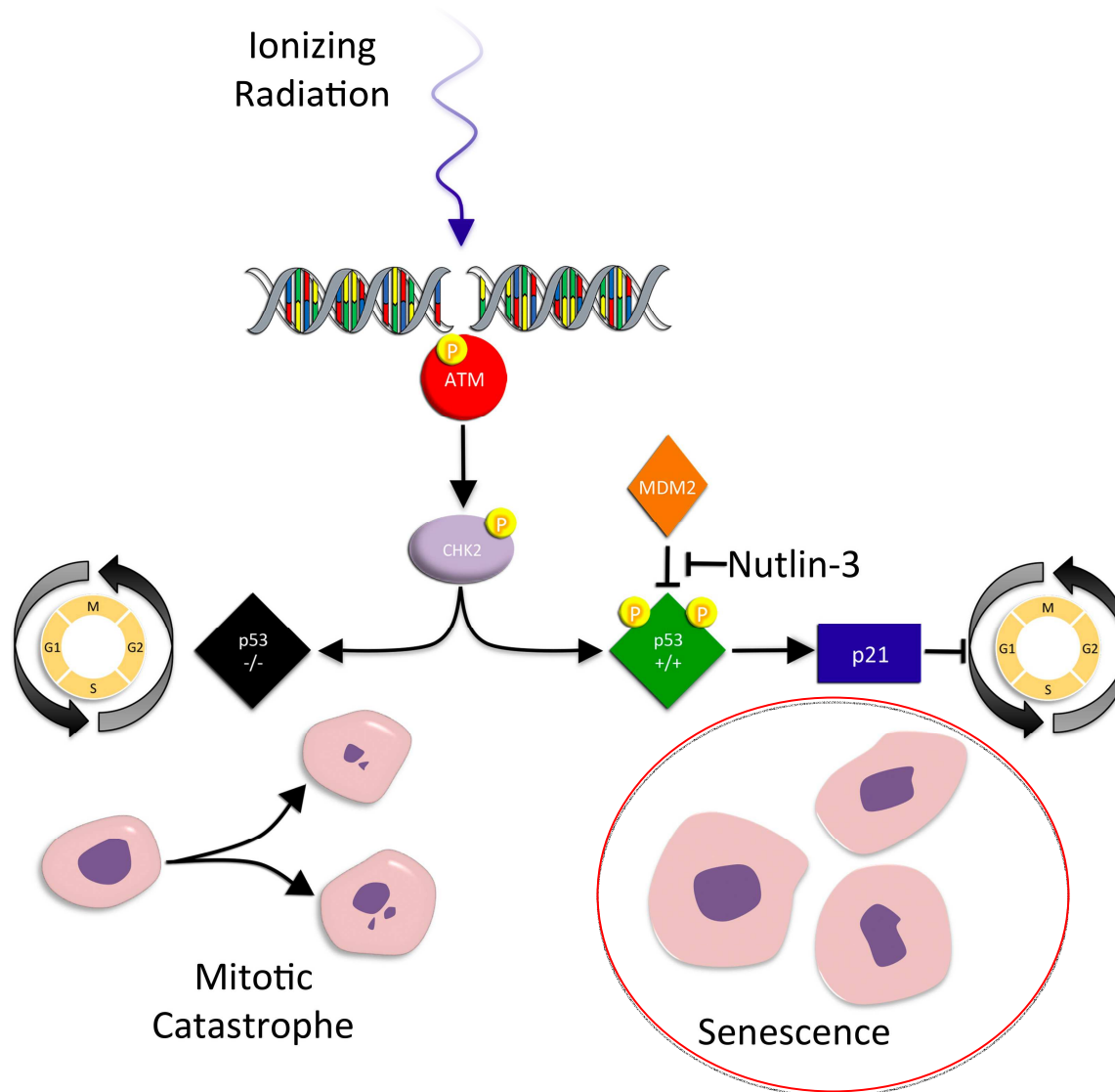
Cell death type depends on:

- intrinsic tumor cell characteristics (p53 status)
- radiation dose and quality (>8 Gy vascular death)
- oxygen tension
- DNA repair capacity
- redox status
- cell cycle phase

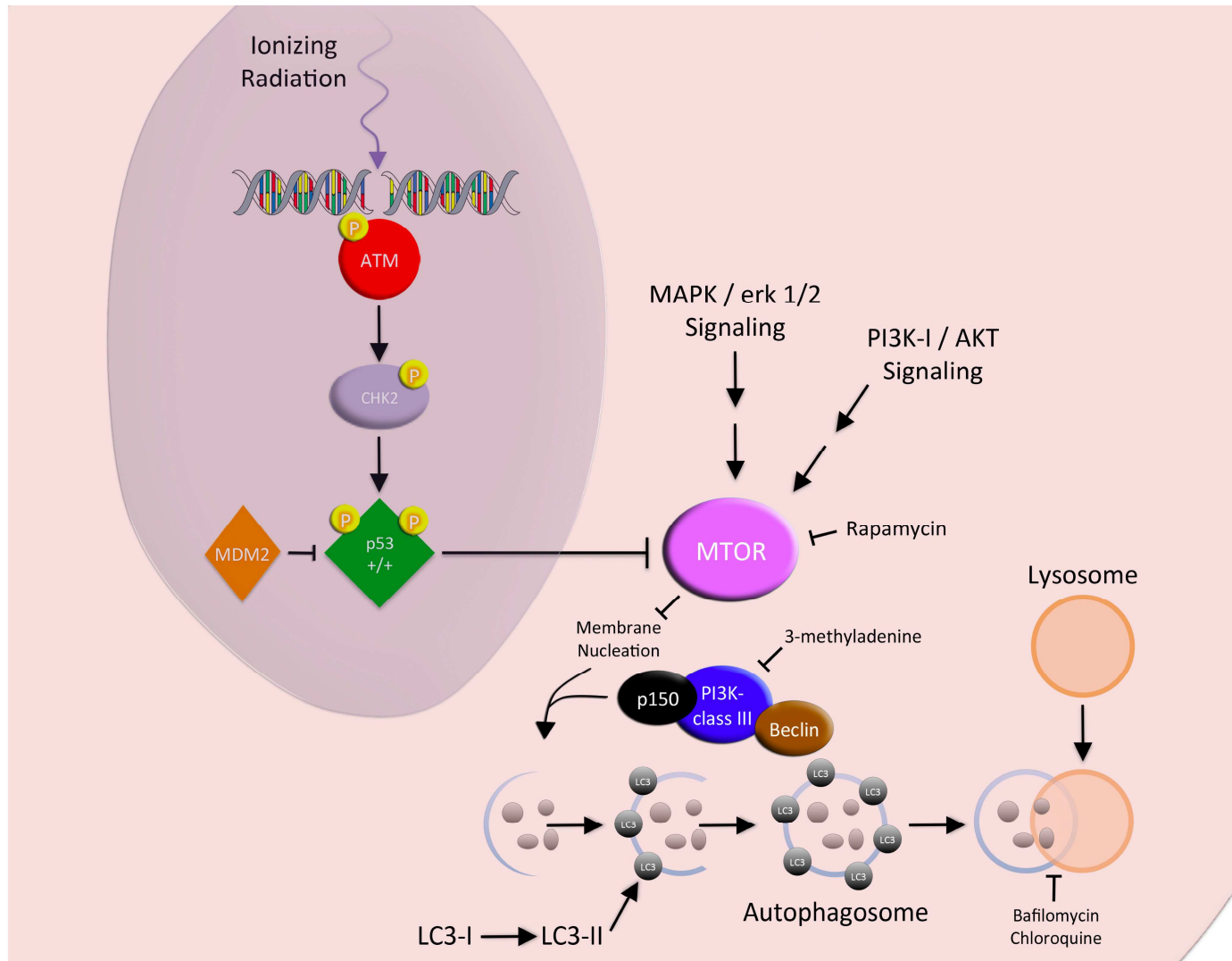
Apoptosis and necrosis



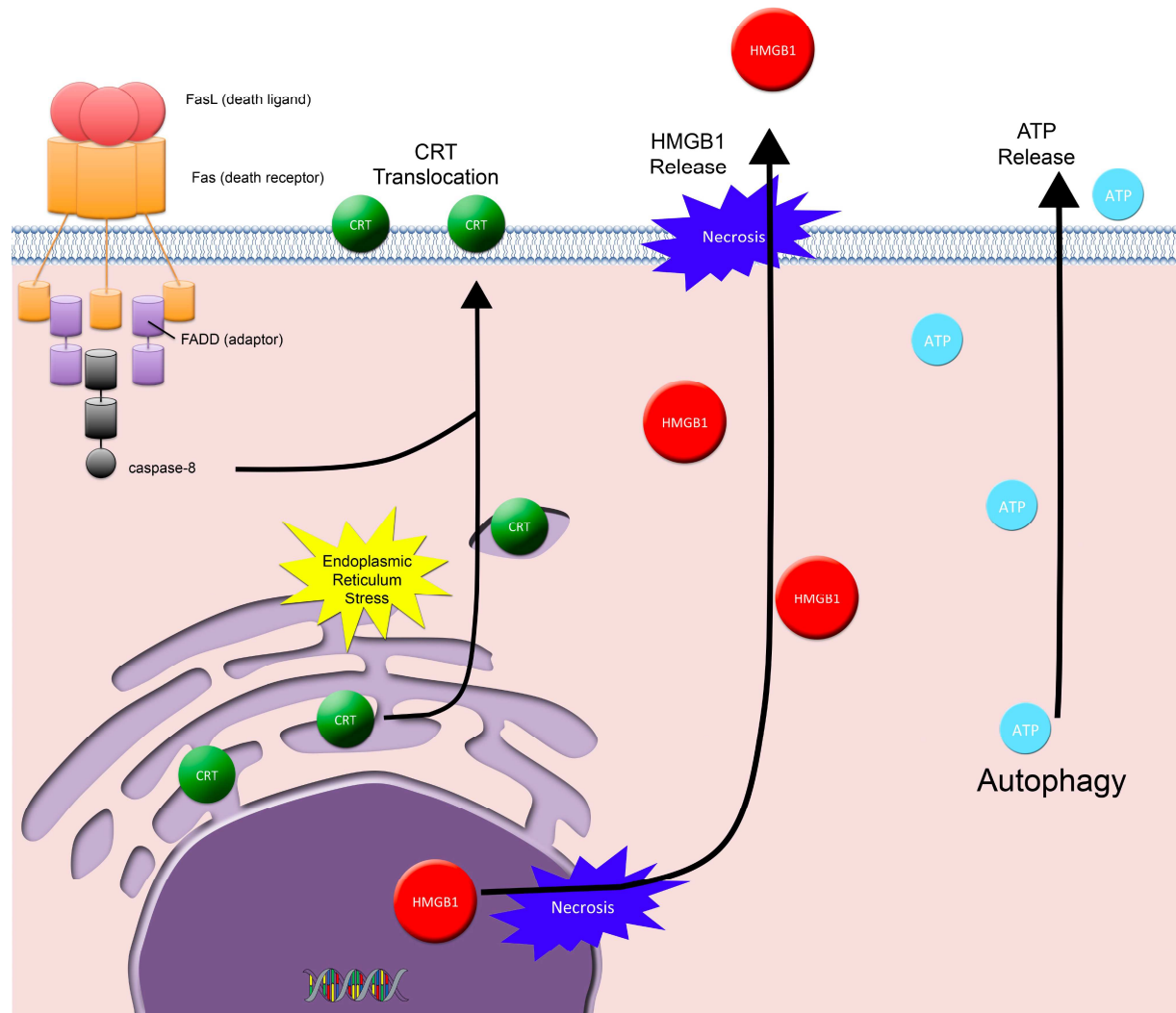
Mitotic catastrophe and senescence



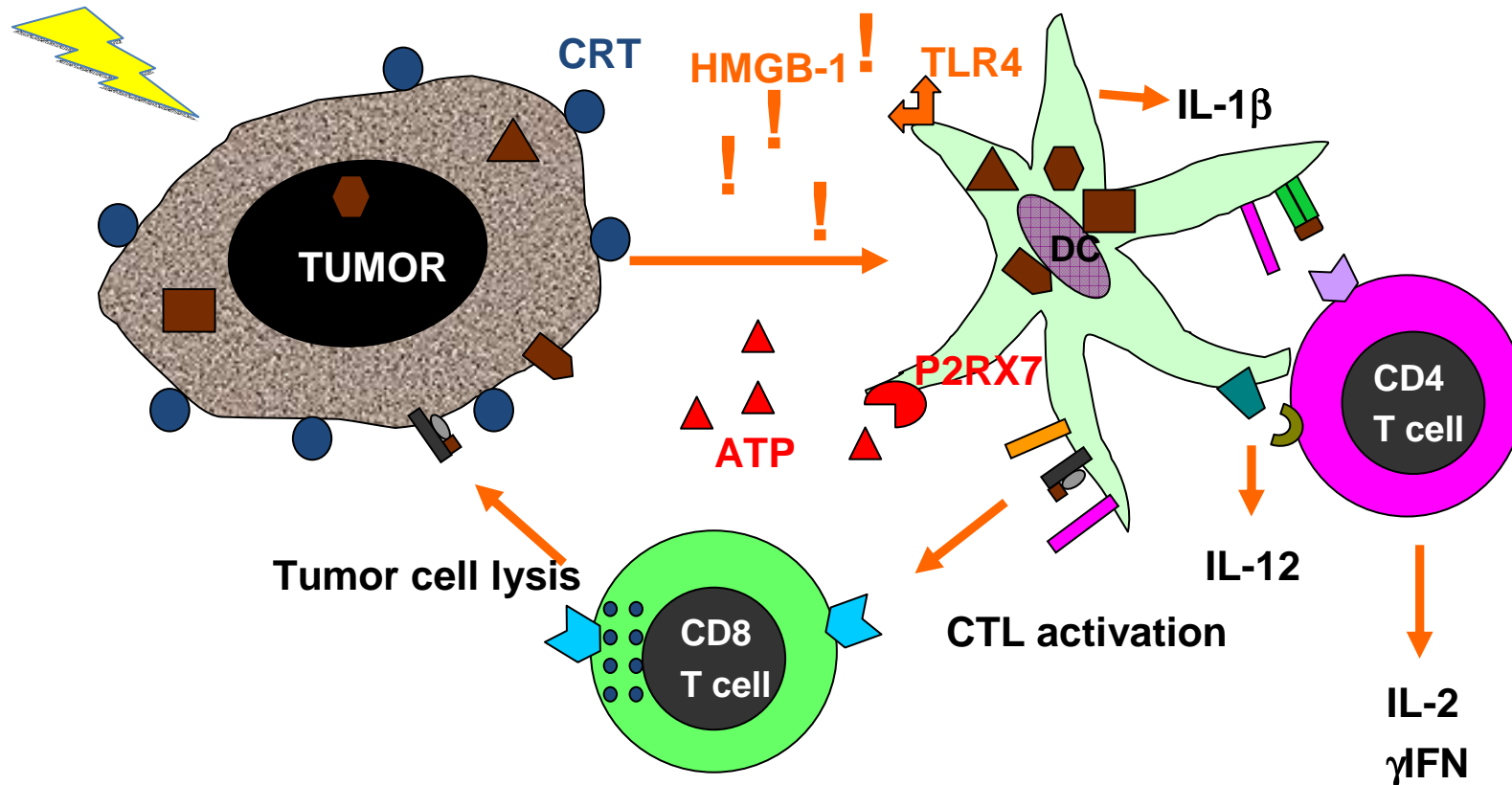
Autophagy



Immunogenic cell death (ICD)



Cross-priming of anti-tumor T cells

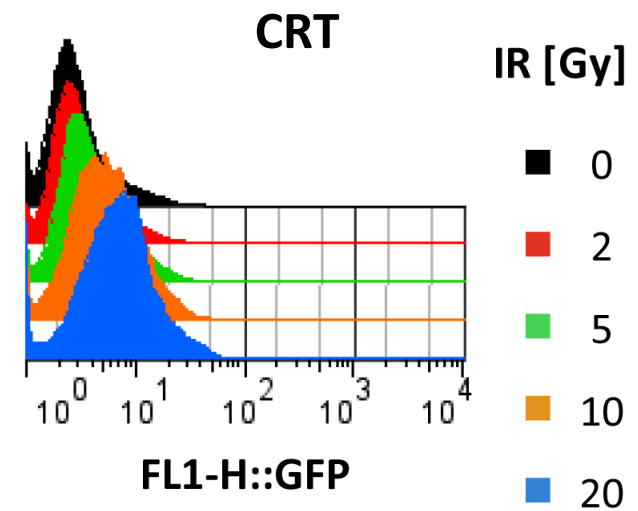
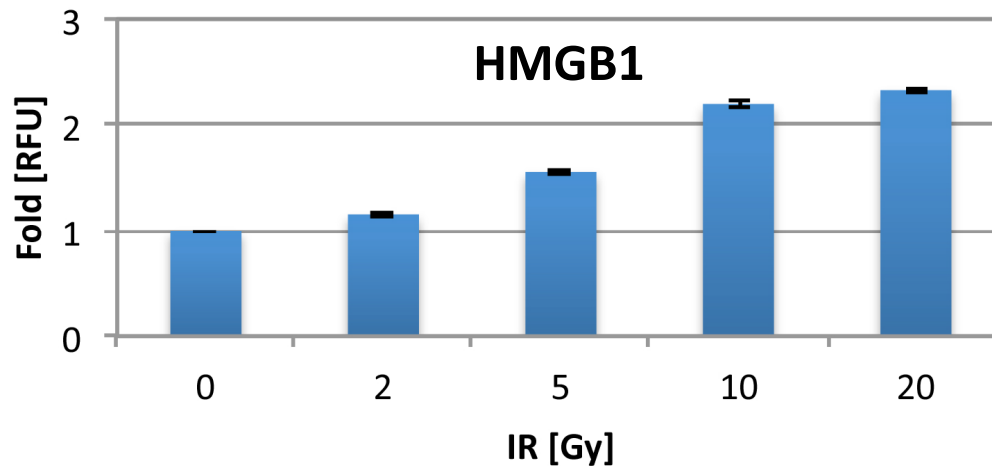
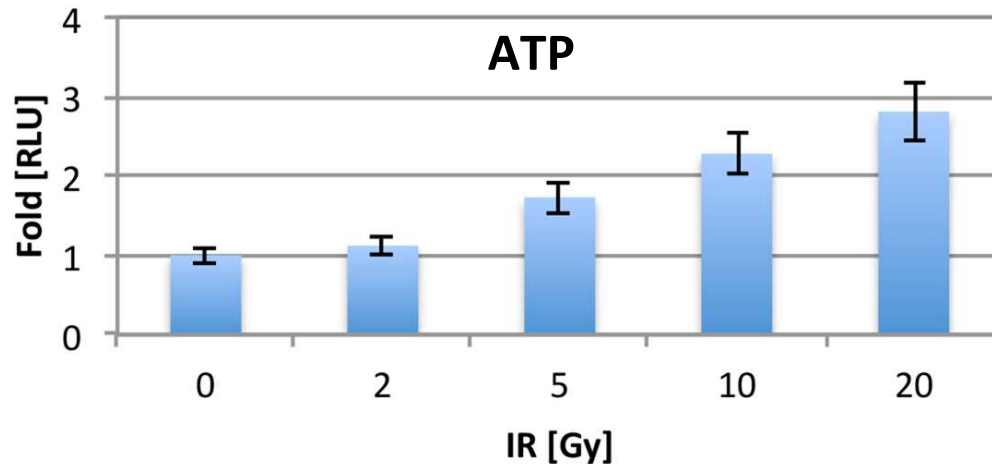


CRT, the “eat me” signal calreticulin translocates to cell surface (Obeid et al., Nat Med 2007, 13:54-61; Cell Death Differ 2007, 14:1848)

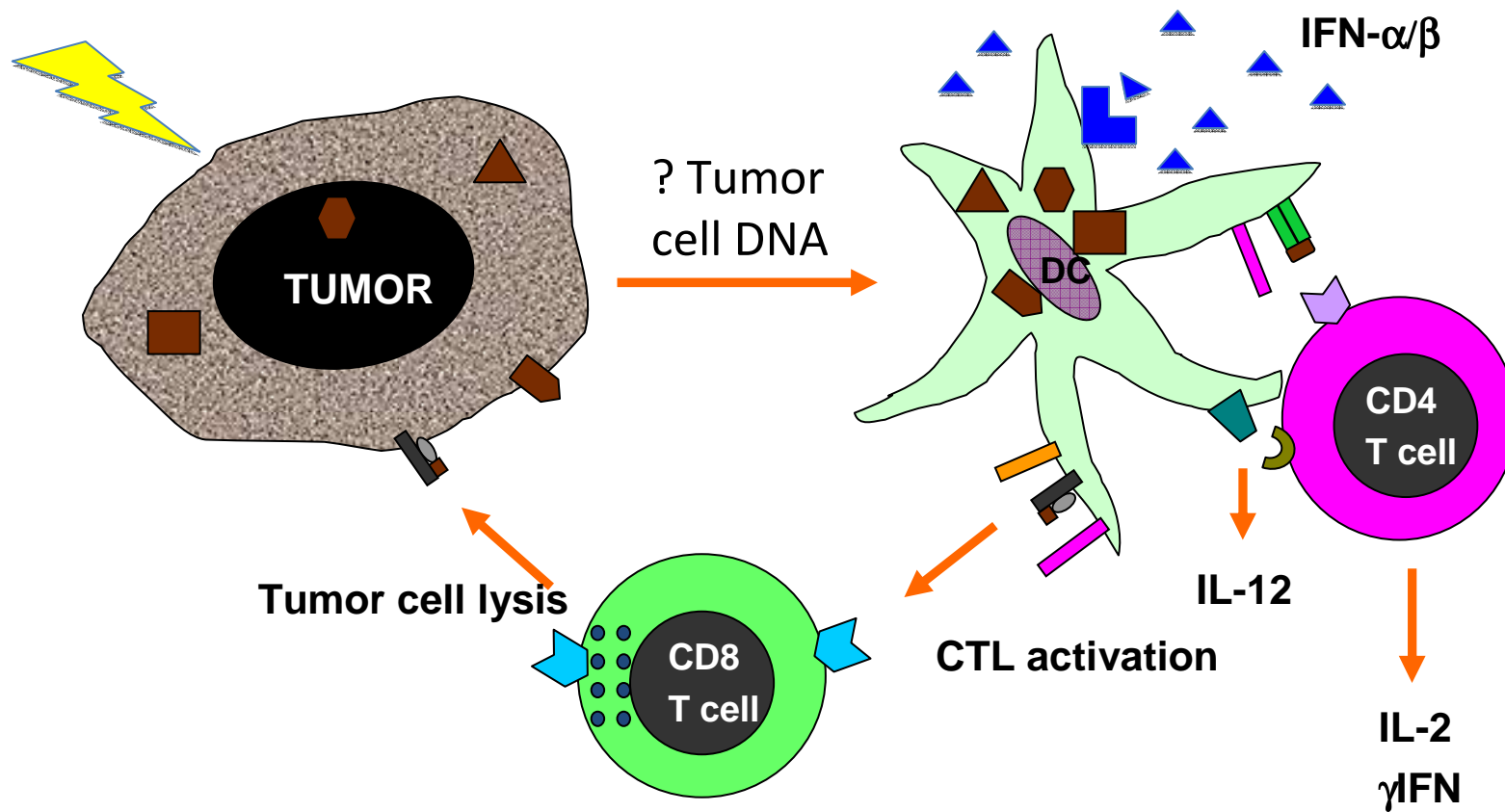
HMGB-1, a damage associated molecular pattern (DAMP) binds to TLR4 to promote cross-presentation of tumor-derived antigens (Apetoh et al., Nat Med 2007, 13:1050)

ATP released by dying cells binds to P2RX7 purinergic receptor leading to inflammasome activation and IL-1 β production (Ghiringhelli et al., Nat Med 2009, 15:1170)

Immunogenic cell death (ICD)



Cross-priming of anti-tumor T cells



IFN type I is produced by DC infiltrating irradiated tumors (Burnette et al., Cancer Res 2011; 71(7); 2488–96)

Effects of radiation on the tumor microenvironment

Increased expression by irradiated tumor/stromal cells of:

MHC class I molecules

Stress-induced ligand (NKG2D ligands)

ICAM-1, VCAM

Co-stimulatory molecules (CD80, CD86)

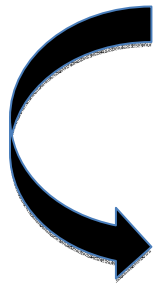
Death Receptors (Fas/CD95)

Tumor antigens (e.g., CEA, MUC-1)

Chemokines (e.g., CXCL16, CXCL10, CXCL9)

Cytokines (e.g., IL-1 β , TNF α , IFN type I)

Hsp70

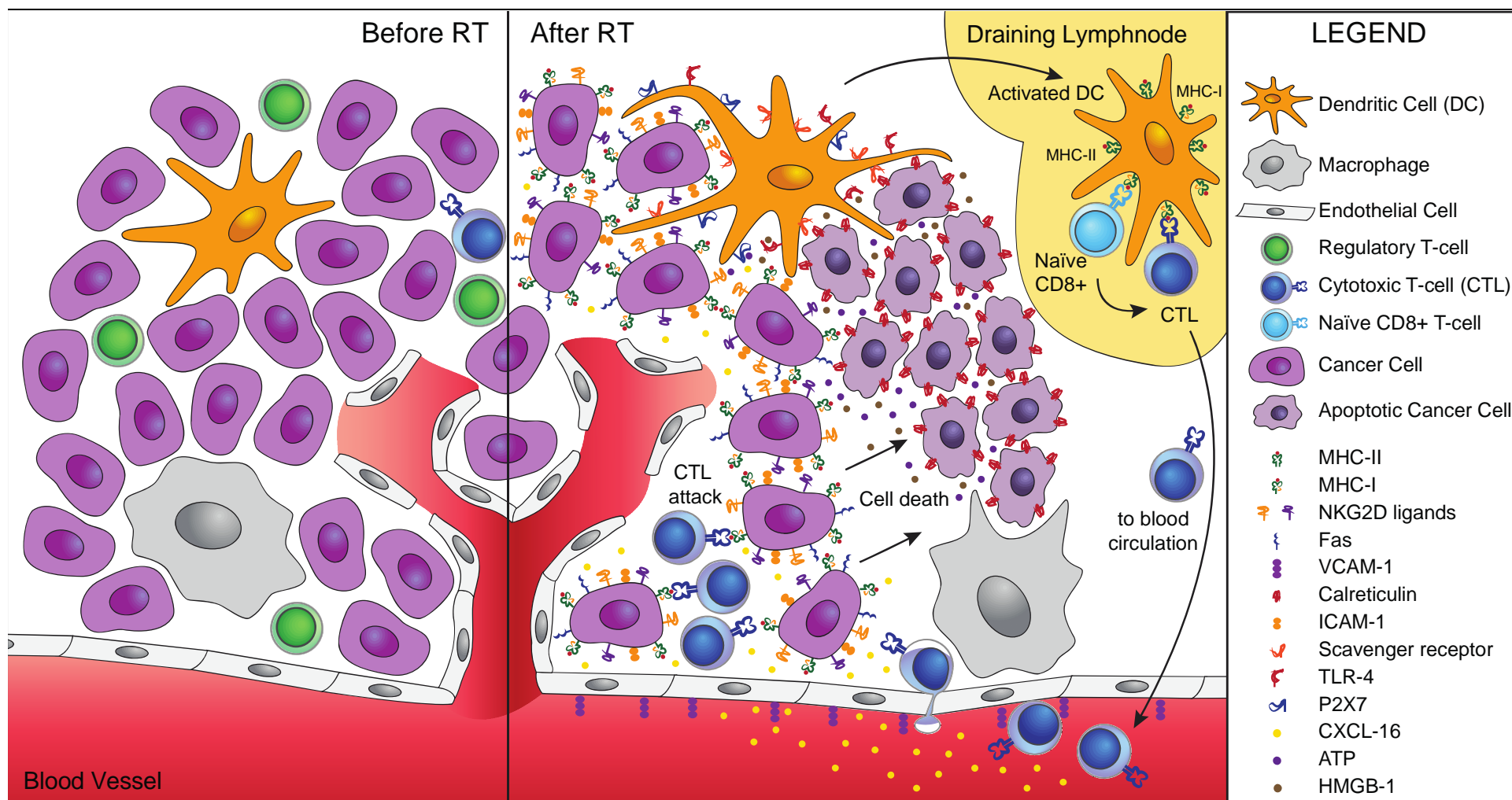


Increased DC and T cell recruitment and infiltration

Increased interactions between effector T cells and tumor cells

Increased tumor cell killing

Radiotherapy converts the tumor into an immunogenic hub



Effects of radiation that can hinder the development of anti-tumor immunity

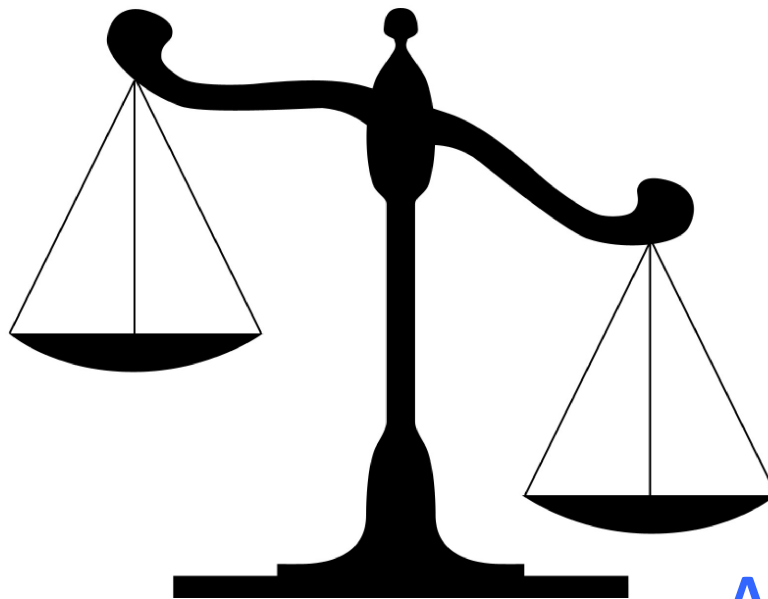
- Induction of Treg cells (*Schaue et al., Front Oncol 2012*)
- Activation of TGF β (*Barcellos-Hoff et al., J Clin Invest 1994*)
- Up-regulation of PDL-1 (*Deng et al., J Clin Invest 2014*)
- Induction of pro-tumorigenic M2 macrophages (*Tsai et al., IJROBP 2007*)
- Induction/activation of STAT-3, Bcl-XL, VEGF (*Ho et al., Cancer Sci 2010; Kargiotis et al., J Neurooncol 2010*)

Acute
inflammation

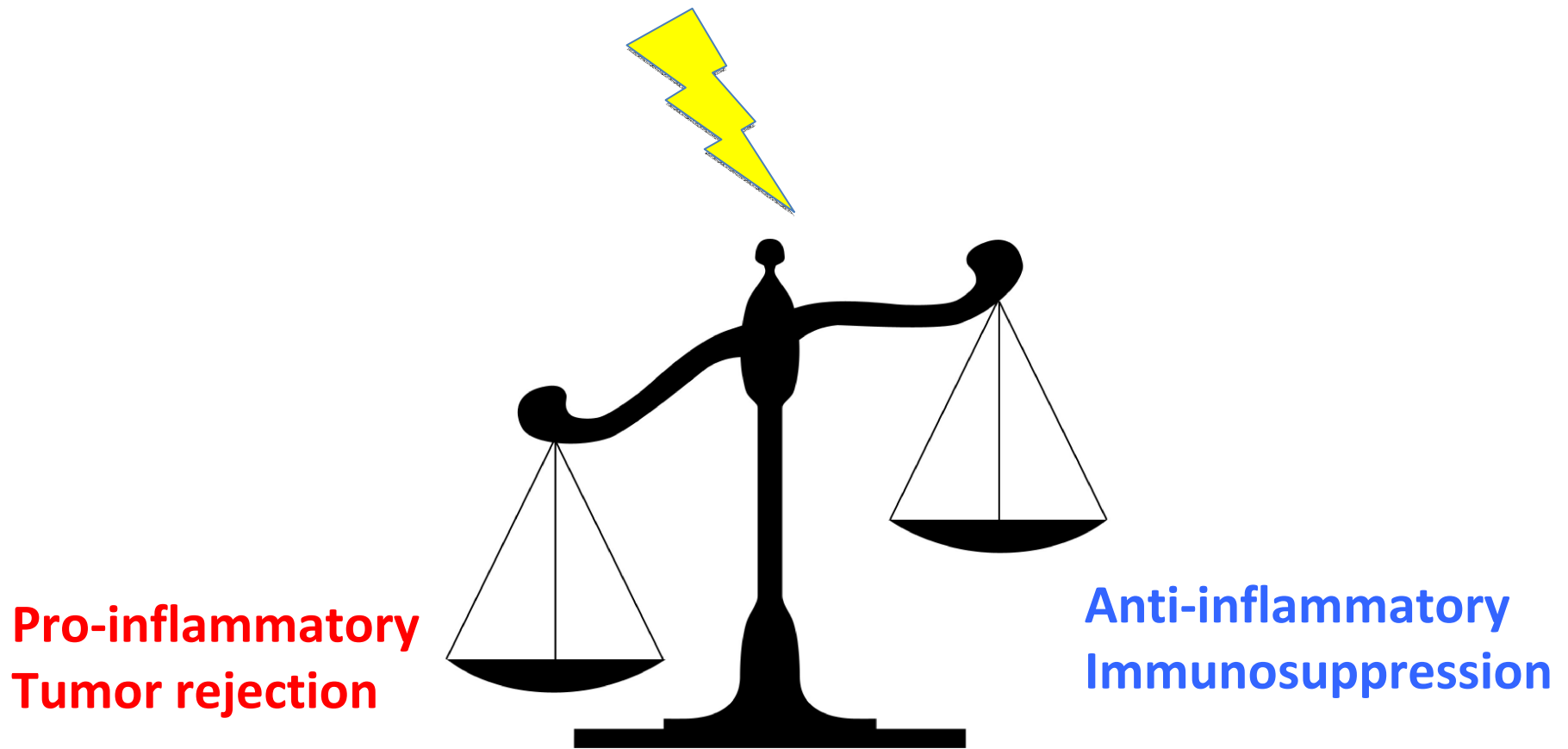


Chronic smoldering
inflammation

Pro-inflammatory
Tumor rejection



Anti-inflammatory
Immunosuppression



? Radiation dose and fractionation

Combination of radiation and immunotherapy

Effective cancer immunotherapy

IMPROVING PRIMING

- APC defects (decreased co-stimulation) Flt3-L, GM-CSF, TLR agonists, exogenous DC
- Active immune suppression (e.g., TGF β , IL-10, IDO) TGF β neutralizing mAb
- Tolerance to TAA Immune checkpoint blockade
- Vaccination GVAX, TRICOM

IMPROVING EFFECTOR PHASE

- Barriers to *homing* of activated T cells to tumors T Cell Therapy
- Barriers to *function* of activated T cells within tumors CD137 costimulation
OX40 costimulation

Pre-clinical combinations of RT and immunotherapy

Increasing APC

Flt3-Ligand

Chakravarty et al, Cancer Res 1999
Demaria et al. IJROBP 2004

Exogenous DC

Nikitina et al., Int J Cancer 2001
Teitz-Tennenbaum et al, Cancer Res 2003
Kim et al., Int J Cancer 2004

Increasing danger signals

TLR9 agonists

Milas et al., Cancer Res 2004
Mason et al., Clin Cancer Res 2005
Zhang et al., PlosOne 2012

TLR7 agonists

Dewan et al., Clin Cancer Res 2012
Dovedi et al., Blood 2013

Pre-clinical combinations of RT and immunotherapy

Improving T cell co-stimulation 2010

Anti-CD137 mAb

Shi et al., Anticancer Res 2006

Newcomb et al., Radiat Res

Anti-OX40 mAb

Gough et al., J. Immunother 2010

Blocking immune checkpoints

Anti-CTLA-4 mAb

Demaria et al., Clin Cancer Res 2005

Dewan et al., Clin Cancer Res 2009

Anti-PD-1 mAb

Verbrugge et al., Cancer Res 2012

Zeng et al., IJROBP 2013

Anti-PDL-1 mAb

Deng et al., J Clin Invest 2014

Pre-clinical combinations of RT and immunotherapy

Priming anti-tumor T cells

Vaccinia and avipox CEA vaccines Chakraborty et al., Cancer Res 2004

GM-CSF tumor cell vaccine (GVAX) Newcomb et al., Clin Cancer Res 2006

Adoptive T cell transfer

Effector CD8 T cells Chakraborty et al., J Immunol 2003
Reits et al., J Exp Med 2006

CD8 and CD4 T cells Klug et al., Cancer Cell 2013

Clinical Translation

DC administration

Hepatocellular carcinoma: DC i.t. 2 days after one dose radiation: partial response in 2/14 patients (*Chi et al., J Immunother 2005*)

Sarcoma: DC i.t. during multi-fraction neoadjuvant radiation: tumor-specific immune responses in 9/17 patients (*Finkelstein et al., IJROBP 2012*)

Vaccination

Prostate carcinoma: Poxviral vaccine expressing PSA with standard multi fraction radiation starting vaccination before radiation: PSA-specific T cell in 13/17 patients plus evidence of antigenic cascade (*Gulley et al., Clin Cancer Res 2005*)

Clinical Translation

TLR9 agonists

Lymphoma: phase I/II study 2 Gy x 2 plus i.t. synthetic CpG. Abscopal responses in 27% with one complete response, three partial responses, and eight patients with stable disease (*Brody et al., JCO 2009*)

Second trial performed in Mycosis fungoides (*Kim et al., Blood. 2012*).

TLR7 agonists

Breast cancer with cutaneous metastases: phase I/II study 6 Gy x 5 topical imiquimod (NCT01421017).

High dose IL-2

Melanoma and RCC (phase I): SBRT followed by high dose IL-2. Response rate of 66% was observed, higher than historical comparisons. Patients who responded showed a higher effector memory T cell phenotype (*Seung et al., Sci Transl Med. 2012*).

Phase II randomized study of SBRT (20 Gy) and high-dose IL-2 versus IL-2 alone in patients with metastatic melanoma is ongoing (NCT01416831).

Phase II in metastatic RCC (NCT01896271).

Clinical Translation

Anti-CTLA-4 (Ipilimumab)

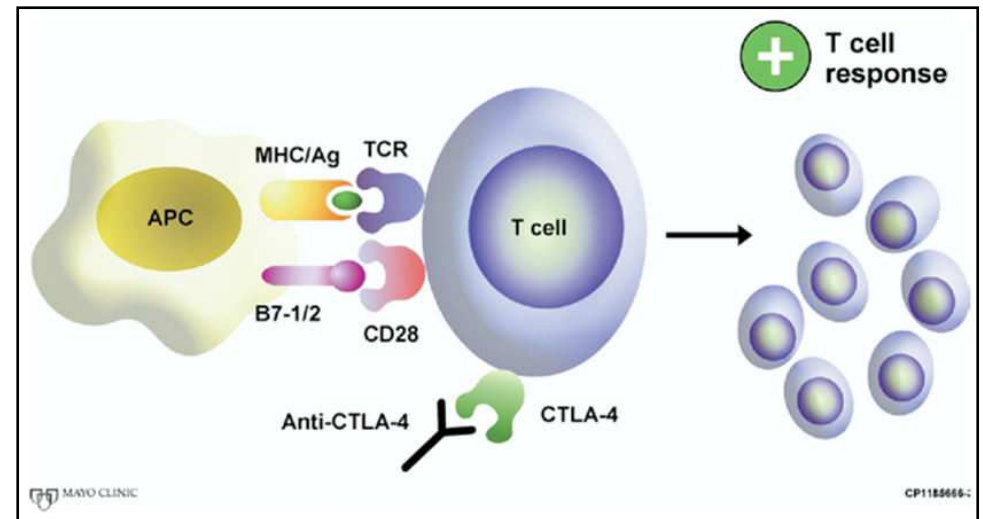
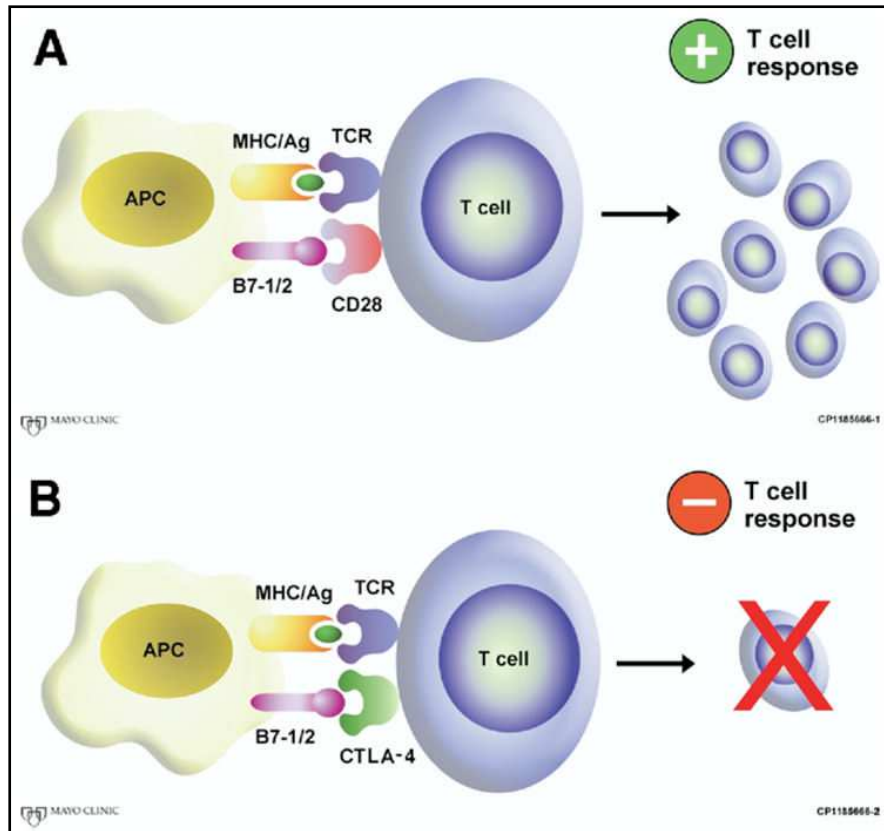
Metastatic melanoma: Case report of a patient with progressive disease on Ipilimumab who showed a dramatic abscopal effect after radiotherapy (9.5 Gy x 3) to one site, accompanied by immunological changes (*Postow et al., NEJM 2012*).

Data are supported by pre-clinical data demonstrating that tumors unresponsive to anti-CTLA-4 show abscopal responses when one site is irradiated (*Dewan et al., Clin Cancer Res 2009*)

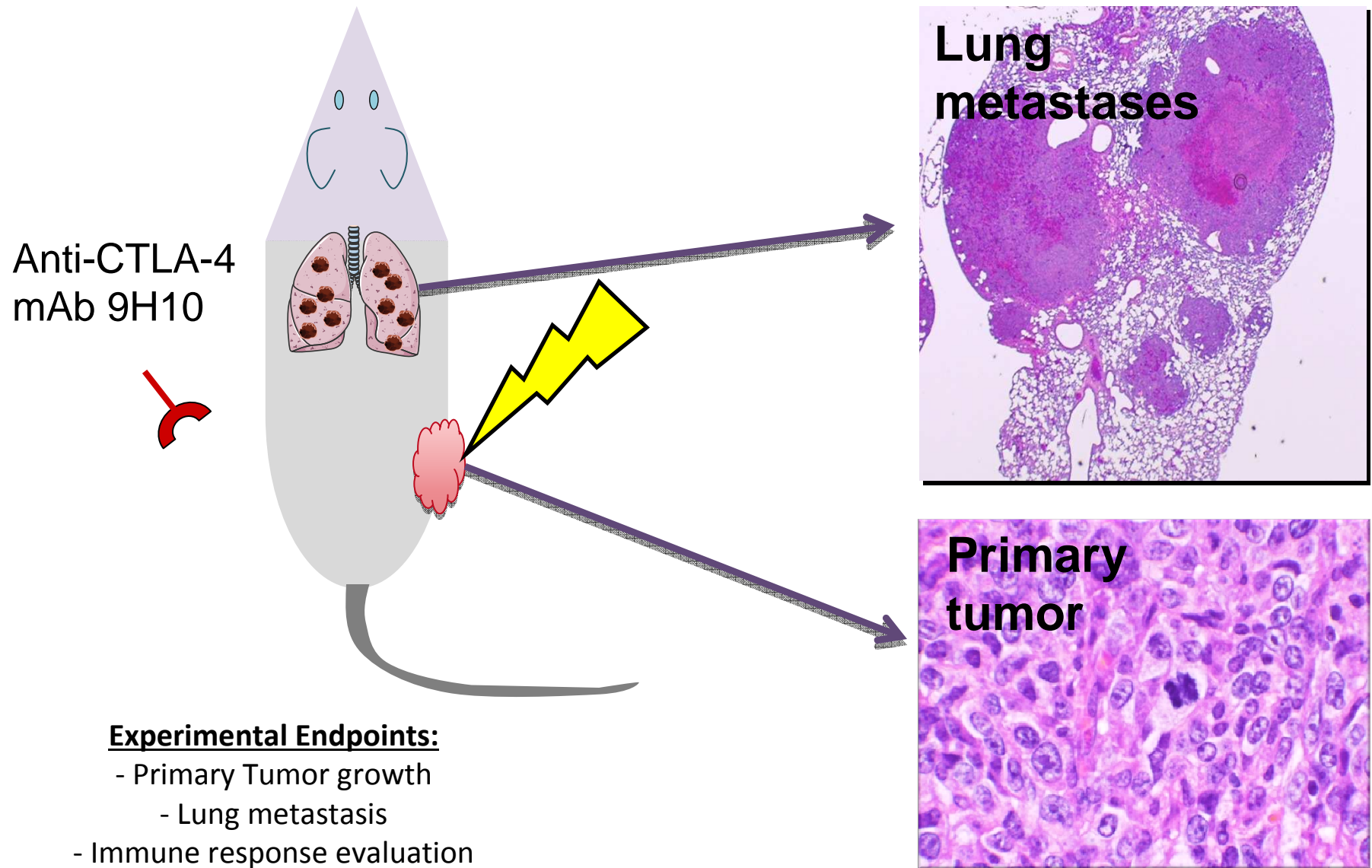
Currently >20 trials testing combination of radiotherapy and Ipilimumab are underway.

Synergy of radiation and anti-CTLA-4
mAb: a complex interaction

CTLA-4 is a master regulator of T cell activation



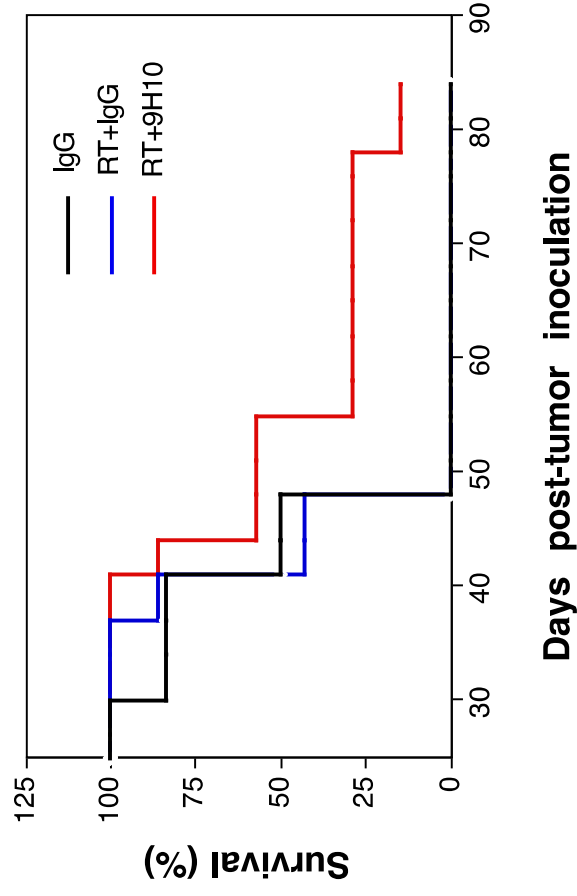
4T1 mouse model of metastatic breast cancer



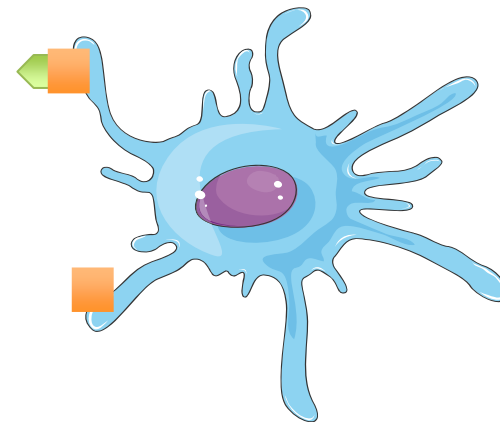
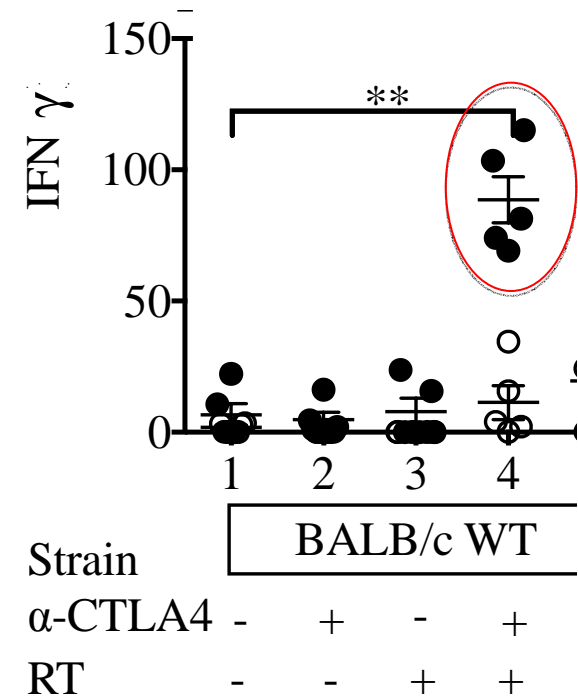
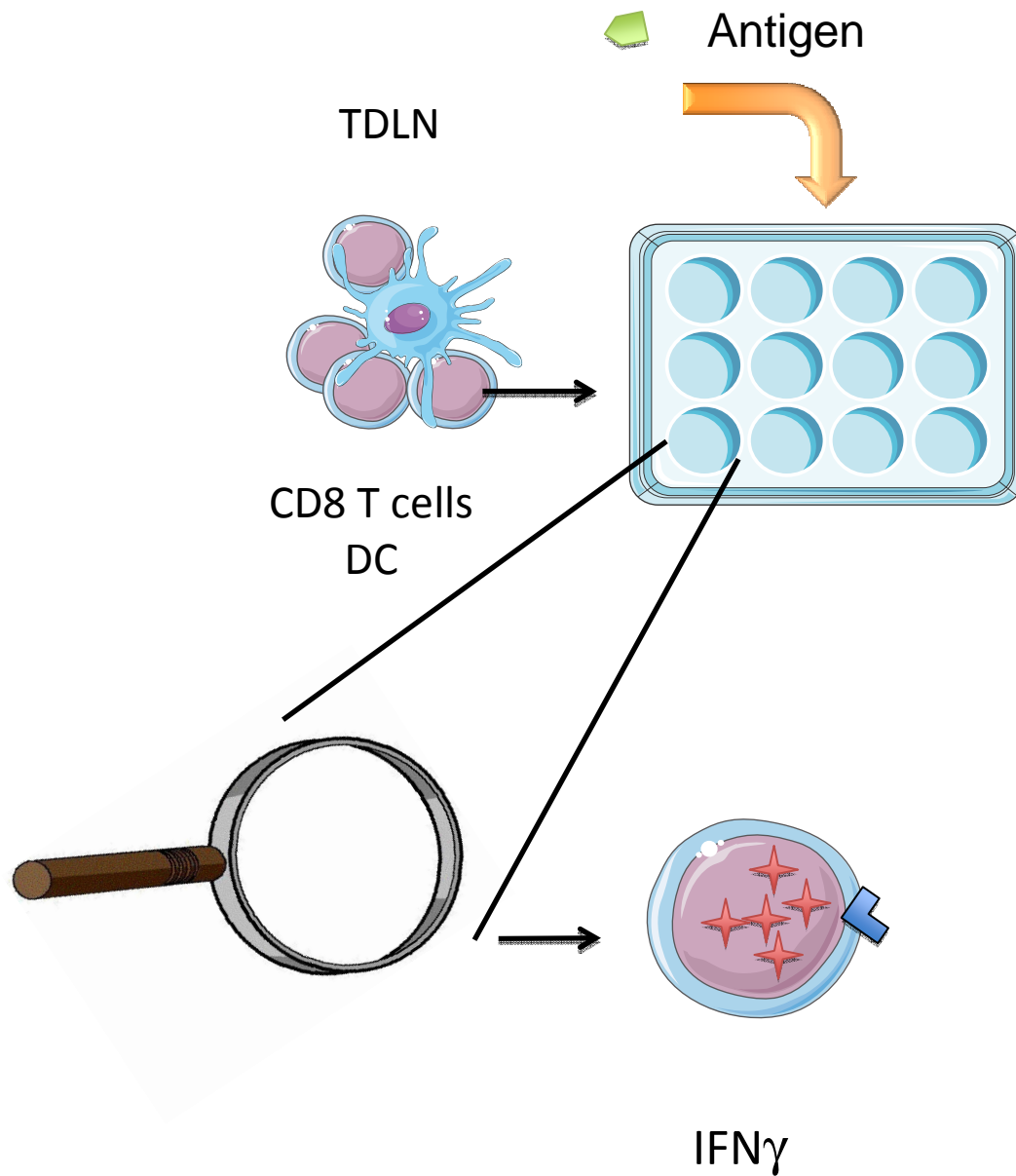
Immune-Mediated Inhibition of Metastases after Treatment with Local Radiation and CTLA-4 Blockade in a Mouse Model of Breast Cancer

Sandra Demaria,¹ Noriko Kawashima,¹
Anne Marie Yang,¹ Mary Louise Devitt,²
James S. Babb,³ James P. Allison,⁴
and Silvia C. Formenti²

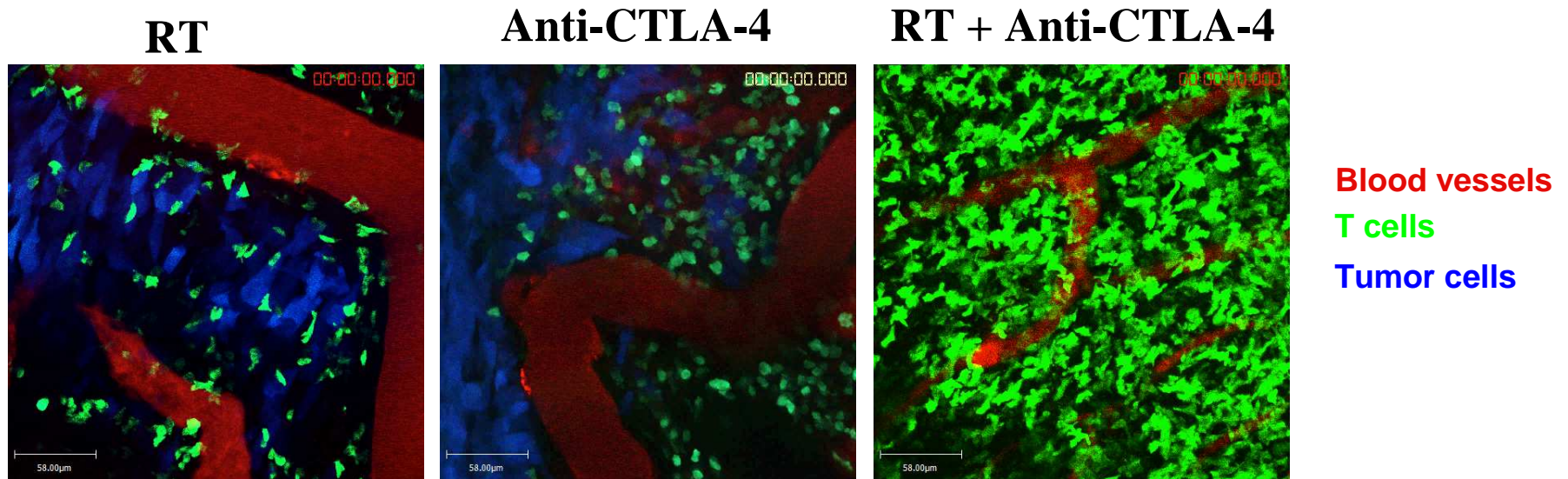
Departments of ¹Pathology, ²Radiation Oncology, and ³Radiology,
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⁴Howard Hughes Medical Institute, University of California at Berkeley,
Berkeley, California



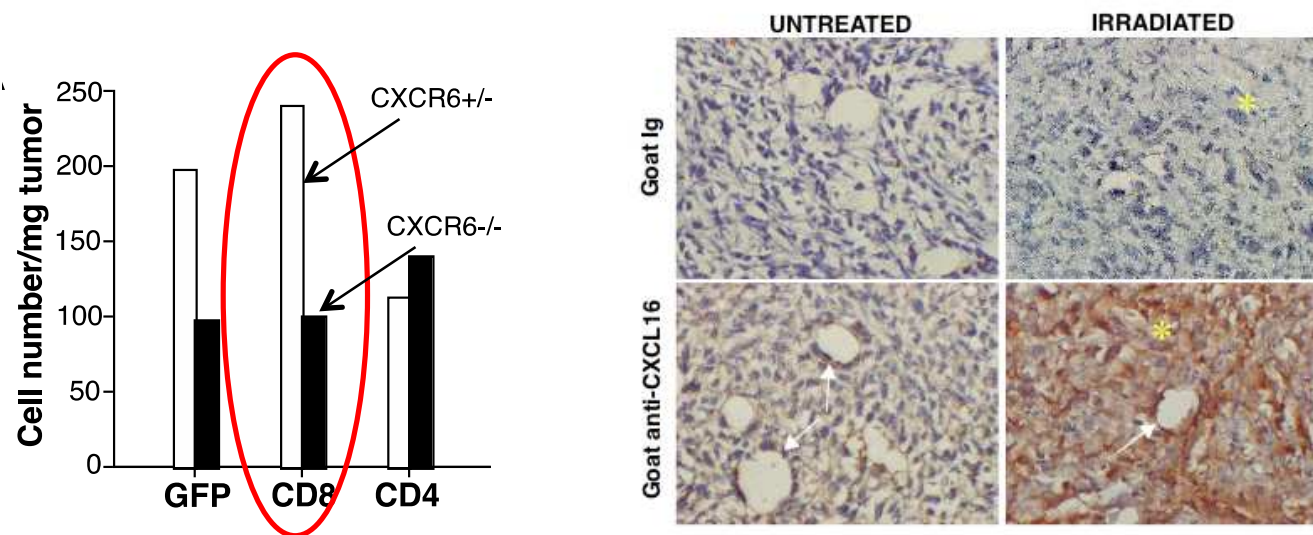
Anti-tumor CD8⁺ T cells priming



CD8⁺ T cells recruitment and tumor infiltration

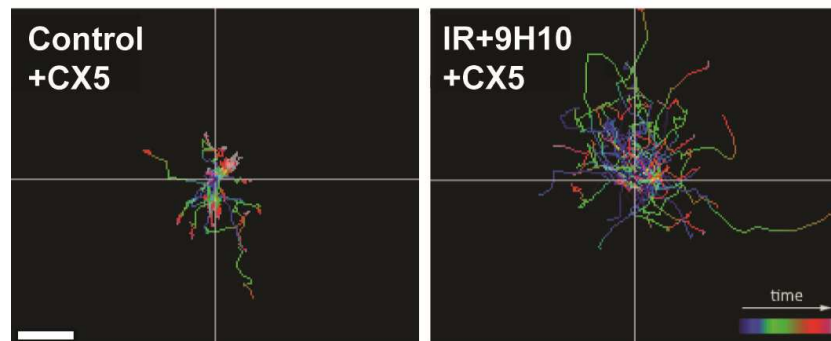
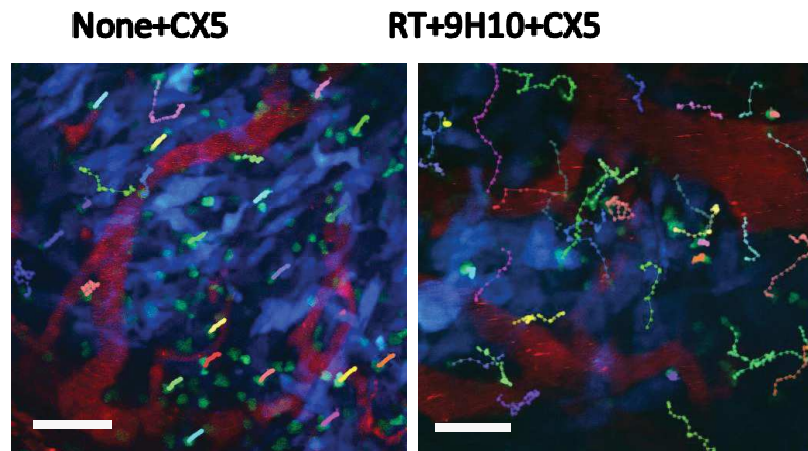


Ruocco et al., *J Clin Invest* 122:3718-30 2012

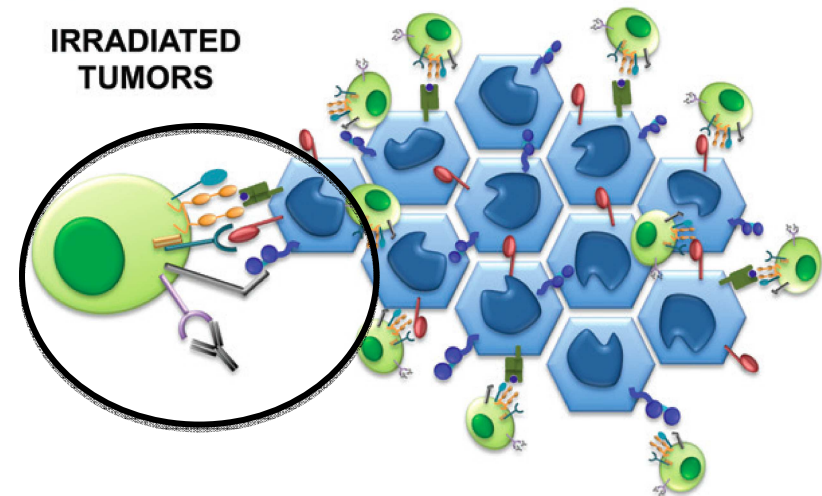
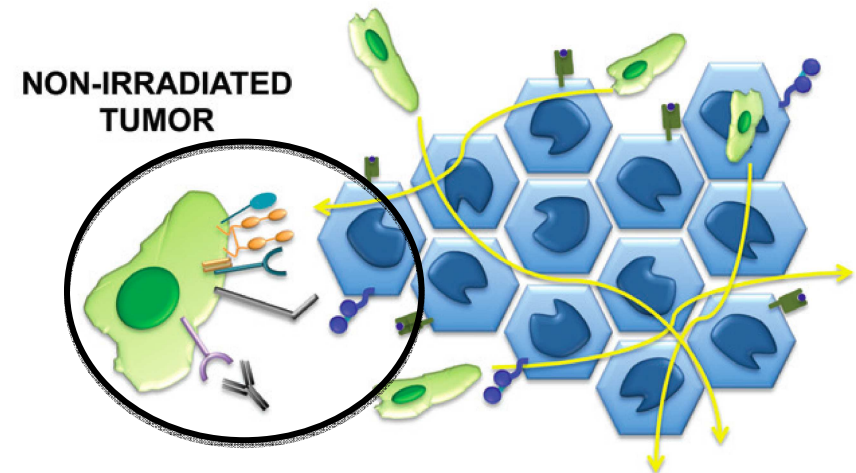


Matsumura et al., *J Immunol* 181:3099-3107 2008

Formation of immune synapse between CD8⁺ T cells and tumor cells



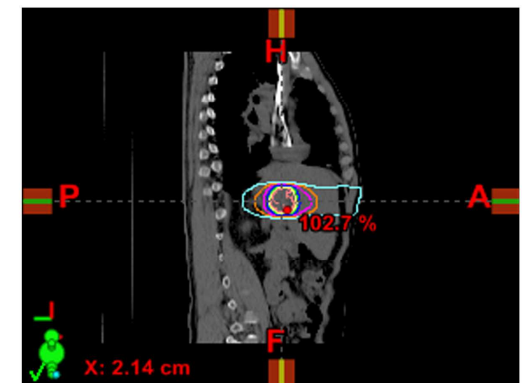
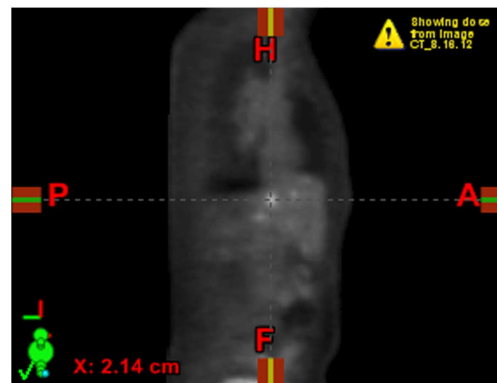
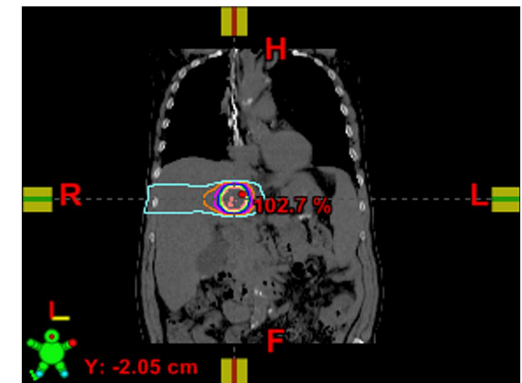
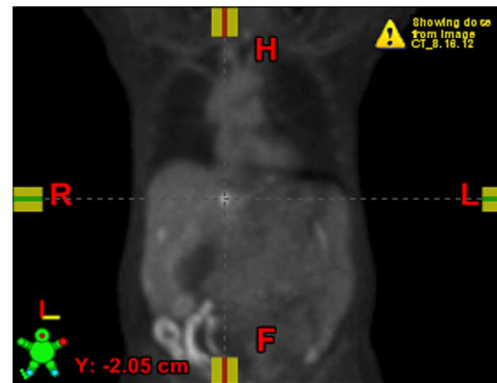
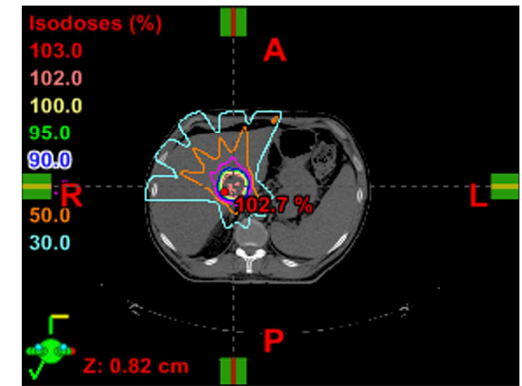
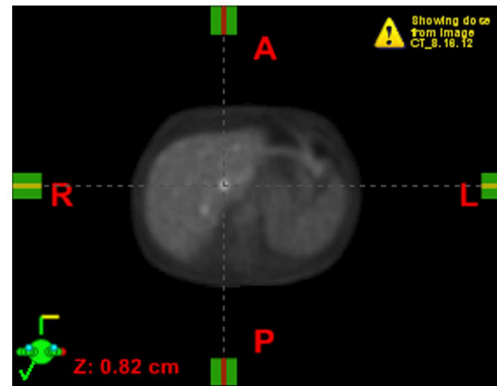
Ruocco et al., *J Clin Invest* 122:3718-30 2012



Demaria et al., *Oncolmunology* 2:3, e23127 2013

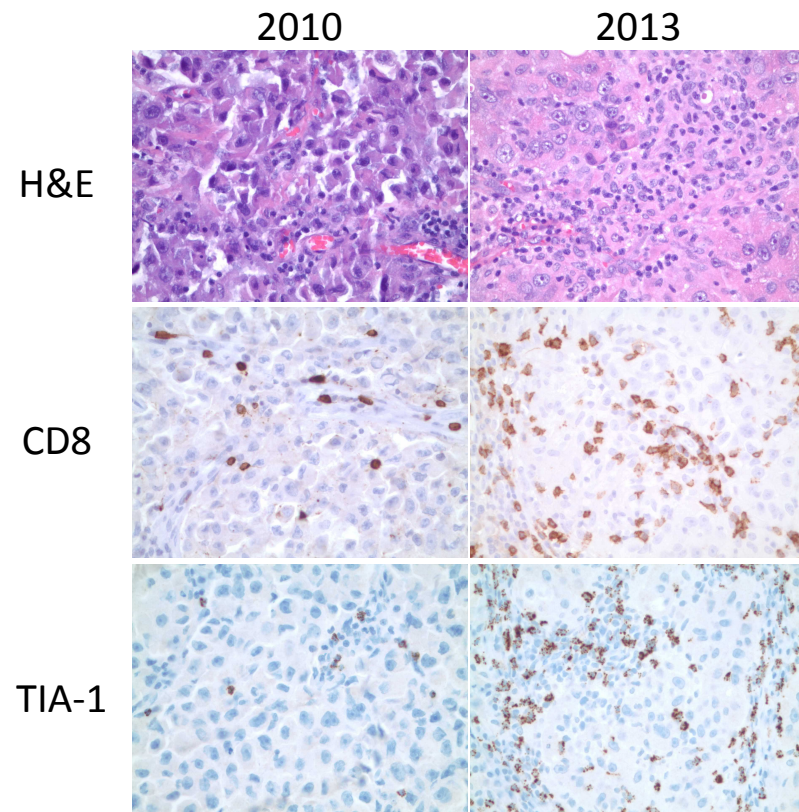
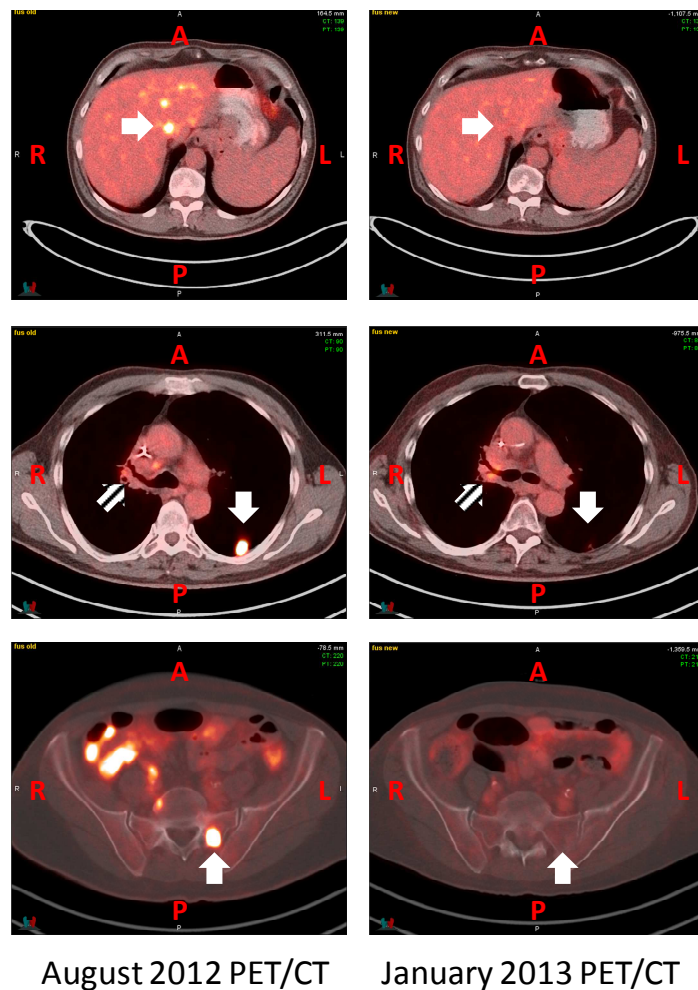
Patient with Metastatic NSCLC

Progressing after 3 lines of chemo and chest RT: Multiple lung, bone and liver metastasis



RT to one liver met
6 Gy X 5 (TD 30 GY)
Ipilimumab, 3 mg/Kg, after first RT
q3 weeks, X 4 cycles

Abscopal response in a NSCLC patient treated with local RT and Ipilimumab



Golden *et al.*, *Cancer Immunology Res.* 2013

CONCLUSIONS

- RT can promote the priming and effector phase of anti-tumor immune responses and may be able to **convert a T-cell poor into T-cell inflammed tumor**
- RT can be successfully combined with different immunotherapy strategies
- The optimal doses and regimens of RT and sequencing with immunotherapy remain to be defined and may depend on the immunotherapy used
- There is at least some evidence that RT can induce anti-tumor immune responses in patients
- Several trials testing combinations of RT and immunotherapy are ongoing