



Adoptively transferred T cells for primary and secondary CNS malignancies

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National Cancer Institute, Bethesda, Maryland

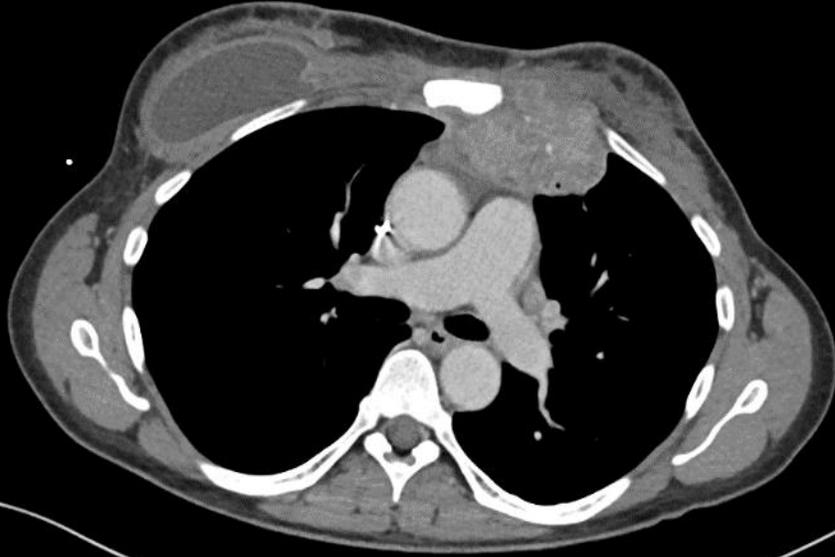
Disclosure

I have no conflicts of interest to disclose

NCI Surgery Branch CRADA: Kite Pharma
 lovance Biotherapeutics
 Ziopharm Oncology

Richard Sherry, MD, FACS, Surgery Branch
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Baseline 12/2015



**49 YO Woman with
advanced BrCa
ER+PR+ her2-**

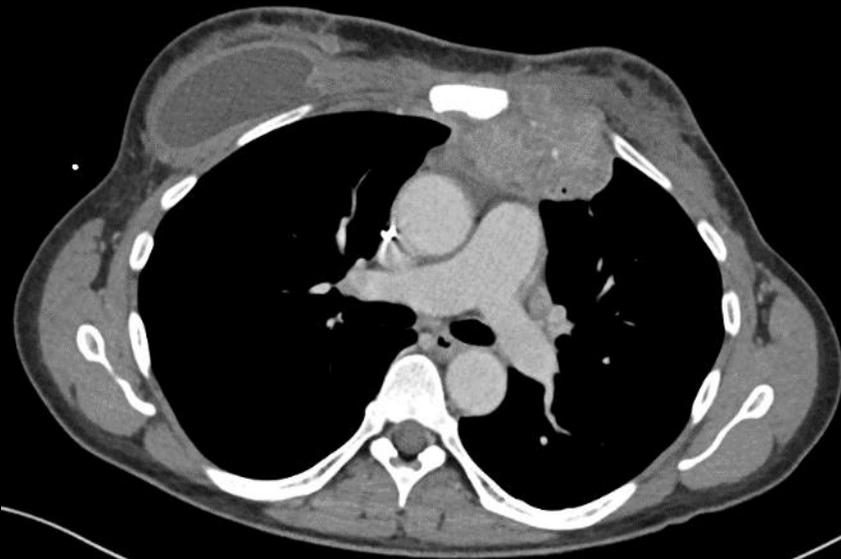
**S/p multiple hormonal
therapies**

**S/p 6 different lines of
Chemotherapy**

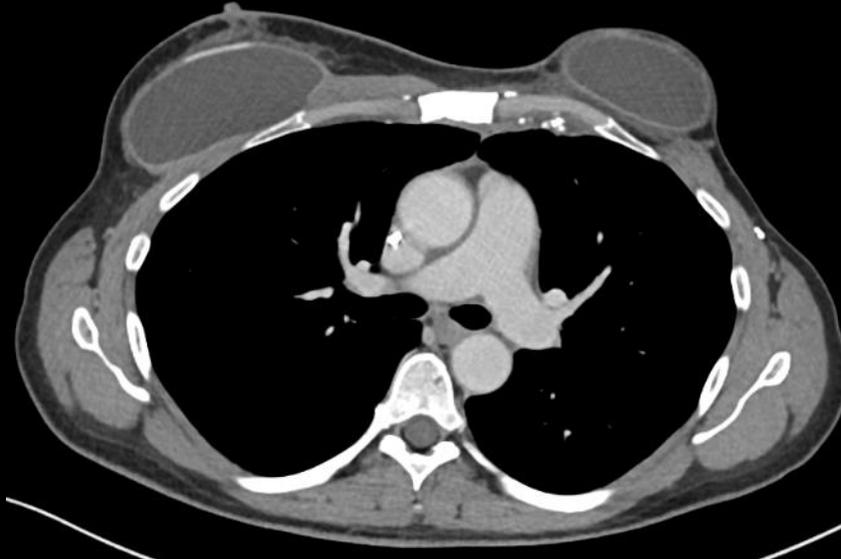


**Received TIL *selected*
for somatic mutation
reactivity
(And 4 doses Pembro)**

Baseline 12/2015



F/U 5/2019 (+40)



Adoptively transferred T cells for primary and secondary CNS malignancies

ACT Background and Principles

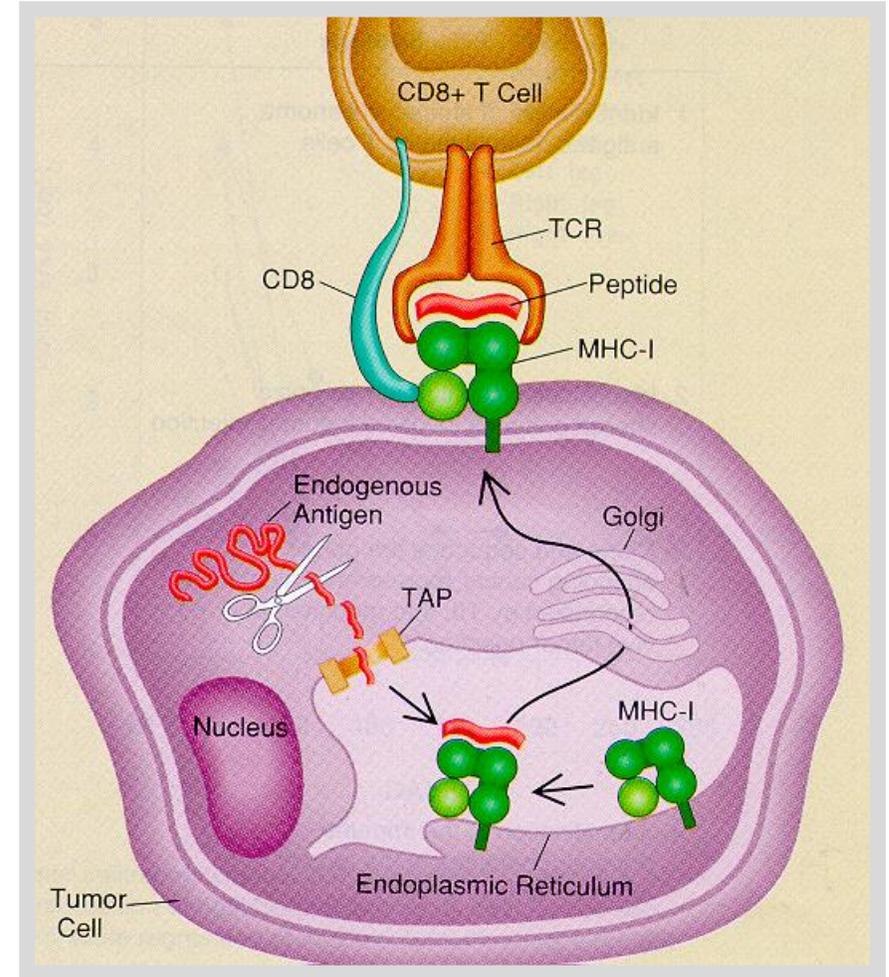
Can infused Lymphocytes effectively traffic to the brain and cause cancer regression ?

Anti-EGFRvIII CAR for patients with GBM

J Immunother 42(4):126, 2019

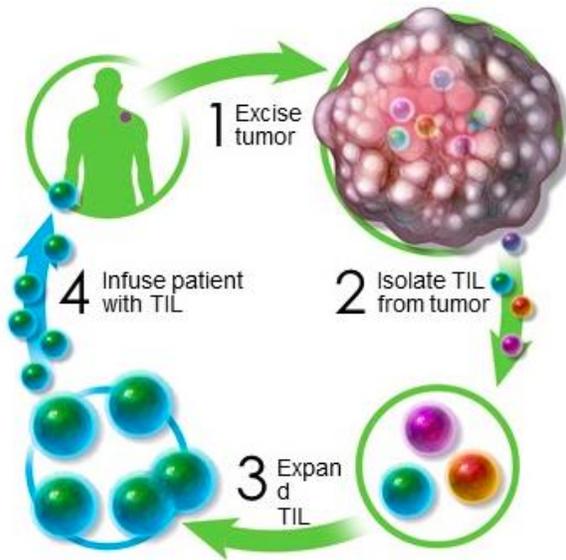
Adoptive Cellular Therapy (ACT) for Cancer

- Infusion of tumor specific T Cells in combination with lymphodepleting chemotherapy given to eliminate cancer.
- Depends on the isolation, identification, or de novo generation of appropriate effector T-Cells.



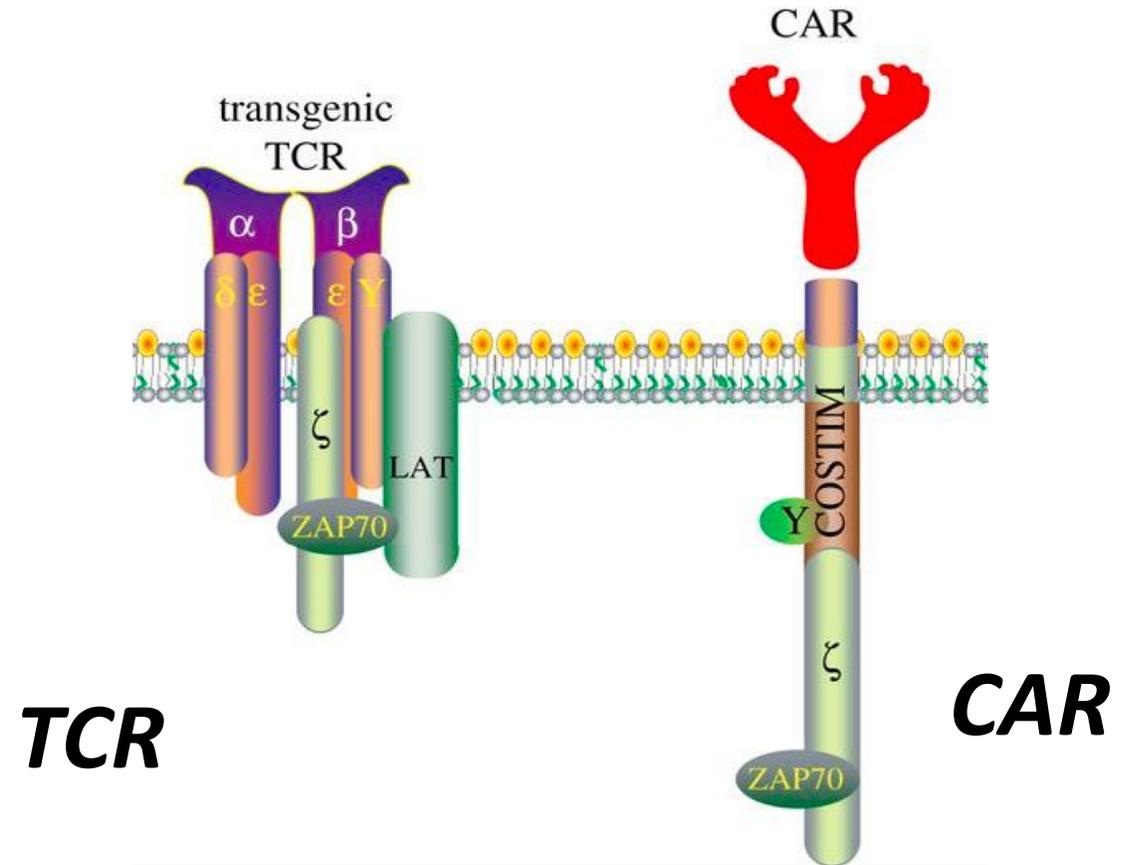
Adoptive T Cell Therapy for Cancer

TIL



Naturally occurring autologous lymphocytes derived from tumor

Gene modified PBL*



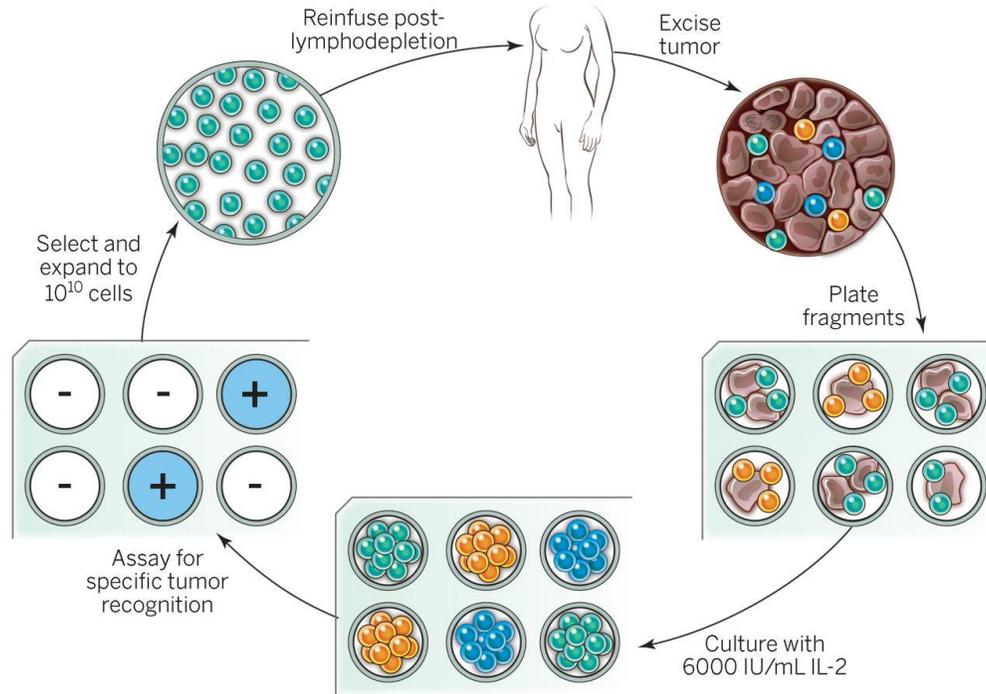
TCR

CAR

Insertion of target receptor

* Peripheral Blood Lymphocytes

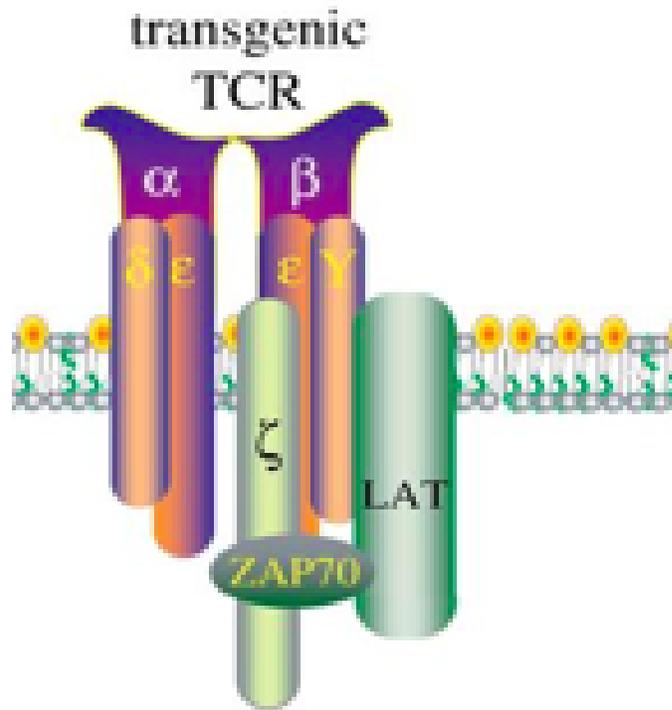
TIL



***Naturally occurring
autologous lymphocytes
derived from tumor***

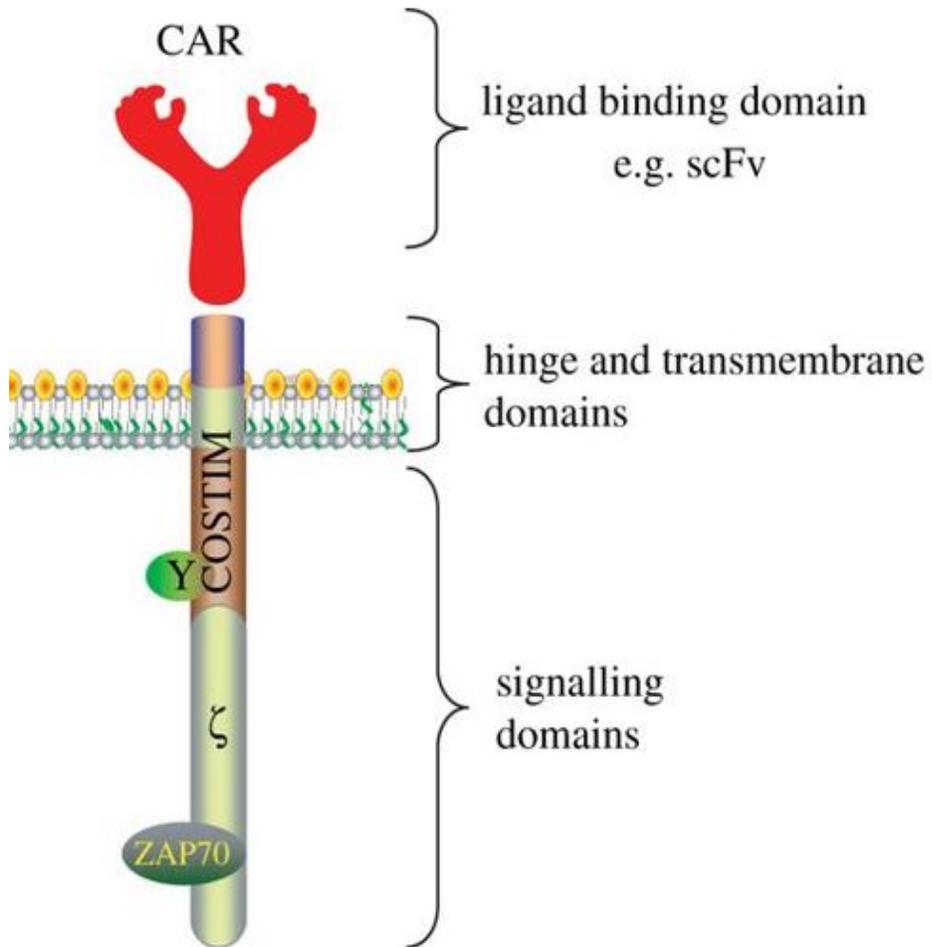
- Polyclonal
- TCR have had normal selection in the thymus
- *TCRs recognize unique mutated peptide products expressed on the cancer cell surface on HLA*

TCR



- Autologous PBL Transduced with TCR
- TCR inserted randomly into an “open repertoire”
- Each T Cells has its native unique TCR *and* the transduced TCR
- HLA presentation required

CAR



- Autologous PBL Transduced with CAR
- Each T Cell has its unique native TCR *and* the Transduced CAR
- CAR inserted randomly into an “open repertoire”
- Recognizes selected cell surface protein without HLA presentation

Adoptive Cell Transfer 2019

- *The T Cell*

Number of Cells impacts efficacy and toxicity

Phenotype of Cells? (CM, EM, N)

Persistence?

- *The Target*

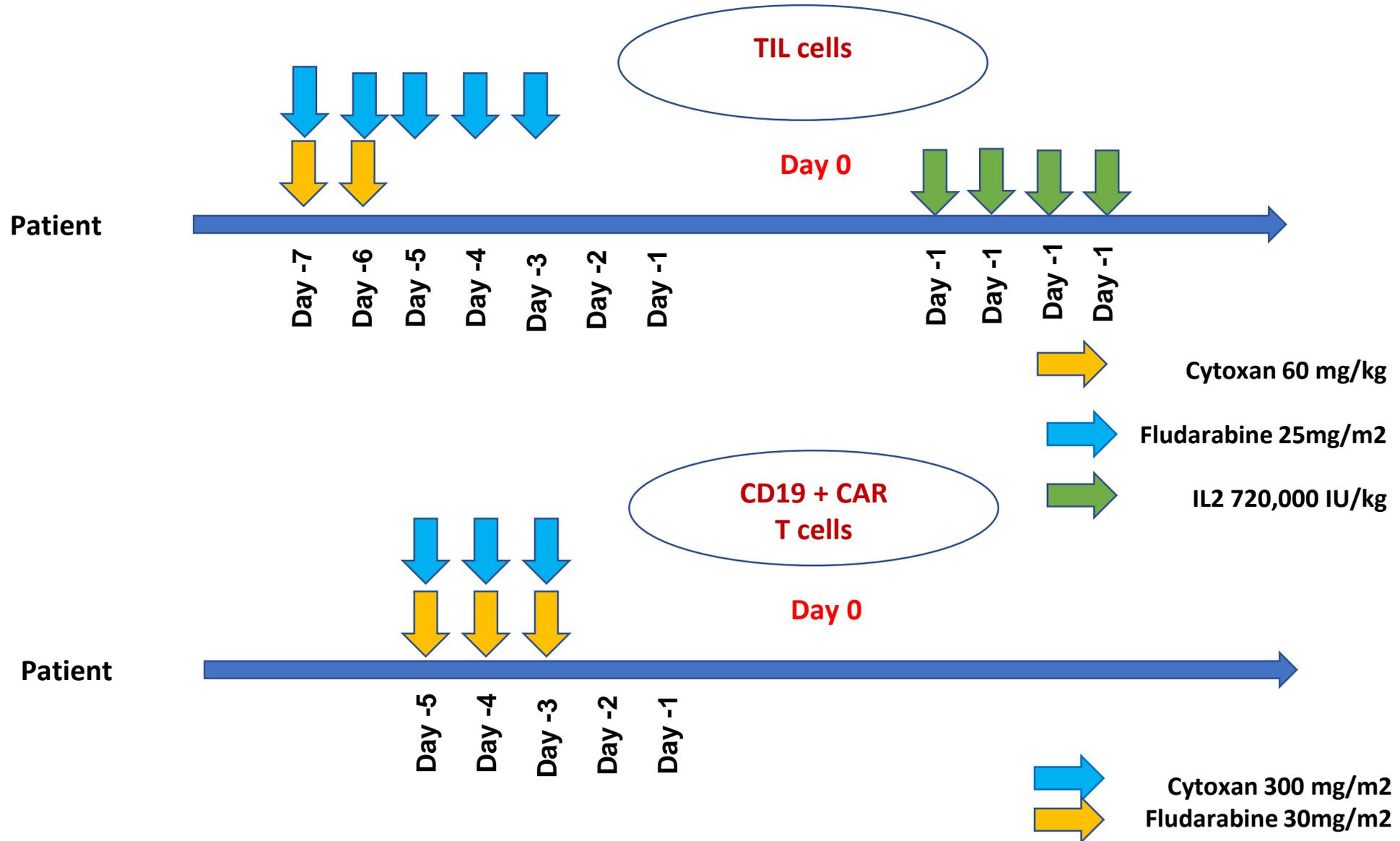
Must be on the Tumor cell surface

Antigen must be processed and presented by HLA

Tumor heterogeneity?, normal tissue expression?, Ag loss?

- *Lymphodepletion*

Typical Conditioning Regimens



Importance of lymphodepletion on ACT

1998

20 Patients advanced MM
TIL (2 E11)
HD IL-2

Cytosin 25 mg/kg X 1

40 % ORR

Durable CR 5%

Responses not durable

2016

101 Patients Advanced MM
TIL (5 E10)
HD IL-2

Cytosin 60 mg/kg X2

Fludarabine 25 mg/m² X5

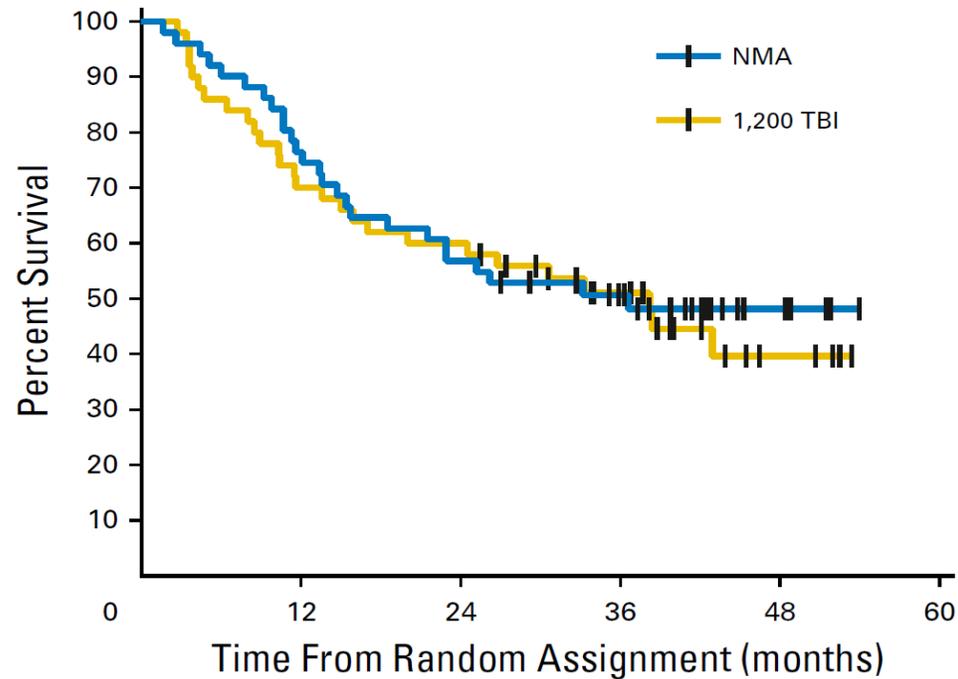
56% ORR

Durable CR/PR 36%

Responses durable

Adoptive Cell Therapy with Autologous Tumor-infiltrating Lymphocytes for Patients with Metastatic Melanoma

A

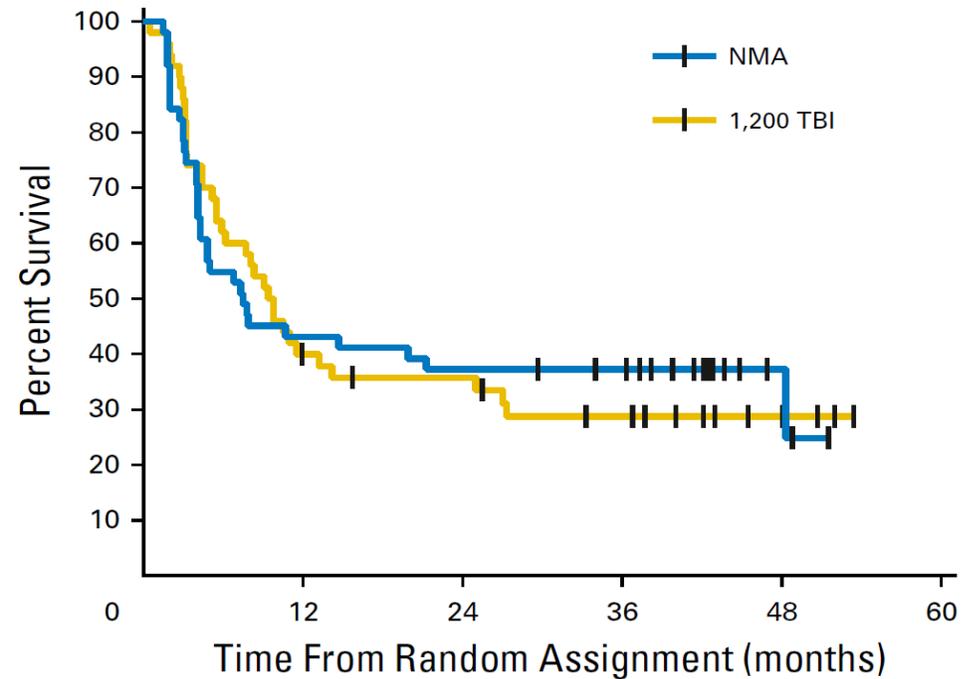


No. at risk

NMA	51	39	30	21	6	0
1,200 TBI	50	35	30	18	4	0

OS

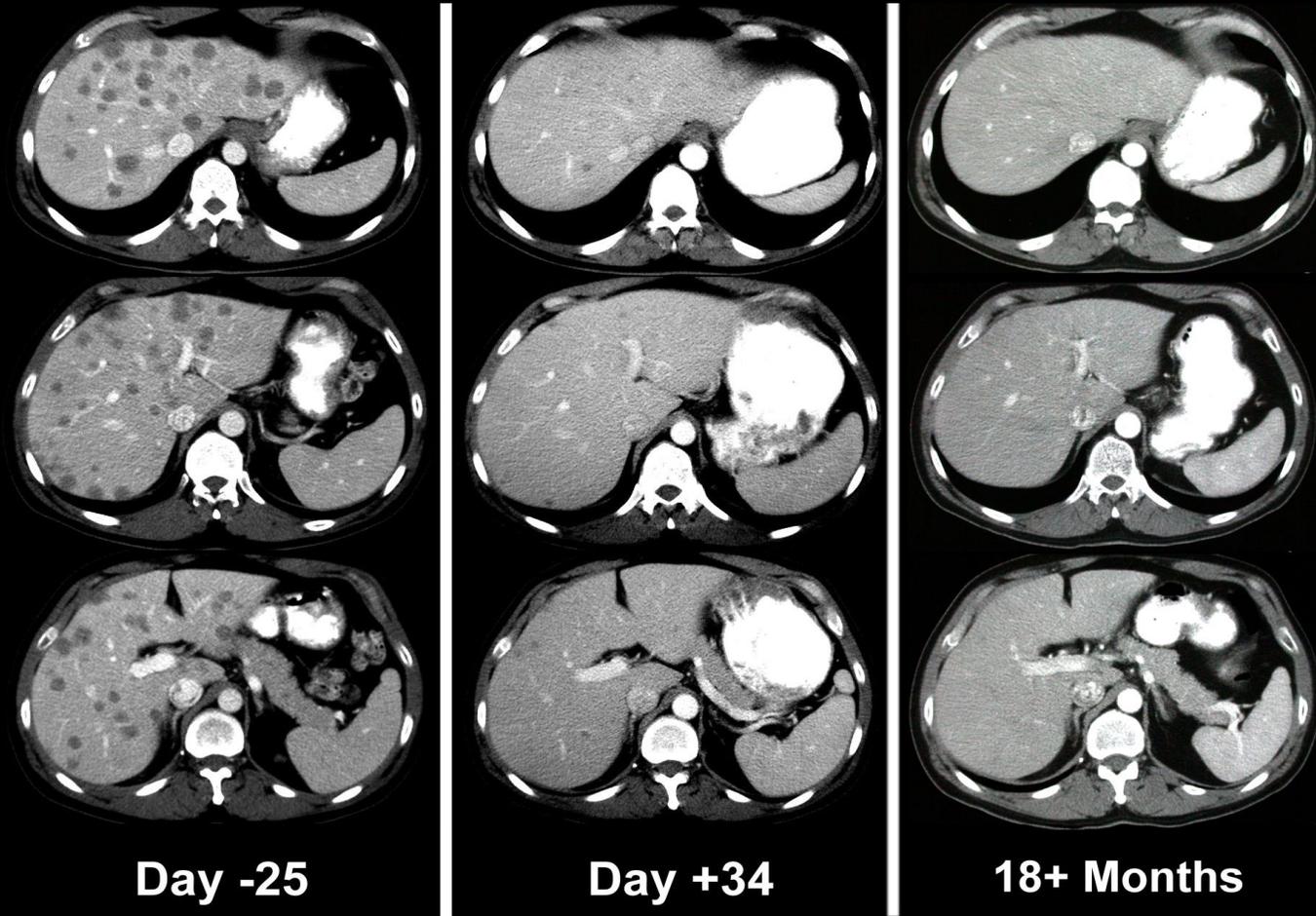
B



NMA	51	22	19	17	3	0
1,200 TBI	50	18	16	10	4	0

PFS

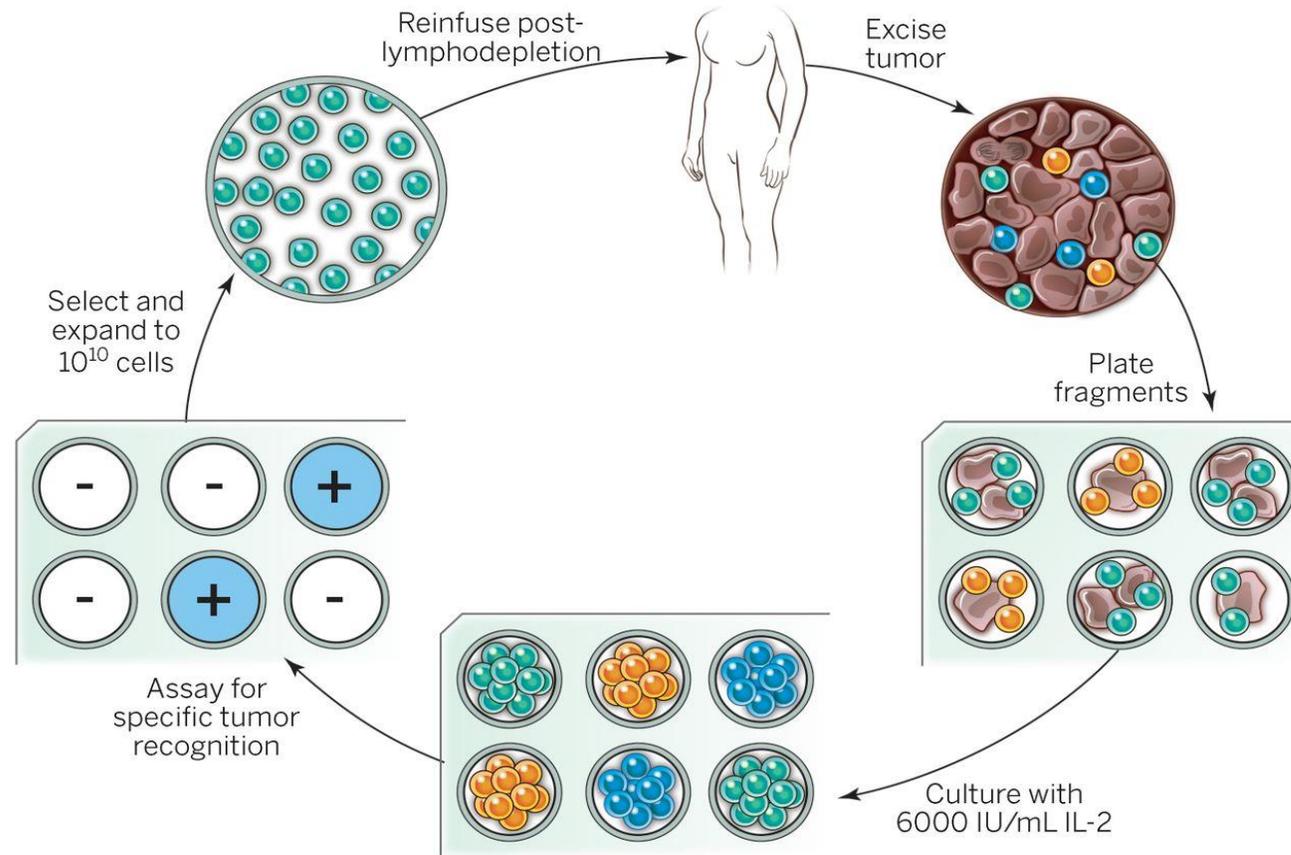
(JCO 2016 34:2389)



**Resolution of liver disease in a patient treated with
TIL and IL-2 after preparative lymphodepleting regimens of Cy/Flu**

What is the evidence that ex vivo manipulated tumor specific lymphocytes can be infused and then traffic to the brain to mediate cancer regression ?

Melanoma TIL production takes weeks



Treatment of melanoma brain metastasis with Tumor infiltrating lymphocytes

Standard eligibility criteria for *ACT Plus*

Synchronous evaluable non CNS disease

≤ 3 *brain lesions*

≤ 10 *mm*

(Excluded protocol exemptions)

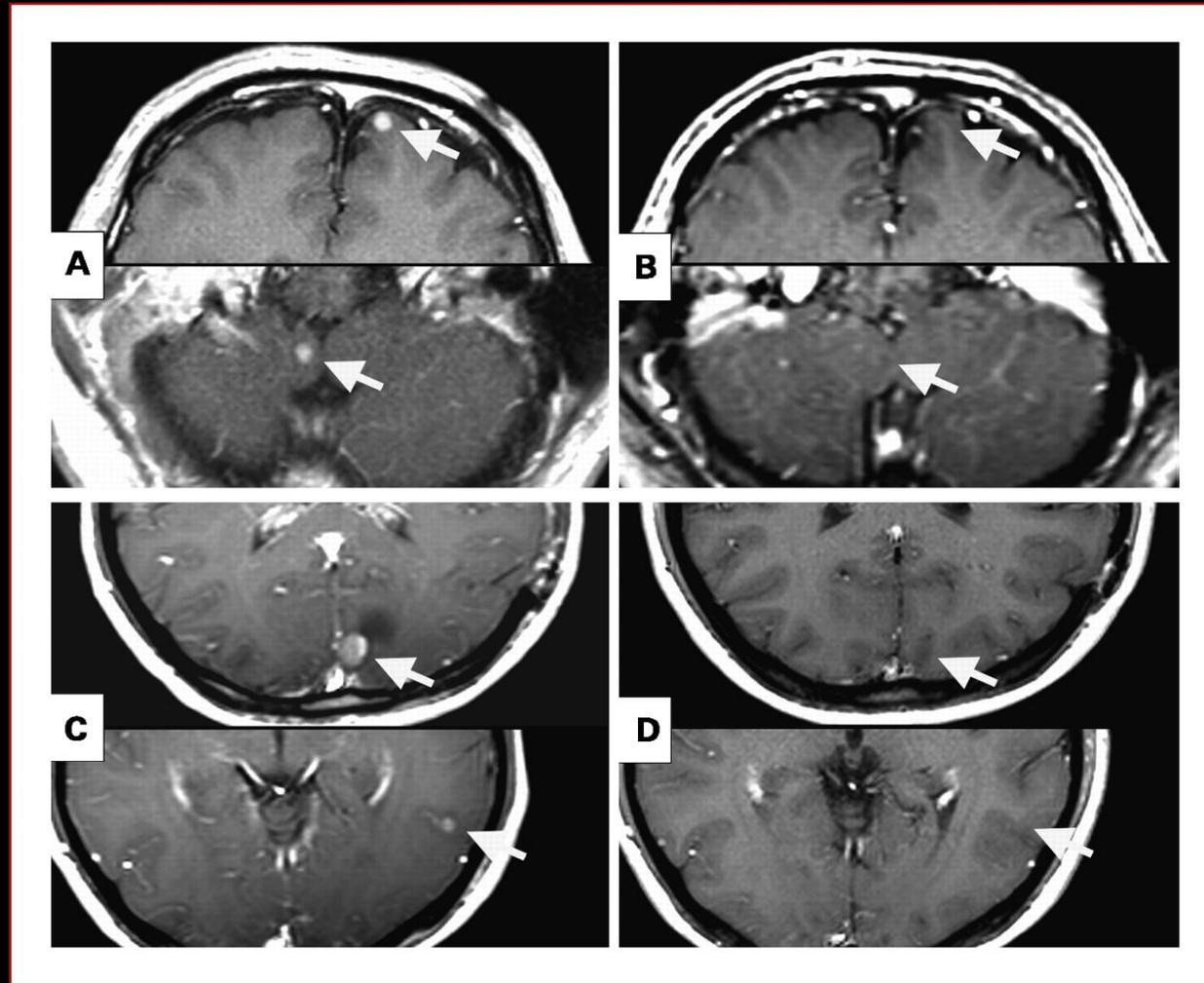
371 patients with advanced melanoma treated with TIL 2001-15

45 pts identified with **untreated** melanoma brain metastasis

Treatment of melanoma brain metastasis with Tumor infiltrating lymphocytes

45 pts with untreated melanoma brain metastasis
and synchronous evaluable non CNS disease

	<i>Overall Systemic Response</i>	<i>In Brain Response</i>
<i>ORR</i>	15 (33%)	11 (24%)
<i>PR</i>	14	1
<i>CR</i>	1	10 (22%)



Resolution of multiple brain lesions in two patients treated with TIL and IL-2 after preparative lymphodepleting regimens of Cy/Flu

Cancer regression and neurologic toxicity following anti-MAGE-A3 TCR gene therapy

Richard A. Morgan^{1,*}, Nachimuthu Chinnasamy¹, Daniel D Abate-Daga, Alena Gros¹, Paul F. Robbins¹, Zhili Zheng¹, Steven A. Feldman¹, James C. Yang¹, Richard M. Sherry¹, Giao Q. Phan¹, Marybeth S. Hughes¹, Udai S. Kammula¹, Akemi D. Miller¹, Crystal J. Hessman¹, Ashley A. Stewart¹, Nicholas P. Restifo¹, Martha M. Quezado², Meghna Alimchandani², Avi Z. Rosenberg², Avindra Nath³, Tongguang Wang³, Bibiana Bielekova³, Simone C. Wuest³, Akula Nirmala⁴, Francis J. McMahon⁴, Susanne Wilde⁵, Barbara Mosetter⁵, Dolores J. Schendel^{5,6}, Carolyn M. Laurencot¹, and Steven A Rosenberg¹

J Immunother 36(2):133, 2013

Cancer regression and neurologic toxicity following anti-MAGE-A3 TCR gene therapy

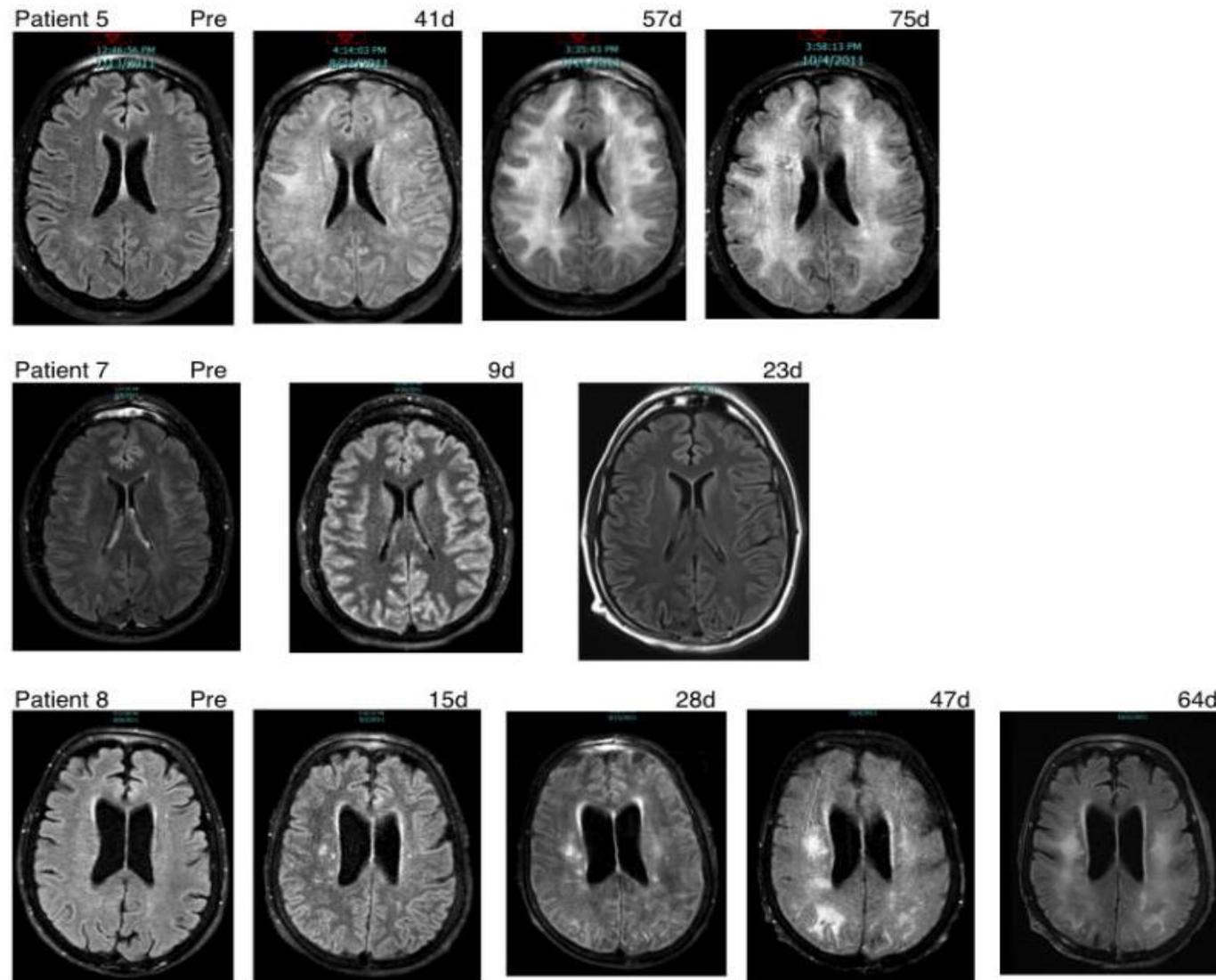
Anti-Mage-A3 TCR

Recognized Epitope KVAELVHFL (Mage A3 protein)

AND

Recognized Epitope KMAELVHFL (Mage A12 protein)

Figure 5



Neurological imaging studies. Shown are MRI scans for patients 5, 7 and 8, with the timing of the images as listed.

BRIEF REPORT | SEPTEMBER 12, 2019

Tisagenlecleucel CAR T-cell therapy in secondary CNS lymphoma

Clinical Trials & Observations

Brief Report

Matthew J. Frigault , Jorg Dietrich , Maria Martinez-Lage , Mark Leick , Bryan D. Choi , Zachariah DeFilipp , Yi-Bin Chen , Jeremy Abramson , Jennifer Crombie , Philippe Armand , Lakshmi Nayak , Chris Panzini , Lauren S. Riley , Kathleen Gallagher , Marcela V. Maus

Blood (2019) 134 (11): 860-866.



Administering T Cells Expressing Anti-EGFRvIII Chimeric Antigen Receptor to Patients with Glioblastoma Expressing EGFRvIII

Phase I Study (NCT01454596)

NCI IRB approved

All patients signed informed consent

Opened 2011

Primary end points:

Maximum safe dose

Determine 6 month PFS

Secondary end points:

CAR persistence

Radiologic response

(Goff et al. J Immunother; 42:126, 2019)

Administering T Cells Expressing Anti-EGFRvIII Chimeric Antigen Receptor to Patients with Glioblastoma Expressing EGFRvIII

Epidermal growth factor receptor (EGFR)

The most frequent genetic alteration associated with GBM

The most frequent variant is EGFRvIII (25-64%)

Driver mutation

Extracellular ligand binding domain is truncated and is constitutively active

EGFRvIII is not present in normal tissue and is tumor specific

Administering T Cells Expressing Anti-EGFRvIII Chimeric Antigen Receptor to Patients with Glioblastoma Expressing EGFRvIII

Protocol

Nonmyeloablative preparative chemotherapy

Cell infusion (1 E7 total cell starting dose)

Low dose IV IL-2 (72,000 IU/kg) q 8 to tolerance

MRI follow up at monthly intervals

Administering T Cells Expressing Anti-EGFRvIII Chimeric Antigen Receptor to Patients with Glioblastoma Expressing EGFRvIII

Eligibility

Recurrent GBM following surgery , XRT , and chemo

18-70 years of age

Karnofsky performance status \geq 60%

Steroids allowed if on a stable dose

EGFRvIII + Determination based on a two step PCR on RNA from in-house or submitted samples (NCI Path Department)

Administering T Cells Expressing Anti-EGFRvIII Chimeric Antigen Receptor to Patients with Glioblastoma Expressing EGFRvIII

Anti-EGFRvIII CAR T Cell Production and Analysis (139-28BBZ)

Single chain human scFV (139)

3rd Generation (CD28 costimulation, CD3Z, 4-1BB costimulation)



Diagram of retroviral construct detailing location of single chain variable fragment of Hu monoclonal Ab 139 CD8 linker domain, CD28 and 4-1BB costim domains and CD3zeta signaling domain

Morgan RA et al.

Hum Gene Ther. 2012,23(10):1043

Administering T Cells Expressing Anti-EGFRvIII Chimeric Antigen Receptor to Patients with Glioblastoma Expressing EGFRvIII

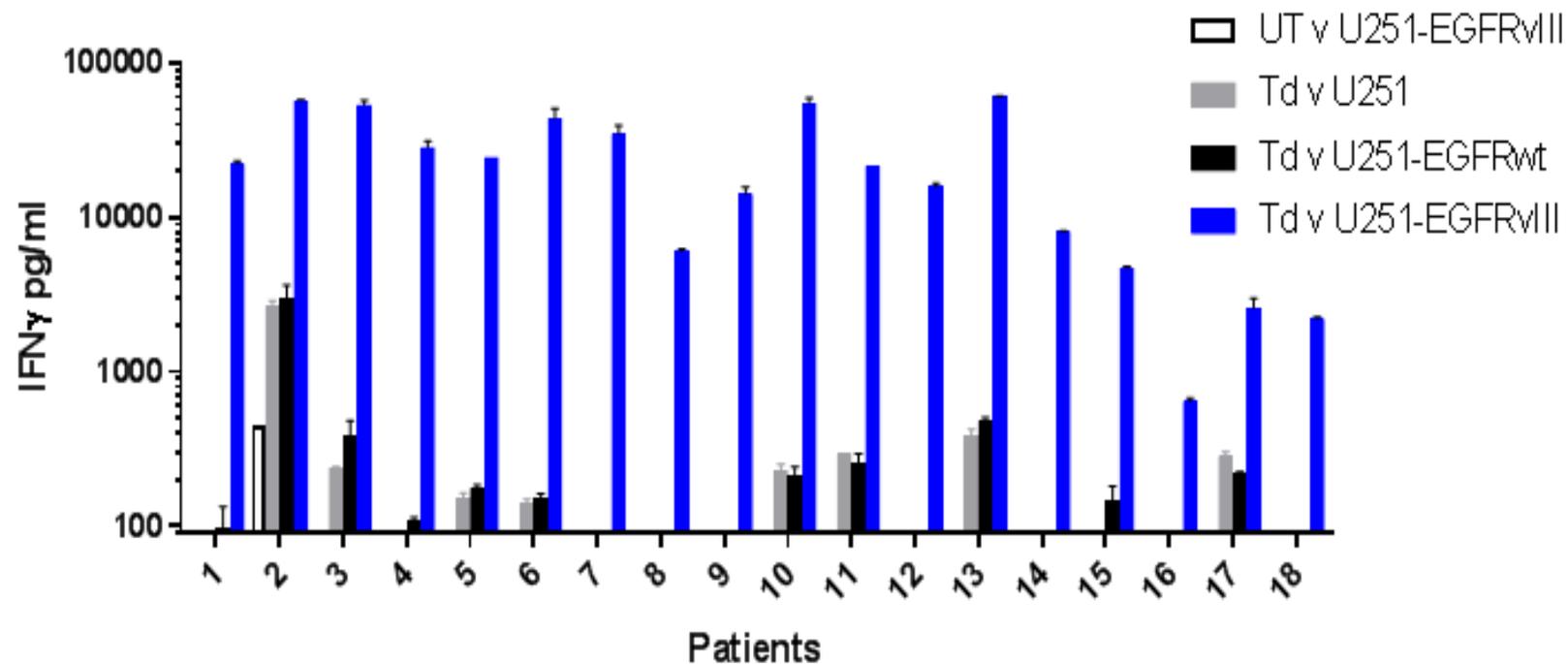
Anti-EGFRvIII CAR T Cell Production and Analysis (139-28BBZ)

OKT3 stimulated PBL transduced with a gamma-retroviral vector
encoding the EGFRvIII CAR

COA included $\geq 10\%$ CAR+ CD3 Cells
Specific INF-g release to EGFRvIII+ cell lines

($\geq 3 \times 10^6$ cells required additional rapid expansion)

B.



EGFRvIII-specific cytokine release of infused cell product as measured by interferon- γ ELISA. UT: untransduced PBL, Td: PBL transduced with CAR-28BBZ, U251: glioblastoma cell line \pm transduction to express EGFRwt or EGFRvIII

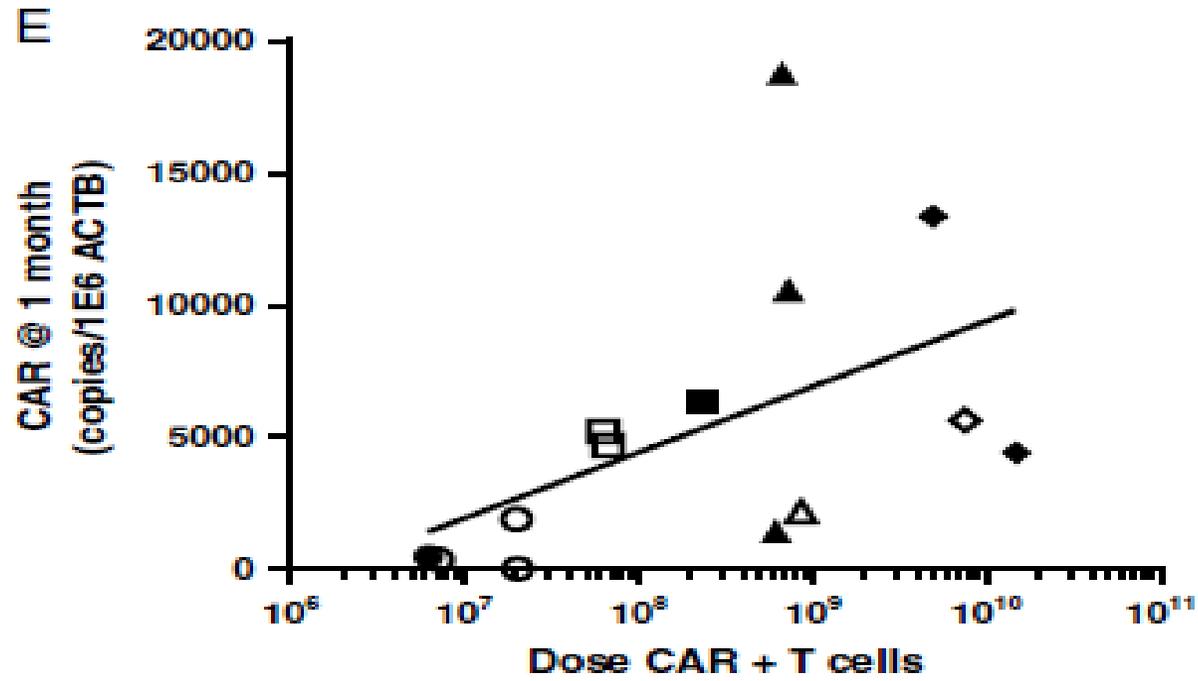
ID	AGE	SEX	PRIOR TREATMENTS
1	45*	M	Surgery, XRT, TMZ, bevacizumab, BCNU
2	43	M	Surgery, XRT, TMZ, bevacizumab
3	52*	M	Surgery, XRT, TMZ, bevacizumab
4	46	M	Surgery, XRT, TMZ
5	55	M	Surgery, XRT, TMZ
6	57*	M	XRT, TMZ, bevacizumab
7	53	M	Surgery, XRT, TMZ
8	56	M	Surgery, XRT, TMZ, AZD7451
9	55	F	Surgery, XRT, TMZ, bevacizumab
10	55	M	Surgery, XRT, TMZ
11*	61*	F	Surgery, XRT, TMZ
12*	66*	M	Surgery, XRT, TMZ, veliparib, bevacizumab
13	60*	M	Surgery, XRYT, TMZ, bevacizumab, EGFRvIII vaccine
14*	64	F	Surgery, XRT, TMZ, IMA950 vaccine
15	61*	M	Surgery, XRT, TMZ, EGFRvIII vaccine vs. placebo trial, bevacizumab, trebananib
16	43*	M	Surgery, XRT, TMZ carotuximab, bevacizumab
17	47*	M	Surgery, XRT, TMZ, EGFRvIII vaccine vs placebo trial, bevacizumab
18*	57	M	Surgery, XRT, TMZ

			ADMINISTERED CELLS						SURVIVAL (MONTHS)		
ID	AGE		DOSE	% CAR (+)	# CAR (+)	# IL-2 DOSES		RESP	PFS	OS	
1	45*	M	1.00E+07	71.0%	7.10E+06	7		NR	1.1	2.2	
2	43	M	1.00E+07	62.6%	6.26E+06	10		NR	1.9	13.1	
3	52*	M	3.00E+07	67.6%	2.03E+07	8		NR	1.1	4.5	
4	46	M	3.00E+07	66.5%	2.00E+07	4		NR	2.0	11.1	
5	55	M	1.00E+08	67.5%	6.75E+07	7		NR	1.5	9.0	
6	57*	M	1.00E+08	62.9%	6.29E+07	6		NR	0.0	6.9	
7	53	M	3.00E+08	76.5%	2.30E+08	8		NR	1.2	9.7	
8	56	M	1.00E+09	62.0%	6.20E+08	6		NR	0.9	10.1	
9	55	F	1.00E+09	67.6%	6.76E+08	6		NR	1.1	2.0	
10	55	M	1.00E+09	73.3%	7.33E+08	5		NR	1.3	4.4	
11*	61*	F	2.48E+09	35.0%	8.68E+08	3		NR	12.5	46.8+	
12*	66*	M	3.00E+09	67.9%	2.04E+09	5		NR	0.9	2.1	
13	60*	M	3.00E+09	79.3%	2.38E+09	1		NR	2.7	4.5	
14*	64	F	1.00E+10	49.4%	4.94E+09	5		NR	1.6	8.9	
15	61*	M	1.00E+10	66.0%	6.60E+09	0		NR	1.1	1.4	
16	43*	M	1.00E+10	75.3%	7.53E+09	1		NR	1.1	6.9	
17	47*	M	6.00E+10	43.2%	2.59E+10	0		NE	TRM	TRM	
18*	57	M	3.00E+10	49.9%	1.50E+10	0		NR	2.0	13.6	

Grade 3 and 4 Serious AEs (excludes CY/Flu Aes)

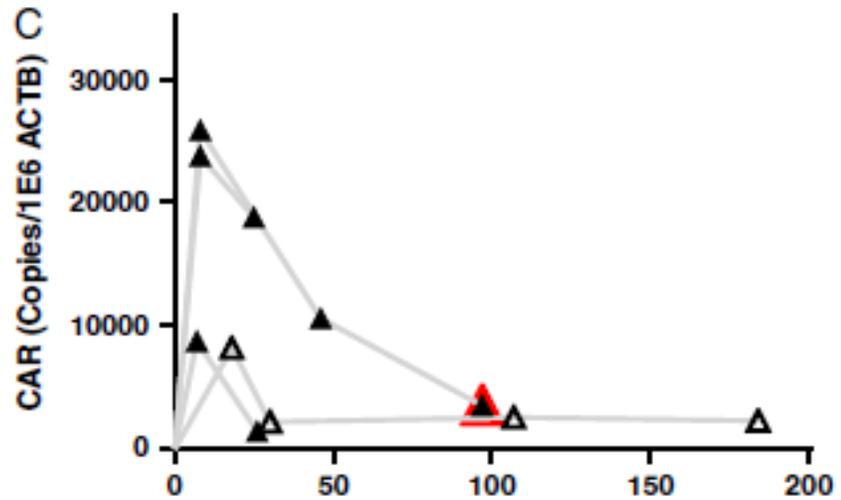
Cardiopulmonary	<i>Dyspnea/Hypoxia</i>	2
	Hypotension(non septic)	2
	Capillary leak	1
Infectious	Febrile Neutropenia	2
	Bacteremia	8
Neurologic	transient motor weakness	1
	transient urinary incontinence	1
Coagulation	Prolonged PTT	1
	DVT	1
	PE	1

Persistence of infused CAR+ cells as measured by qPCR

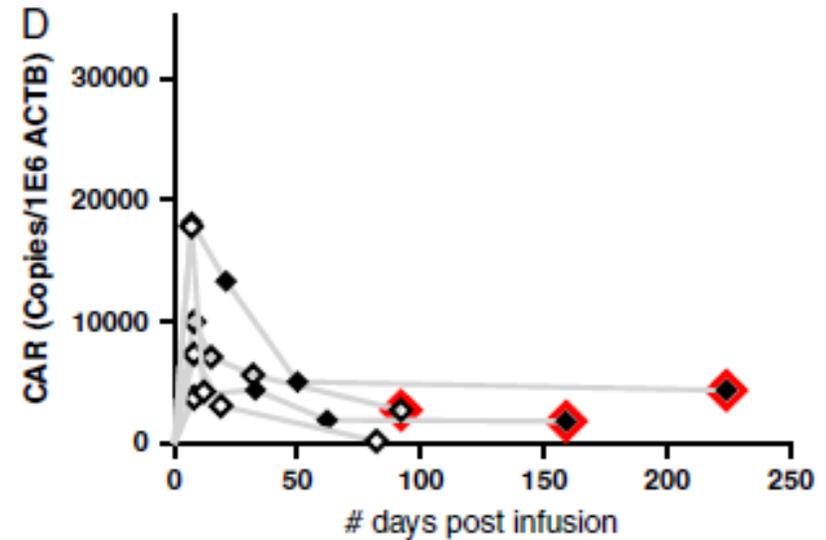


Persistence at one month (median day 32, n=14) was correlated with CAR+ cell dose ($r=0.6615$, $p=0.0121$), but not survival (not shown).

Persistence of infused CAR+ cells as measured by qPCR



Patients who received $< 3 \times 10^7$ CAR+ cells



Patients who received $> 3 \times 10^8$ CAR+ cells

The Safety and Feasibility of Administering T Cells Expressing Anti-EGFRvIII Chimeric Antigen Receptor to Patients with Malignant Gliomas Expressing EGFRvIII

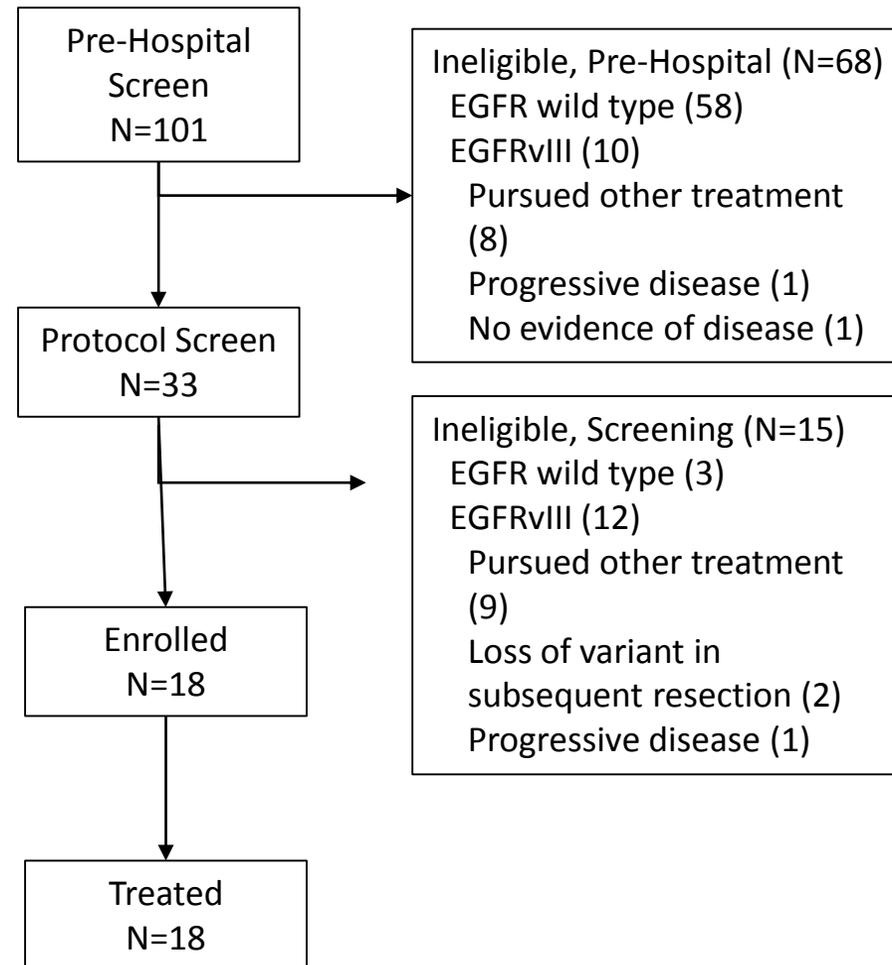
Conclusions *Safe cell dose (1 -3 E 10 total cells)*
Patients with GBM can tolerate CAR T-Cells/NMA/LD IL-2
Persistence of CAR T cells appeared to be dose related
Low level CAR T cells could be detected months following infusion
CAR Persistence did not correlate with survival

Administration of anti-EGFRvIII Car T cells did **not** mediate a clinically meaningful effect in patients with GBM in this Phase 1 Study

(Goff et al. J Immunother; 42:126, 2019)



NCI Surgery Branch



CONSORT Diagram of Trial Enrollment.