

Induction of CD8+ T Cell Responses Against Novel Glioma-Associated Antigen Peptides and Clinical Activity by Vaccinations with α -Type-1-Polarized Dendritic Cells and Poly-ICLC in Patients with Recurrent Malignant Glioma



Hideho Okada
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iSBTC 25th Annual Scientific Meeting Oct 3, 2010



Conflicts of Interests (COI)

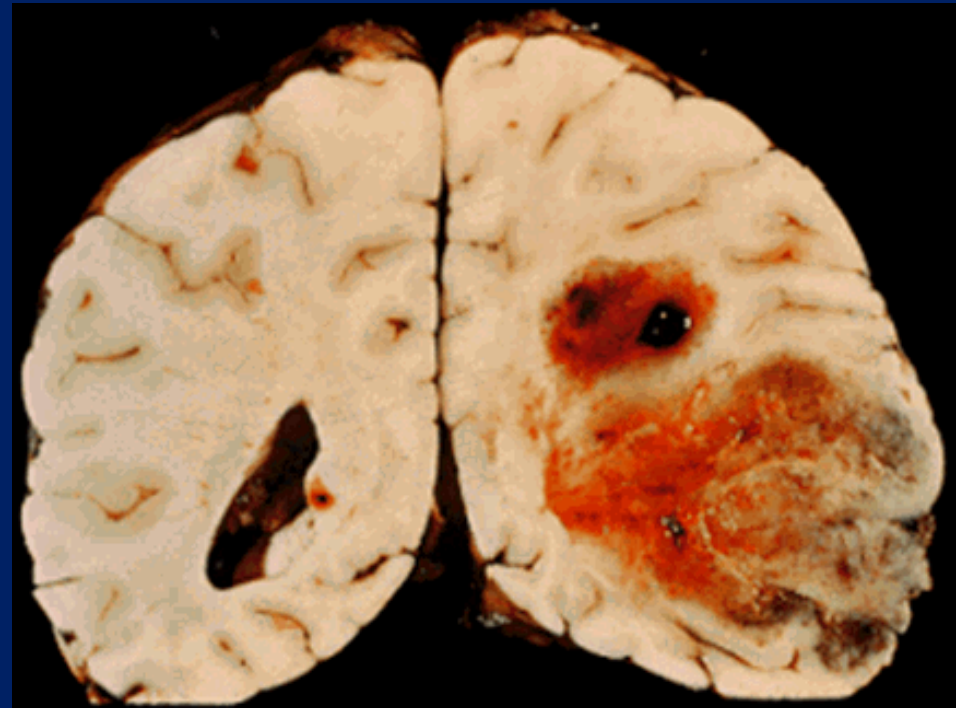
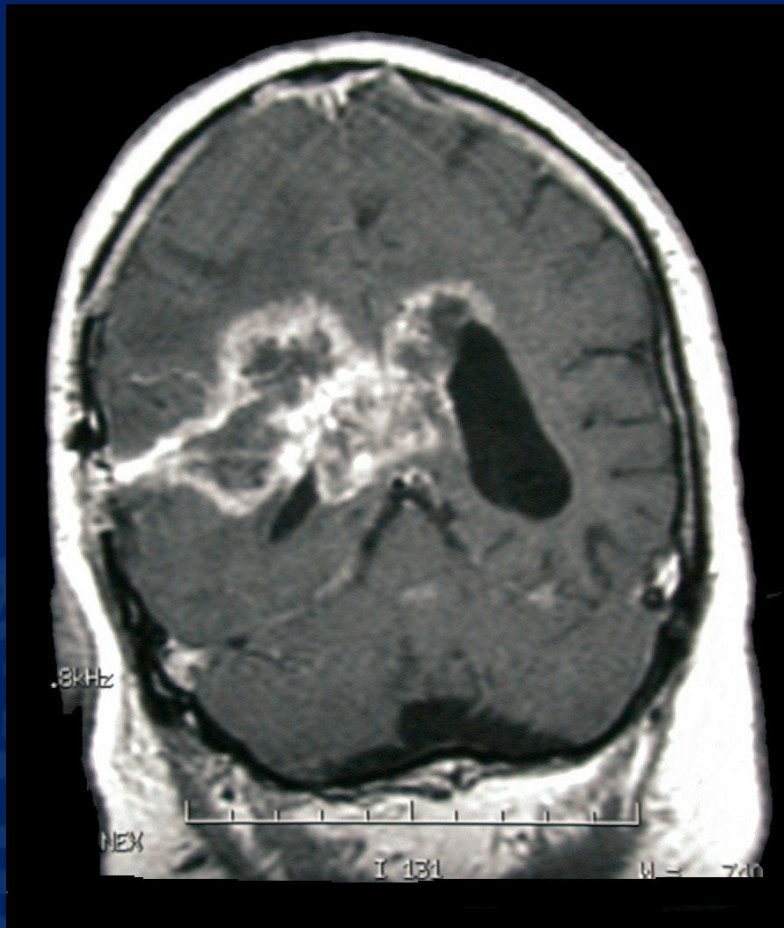
Hideho Okada is one of the inventors of the IL-13R α 2 (345-353:1A9V) and EphA2 (883-891) peptides, for which an exclusive licensing agreement has been executed with Stemline, Inc.

Per the University of Pittsburgh COI policy, interpretation of presented data was not performed solely by Hideho Okada, but by the investigator team.



Malignant Gliomas

- WHO grade 3 anaplastic glioma
- WHO grade 4 glioblastoma multiforme (GBM)



Critical Aspects/Factors for Potent Dendritic Cell Vaccines (05-115)

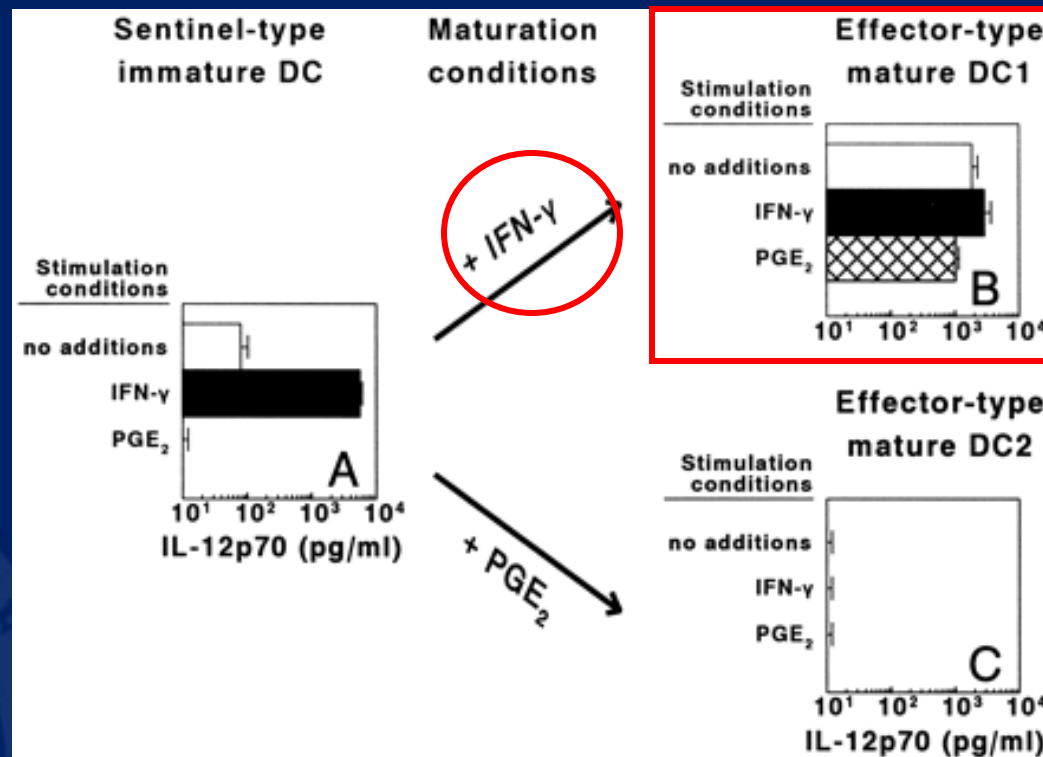
- Type of DCs
 - Type-1 DCs (alphaDC1)
- Target Antigens
 - Multiple CTL epitopes from 4 GAAs
- Administration Route
 - Intranodal administration (superior to s.c)
- Adjuvant
 - Poly-ICLC as a ligand for intracellular dsRNA receptors



Background

Glioma Vaccines with Type 1 DCs

Cytokines modulate the IL-12 production in DC and promote the development of stable, polarized DC (DC1).



Type-1
"Polarizing"
cocktails

IFN- γ
IFN- α
TNF- α
IL-1 β
Poly-IC

Kalinski P *et al.* The J. Immunol. 2000, Cancer Res 2004

Kalinski P and Okada H. Seminars in Immunology 2010



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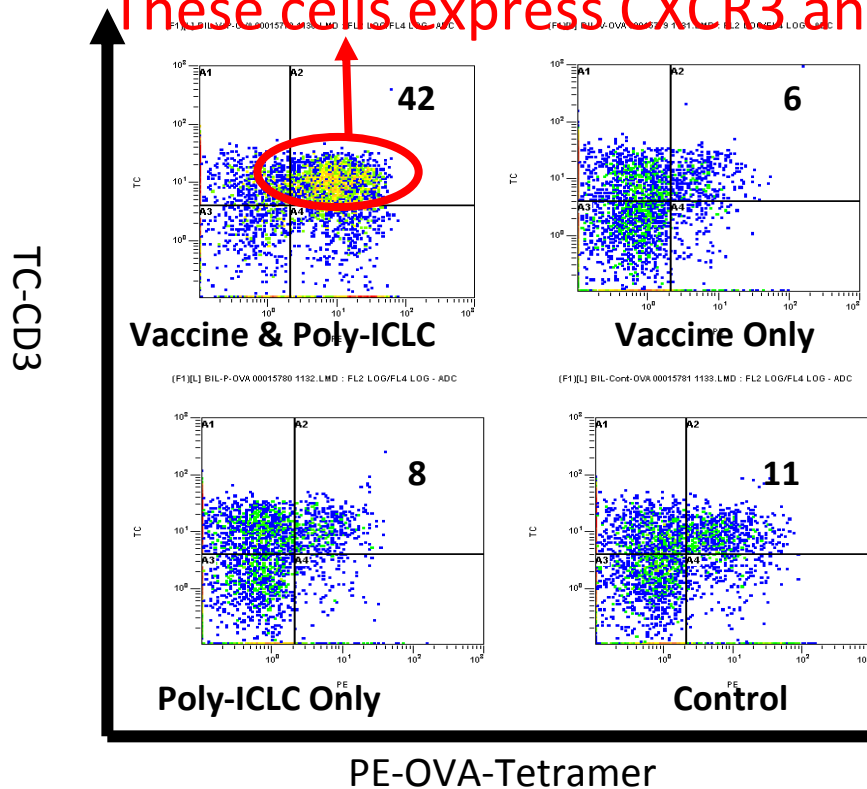


Poly-ICLC administration enhances the infiltration of OVA-Specific T cells and therapeutic efficacy

Brain-Infiltrating T cells

Anti-Tumor Effect

These cells express CXCR3 and VLA-4



Mock Tx Alone

Poly-ICLC Alone

Vaccine Alone

Vaccine plus Poly-ICLC

Zhu X. *et al.*

Objectives (UPCI 05-115)

- Primary Objective
 - Safety - to determine the maximal tolerated dose of DC1 and evaluation of toxicities
- Secondary Objectives
 - Assess immunological response against GAAs using ELISPOT and tetramer assays
 - Assess the preliminary anti-tumor clinical activity of the vaccines as measured by radiological response (MRI), overall survival as well as 6 month-progression free survival (PFS).



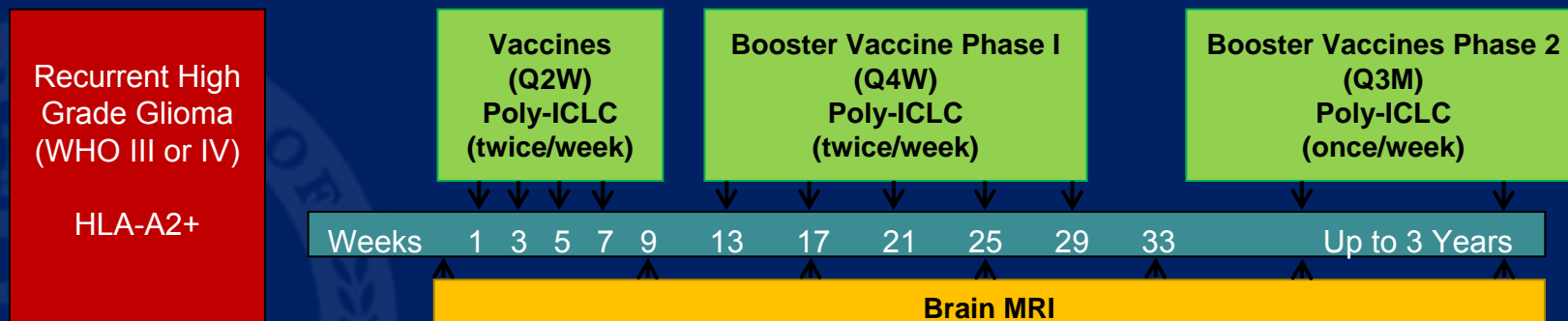
Eligibility

- Adult patients with recurrent GBM or WHO grade 3 AG
- HLA-A2+ based on flow-cytometry
- Minimum corticosteroid (4 mg/day or less for Dexamethasone)
- Maximum 2 previous recurrences



Treatment

- Ultrasound-guided intranodal injections of **type-1 DC1** (1 or 3 x 10⁷ /injection with dose escalation) loaded with **4 glioma-associated antigen (GAA)-derived HLA-A2-restricted CTL epitopes** (IL-13Ra2_{345-353:1A9V}, gp100_{209-217:2M}, EphA2₈₈₃₋₈₉₁ and YKL-40₂₀₁₋₂₁₀)
- Intramuscular injections of **poly-ICLC** (20 mcg/kg; Twice/week)



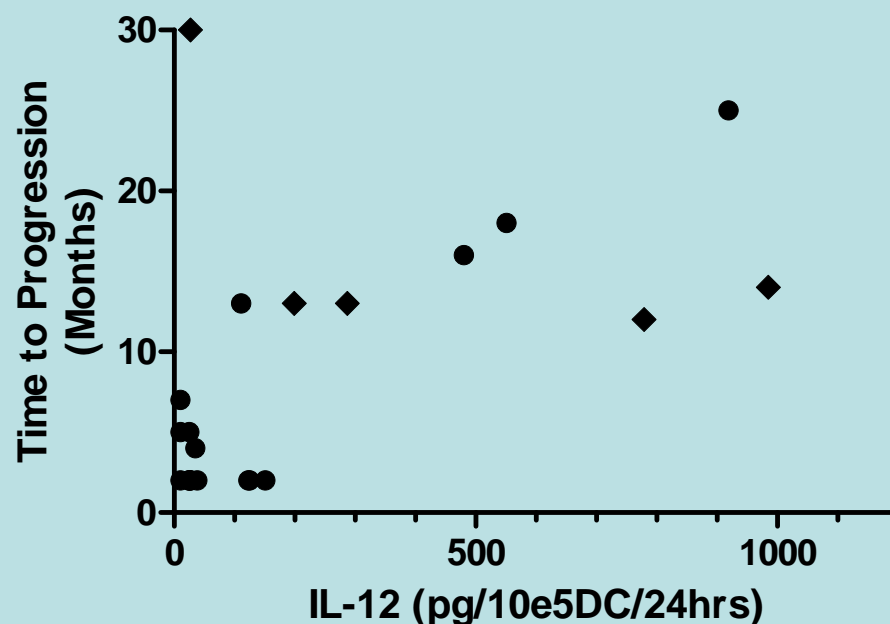
SL-701 Adverse Events

No grade 3 or 4 toxicities observed

Adverse Event	Grade 1		Grade 2	
	No.	%	No.	%
Blood/Bone Marrow				
Leukocytopenia			1	5
Injection site reactions				
Redness, induration, pruritis, pain	17	77	1	5
Constitutional symptoms				
Fatigue (lethargy, malaise, asthenia)	16	73		
Fever	5	23		
Chills/Rigors	4	18		
Nausea	7	32		
Vomiting	1	5		
Headache	5	23	2	9
Insomnia	1	5		
Light headed/dizziness	2	9		
Myalgia	7	32		
Body ache	6	27		
Dermatological				
Skin rash	3	14		
Dry skin	1	5		
Bruising	2	9		
Urticaria	1	5		
Pulmonary/Upper Respiratory				
Rhinitis/Runny nose	1	5		



IL-12 production levels positively correlated with PFS ($p=0.0255$ based on Cox regression test)

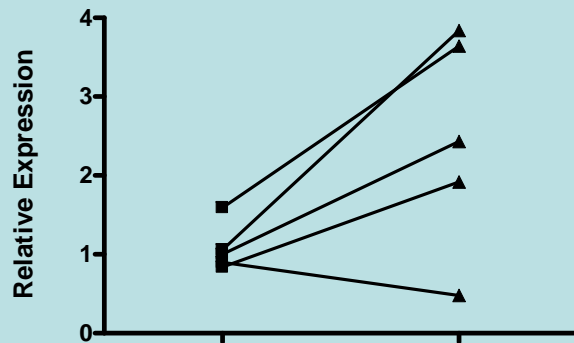


Closed circles indicate patients who have already progressed, whereas closed diamonds represent patients who have not recurred to date.

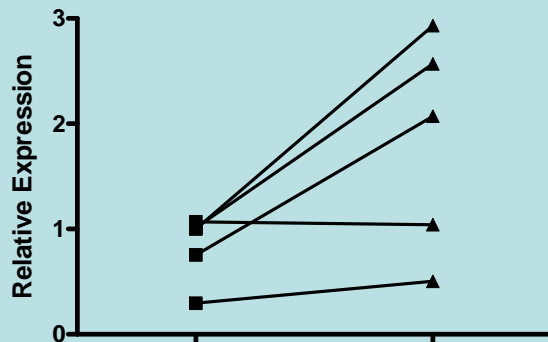


Promotion of type-1 responses detected in post-vaccine PBMCs by RT-PCR

IFN α 1 (p=0.0306)

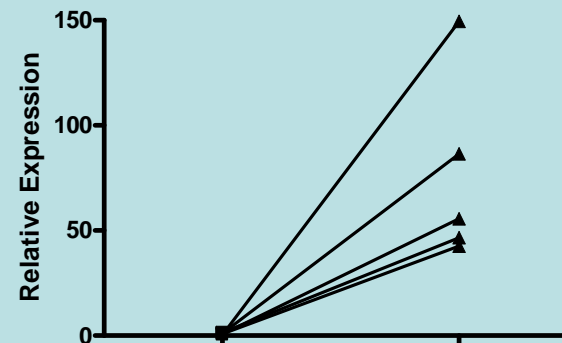


Pre- vs. Post-first vac/poly-ICLC
TLR3 (p=0.0303)

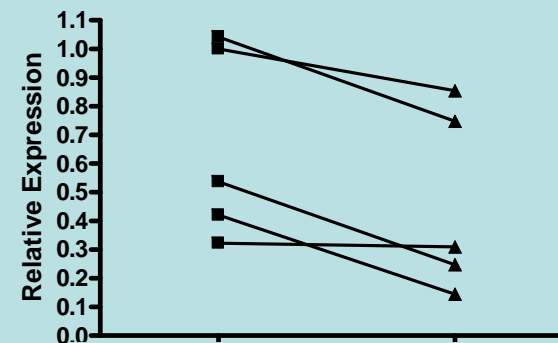


Pre- vs. Post-first vac/poly-ICLC

CXCL10 (p=0.0098)



Pre- vs. Post-first vac/poly-ICLC
CCL22 (p=0.0105)

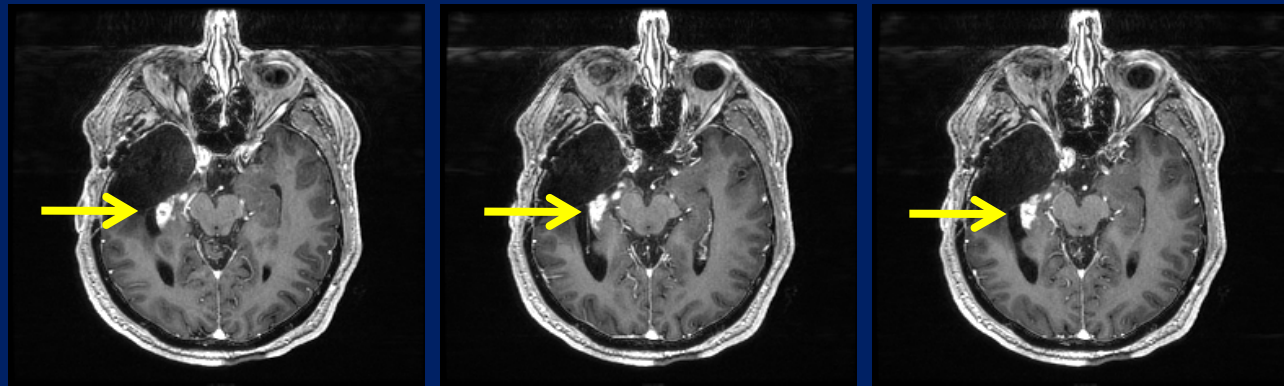


Pre- vs. Post-first vac/poly-ICLC

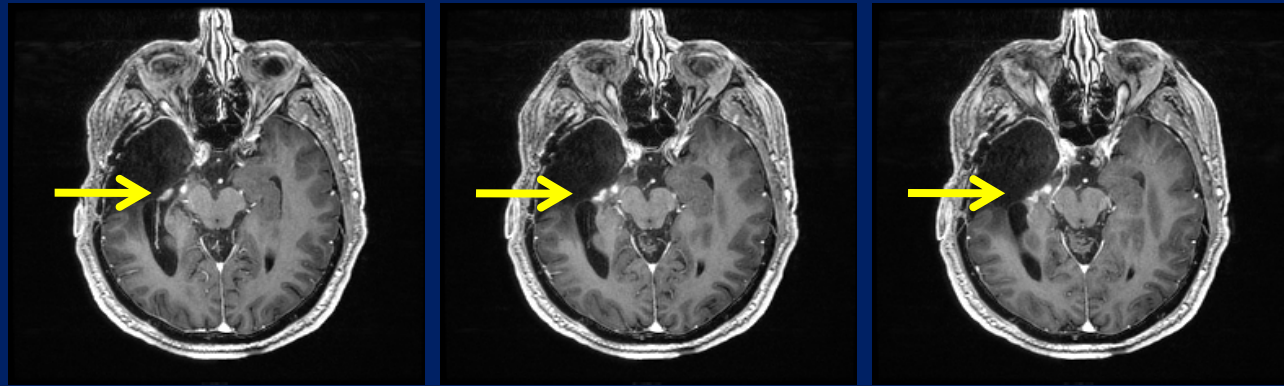


Complete Radiological Response in A Patient with Recurrent GBM (Pt #20)

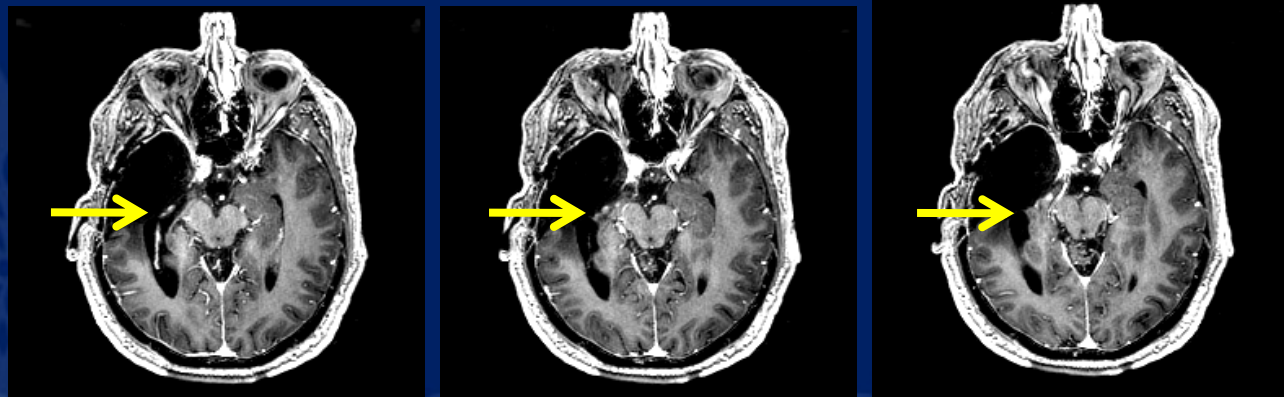
Pre-Vaccine
Week 0



Post-Vaccine
Week 9

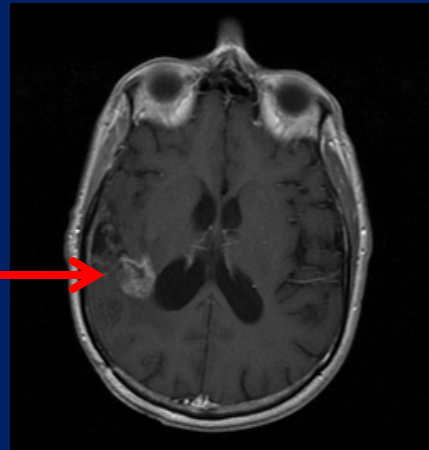


Post-Vaccine
Week 17

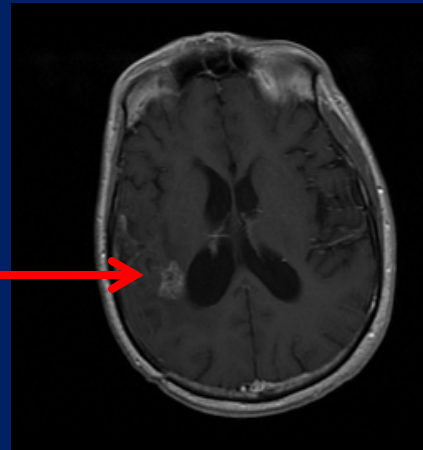


PR Patient with subsequent pseudo-tumor progression

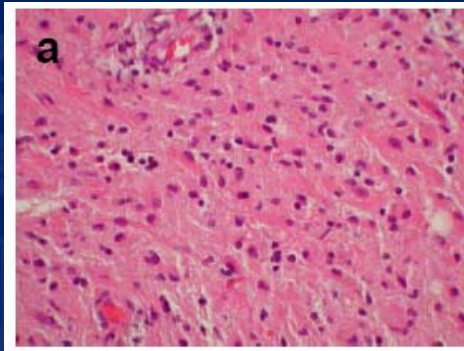
Pre-Vaccine



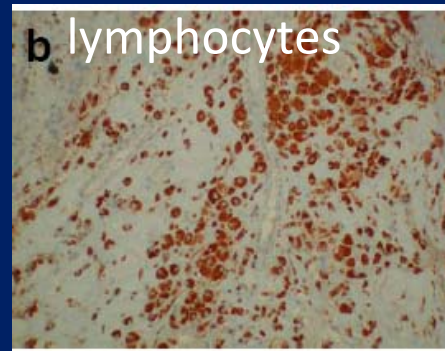
Post-Vaccine (wk 9)



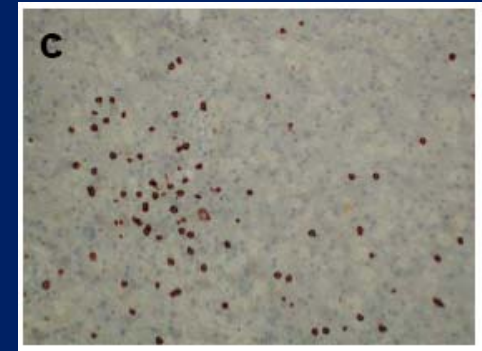
Biopsy demonstrates intratumoral infiltration of macrophages and CD8⁺ T



Reactive gliosis,
possible residual glioma



Numerous
CD68⁺ macrophages



CD8⁺ T cells



Summary 05-115; Phase I/II Vaccine Study in Adult Recurrent Malignant Glioma (JCO In Press)

- The regimen was well tolerated in 22 patients.
- Immune responses against at least one of the vaccination-targeted GAAs were detected in post-vaccine PBMC in 11 of 19 patients.
- Analyses of PBMC demonstrated significant up-regulation of type-1 cytokines and chemokines, including IFN- α and CXCL10.
- Nine (4 GBM, 2 AA, 2 AO and 1 AOA) achieved progression free status lasting at least 12 months. One patient with recurrent GBM demonstrated sustained complete response.
- IL-12 production levels by aDC1 positively correlated with progression-free survival.



Primary vs. Secondary GBM

p53 (17p13) mutation (>65%)

PDGF, FGF2
overexpression (~60%)

Low grade astrocytoma

CDK4 (12q13) amplification
RB (13q13) alteration (~25%)
LOH 19q (~50%)

Anaplastic astrocytoma

PTEN (10q23) loss (~4%)
PDGFR-α amplification (<10%)

Secondary glioblastoma

mean = 45 yrs

EGFR (7p12)
amplification (~40%)
overexpression (~60%)

MDM2 (12q14)
amplification (~8%)
overexpression (~50%)

CDKN2A (9p21) loss (~50%)

PTEN (10q23) loss (~70%)

**Primary glioblastoma
*de novo***

mean = 55 yrs

Contributors

Co-Investigators

Pawel Kalinski – development of alphaDC1

Ryo Ueda, Aki Hoji and Gary Kohanbash – cytokine and tetramer assays

Frank S. Lieberman and Teresa E. Donegan – patient management

Arlan H. Mintz, Johnathan A. Engh, David L. Bartlett, Charles K. Brown, Herbert Zeh, Matthew P. Holtzman and Ian F. Pollack – surgical aspects

Todd A. Reinhart – *in situ* hybridization

Theresa L. Whiteside and Lisa H.

Butterfield – immuno-monitoring

Ronald L. Hamilton – neuro-pathology

Douglas M. Potter – biostatistics

Andres M. Salazar - provision of poly-ICLC

Brain Tumor Program
Clinical Research Services
IMCPL of the UPCI

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Musella Foundation

Pittsburgh Foundation

The Brain Tumor Society

Participants and their families



I am stopping my talk



- though our work will
never stop at any time!
THANK YOU!



A Bi-Institutional Pilot Study of Vaccinations with GAA-peptides in Adult Patients with High-Risk LGG – University of Pittsburgh and Wake Forest University

Primary Objectives – To determine safety and glioma-associated antigen (GAA)-specific immune responses of the regimen.

Rationale –

- 1) The slow growth rate of LGG should allow sufficient time to repeat multiple immunizations, and the induction of high levels of GAA-specific immunity
- 2) minimal toxicity allows for maintenance of high quality of life
- 3) SL-701 could delay the use of RT in this patient population

Eligibility - HLA-A2+ adult patients WHO grade II astrocytoma or oligoastrocytoma with “high-risk” factors - defined as at least one of the following conditions: 1) age ≥ 40 with any extent resection
2) age 18-39 with incomplete resection (post-op MRI showing $>1\text{cm}$ residual disease) or
3) the tumor size is $\geq 4\text{ cm}$



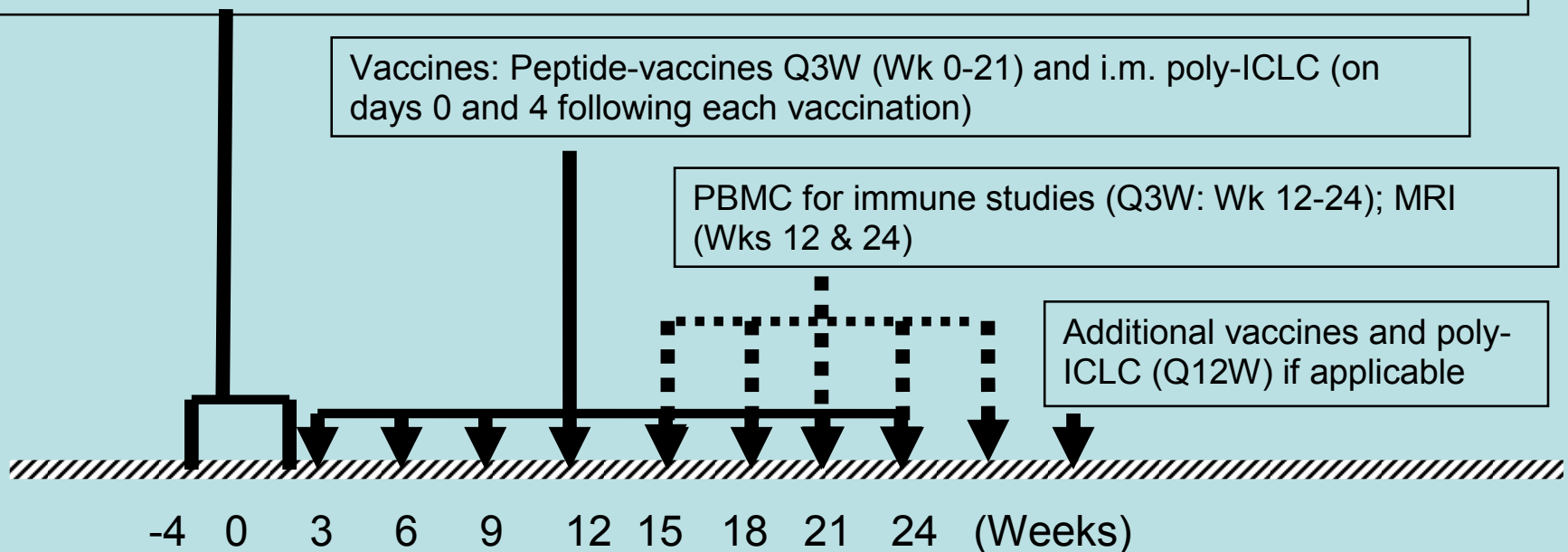
Treatment Schema

No corticosteroid will be allowed within 4 weeks prior to the first vaccine. Baseline MRI and other screening procedures will be done within 4 weeks prior to the 1st vaccination

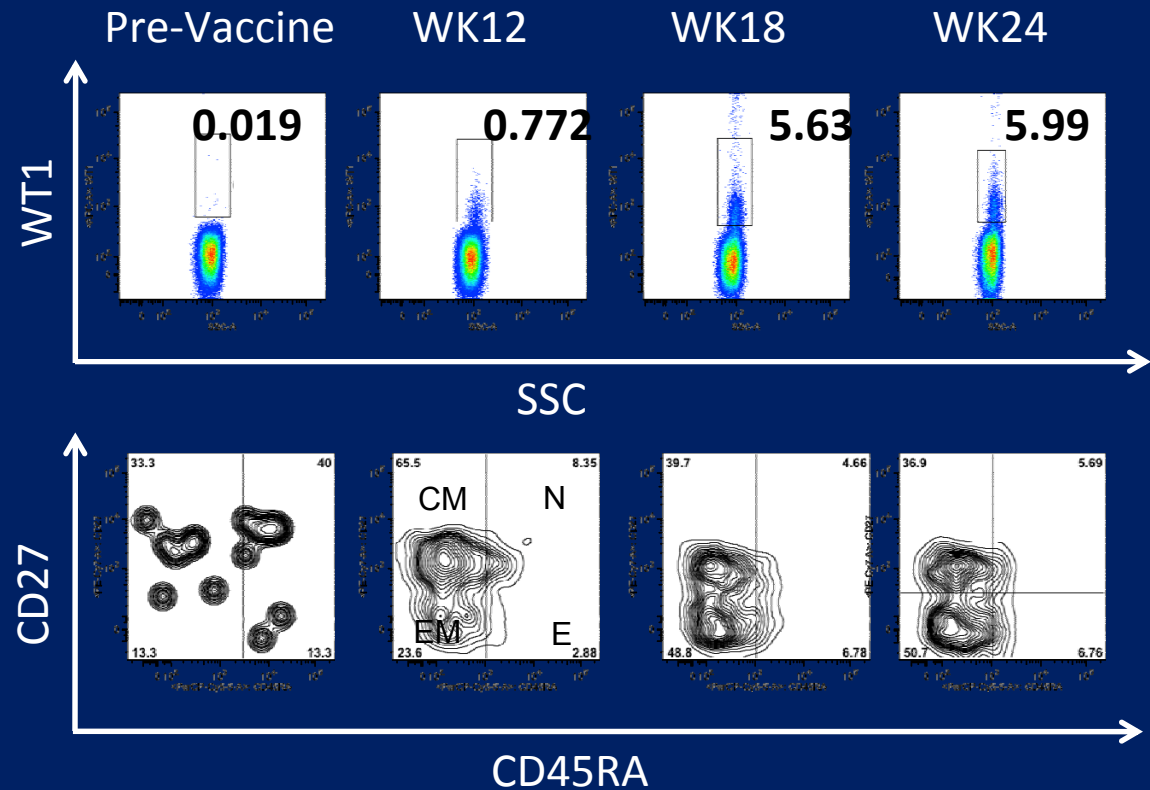
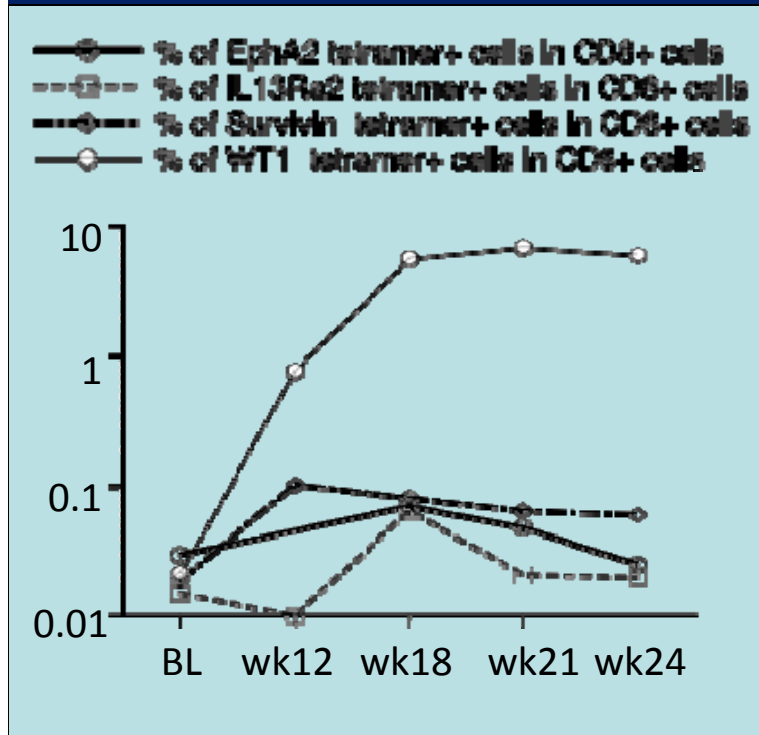
Vaccines: Peptide-vaccines Q3W (Wk 0-21) and i.m. poly-ICLC (on days 0 and 4 following each vaccination)

PBMC for immune studies (Q3W: Wk 12-24); MRI (Wks 12 & 24)

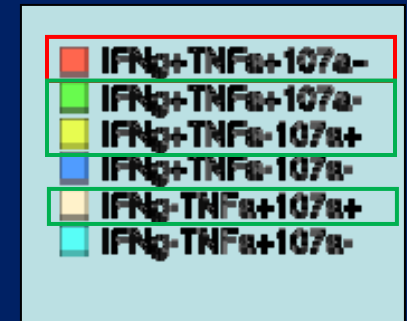
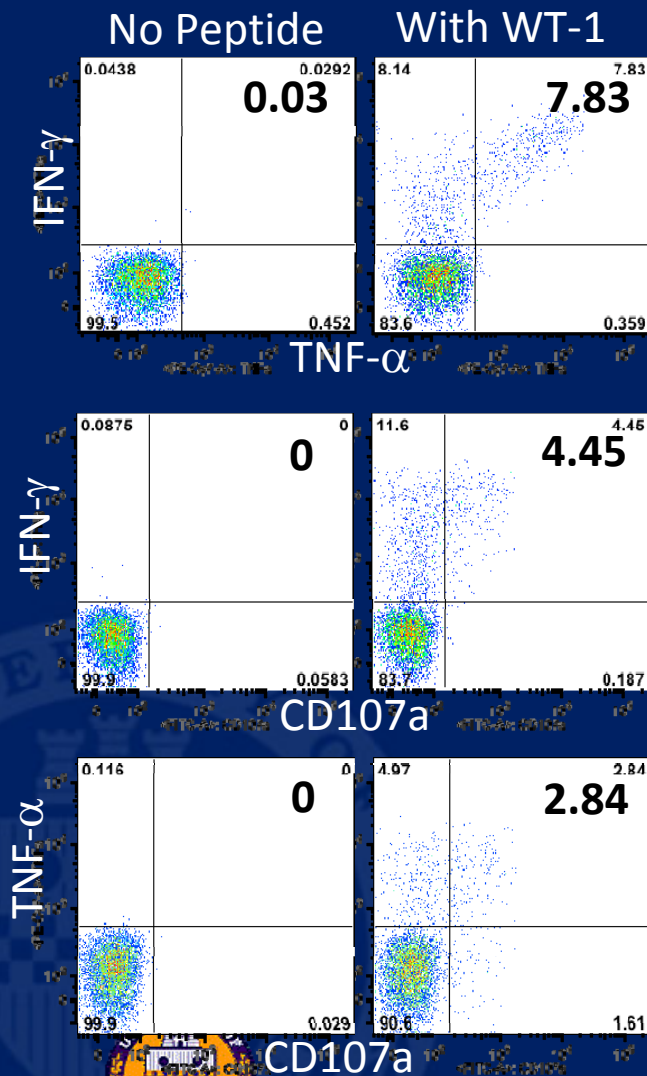
Additional vaccines and poly-ICLC (Q12W) if applicable



Robust Induction of GAA-Specific CD8+ cell Response in a Subject with WHO Grade 2 Low Grade Glioma



Type-1 effector function of CD8+ T cells in response to brief *ex vivo* stimulation with the WT-1 peptide



Acknowledgement

Co-Investigators

Frank S. Lieberman, Ryo Ueda, Aki Hoji,
Pawel Kalinski, Arlan H. Mintz, Johnathan
A. Engh, David L. Bartlett, Herbert Zeh,
Teresa E. Donegan, Theresa L. Whiteside,
Lisa H. Butterfield, Walter J. Storkus,
Douglas M. Potter, Ronald L. Hamilton,
Regina Jakacki and Ian F. Pollack

Key External Collaborators

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Michael E. Scheurer (Baylor)
John H. Ohlfest (U. Minnesota)
Andres M. Salazar (Oncovir, Inc)

Brain Tumor Program
Clinical Research Services
IMCPL
Of the UPCI

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Pittsburgh Foundation
The Brain Tumor Society

Participants and their families



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Brain Tumor Program Pittsburgh Cancer Institute

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Streets around Univ. Pittsburgh



University of Pittsburgh Medical Center University of Pittsburgh Cancer Institute



Central Nervous System (CNS) immunology

Concept of CNS as immunologically privileged

1. Blood Brain Barrier/No lymphatic system
2. No dendritic cell (DC) distribution
3. Roles of microglia/brain macrophages as antigen-presenting cells (APC) are not clear



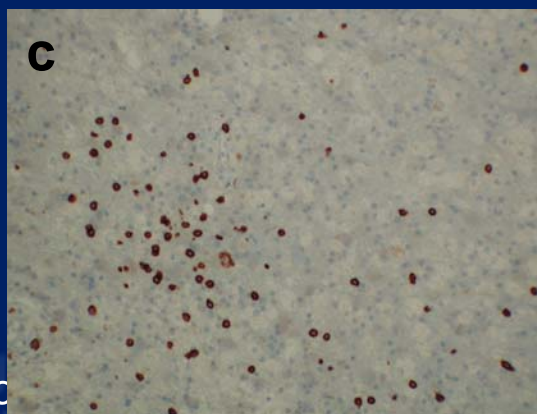
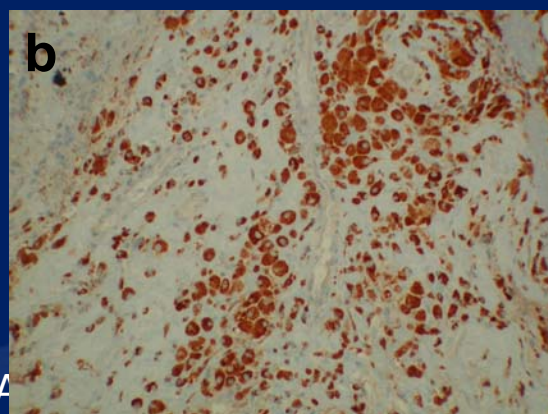
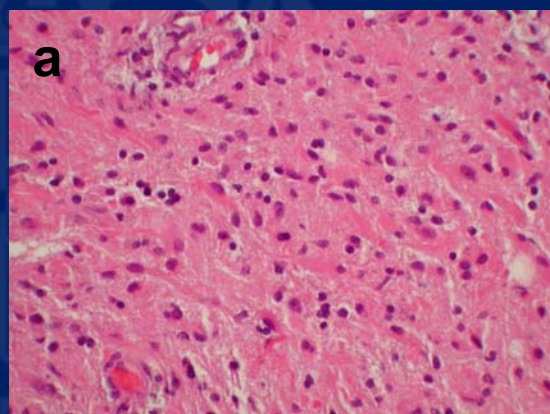
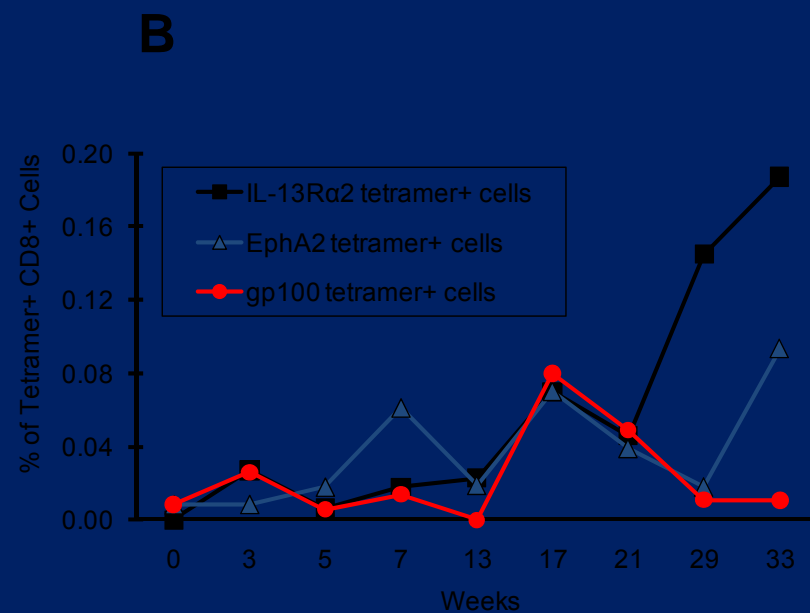
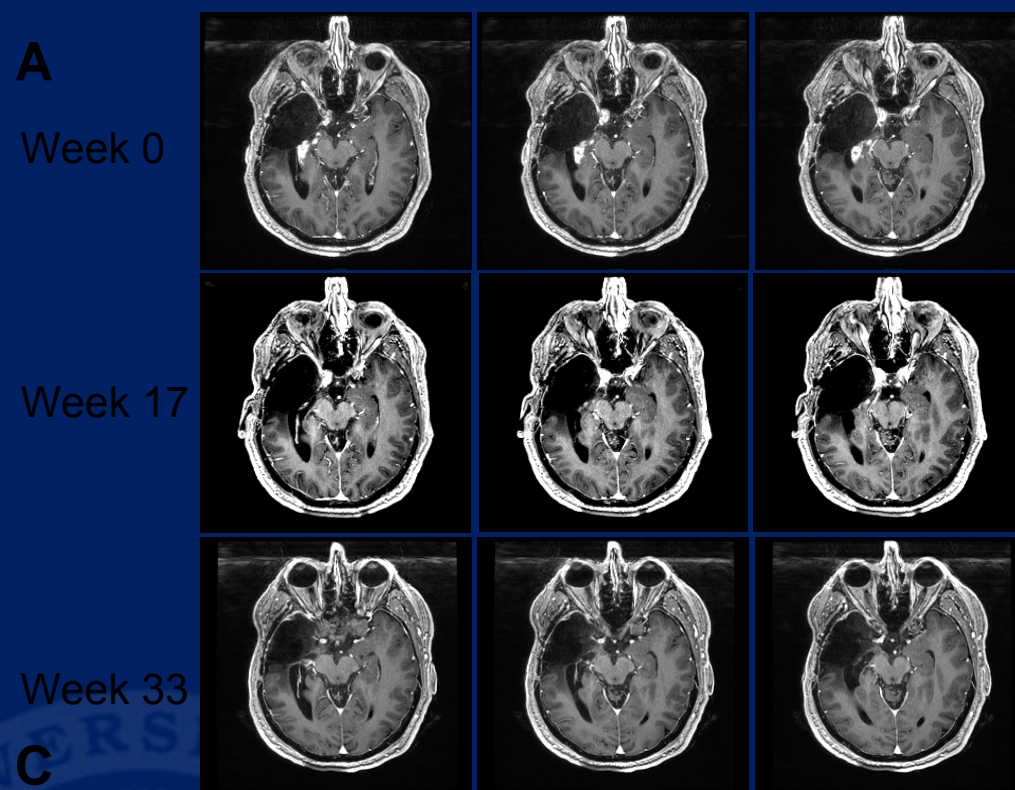
Central Nervous System (CNS) immunology

- “Privileged” status has been revised -

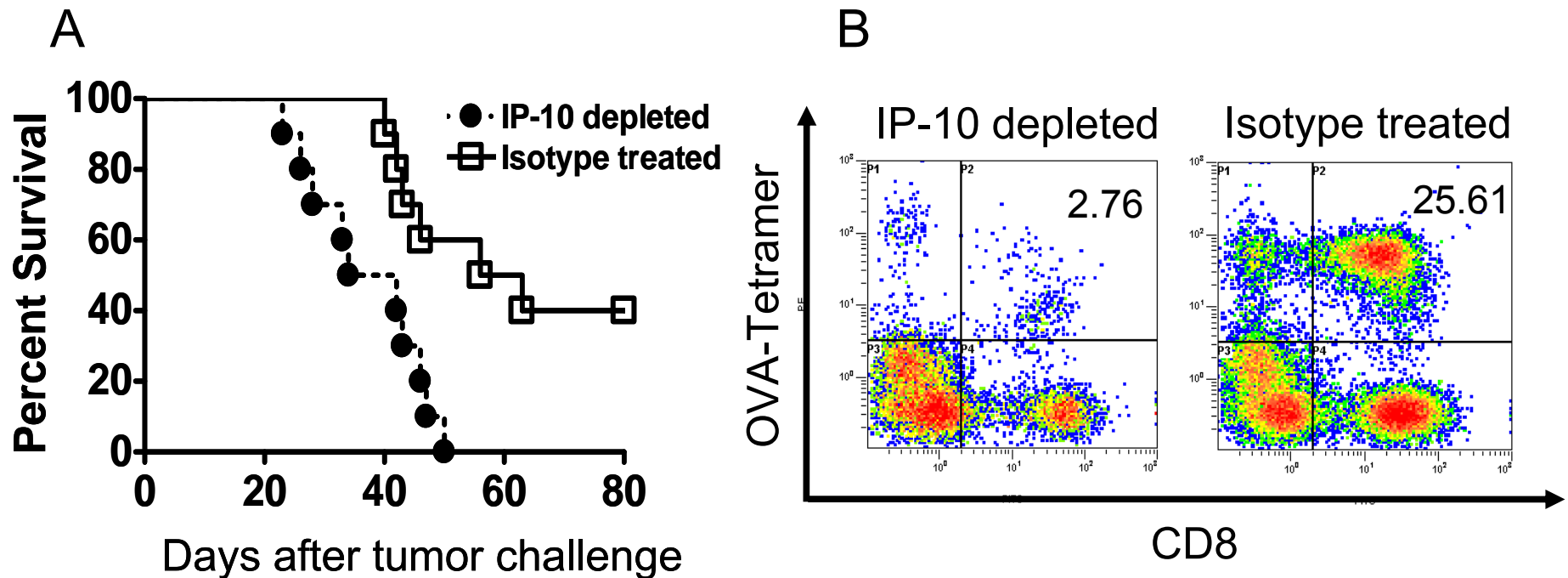
- Autoimmune diseases in the CNS
(Experimental Autoimmune Encephalitis ,
Multiple Sclerosis, Paraneoplastic Cerebellar
Degeneration [PCD] against cdr2)
- A-beta1-40/42 formulated in QS-21 as a
vaccine for **Alzheimer’s disease**- a few
treated patients developed signs of **aseptic
encephalitis/meningitis** following the second
administration of the vaccine.



Figure 4



IFN-inducible protein (IP)-10/CXCL10 plays a critical role in the recruitment of Tc1 cells to the CNS tumor site



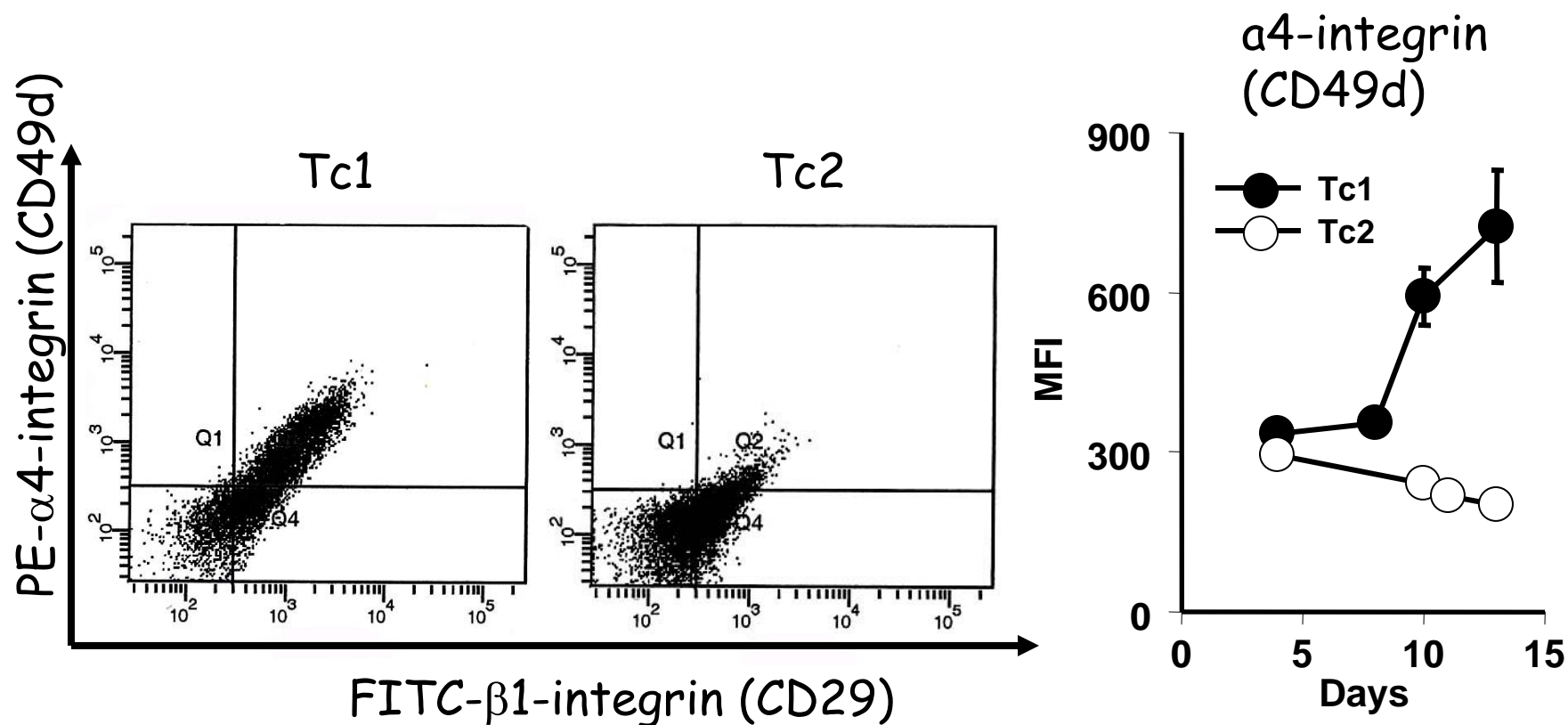
Nishimura F. *et al.* Cancer Res. 2006 66(8):4478-87

IP-10 deficient mice bearing day 7 i.c. M05 received i.v. adoptive transfer of 3×10^6 Tc1 cells and anti-IP-10 i.p. ($250 \mu\text{g}$ on day 6 and $100 \mu\text{g}$ on days 7, 8, and 9). On the same day (day 7), $50 \mu\text{g}$ anti-IP-10 was i.t. co-injected with 1×10^5 DC-IFN- α . BILs were harvested 3 days after adoptive transfer

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High level expression of VLA-4 (heterodimer of $\alpha 4$ -integrin [CD49d] and $\beta 1$ -integrin [CD29]) on Tc1



Sasaki et al. Cancer Res, 2007



Pittsburgh Night View



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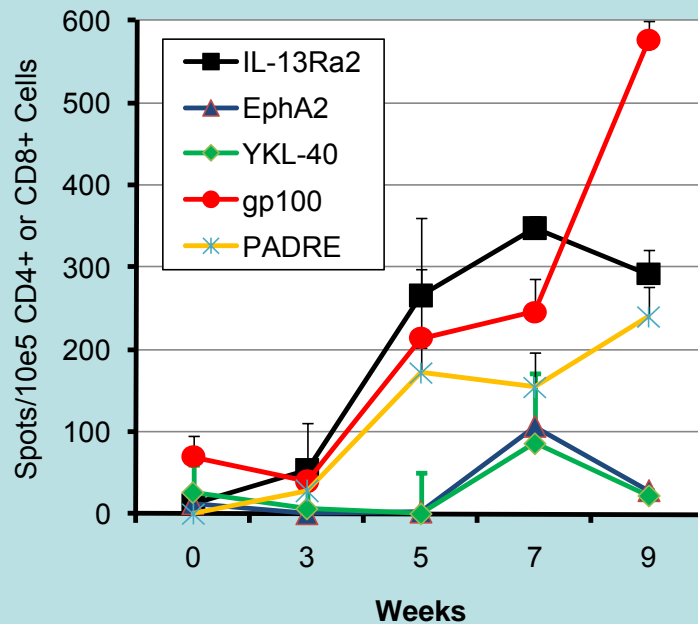


Pittsburgh from Mt. Washiongton

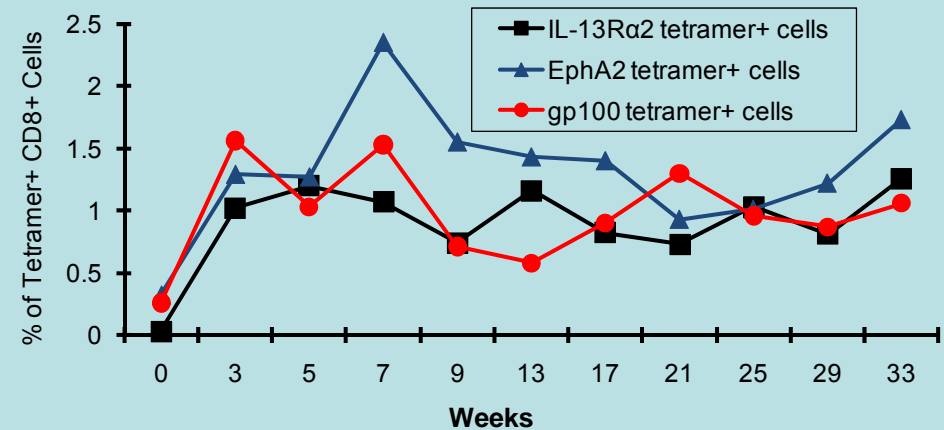
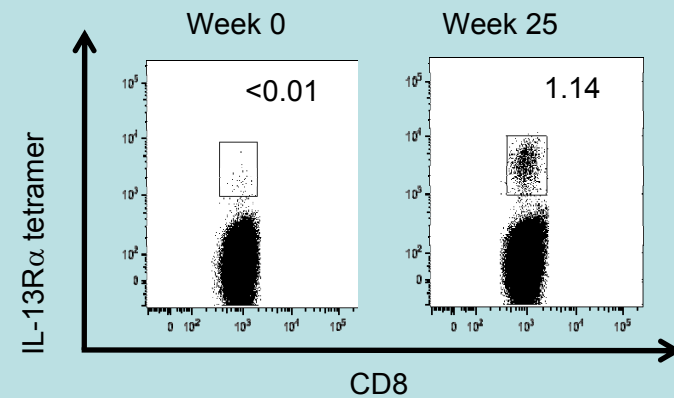


Representative ELISPOT and Tetramer Responders

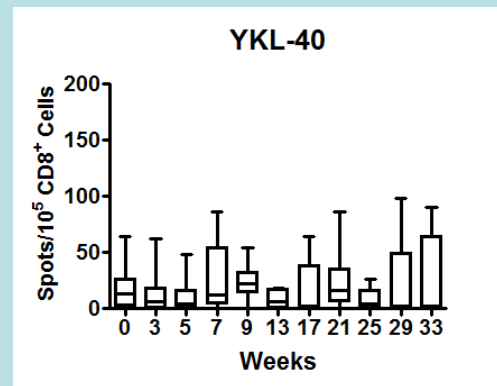
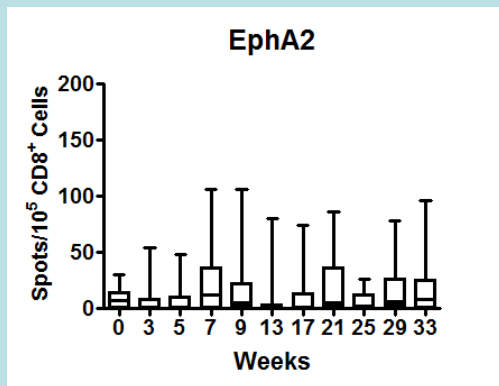
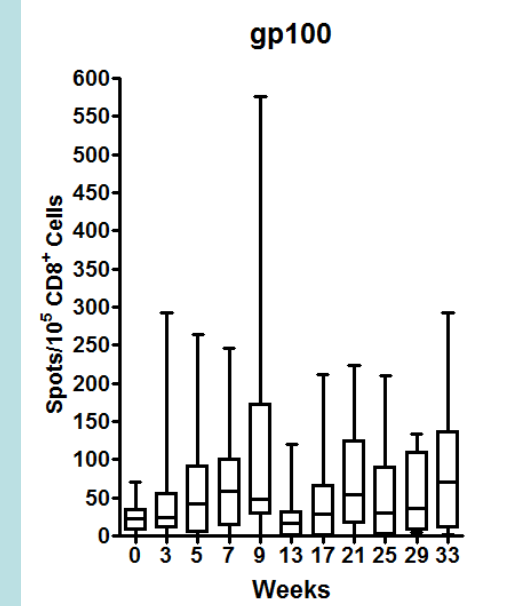
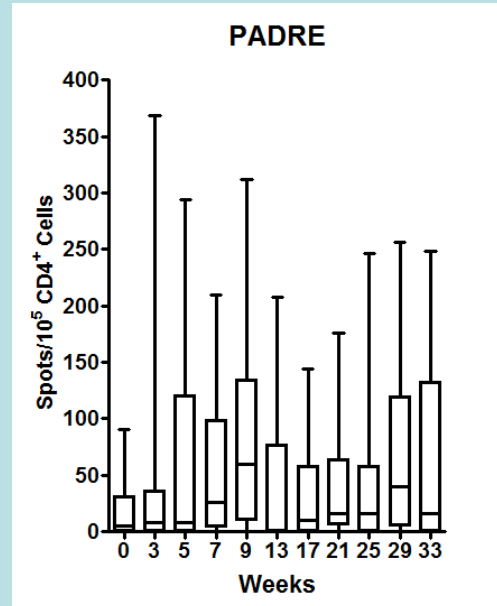
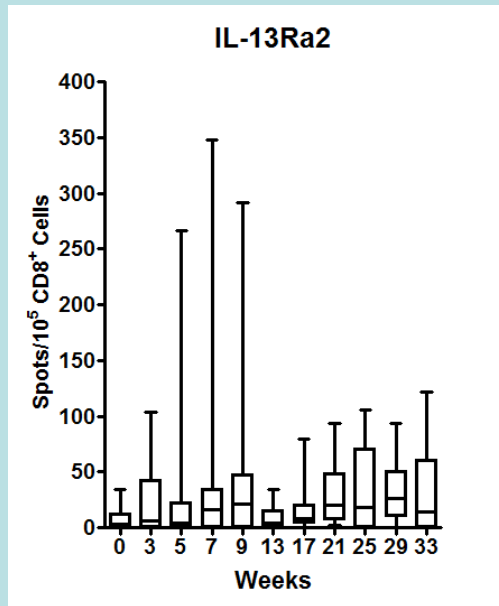
Patient 10 (GBM)



Patient 6 (AOA)

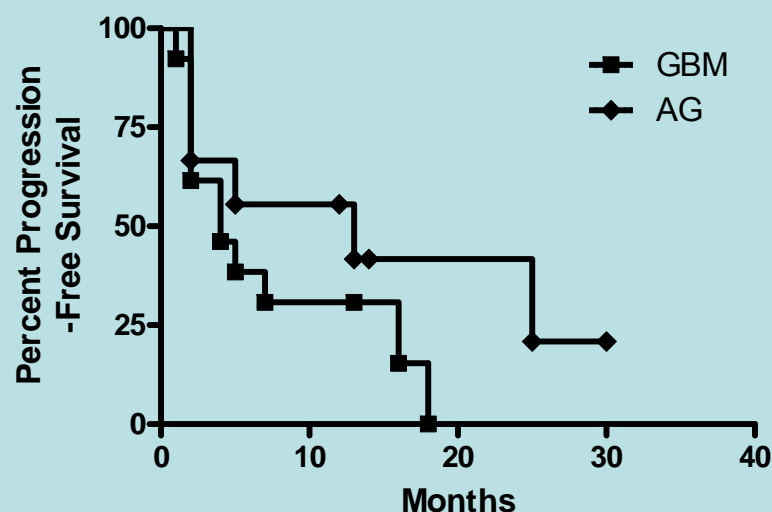


Time course for IFN- γ ESLIPOT assays for all evaluated patients with box plots



TTP and OS for Each of the Two Tumor Grades (WHO Grade 3 vs. 4)

Time to Progression (TTP)

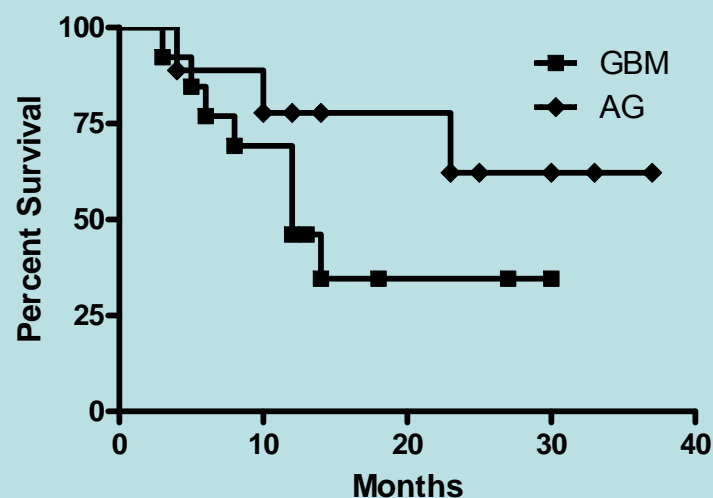


Median TTP

4 months for GBM

13 months for AG

Overall Survival (OS)



Median OS

12 months for GBM

Undefined for AG (as 5 of 9 patients are still alive with the median follow-up period for 23 months)



	Pt ID	Age/ Gender	Tumor Histol.	Location of Tumor	Prior Therapy	No. Prev Rec.	DC IL-12 (pg)	ELISPOT					Tetramer			RR at Week 9	TTP (Mo)	OS (Mo)
								I	E	Y	G	Pa	I	E	G			
Dose Level 1 (1 x 10 ⁷ DC/dose)	1	57/M	GBM	Rt. Temp/Pa	Res/RT/TMZ/Mol	1	10					@	P	P	P	PR	7	14
	2	52/M	GBM	Rt. Temporal	Res/RT/TMZ	1	25					@	P	P	P	PD	<2	12
	3	57/M	AA	Rt. Parietal	Resx2/RT/TMZ/Mol	2	25						N	N	N	SD	5	10
	4	53/M	AA	Rt. Frontal	SB/RT/TMZ	0	<10	Not Tested					Not Tested			ND*	<2	>35
	5	63/M	GBM	Rt. Parietal	SB/RT/TMZ	0	26						N	N	N	PD	<2	5
	6	51/M	AO	Lt. Temporal	SB/RT/TMZ	0	27	\$		\$			P	P	P	SD	>28	>28
	7	37/F	AA	Rt. Temporal	SB/RT/TMZ	0	919	\$			\$		P	P	P	SD	25	>31
	8	45/F	GBM	Rt. Frontal	SB/RT/TMZ/Mol	0	480	\$		\$	\$		P\$	P\$	N	SD	16	>29
	9	43/F	AO	Rt. Frontal	Res/RT/TMZ/SR	0	24			@			N	N	N	PD	<2	>25
	10	71/F	GBM	Lt. Parietal	Resx2/RT/TMZ/CE	2	<10		@	@			N	N	N	SD	5	>25
Dose Level 2 (3 x 10 ⁷ DC/dose)	11	54/M	GBM	Rt. Temporal	Res/RT/TMZ/Mol	2	38	Not Tested					Not Tested			ND*	<2	3
	12	33/F	AO	Lt. Frontal	SB/RT/TMZ	0	111	\$	\$	\$			P\$	N	N	SD	13	>23
	13	46/M	AA	Rt. Parietal	Res/RT/TMZ	1	151	Not Tested					Not Tested			ND*	<2	4
	14	54/M	GBM	Multiple	Res/RT/TMZ	0	35				@		N	N	N	SD	4	>15
	16	33/M	AO	Rt. Frontal	Res/RT/TMZ/Mol	2	985						N	P\$	P\$	SD	>12	>12
	17	30/F	GBM	Lt. Parietal	Resx2/RT/TMZ/CW	1	123	@		@			Not Tested			PD	<2	6
	18	61/F	GBM	Bil. Occipital	Res/RT/TMZ/BI	1	125						N	N	N	PD	<2	5
	19	63/M	GBM	Lt. Temporal	Res/RT/TMZ/SR	1	199						P\$	P\$	P	SD	>11	>11
	20	62/M	GBM	Rt. Temporal	Res/RT/TMZ	0	287						P\$	P\$	P\$	PR	>11	>11
	21	38/F	GBM	Rt. Hemi	Res/RT/TMZ/CPT-Bev	1	27			@			Not Tested			PD	<2	>11
	22	28/M	AA	Brain Stem	SB/RT/TMZ/Res	0	779	P					Not Tested			SD	>9	>9

Please, do not read – I will summarize these findings for you –

<24

25-49

50-99

100-199

200≤

Spots/
Per 10e5
Cells



Poly-ICLC as the Key Adjuvant

- Polyinosinic-polycytidylic acid (poly-IC) stabilized with poly-lysine and carboxymethylcellulose (Poly-ICLC), has been used as a single agent or combo with TMZ for treatment of malignant glioma (provided by Dr. Andres Salazar [Oncovir]).
- Poly-ICLC stimulates TLR3 RIG-I and MDA-5
- Among the TLRs, TLR3 is most abundantly expressed in the CNS.



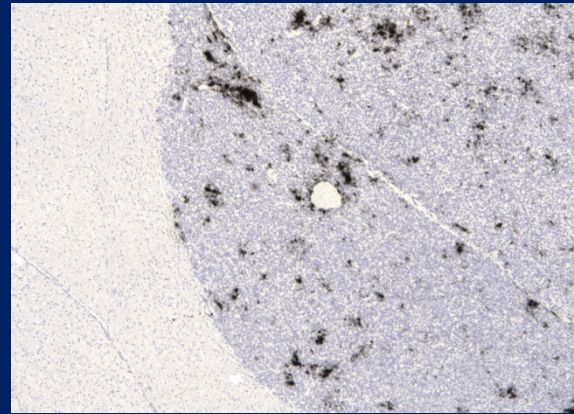
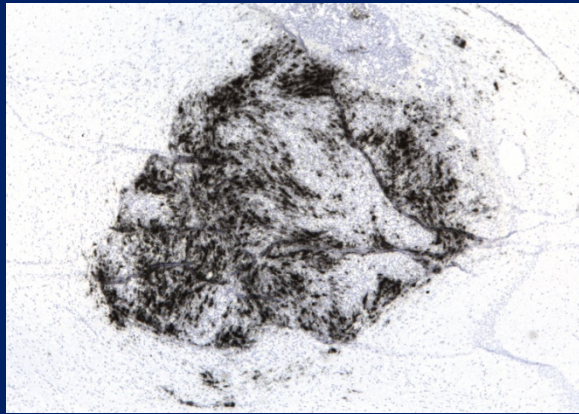
Demographics and Clinical Characteristics of Participating Patients

Characteristics	<u>DC Dose Level (No. of DC/dose)</u>		<u>Total (n=22)</u>	
	1 (1 x 10 ⁷)	2 (3 x 10 ⁷)	No. of Patients	%
Received at least one vaccine	11	11	22	
Completed at least 4 vaccines	10	9	19	86
Female (received at least 4 vaccines)		5	4	9
47				
Median age, years	52	46	48	
Range	37-71	28-63	28-71	
Tumor Histology				
AA	3	2	5	23
AO	1	2	3	14
AOA	1	0	1	4
GBM	6	7	13	59
No. of Previous Recurrences				
0	7	4	11	50
1	2	5	7	32
2	2	2	4	18

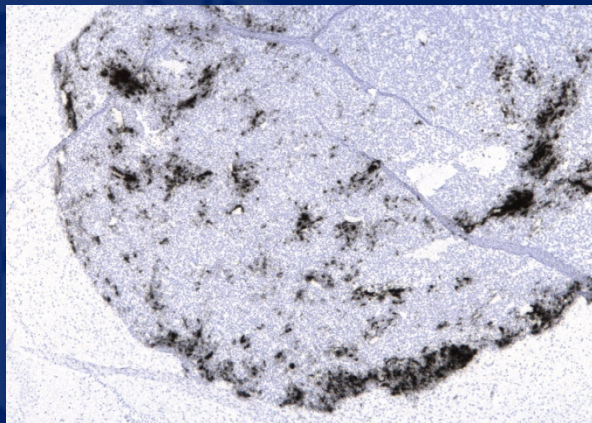


Upregulated expression of CXCL10 mRNA in murine GL261 glioma treated with GAA-vaccines and i.m. poly-ICLC (*In situ* hybridization)

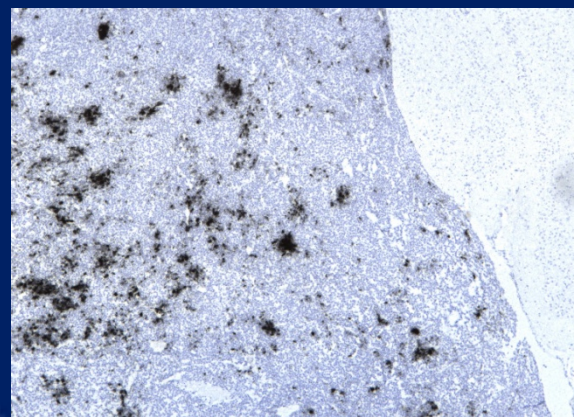
A Vaccine Plus Poly-ICLC B Vaccine Alone



C Poly-ICLC Alone



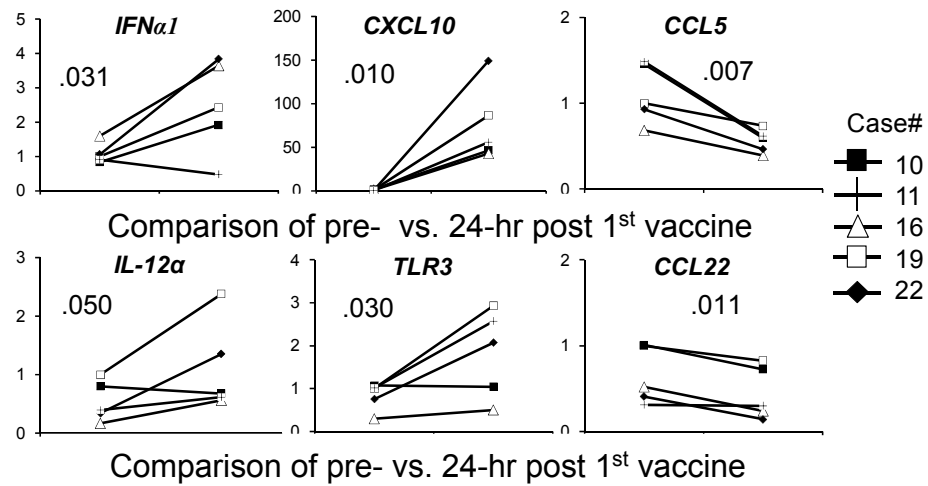
D Mock-Treatment



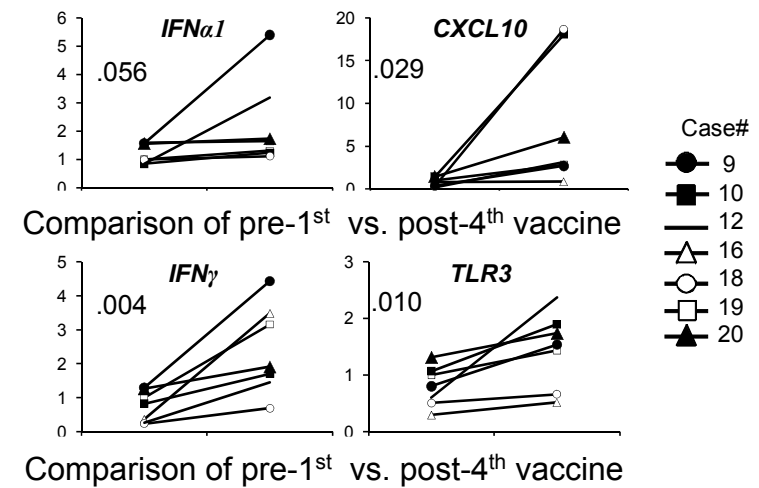
Zhu X. *Et al*
Cancer Immunol.
Immunother.
2010



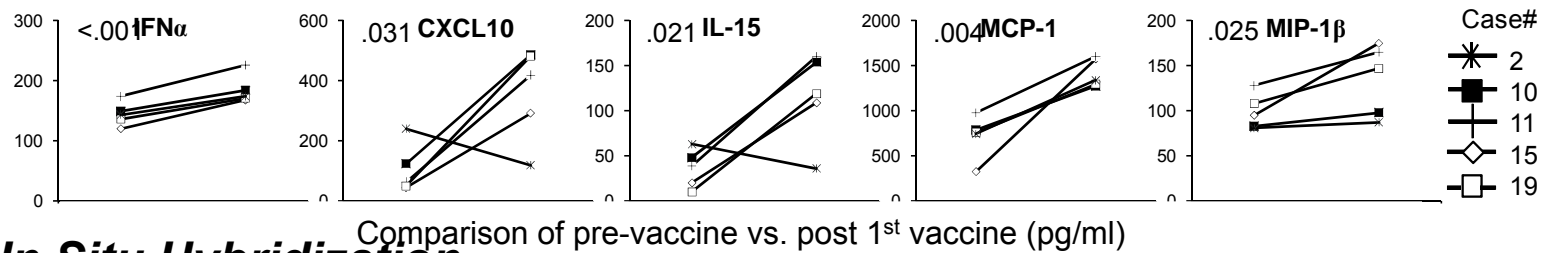
RT-PCR (Pre-vac vs. 24-hr Post 1st Vac



RT-PCR (Pre-vac vs Post-4th Vac



C Luminex



D In Situ Hybridization

