

SITC 2019

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Gaylord National Hotel
& Convention Center

NATIONAL HARBOR, MARYLAND



Society for Immunotherapy of Cancer



Toll Like Receptors 7/8 in Cancer Therapy

Willem Overwijk, PhD

NEKTAR Therapeutics



Society for Immunotherapy of Cancer

#SITC2019

Presenter Disclosure Information



Willem Overwijk, PhD
Nektar Therapeutics

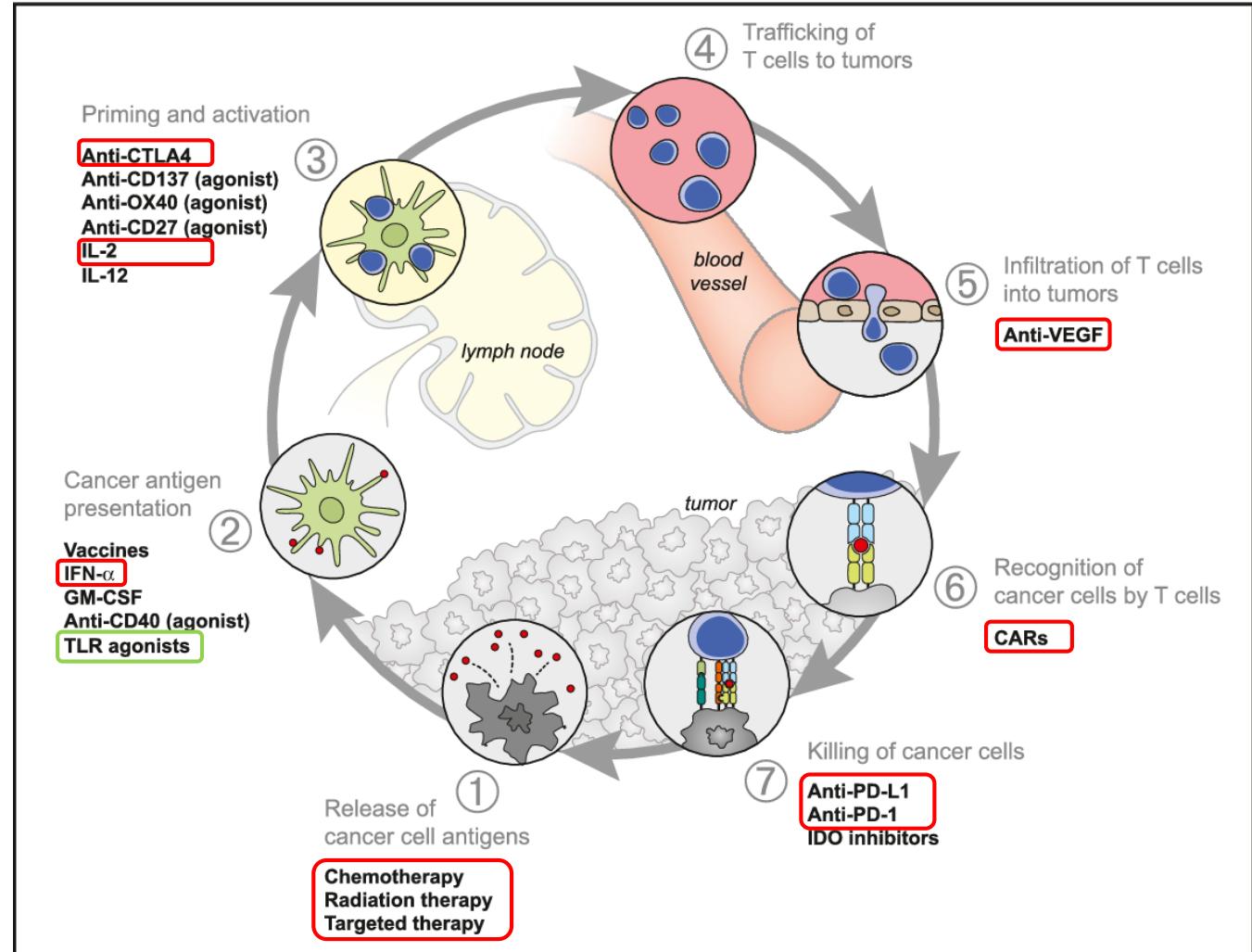
The following relationships exist related to this presenter:

- Employee and Shareholder of Nektar Therapeutics
- Nektar Therapeutics develops the intratumoral TLR7/8 agonist, NKTR-262

A Place for TLR Agonists in the Cancer Immunity Cycle

Ideal Scenario:

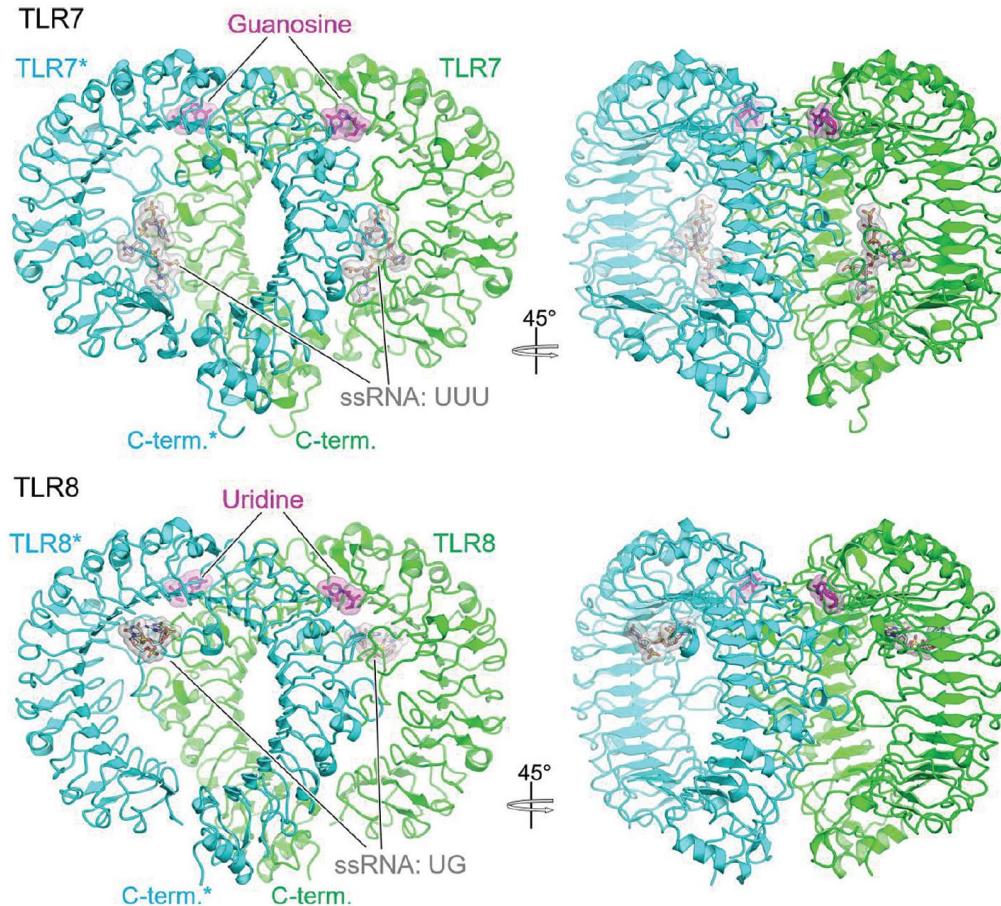
1. Intratumoral presence of TLR agonist
2. Antigen presenting cell maturation
3. Tumor antigen presentation to T cells
4. T cell (re-)activation
5. Systemic T cell response
6. Systemic tumor eradication



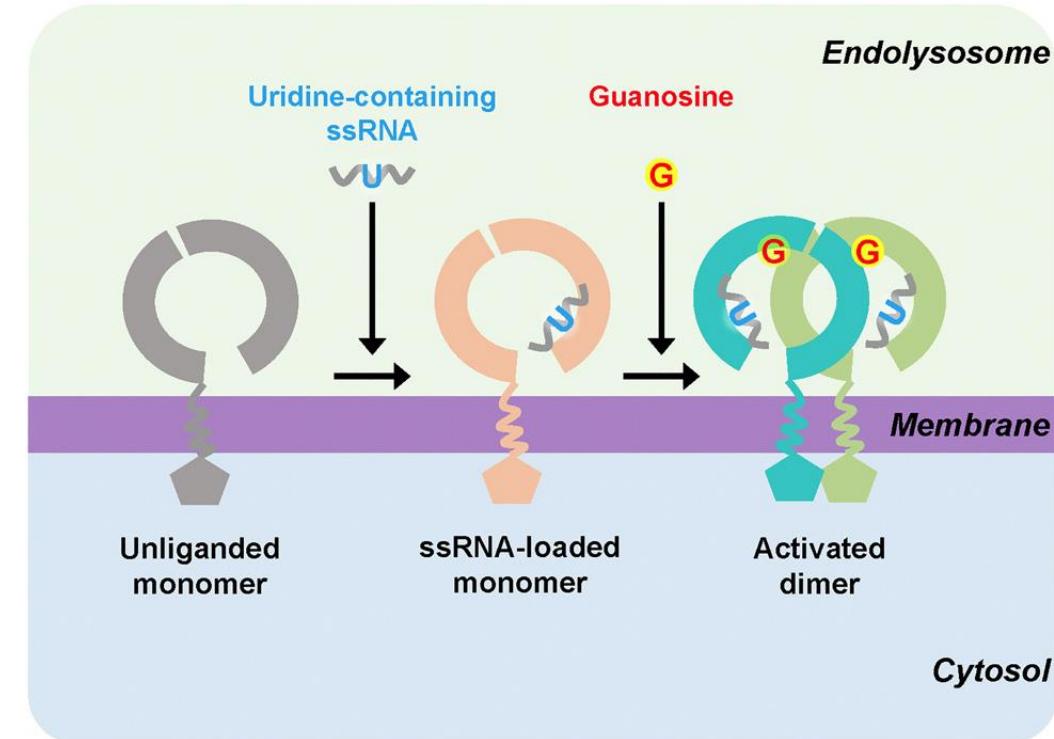
Chen and Mellman. *Immunity*. 2013;39:1-10.

TLR7 and TLR8 Structure and Signaling

TLR 7/8 Molecular Structure and Natural Ligand Binding Sites

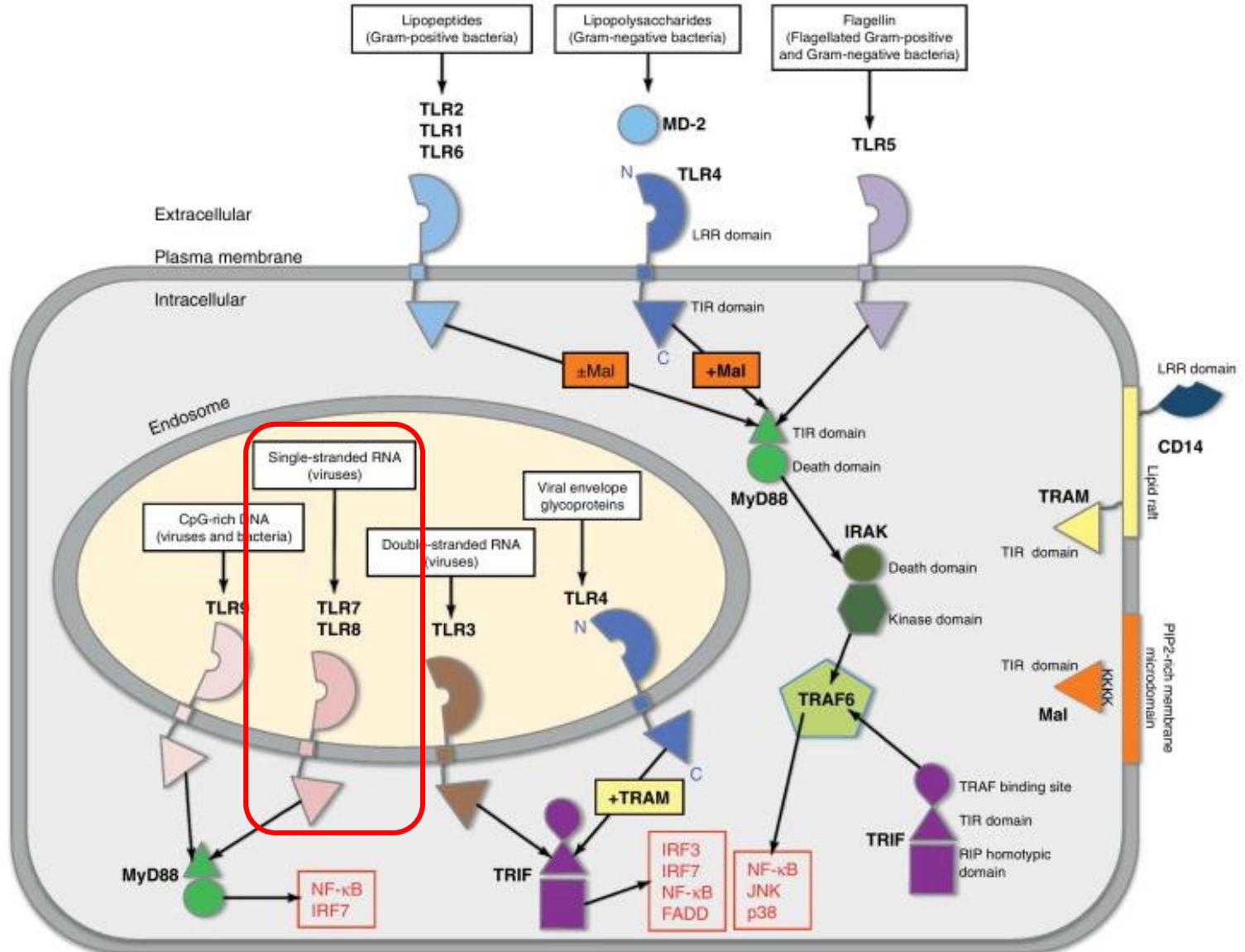


TLR7/8 ligands: ssRNA and ssRNA-derived guanosine (G) and uridine (U)



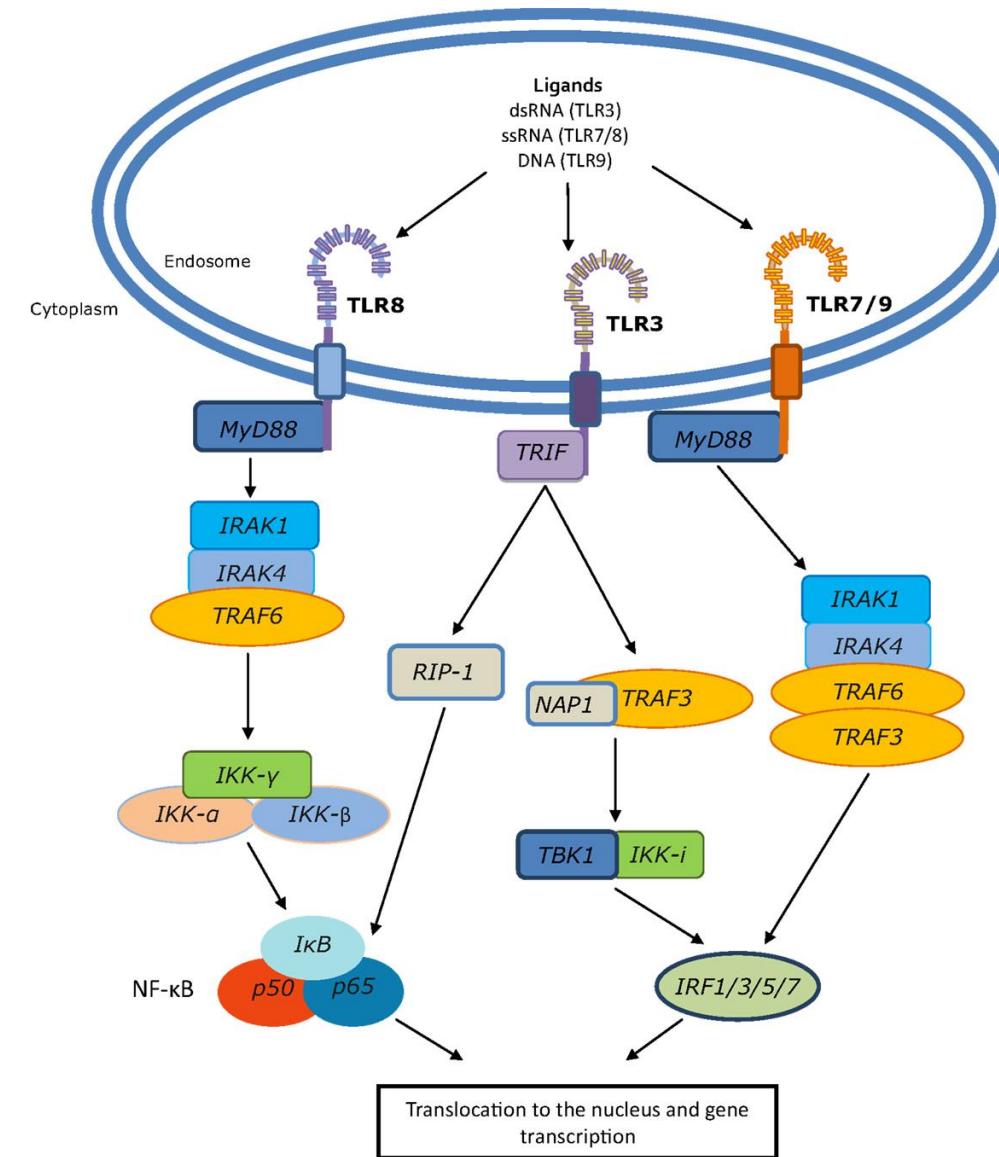
Miyake *et al.*, Immunity 2016

TLR7/8 Are Expressed in Endosomes



Gangloff. *Trends Biochem Sci.* 2012;37:92-8.

Consequence of TLR7 vs. TLR8 Signaling



Thwaites et al. *Front Immunol.* 2014 Jan 16;5:1. doi: 10.3389/fimmu.2014.00001.

Expression of TLR 7/8 and Consequences of Triggering

Table 1 TLR expression and functionality on DC subtypes

DC subtype	TLR	Expression	Effects of activation	DC subtype	TLR	Expression	Effects of activation
moDC	1	+	See TLR2	pDC	1	±	
	2	++	Increased IL-6, IL-8, IL-10, IL-12, TNF α Low IFN β response and no IFN γ response		2	-	
	3	+	Specific IFN β mRNA upregulation (not IFN γ)		3	-	
	4	++	Upregulation of CD80, CD86, CD83, CCR7 Secretion of IFN β , IFN γ , IL-1 β , IL-12p70, IL-13, IP-10 Decreased endocytic capacity		4	-	
	5	+	Upregulation of CD80, CD86, CD83, CCR7 Secretion of IFN γ , IL-1 β , TNF, IL-8, IL-12p40 (not IL-12p70), IL-13 Decreased endocytic capacity		5	-	
	6	±	See TLR2		6	-	
	7	±	-		7	++	Upregulation of CD40, CD80, CD86, CCR7 Very high IFN γ response No IL-12p70 response
	8	+	Increased TNF α , IL-8, IL-12p40, MCP-1, CCL2, CCL3, CCL4, CCL5		8	-	
	9	-			9	+++	Upregulation of CD40, CD80, CD86, CD83, HLA-DR, CCR7 Upregulation of IFN γ (very high), IFN β (lower), IL-6, TNF α (low), IL-8, IP-10 No IL-10 secretion No ligand known
	10	-			10	+	
mDC	1	+	See TLR2				
	2	++	Upregulation of CCR7, IL-6, IL-10, IL-12p70, TNF α , no INF α				
	3	++	IFN α (intermediate), IL-12p70 (high) No TNF α or IL-6				
	4	+	Upregulation of CD80, CD86, CD83, CD40, CCR7 Secretion or upregulation of CCR7, IL-6, IL-8, IL-10, IL-12p70 No IFN α response				
	5	+	Upregulation of CD80, CD86, CD83, CCR7 Secretion of TNF and IL-8 Upregulation of CCR7				
	6	+	See TLR2				
	7	+	Upregulation of CD40, CD80 and CD86 Secretion of IL-12p70 No IFN α response				
	8	±	See TLR7				
	9	-					
	10	+	No ligand known				

Schreiberl et al. *Cancer Immunol Immunother.* 2010;59:1573-82.

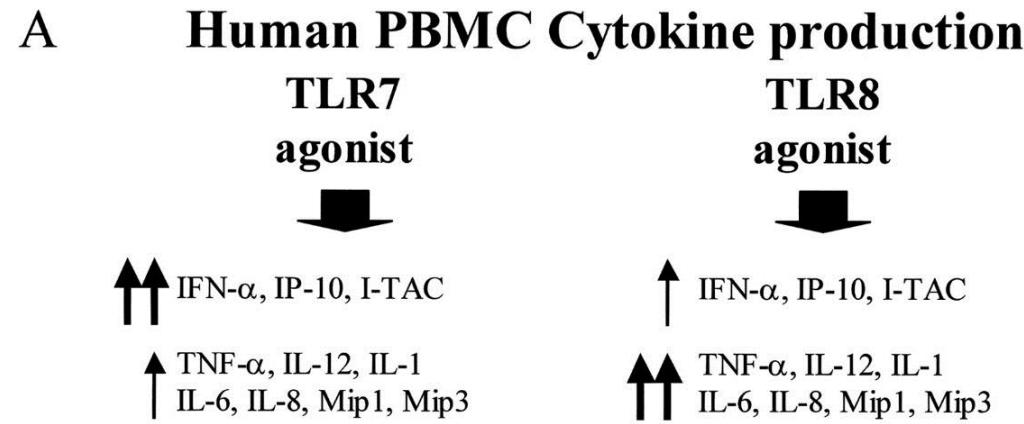
Expression of TLR 7/8 and Consequences of Triggering

Table 1 Phenotype and function of human DC subsets

	CD1c ⁺ DC	CD141 ⁺ DC	pDC	MoDC	LC
Mouse equivalent	CD11b ⁺ cDC	CD8 ⁺ cDC	pDC	Inf DC	LC
Location	Blood, lymphoid, non-lymphoid organs	Blood, lymphoid, non-lymphoid organs	Blood, lymphoid organs	Skin (steady state), inflamed tissues (inflammation)	Skin
Phenotype	Lin ⁻ CD11c ⁺ DR ⁺ Sirp α (CD172a) ⁺ CD1c ⁺ CD14 ^{low} CD11b ⁺ CD103 ⁺ (intestine only)	Lin ⁻ CD11c ⁺ DR ⁺ CD141 ⁺ XCR1 ⁺ CLEC9A ⁺ Necl2 ⁺	Lin ⁻ CD11c ⁻ DR ⁺ BDCA-2 ⁺ BDCA-4 ⁺ CD123 ⁺	Lin ⁻ CD11c ⁺ DR ⁺ CD1a ⁺ CD1c ⁺ CD14 ⁺ CD11b ⁺ Fc ε RI	Lin ⁻ CD11c ⁺ DR ⁺ CD1c ⁺ Langerin ⁺ E-cadherin ⁺
TLR expression	3 (low), 4 (low), 8	3, 8	7, 9	3 (low), 4, 7 (low), 8	3
Cytokine profile	IL-12, IL-23, IL-10	Type III	Type I and III IFN	IL-1 β , IL-6, IL-10, IL-23	ND
Functions to date	Th2 induction in response to allergen, Th17 induction in response to fungal infection, immune regulation	Cross presentation of cellular Ag and immune complexes, CTL priming.	Type I IFN production against viral and fungal infections, immune regulation	Induction of Th1 and Th17 responses	Induction of Th2, CTL responses

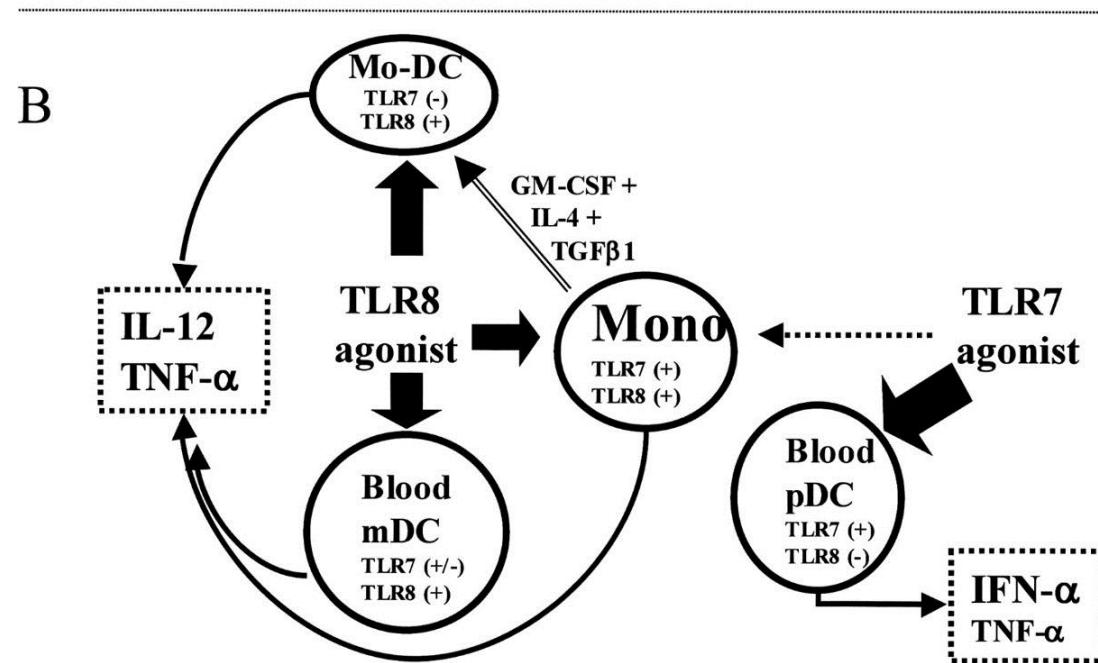
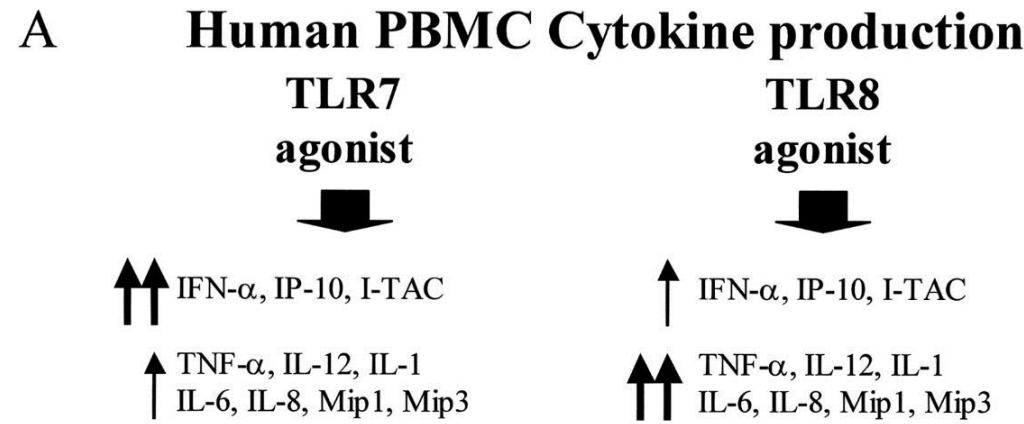
O'Keeffe et al. *Cell Mol Life Sci.* 2015;72:4309-25.

TLR7 vs TLR8 Response in Whole PBMCs



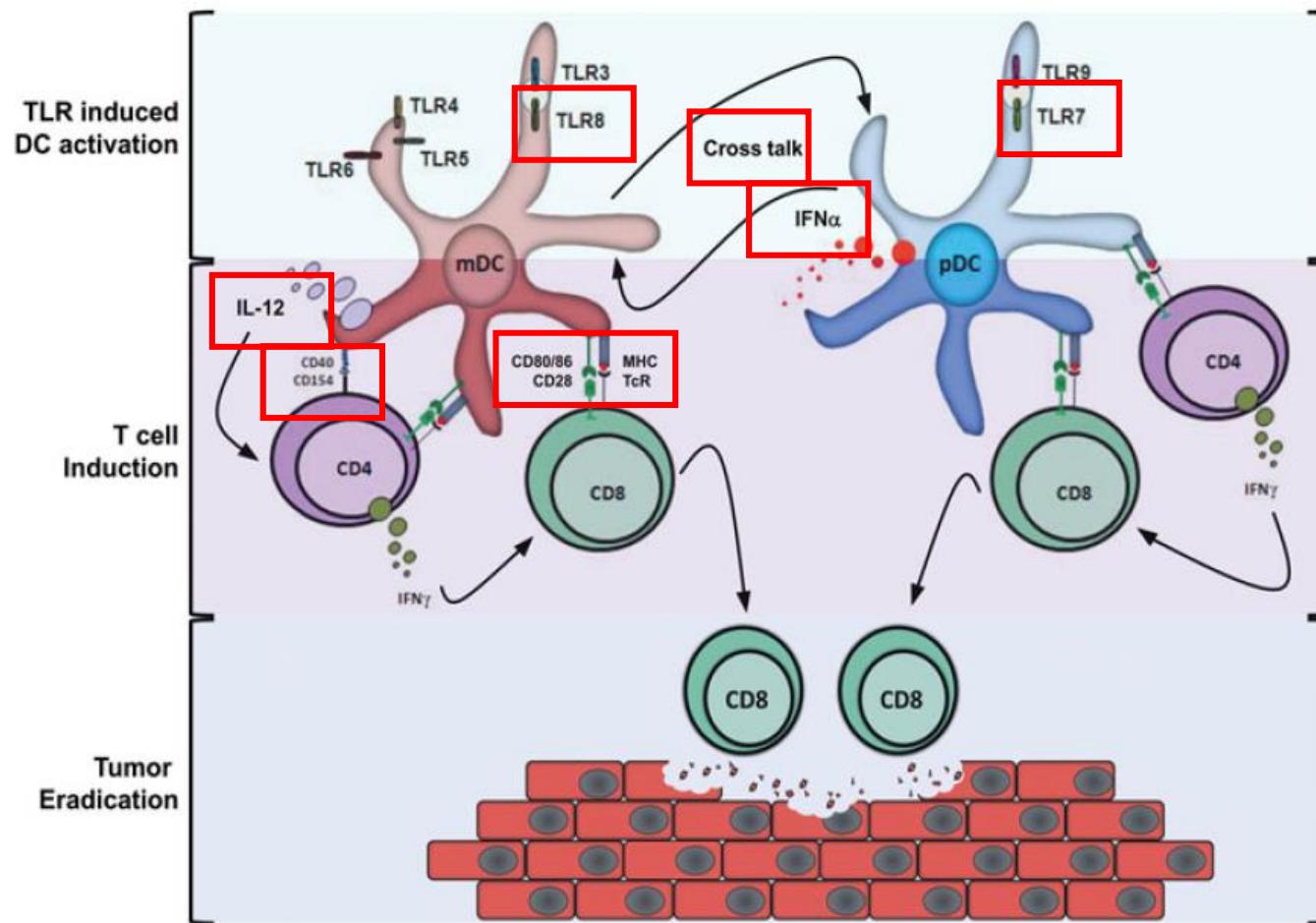
Keith B. Gorden et al. *J Immunol* 2005;174:1259-1268

TLR7 vs TLR8 Response in Whole PBMCs



Keith B. Gorden et al. *J Immunol* 2005;174:1259-1268

TLR7 on pDCs and TLR8 on mDCs: Consequences

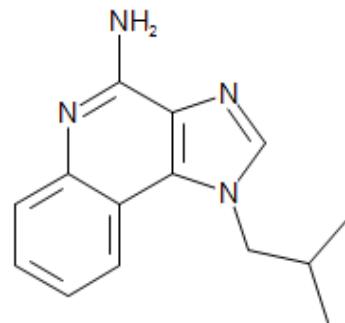


Schreiberl et al. *Cancer Immunol Immunother.* 2010;59:1573-82.

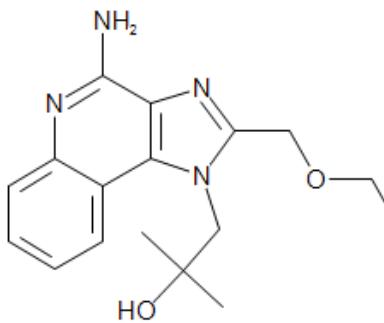
Preclinical Studies: MOA and Potential for Combinations

Structural Similarity Among Some Imidazoquinoline Compounds Under Investigation

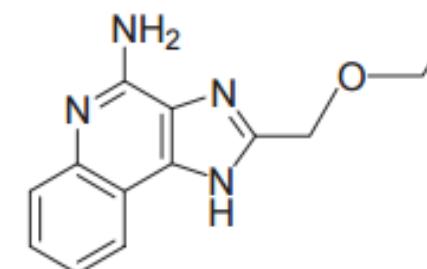
Imiquimod (TLR7)



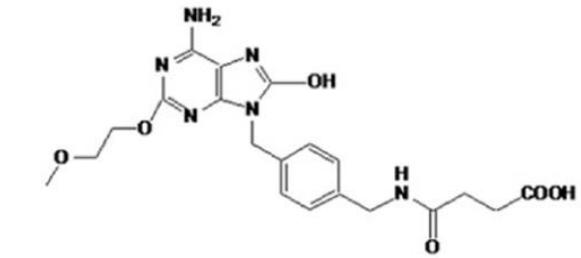
Resiquimod/R848 (TLR7/8)



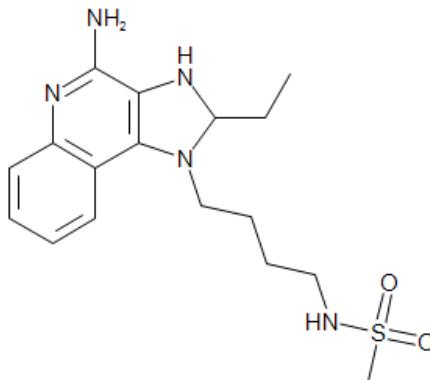
CL097 (TLR7/8)



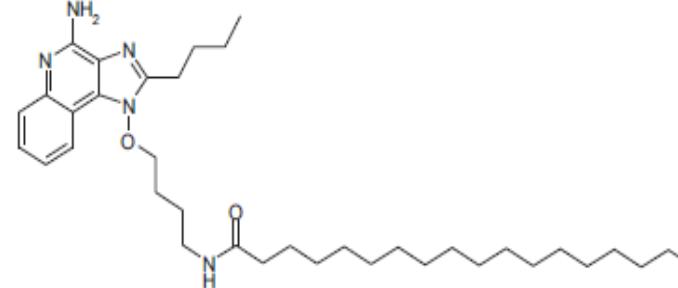
SZU-101 (TLR7)



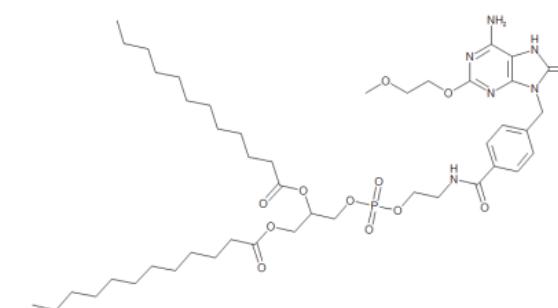
852A (TLR7)



MEDI9197/3M-052 (TLR7/8)



TMX-202 (TLR7)



The chemical structure of Guanosine (TLR7) is shown. It consists of a purine ring system fused to a pyrimidine ring. The purine ring has an amino group (-NH₂) at position 2 and a carbonyl group at position 6. The pyrimidine ring has a carbonyl group at position 4 and an amino group (-NH₂) at position 2'. The nucleoside is linked to a glucose moiety via its 2' hydroxyl group. The glucose moiety has an oxygen atom at position 1' and two hydroxyl groups at positions 2' and 3'.

Preclinical and Clinical Oncology Studies with TLR7/8 Agonists

	Agonist	Tumor type	<i>In vitro</i>	<i>In vivo</i>	References
TLR7	Imiquimod	Squamous carcinoma Prostate cancer Bladder cancer Breast cancer Melanoma	YS-10B, FaDu, TRAMP-C2, PC3 MB49 TSA /	/ C57BL/6 C57BL/6 BALB/c Human (preclinical) Human (preclinical)	Ahn et al., 2012 Han et al., 2013 Hayashi et al., 2010 Adams et al., 2012; Dewan et al., 2012 Narayan et al., 2012
	TLR7/8	Gliomas Acute myeloid leukemia	GL261 HL60, THP1, OCI-AML3, HCT116, 293T	C57BL/6 Nod/SCID/IL2R γ -/(NSG)	Grauer et al., 2008 Smits et al., 2010; Ignatz-Hoover et al., 2015
				BALB/c Human (phase I)	Yin et al., 2015 Rook et al., 2015
		Breast cancer T-cell lymphoma	4T1 /		
	Gardiquimod	Melanoma Pancreatic cancer	B16 BxPC-3	C57BL/6 /	Ma et al., 2010 Zou et al., 2015
TLR7	852A	Ovarian cancers Cervix cancer Breast cancer Melanoma Lymphocytic leukemia	/ / / / /	Human (preclinical) Human (preclinical) Human (preclinical) Human (phase II) Human (phase I/II)	Geller et al., 2010 Geller et al., 2010 Geller et al., 2010 Dummer et al., 2008 Spaner et al., 2010
	Loxoribine	Melanoma B-chronic leukemia	B16 /	C57BL/6 Human (preclinical)	Pope et al., 1994 Tosi et al., 1997; Pellacani et al., 1999
	TLR7	Bladder tumor Prostate cancer Renal-cell carcinoma	KK-47 724 MBT-2 Renca	/ / BALB/c	Tei et al., 2002 Sarosdy, 1997 Fujioka et al., 1995
		Pancreatic cancer Colon cancer	BxPC3 Panc-1 HT29 HCT-116	BALB/c C57BL/6 BALB/c C57BL/6	Scholch et al., 2015 Scholch et al., 2015

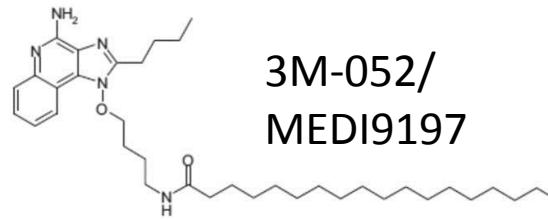
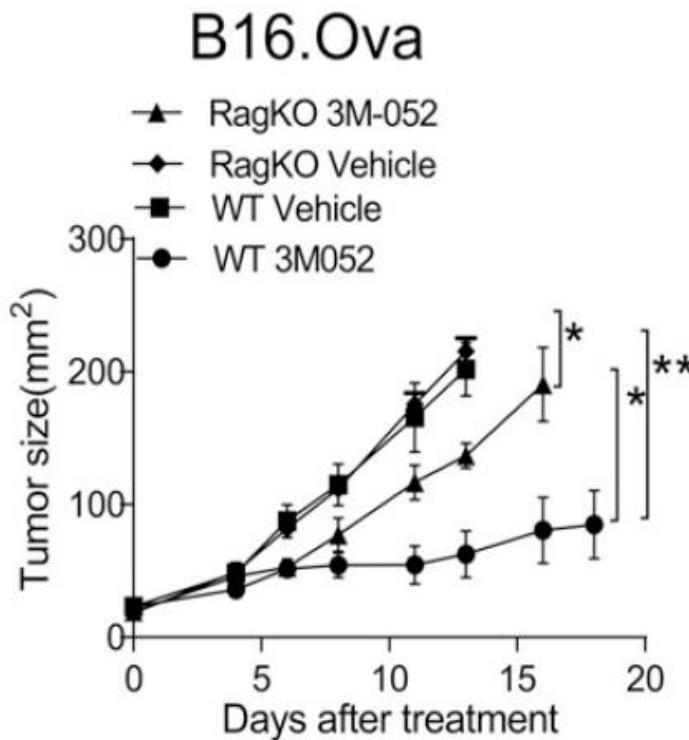
Chi et al., Front. Pharm. 2017

Preclinical and Clinical Oncology Studies with TLR7/8 Agonists

	Agonist	Tumor type	In vitro	In vivo	References
TLR7/8	3M-052	Melanoma	B16.F10, B16.OVA, BP	C57BL/6	Singh et al., 2014
TLR7	DSR-6434	Colon cancer Renal cell carcinoma	CT26 Renca	C3H BALB/c Balb/c C57BL/6	Adlard et al., 2014 Koga-Yamakawa et al., 2015
TLR7	DSR-29133	Colon cancer Osteosarcoma Renal cell carcinoma	CT26 LM8 Renca	Balb/c C3H Balb/c	Dovedi et al., 2016 Dovedi et al., 2016 Dovedi et al., 2016
TLR7	SC1	Lymphoma Renal cell carcinoma	RMA-S Renca	C57BL/6 Balb/c	Wiedemann et al., 2016 Hamm et al., 2009
TLR7	SZU-101	Breast carcinoma Gastric cancer T cell lymphoma	4T1 EAC EL4	Balb/c Balb/c C57BL/6	Diao et al., 2016 Wang et al., 2015 Zhu et al., 2015
TLR7	SM-360320	Colon cancer	MC38	BALB/c	Dharmapuri et al., 2009
TLR7	SM-276001	Renal cell carcinoma Colon cancer	Renca CT26	Balb/c Balb/c	Koga-Yamakawa et al., 2013 Koga-Yamakawa et al., 2013
TLR7/8	Resiquimod	Gliomas Acute myeloid leukemia Breast cancer T-cell lymphoma	GL261 HL60, THP1, OCI-AML3, HCT116, 293T 4T1 /	C57BL/6 Nod/SCID/IL2Rγ ^{-/-} (NSG) BALB/c Human (phase I)	Grauer et al., 2008 Smits et al., 2010; Ignatz-Hoover et al., 2015 Yin et al., 2015 Rook et al., 2015
TLR8	VTX-2337	Lymphoma	/	Human (phase I)	Northfelt et al., 2014
TLR7/8	3M-011	Pancreatic cancer Colon cancer	BxPC3 Panc-1 HT29 HCT-116	BALB/c C57BL/6 BALB/c C57BL/6	Scholch et al., 2015 Scholch et al., 2015

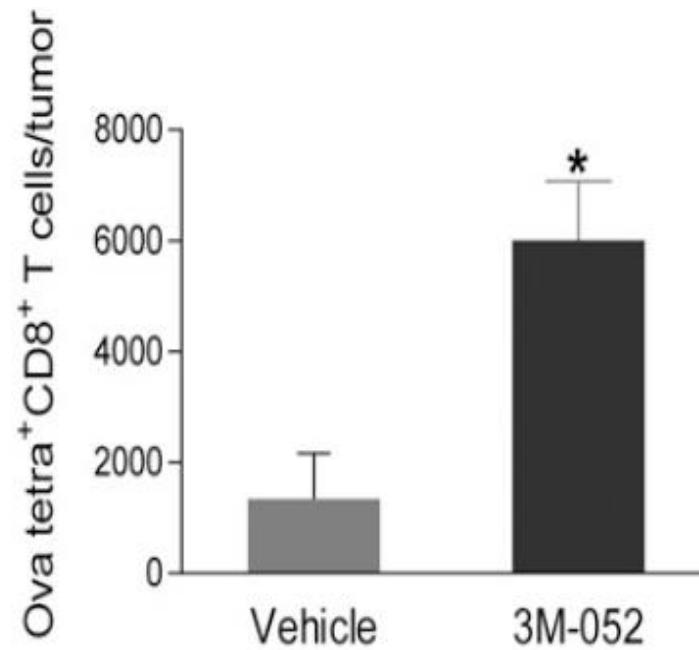
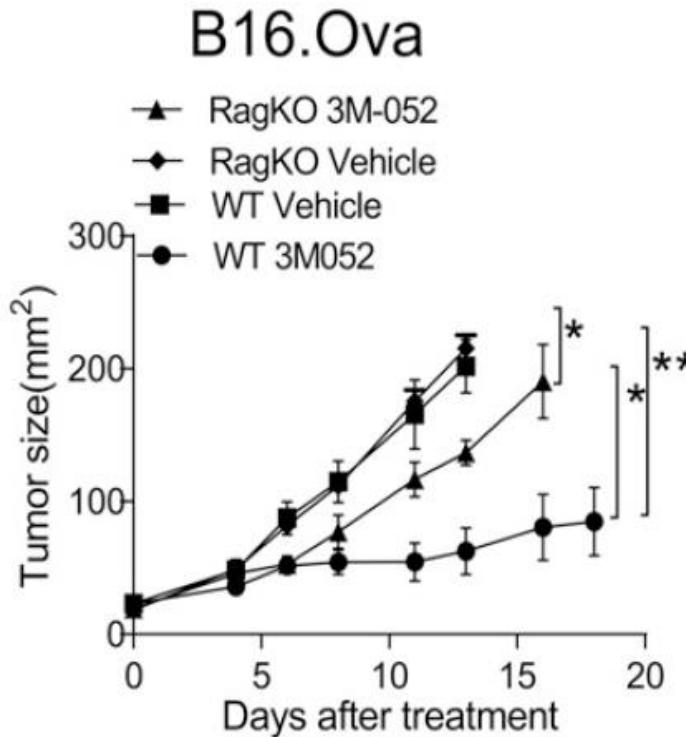
Chi et al., Front. Pharm. 2017

TLR7/8 Agonist Efficacy Depends on CD8⁺ T cells ...



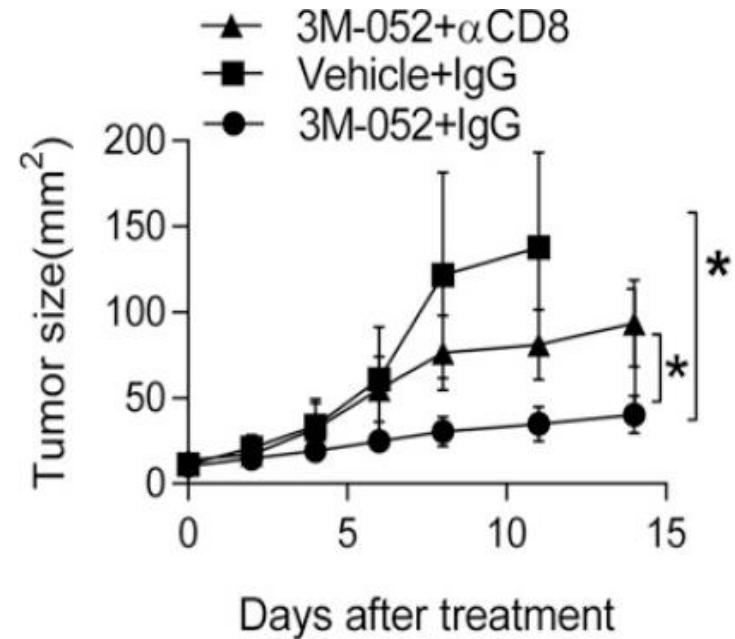
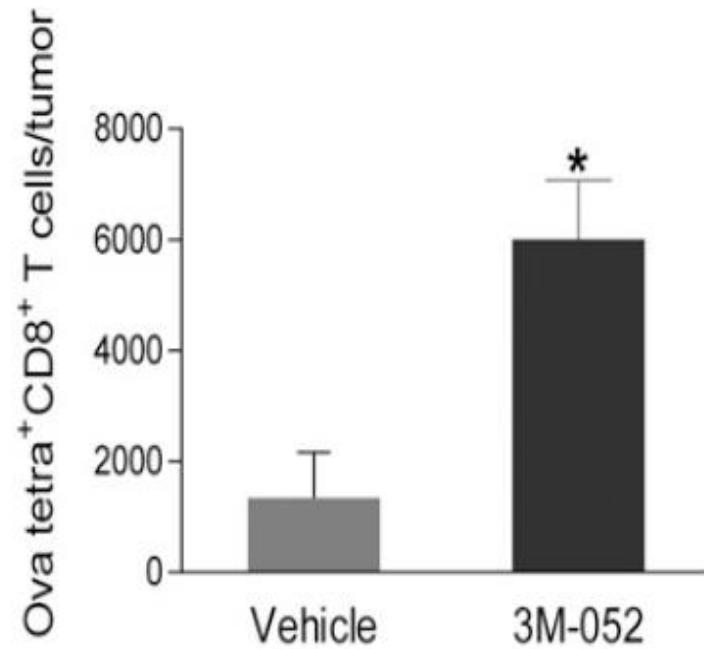
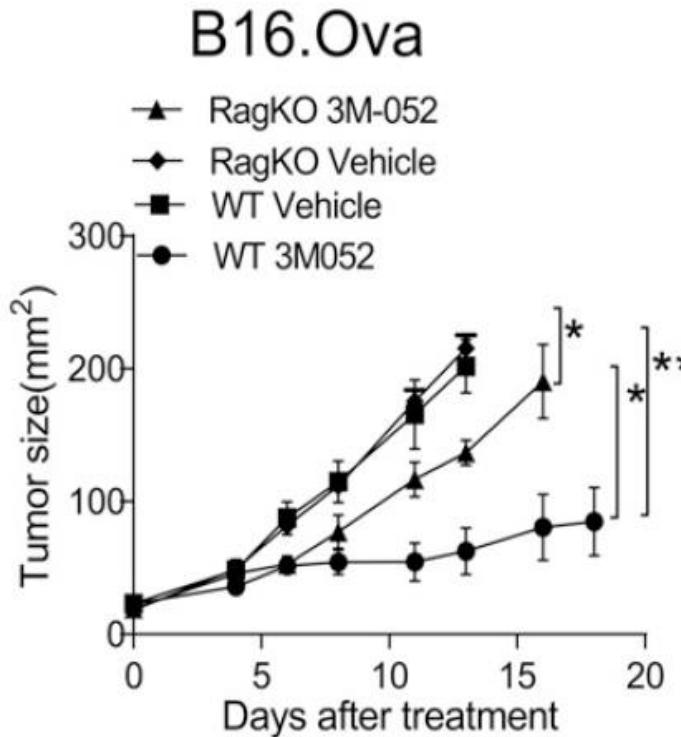
Singh et al., *J. Immunol.* 2014

TLR7/8 Agonist Efficacy Depends on CD8⁺ T cells ...



Singh et al., *J. Immunol.* 2014

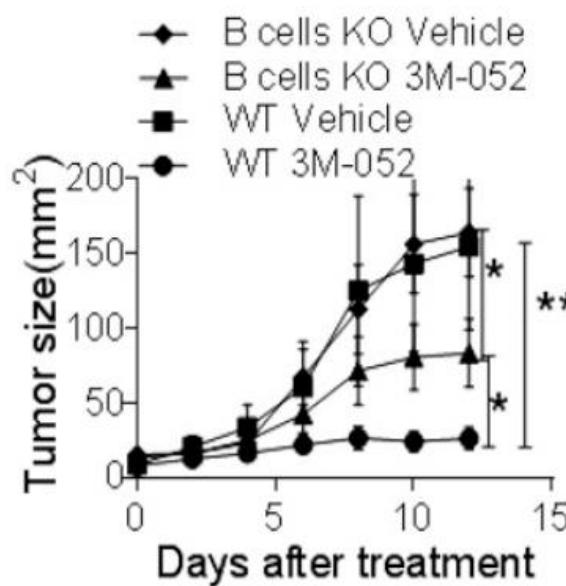
TLR7/8 Agonist Efficacy Depends on CD8⁺ T cells ...



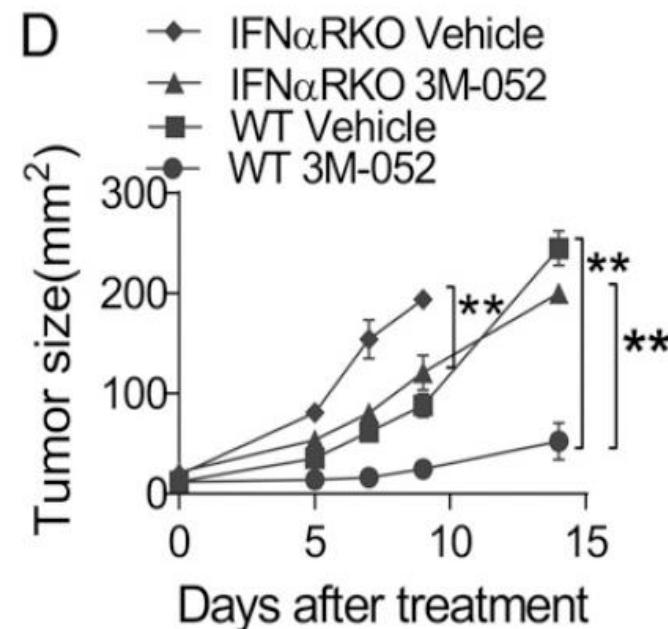
Singh et al., *J. Immunol.* 2014

... and Others!

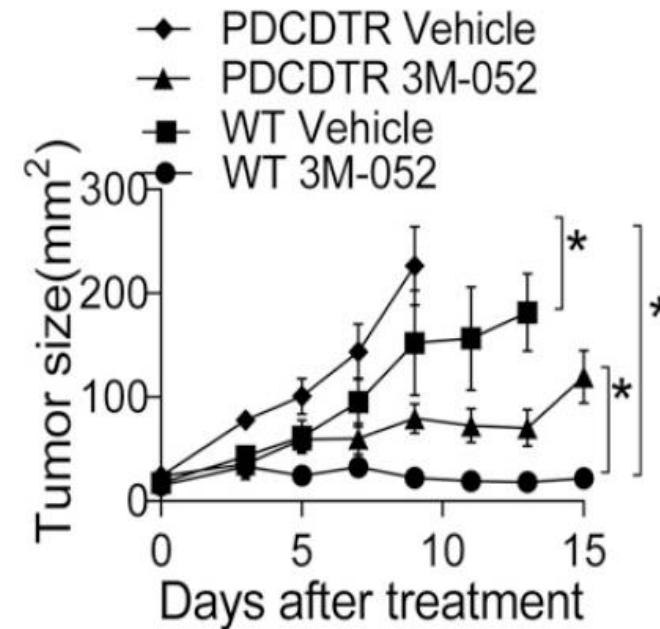
B cells



Type I IFN



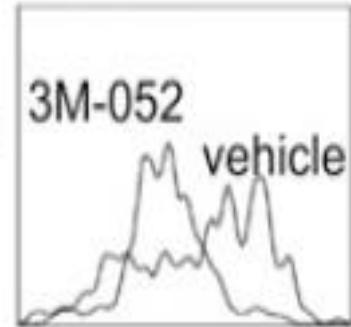
pDCs



But not CD4 $^+$ T cells and NK cells

Singh et al., *J. Immunol.* 2014

Shift from Intratumoral M2 → M1 Macrophages

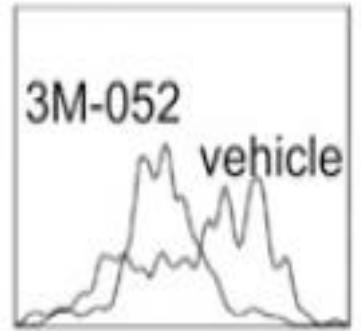


CD206 MFI
(Vehicle VS 3M-052)

1987 \pm 774
VS
659 \pm 156] P=0.04

Singh et al., *J. Immunol.* 2014

Shift from Intratumoral M2 → M1 Macrophages

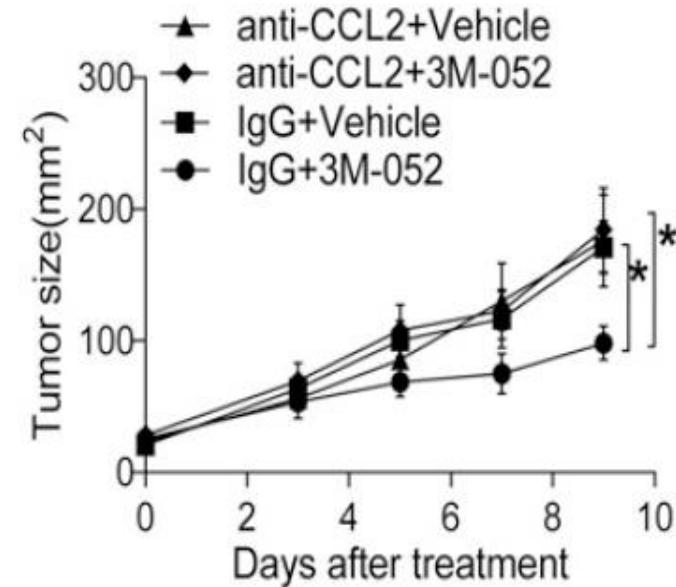


CD206 MFI
(Vehicle VS 3M-052)

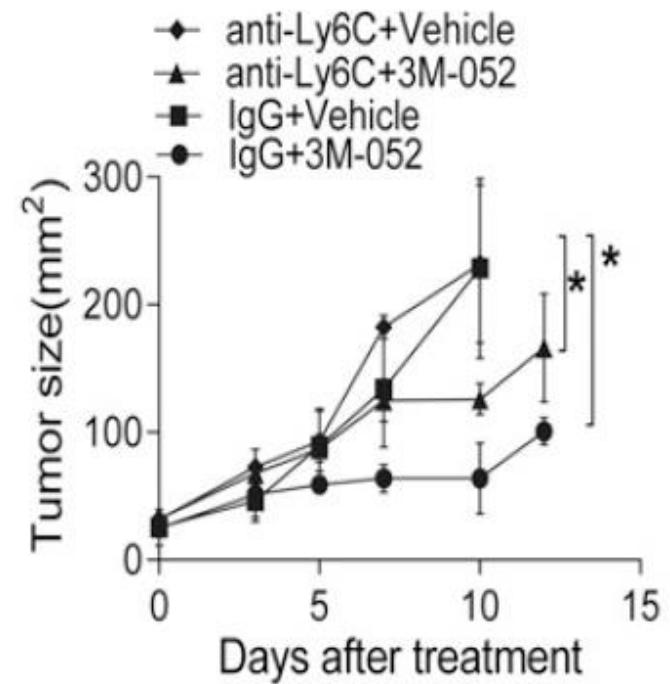
1987 ± 774
VS
 659 ± 156

$\left. \right\} P=0.04$

MAC reduction method 1

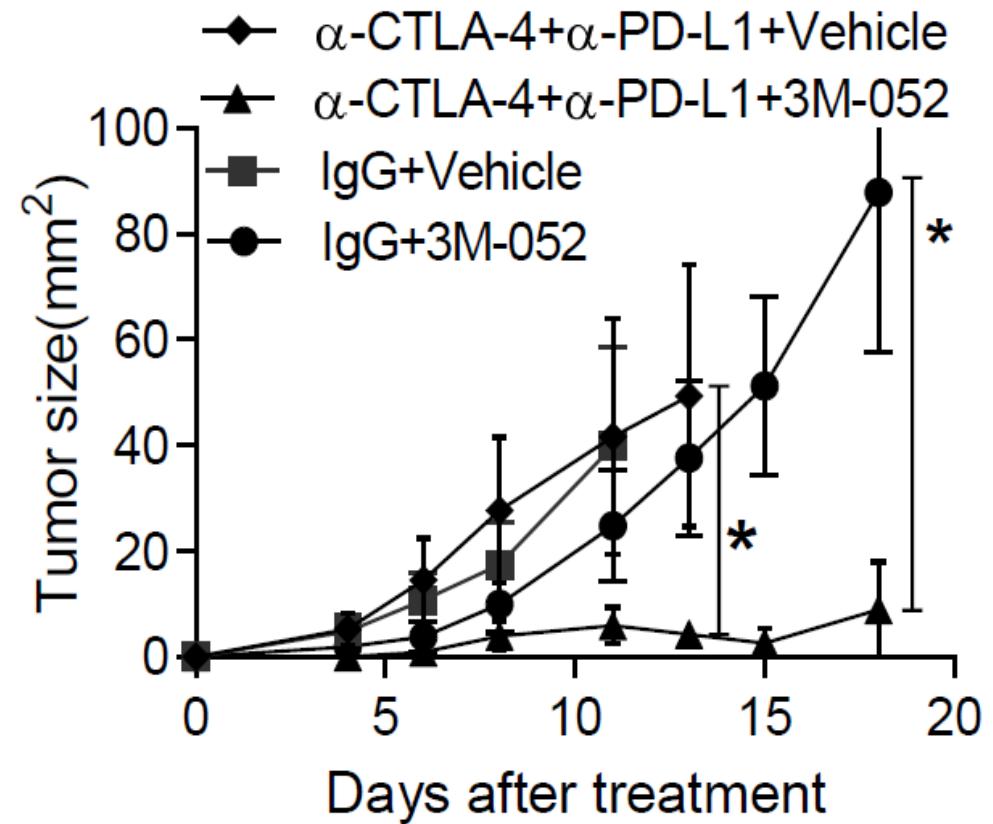
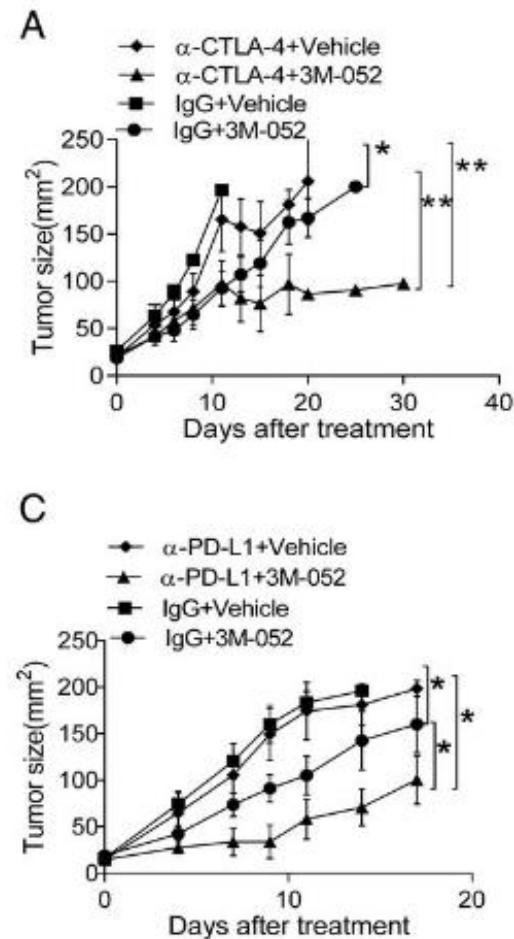


MAC reduction method 2



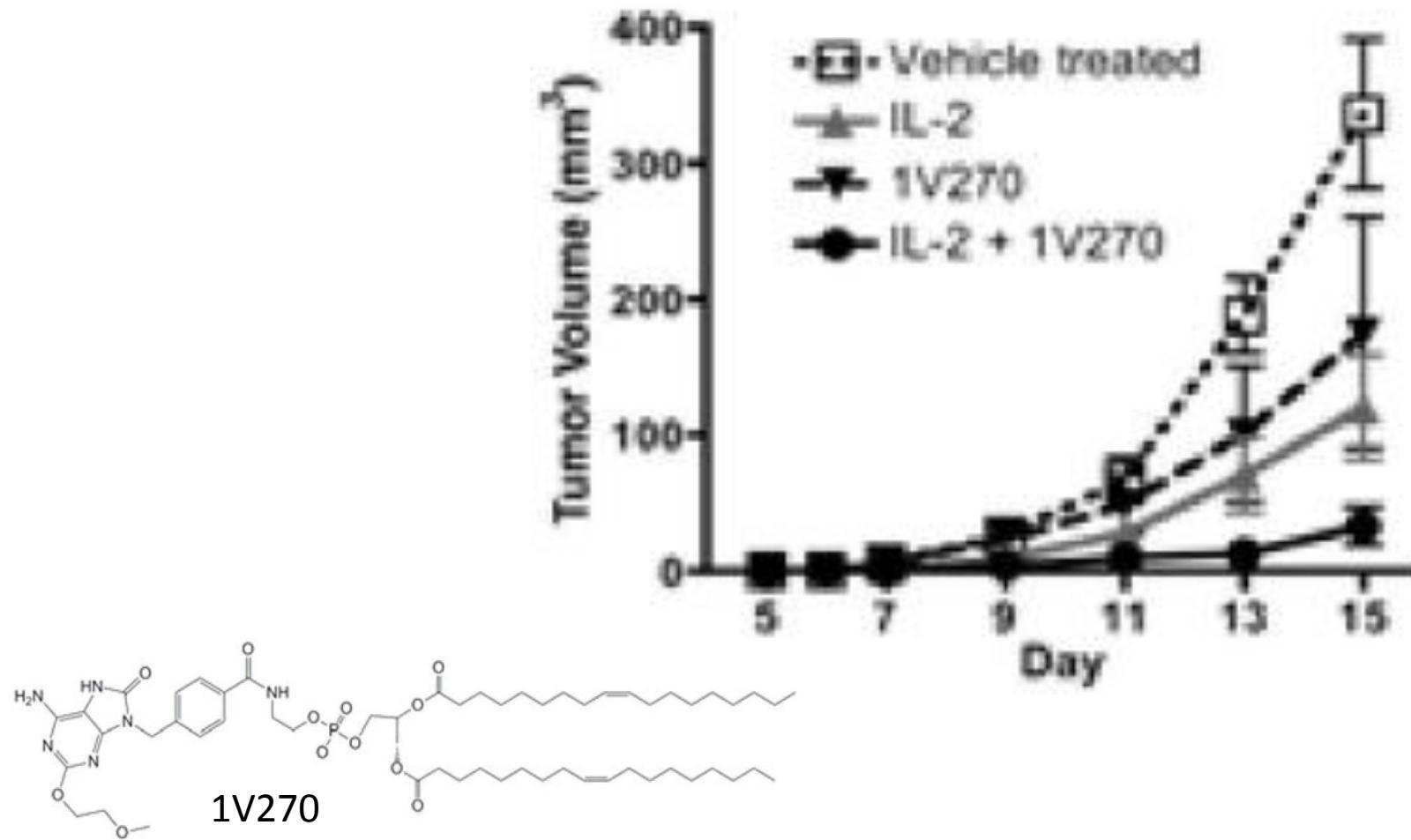
Singh et al., *J. Immunol.* 2014

Combination of CTLA-4 and PD-L1 Checkpoint Inhibitors with Intratumoral TLR7/8 Agonist



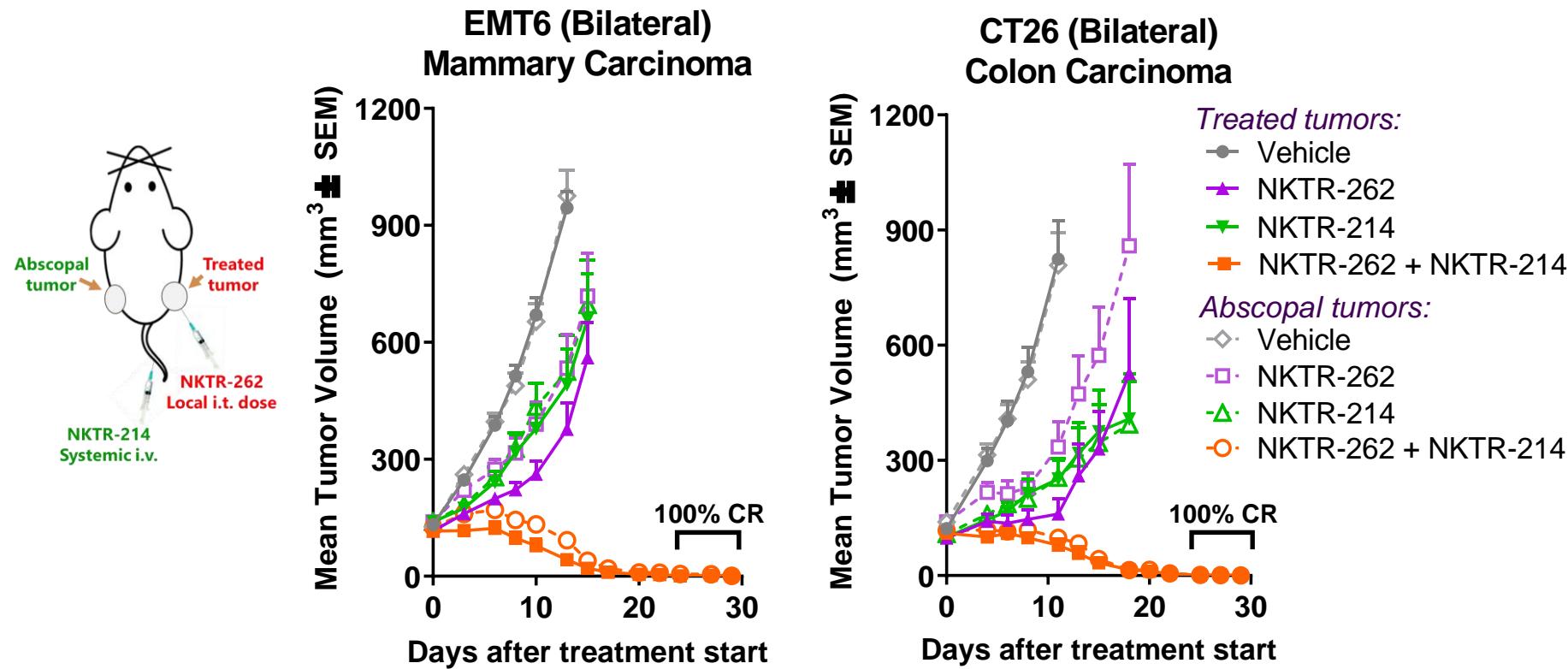
Singh et al., *J. Immunol.* 2014

Synergy Between IL-2 and Tumor-retained TLR7 Agonist



Hayashi et al., Melanoma Res. (2011)

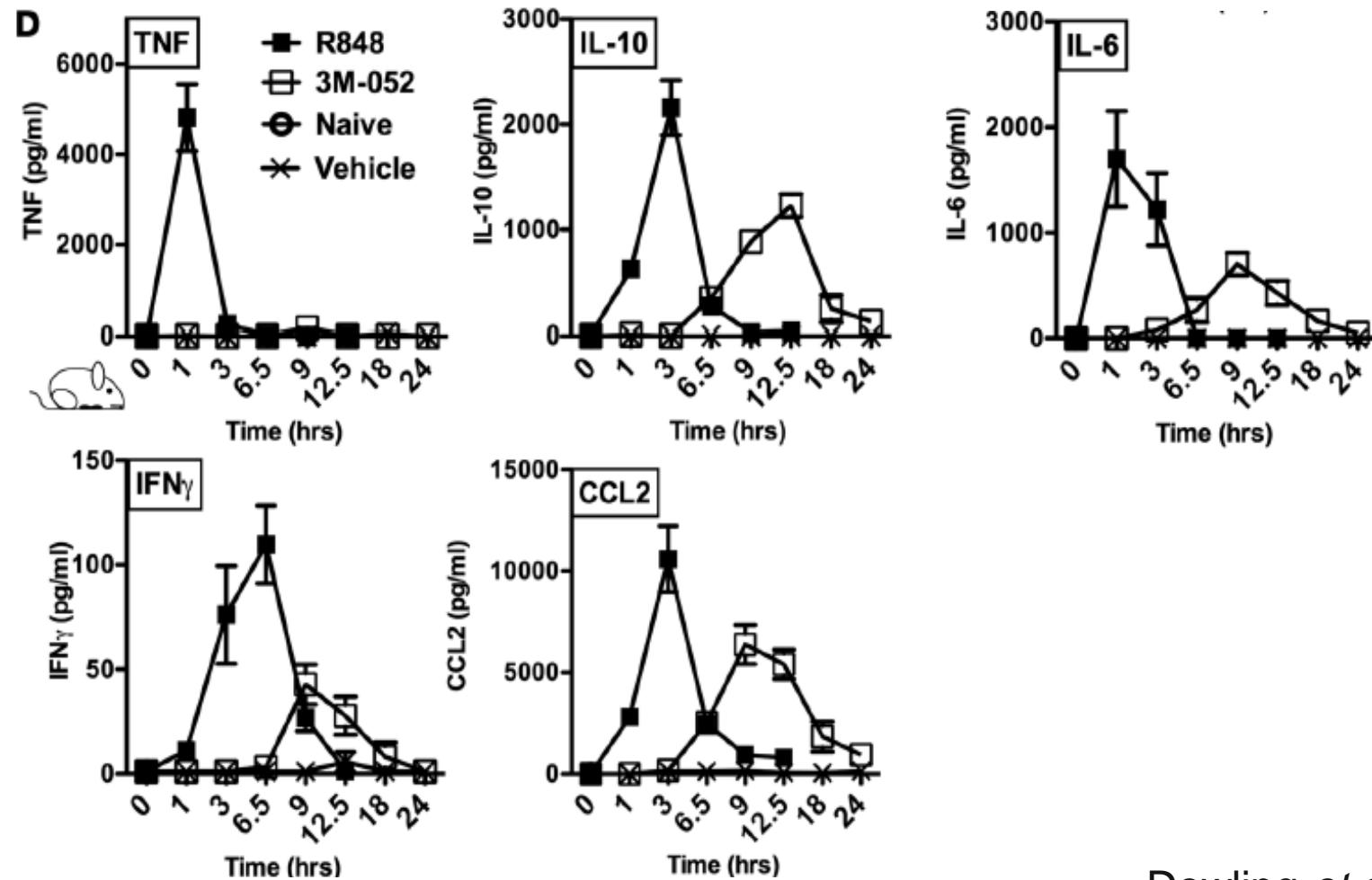
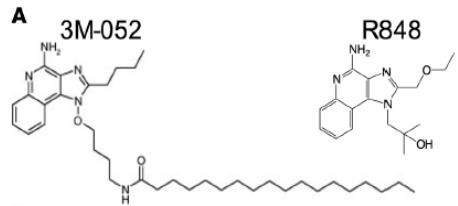
Combination Therapy with Tumor-Retained, PEGylated TLR7/8 Agonist and IL-2 Prodrug



Kivimae *et al.*, SITC 2017

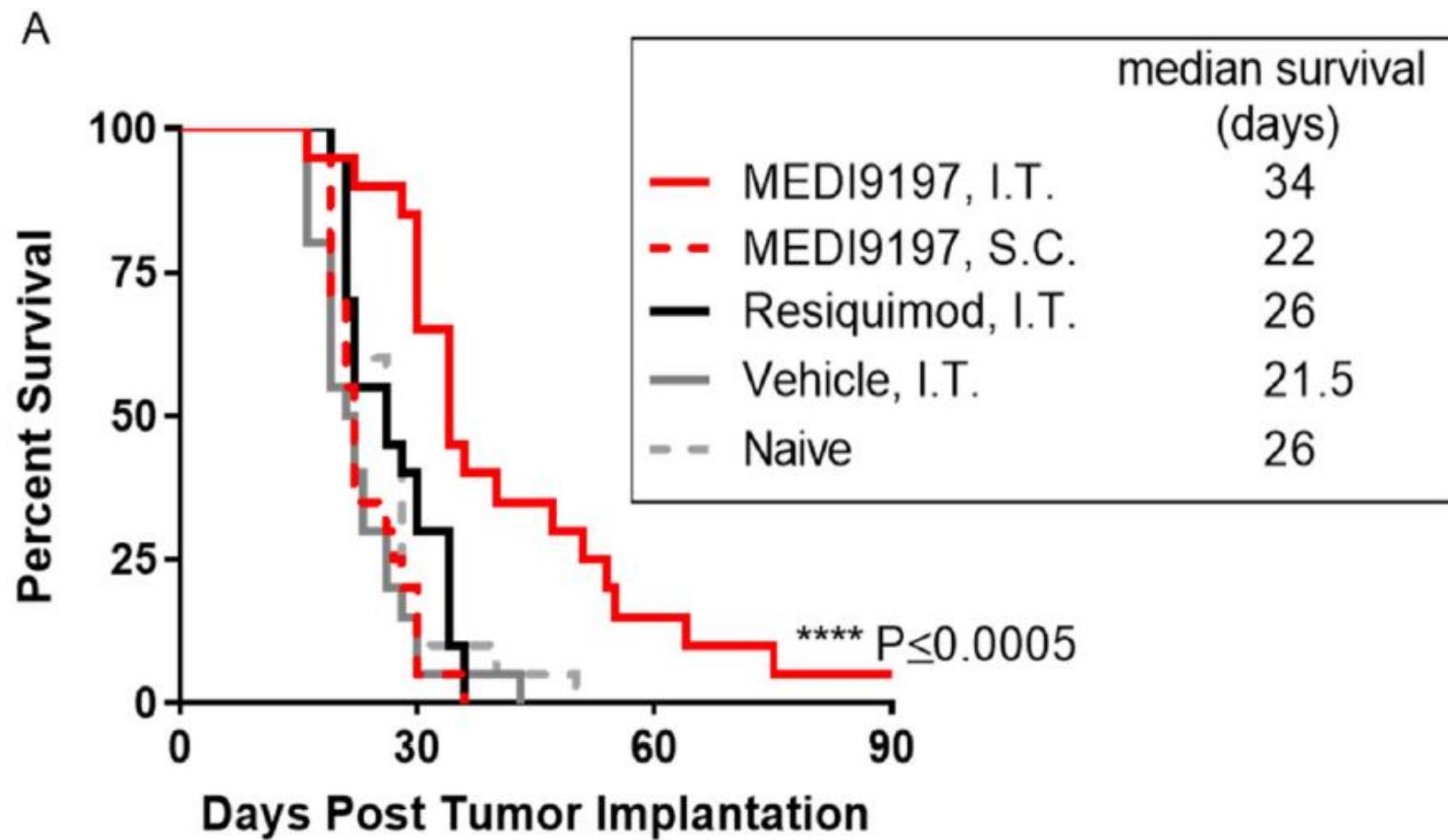
Why go Intratumoral and not Systemic?

Tissue-retained TLR7/8 Agonist: Increased Tolerability



Dowling *et al.* JCI Insight 2017

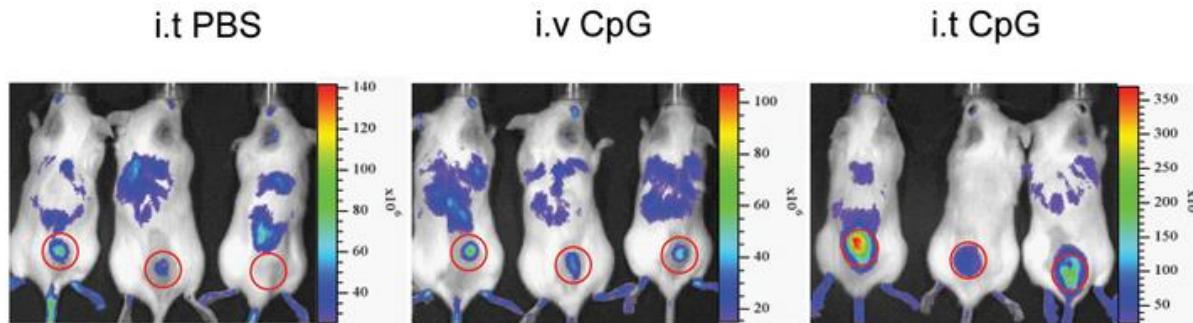
Intratumoral vs. Subcutaneous TLR7/8 Agonist: Route of Administration Is Critical



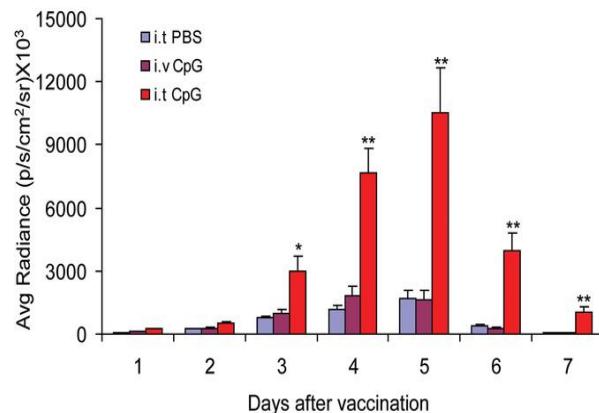
Mullins *et al.* JITC 2019

Intratumoral vs. Intravenous TLR9 Agonist: Route of Administration Is Critical

intratumoral Luciferase⁺ tumor-specific CD8+ T cells



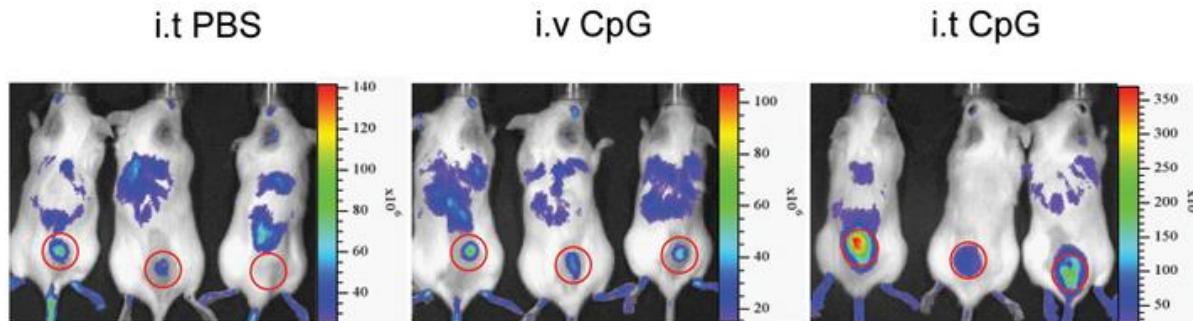
intratumoral T cells



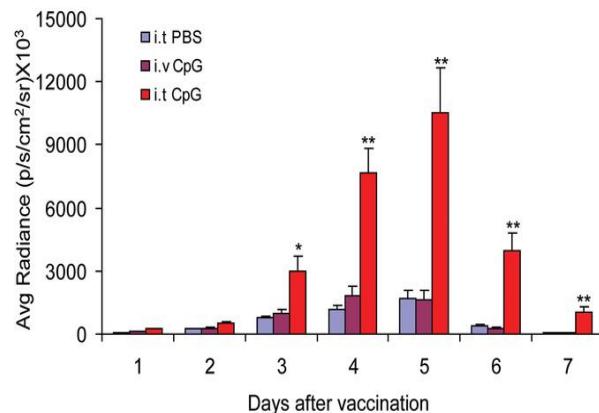
Lou et al., J Immunother. 2011

Intratumoral vs. Intravenous TLR9 Agonist: Route of Administration Is Critical

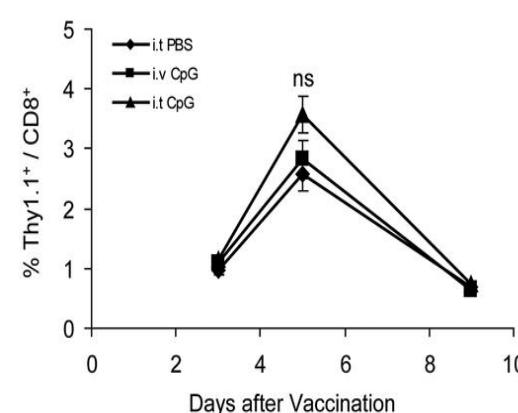
intratumoral Luciferase⁺ tumor-specific CD8+ T cells



intratumoral T cells



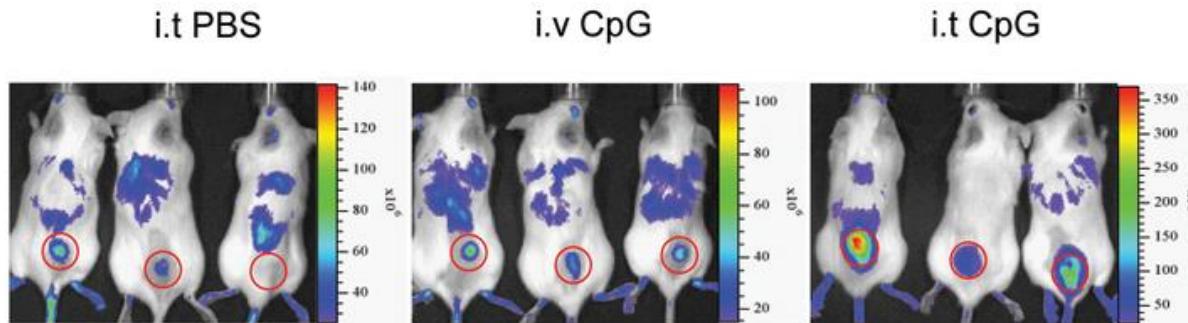
circulating T cells



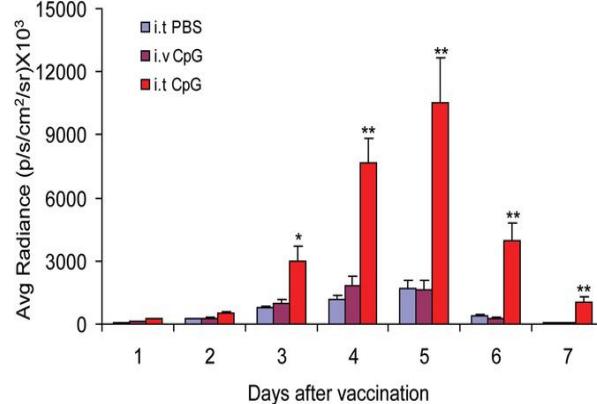
Lou et al., J Immunother. 2011

Intratumoral vs. Intravenous TLR9 Agonist: Route of Administration Is Critical

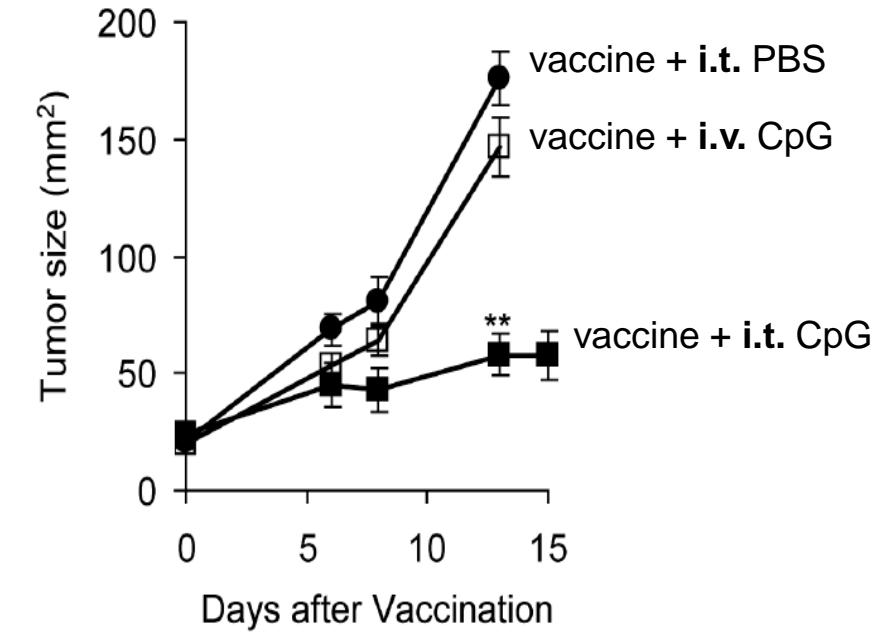
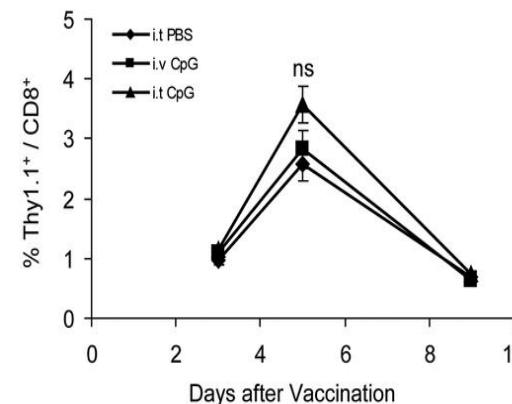
intratumoral Luciferase⁺ tumor-specific CD8+ T cells



intratumoral T cells

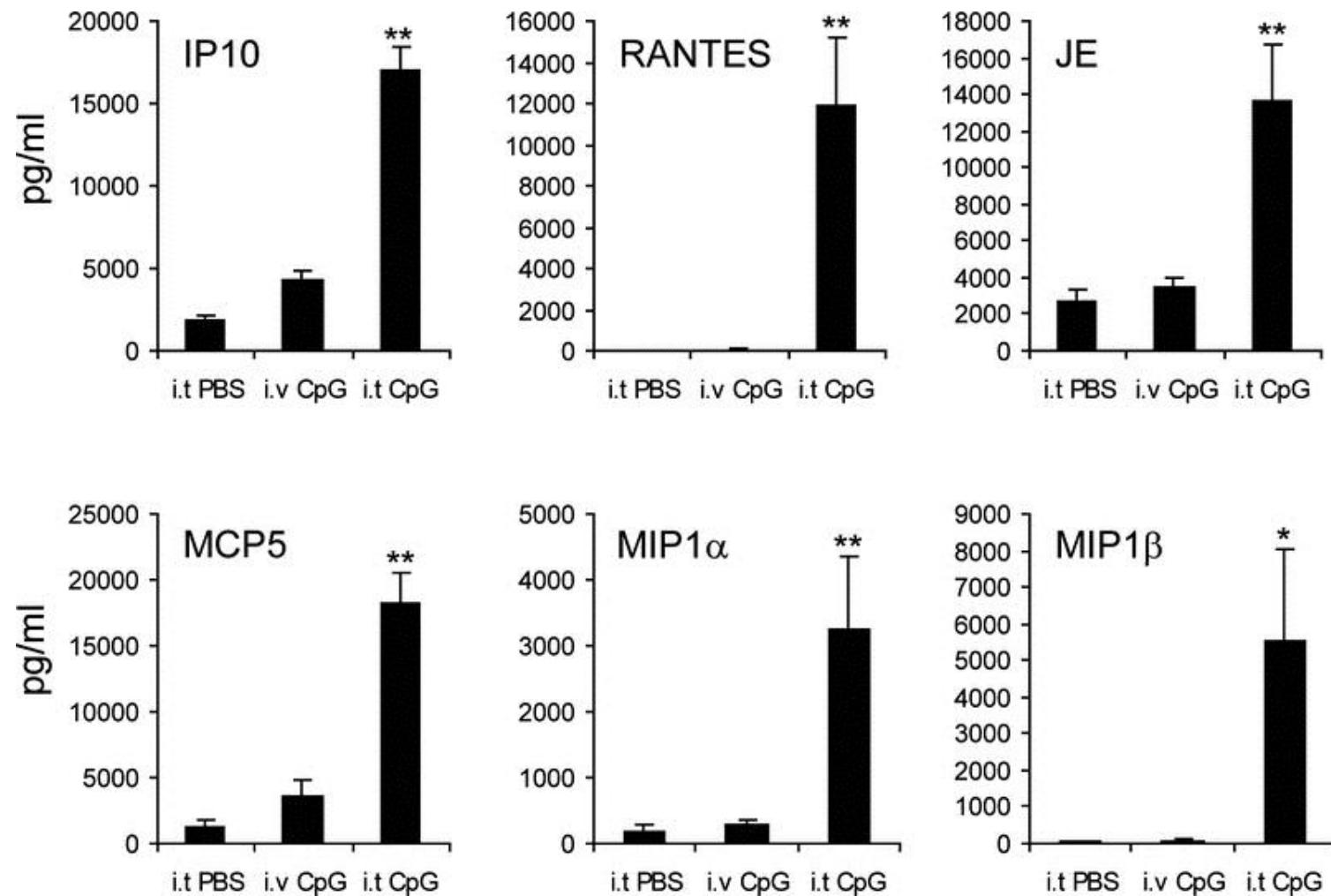


circulating T cells



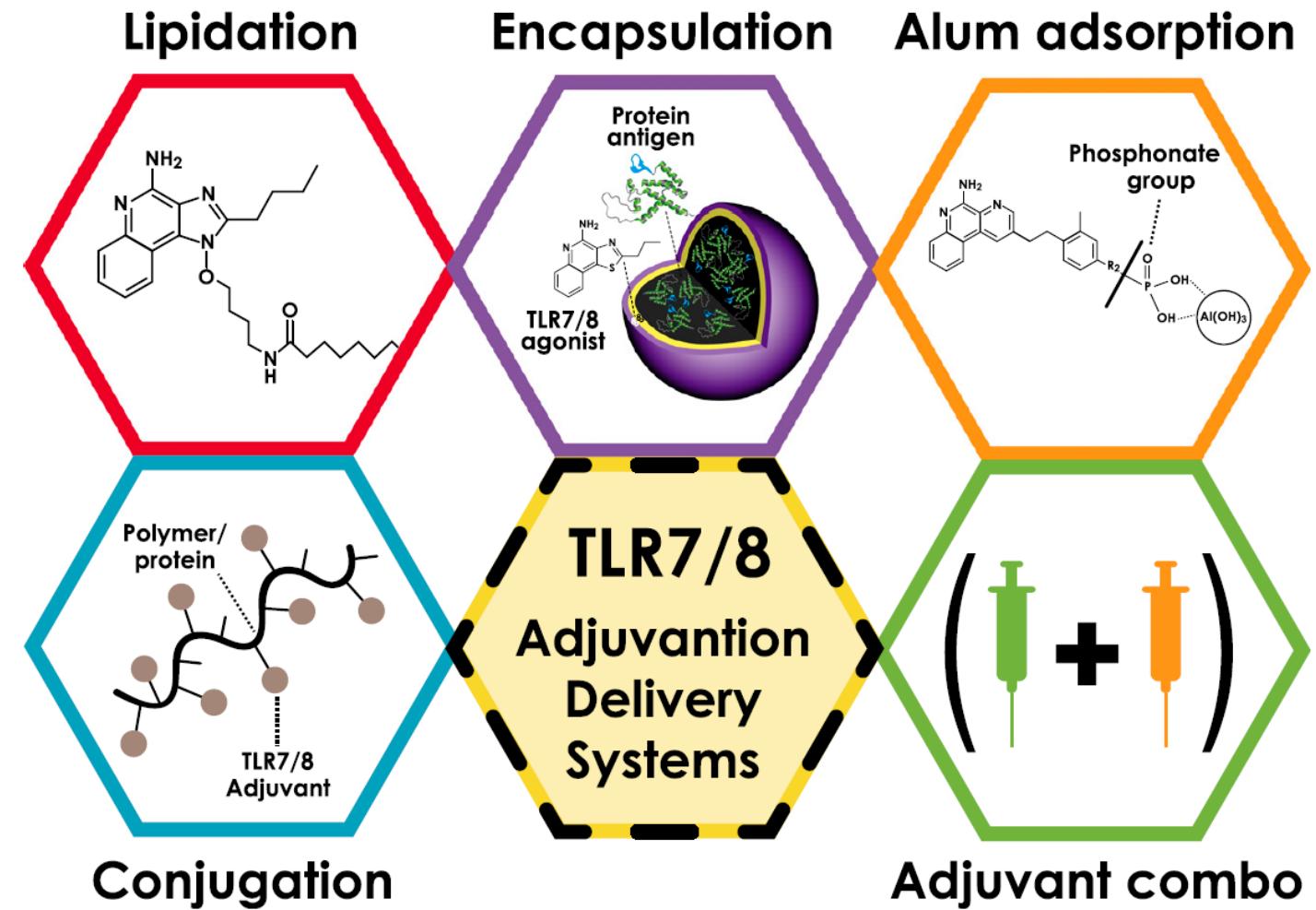
Lou et al., J Immunother. 2011

Intratumoral vs. Systemic TLR9 Agonist: Intratumoral Chemokines



Lou et al., J Immunother. 2011

Approaches to TLR 7/8 Agonist Delivery



Dowling et al., *Immunohorizons* 2018

Clinical Studies

Research

Open Access

Immune-mediated changes in actinic keratosis following topical treatment with imiquimod 5% cream

Abel Torres¹, Leslie Storey¹, Makala Anders¹, Richard L Miller², Barbara J Bulbulian², Jizhong Jin², Shalini Raghavan², James Lee³, Herbert B Slade³ and Woubalem Birmachu*²

Journal of Translational Medicine 2007,

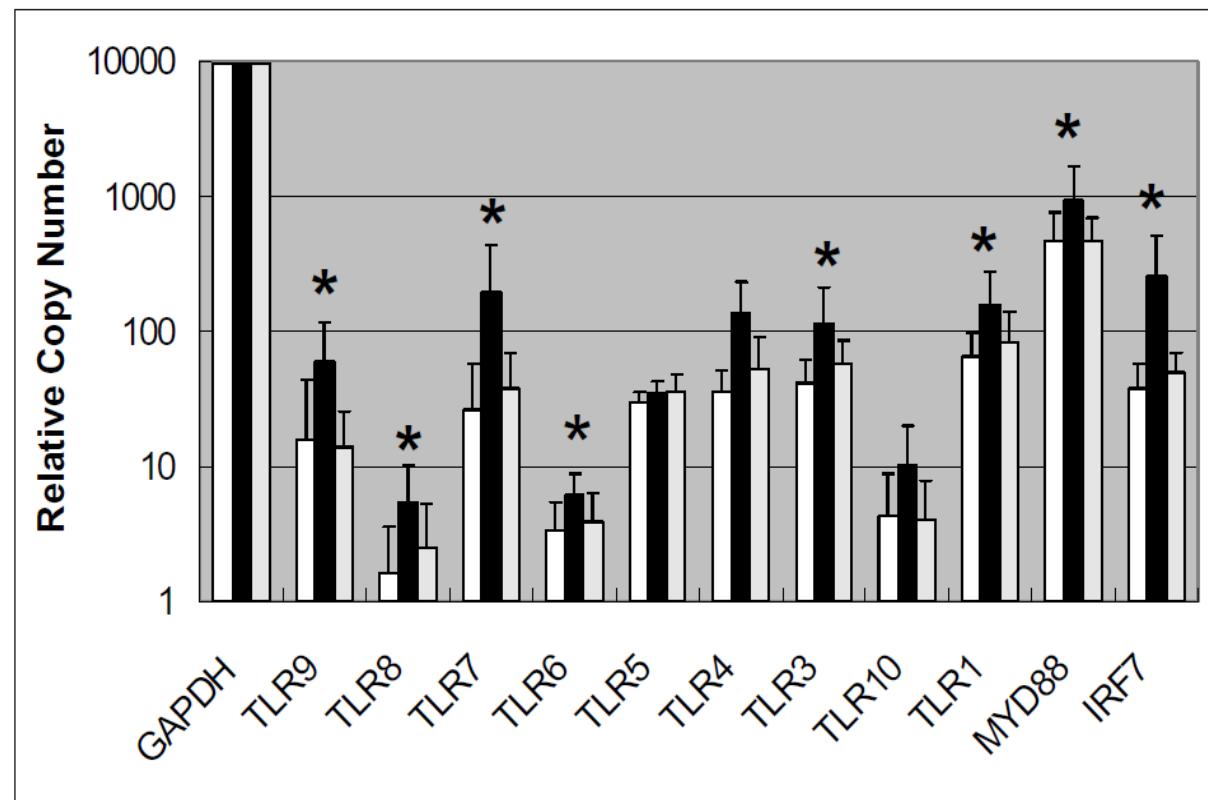


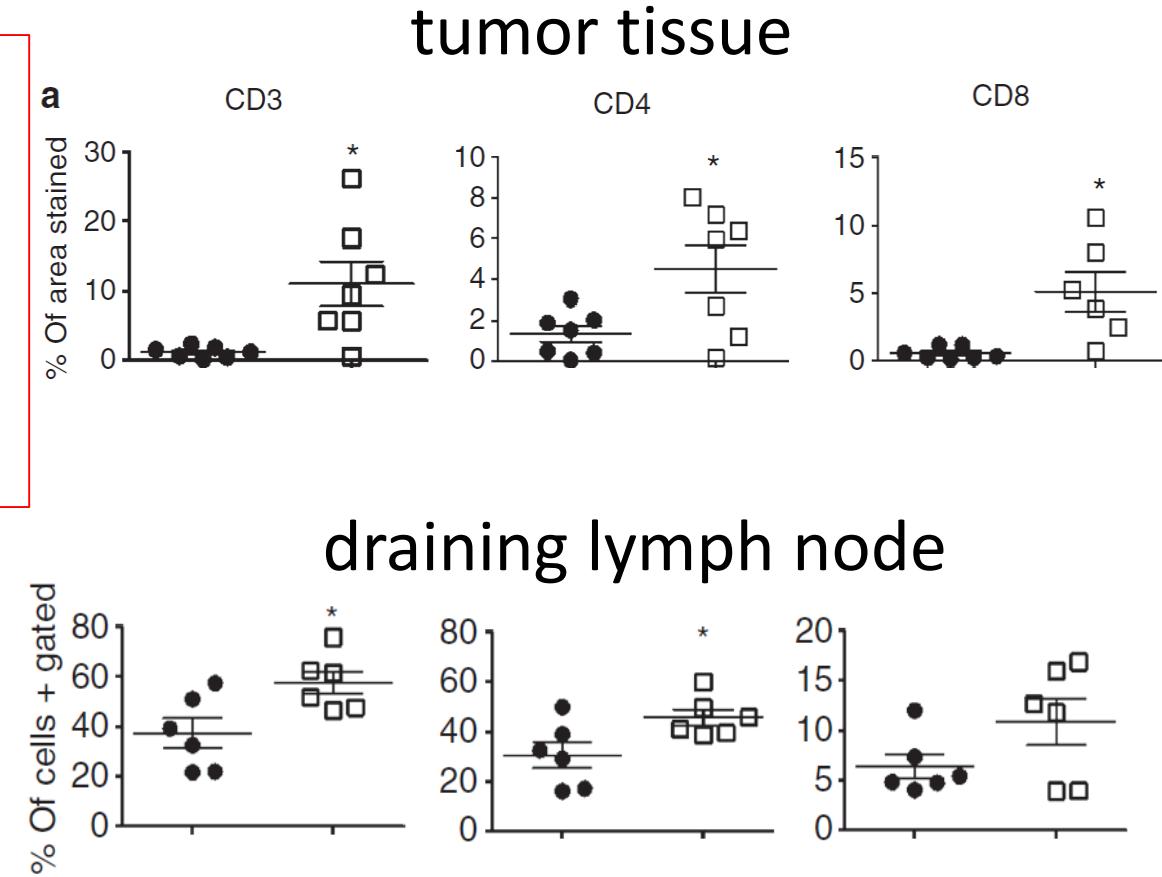
Figure 3

Basal TLR, IRF7, and MyD88 gene expression in skin biopsies as determined by real time RT-PCR. White bars represent pre-treatment AK, black bars represent during imiquimod treatment (maximum response value from week 1, week 2 and week 4 treatment times), and hatched bars represent 4-weeks post treatment. Relative copy number was determined as outlined in Methods and Materials section. Asterisks indicate those genes that had p-values < 0.05 in the ANOVA, comparing expression in pretreatment AK samples to the maximum response expression in samples from subjects (n = 13) during imiquimod treatment. [See Additional file 2].

Imiquimod-induced Regression of Superficial Melanoma



Figure 1. Melanoma site of “responder”, before and after treatment with imiquimod. Patient applied topical imiquimod to tumor site starting on day -14, and ending on day -1, before surgery on day 0.



Narayan et al., Journal of Investigative Dermatology (2012)

Topical TLR7 Agonist Imiquimod Can Induce Immune-Mediated Rejection of Skin Metastases in Patients with Breast Cancer

Sylvia Adams¹, Lina Kozhaya², Frank Martiniuk¹, Tze-Chiang Meng⁷, Luis Chiriboga³, Leonard Liebes¹, Tsivia Hochman⁴, Nicholas Shuman¹, Deborah Axelrod⁵, James Speyer¹, Yelena Novik¹, Amy Tiersten¹, Judith D. Goldberg⁴, Silvia C. Formenti⁶, Nina Bhardwaj³, Derya Unutmaz², and Sandra Demaria³

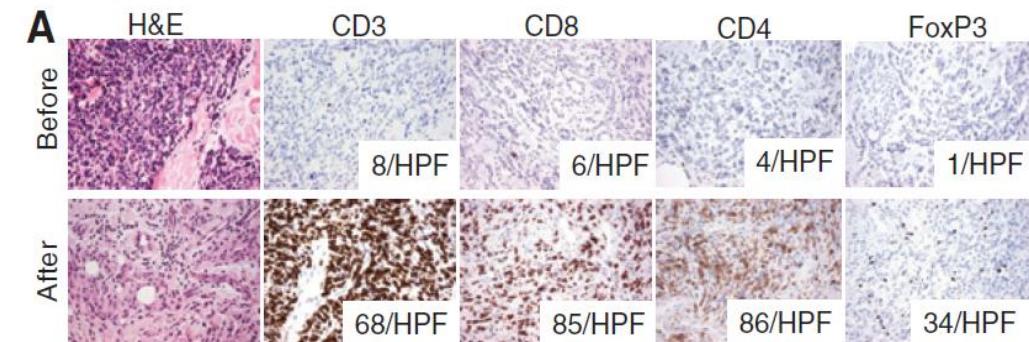
Table 3. Local antitumor response

Response	$ROI^{change} = (ROI^{posttreatment}/ROI^{pretreatment}) \times 100\%$	Patients (%)
CCR	Absence of any detectable residual disease	None
PR	>0% to <50%	2 (20%)
SD	≥50% to <100%	5 (50%)
NR	≥100% to <125%	1 (10%)
PD	≥125% or new skin lesions	2 (20%)

NOTE: Percentage change in ROI after 8-week imiquimod treatment ($n = 10$).

Abbreviations: ROI, region of interest; CCR, complete clinical response; PR, partial response; SD, stable disease; NR, no response; PD, progressive disease.

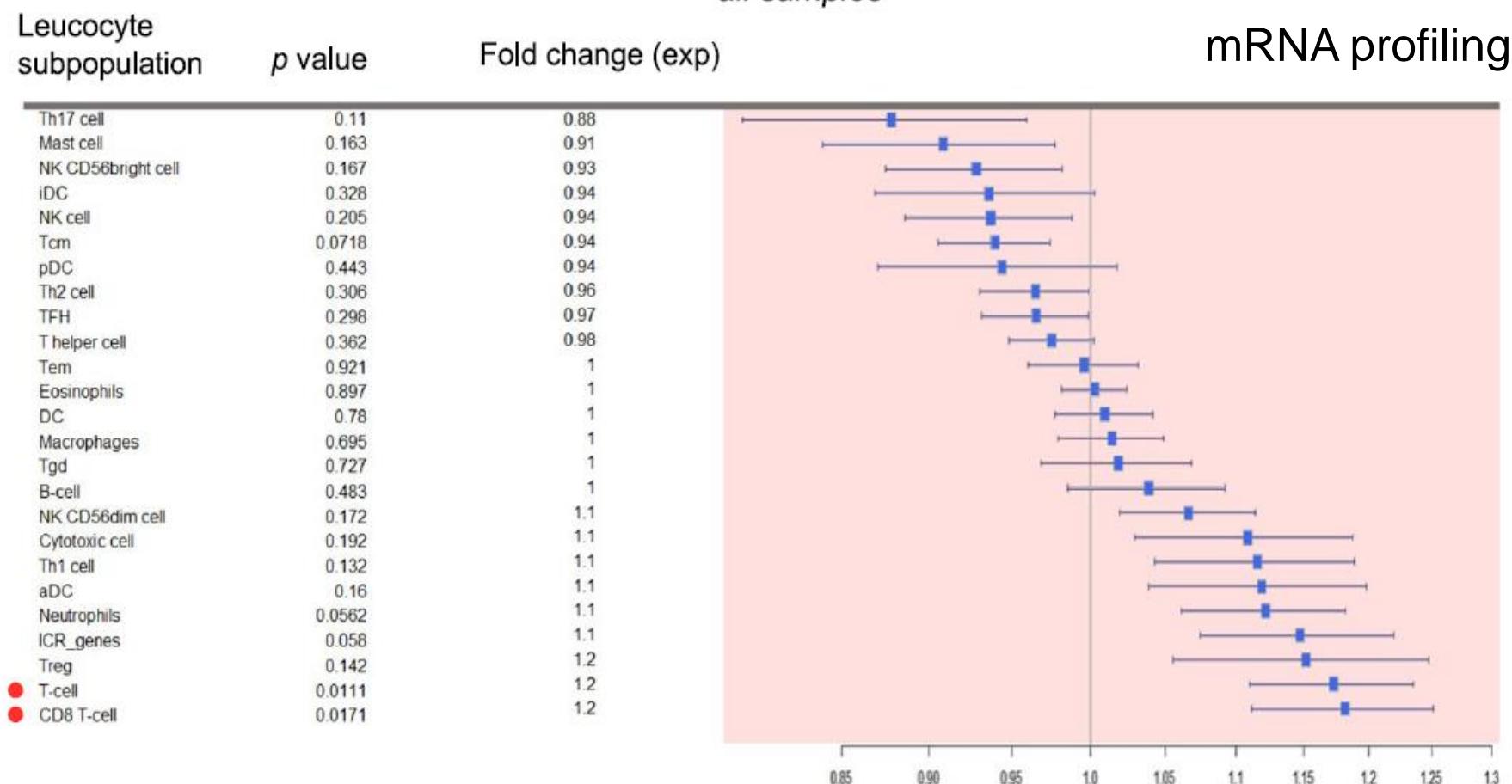
responder biopsy after 8 weeks of imiquimod



Imiquimod-induced Regression of Breast Cancer Skin Metastases

A.

Post- vs Pre-treatment
all samples



Rozenblit et al., Scientific Reports, 2019

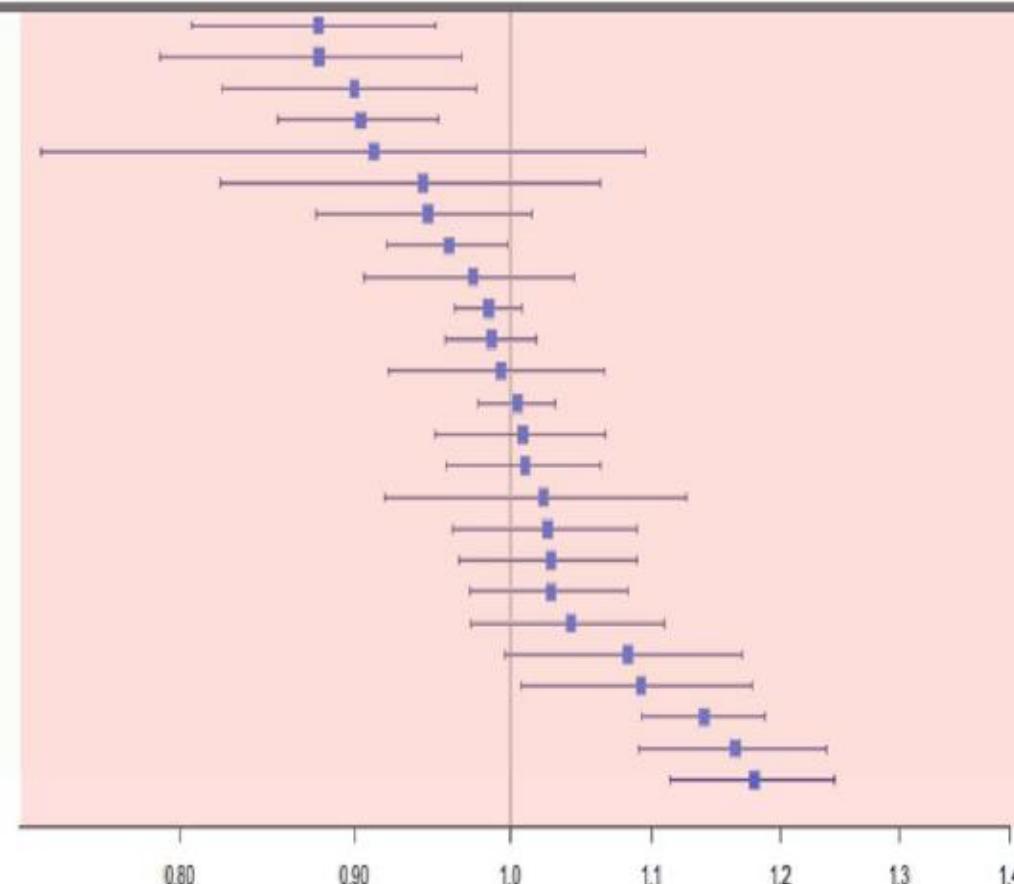
Imiquimod-induced Regression of Breast Cancer Skin Metastases

B.

Complete Responder vs Non Responder
pre-treatment samples

mRNA profiling

aDC	0.0719	0.88
Mast cell	0.147	0.88
TFH	0.171	0.9
Tcm	0.0376	0.9
pDC	0.626	0.91
Tgd	0.634	0.94
DC	0.428	0.95
T helper cell	0.293	0.96
Th1 cell	0.725	0.97
Tem	0.519	0.99
Th2 cell	0.684	0.99
Neutrophils	0.933	0.99
Eosinophils	0.875	1
NK CD56dim cell	0.893	1
B-cell	0.861	1
Treg	0.842	1
iDC	0.71	1
NK cell	0.673	1
ICR_genes	0.633	1
Cytotoxic cell	0.561	1
CD8 T-cell	0.365	1.1
Th17 cell	0.307	1.1
Macrophages	0.00573	1.1
T-cell	0.0393	1.2
NK CD56bright cell	0.0119	1.2



Rozenblit et al., Scientific Reports, 2019

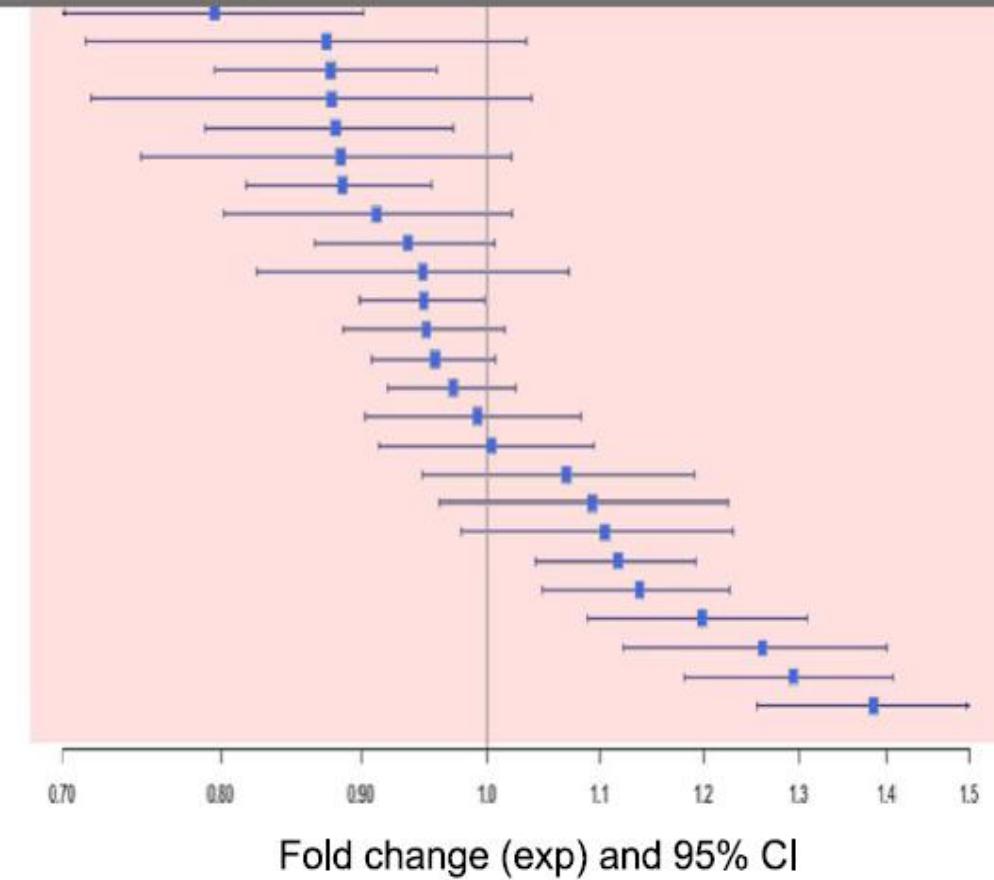
Imiquimod-induced Regression of Superficial Melanoma

C.

Complete Responder vs Non Responder
post-treatment samples

mRNA profiling

● IDC	0.0304	0.8
Treg	0.406	0.87
DC	0.109	0.88
Th17 cell	0.425	0.88
Tem	0.168	0.88
Tgd	0.375	0.88
Mast cell	0.0785	0.89
NK cell	0.408	0.91
Th2 cell	0.355	0.94
B-cell	0.678	0.95
Macrophages	0.293	0.95
T helper cell	0.439	0.95
Tcm	0.391	0.96
Eosinophils	0.6	0.97
NK CD56bright cell	0.937	0.99
TFH	0.969	1
Neutrophils	0.595	1.1
pDC	0.51	1.1
CD8 T-cell	0.437	1.1
T-cell	0.143	1.1
NK CD56dim cell	0.149	1.1
Th1 cell	0.101	1.2
aDC	0.096	1.3
● ICR_genes	0.0228	1.3
● Cytotoxic cell	0.0119	1.4



Rozenblit *et al.*, Scientific Reports, 2019

Some TLR7 & 8 Agonists In (Pre-)Clinical Development

Marketed	Imiquimod/Aldara (Medicis Pharmaceuticals) TLR7 Agonist Actinic Keratosis, Genital Warts, Basal Cell Carcinoma of Skin							
Registration	Imiquimod/Aldara (Medicis Pharmaceuticals) TLR7 Agonist Anal Cancer, Cervical Cancer, Vulvar Intraepithelial Neoplasia							
Phase II	Imiquimod/Aldara (Medicis Pharmaceuticals) TLR7 Agonist Liquid and Solid Tumors	Resiquimod (Spirig Pharma) TLR7/8 Agonist Anaplastic Astrocytoma, Anaplastic Astro-oligodendrogloma, Glioblastoma, Melanoma	DPV 001 (UbiVac) TLR 2,3,4,7,9 Agonist Non Small Cell Lung Cancer	VTX-2337/Motolimod (Celgene, VentiRx) TLR8 Agonist Cancer	CV 9202 (CureVac) TLR7/8 Modulator Non Small Cell Lung Cancer	Vesimune (UroGen Pharma) TLR7 Agonist Bladder Cancer		
Phase I	Imiquimod/Aldara (Medicis Pharmaceuticals) TLR7 Agonist Liquid and Solid Tumors	CV 8102 (CureVac) TLR7/8 Agonist Adenoid-Cystic-Carcinoma, Head and Neck Cancer, Melanoma, Squamous Cell Carcinoma of Skin	DPV 001 (UbiVac) TLR 2,3,4,7,9 Agonist Prostate Cancer	DSP 0509 (Boston Biomedical; Sumitomo Dainippon Pharma) TLR7 Agonist Solid Tumors	Telratolimod/MEDI9197/3M-052 (3M Drug Delivery Systems/MedImmune) TLR7/8 agonist Cutaneous T cell lymphoma, Solid Tumors	BDB001 (Birdie Biopharmaceuticals) TLR Agonist Solid Tumors	NKTR 262 (Nektar Therapeutics) TLR7/8 Agonist Bladder Cancer, CRC, RCC Melanoma, Merkel Cell Carcinoma, Ovarian Cancer, Soft Tissue Sarcoma, TNBC	LHC165/NJH395 (Novartis) TLR7 agonist Solid Tumors
Preclinical	1V270 (University of California, San Diego, La Jolla, USA; Graduate School of Tokyo Medical and Dental University, Japan) TLR7 Agonist Lung metastases, Breast cancer, melanoma, Pulmonary metastatic cancers	DV 1001 (Dynavax Technologies) TLR7/8 Agonist Multiple malignancies	3M-011 (Technische Universität Dresden, Germany) TLR7/8 Agonist Pancreatic cancer, Colon cancer	DSR-6434 (University of Manchester, Manchester Academic Health Sciences Centre, United Kingdom) TLR7 Agonist Colon cancer, Renal cell carcinoma	DSR-29133 (Sumitomo Dainippon Pharma, Osaka, Japan; AstraZeneca Pharmaceuticals Ltd.) TLR7 Agonist Colon cancer, Renal cell carcinoma, Osteosarcoma	SC1 (BioNTech AG , 4SC Discovery GmbH) TLR7 Agonist Lymphoma, Renal cell carcinoma, Cancer	SZU-101 (Shenzhen University, Shenzhen, China) TLR7 Agonist Breast carcinoma, Gastric cancer, T cell lymphoma	SM-276001 (Dainippon Sumitomo Pharmaceuticals) TLR7 Agonist Tumor

Conclusions

- Intratumoral TLR7/8 agonists stimulate APCs to activate tumor-specific T cells
- Other immune cells are also activated/attracted, and contribute to tumor killing
- Selective intratumoral localization of TLR agonist is important
- Intratumoral TLR7/8 agonists combine well with other immunological and non-immunological therapeutics, including checkpoint blockade
- Multiple agonists of TLR7 and/or TLR8 are under clinical development

Thank You