



Is there a role for radiation therapy and immunotherapy?

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The following relationships exist related to this presentation:

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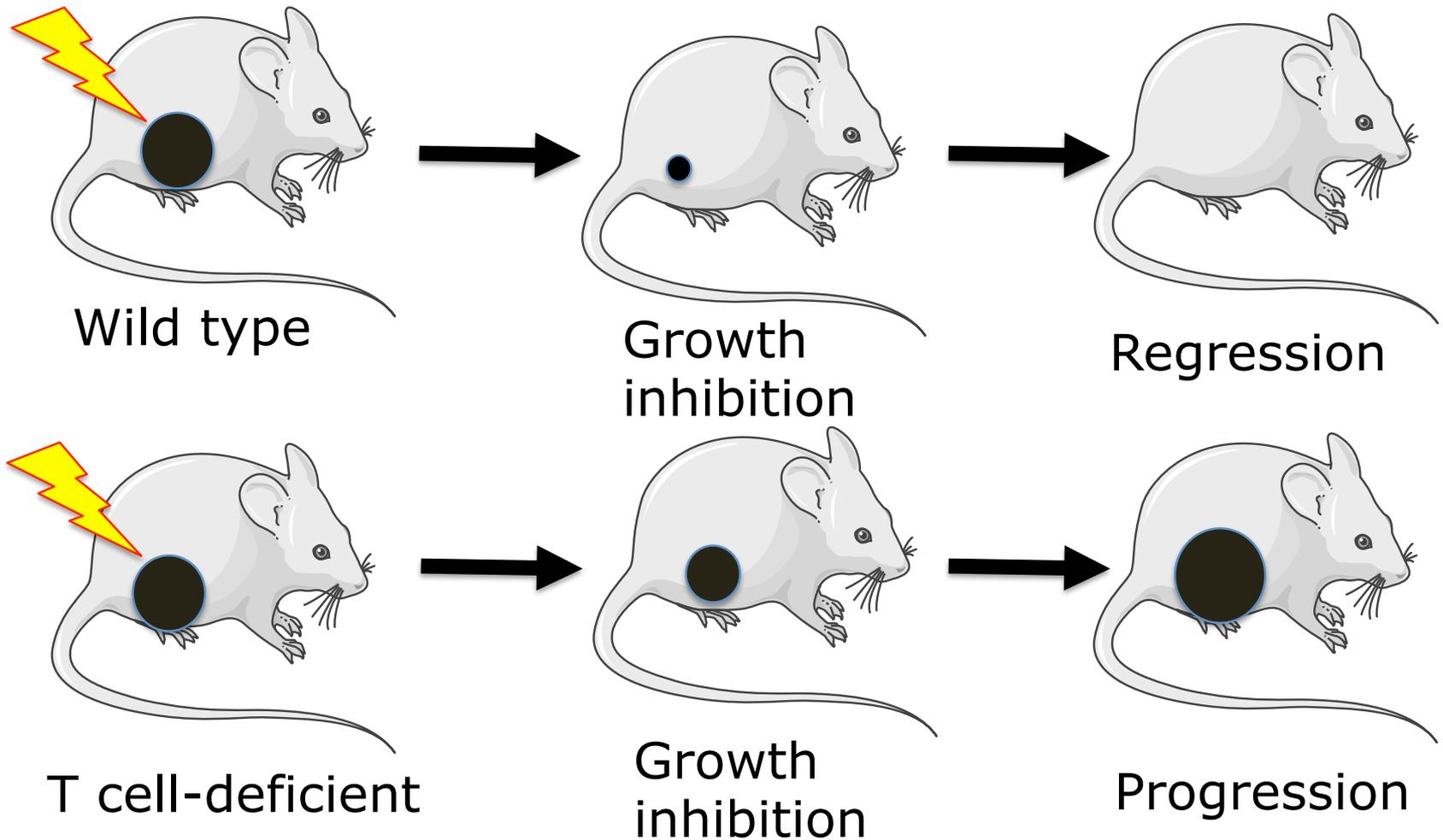
Outline

- Background: Evidence for a role of the immune system in response to ionizing radiation
- In situ vaccination by radiation-induced tumor cell death
- Overcoming T cell exclusion: Effects of radiation on the tumor microenvironment
- Exploiting radiation effects to improve response to immunotherapy

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Immunocompetence of the host affects the response to local radiotherapy



T cells contribute to the response to radiotherapy

<u>FSA Tumor</u>	<u>TCD₅₀ values</u>	<u>Metastases</u>
Normal mice	30.0Gy (28.5-32.4)	1%
Immunosuppressed (6Gy)	50.8Gy (47.6-54.3)	4%
T cell deprived mice	64.5Gy (62.0-67.1)	79%

Evidence for induction of tumor-specific immune responses by radiation

IN MICE

Lugade et al., J Immunol 2005

B16-OVA model, induction of CD4 and CD8 T cells after irradiation with 15 Gy x 1 or 3 Gy x 5,

Lee et al, Blood 2009

B16-SIY model, induction of CD8 T cells after irradiation with 20 Gy x 1

Schaue et al., IJROBP 2012

B16-OVA, best induction of CD8 T cells with 7.5 Gy x 2

Evidence for induction of tumor-specific immune responses by radiation

& HUMAN STUDIES

Nesslering et al., Clin Cancer Res 2007

Tumor-specific antibodies in 14% of prostate cancer patients treated with EBRT and in 25% receiving brachytherapy

Schaue et al, Clin Cancer Res 2008

T cell responses to survivin in prostate and colorectal patients after radiotherapy

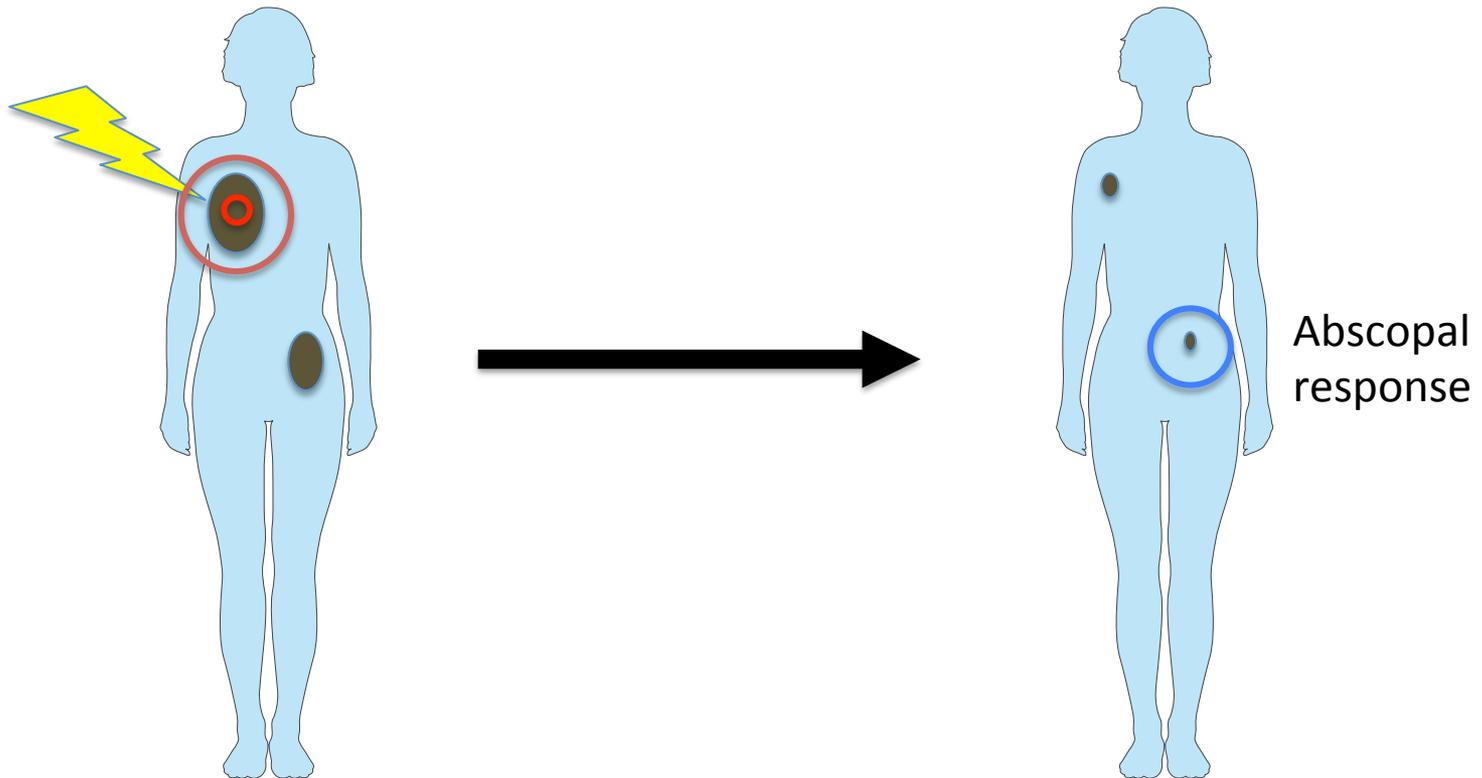
Teulings et al., Br J Dermatol 2013

T and B cell responses to MART-1 and gp100 in melanoma patient with complete durable regression after WBRT

Abscopal effect

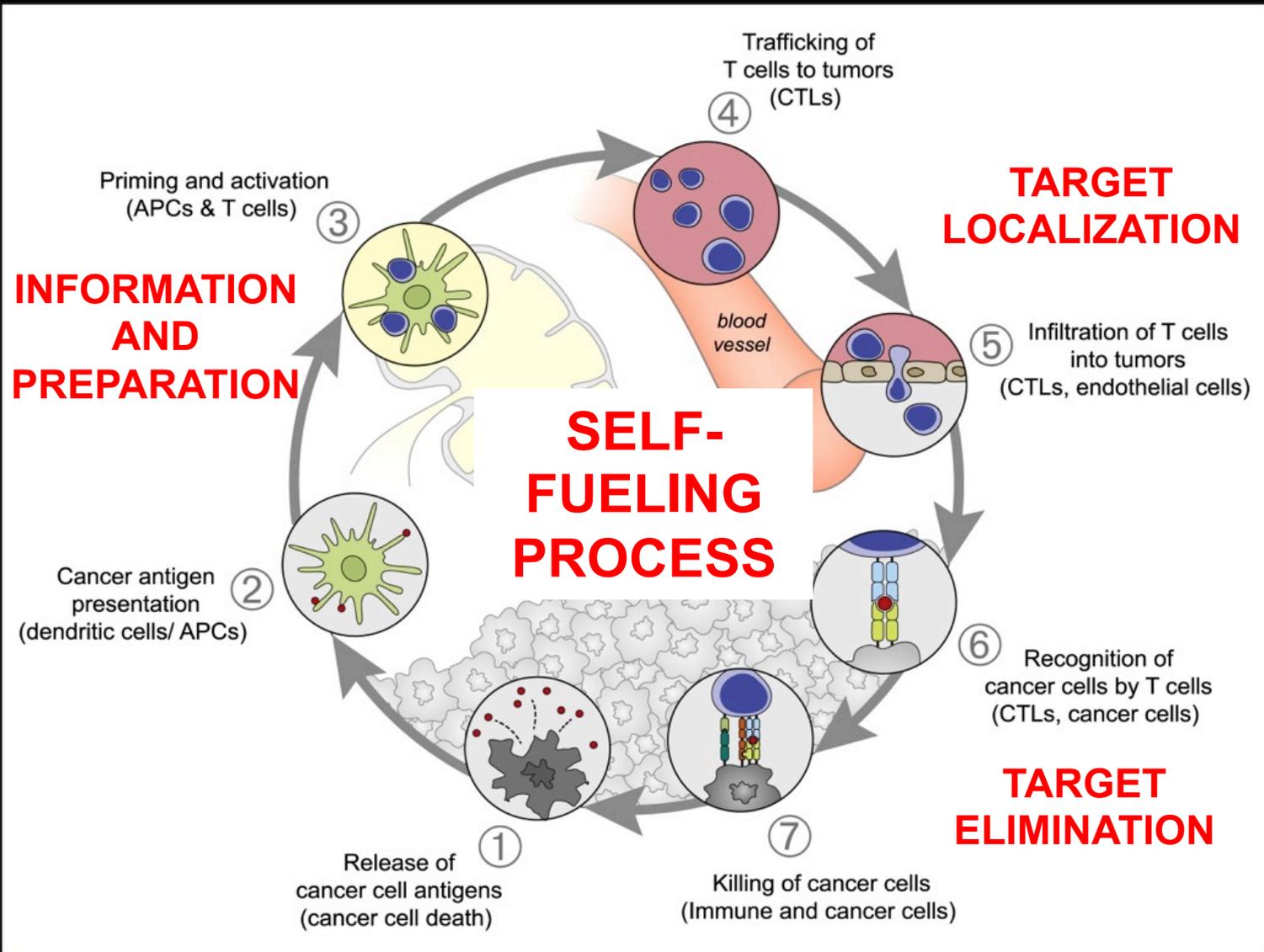
Effect of ionizing radiation on cancer
outside the radiation field

Latin *ab* (position away from) and *scopus* (mark or target)



The Cancer-Immunity Cycle

(Chen and Mellman, *Immunity* 2013)



Outline

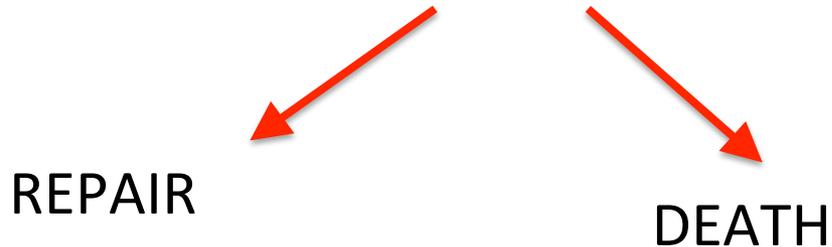
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Radiation-induced tumor cell death

Radiation Damage → DNA, proteins, lipids

DIRECT energy deposition

INDIRECT free radicals (H₂O radiolysis)

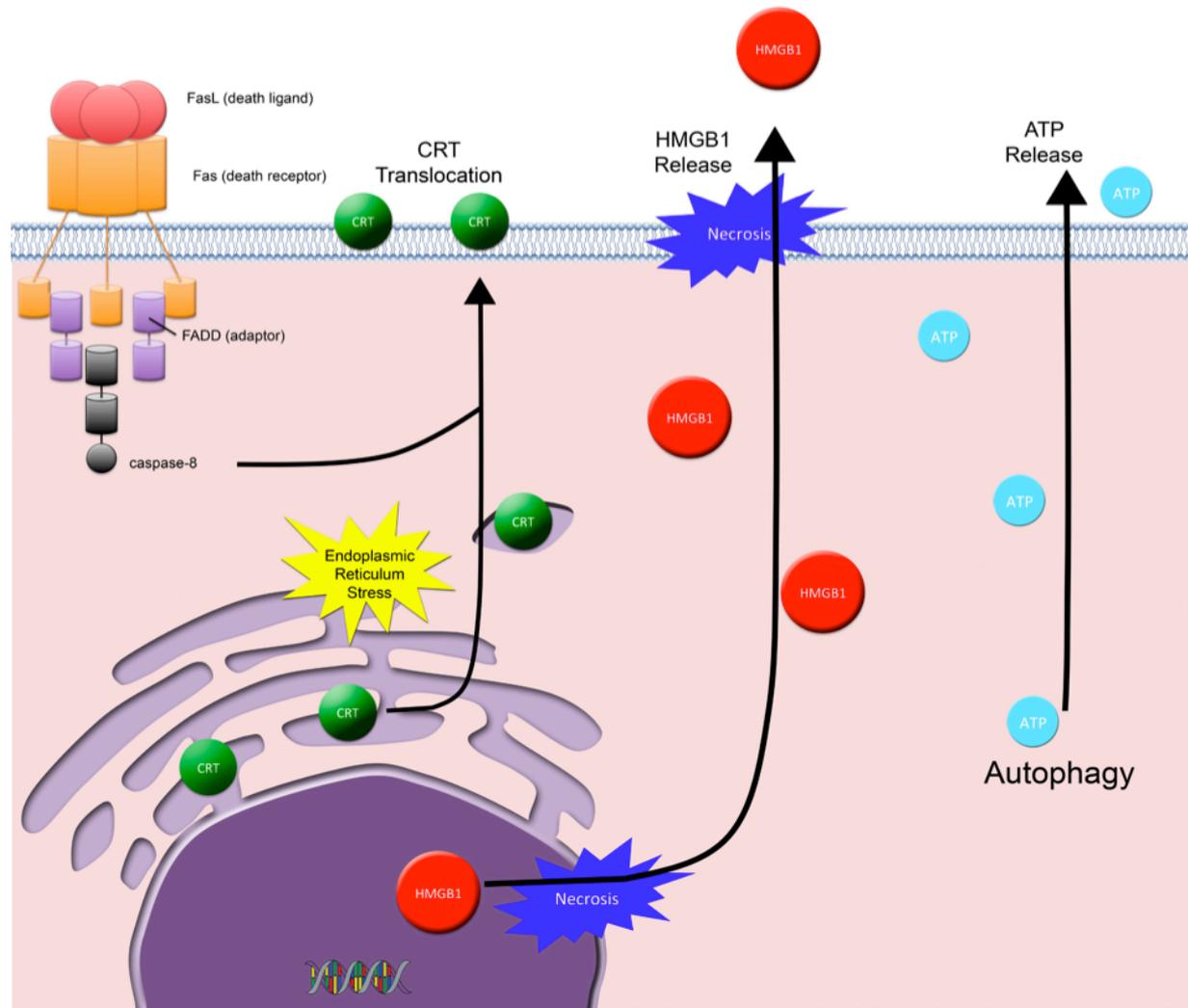


Cell death type depends on:

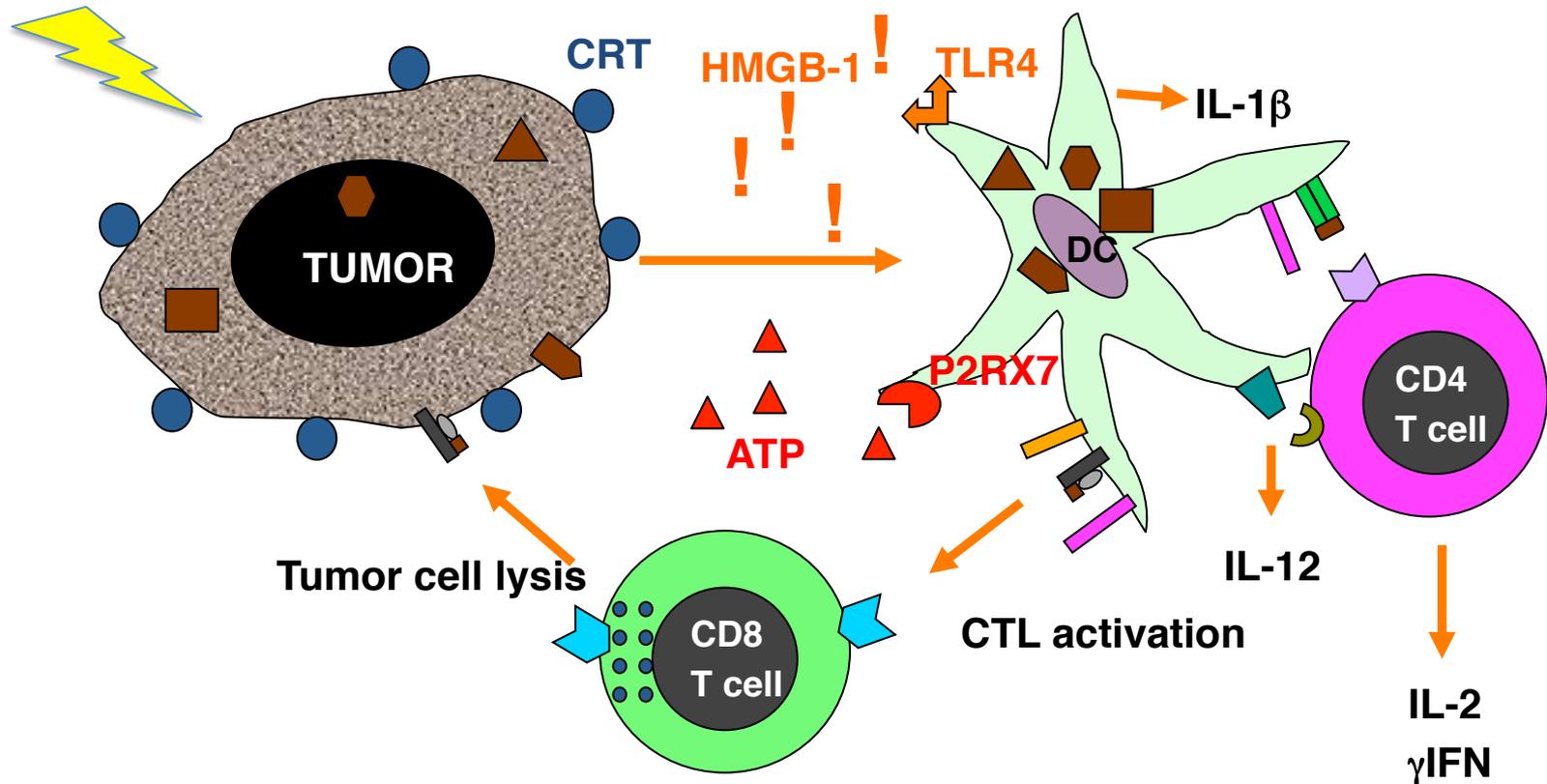
- intrinsic tumor cell characteristics (p53 status)
- radiation dose and fractionation
- oxygen tension
- DNA repair capacity
- redox status
- cell cycle phase

- APOPTOSIS
- NECROSIS
- MITOTIC CATASTROPHE
- SENESENCE
- AUTOPHAGY
- ✧ **IMMUNOGENIC CELL DEATH**

Immunogenic cell death (ICD)



Cross-priming of anti-tumor T cells

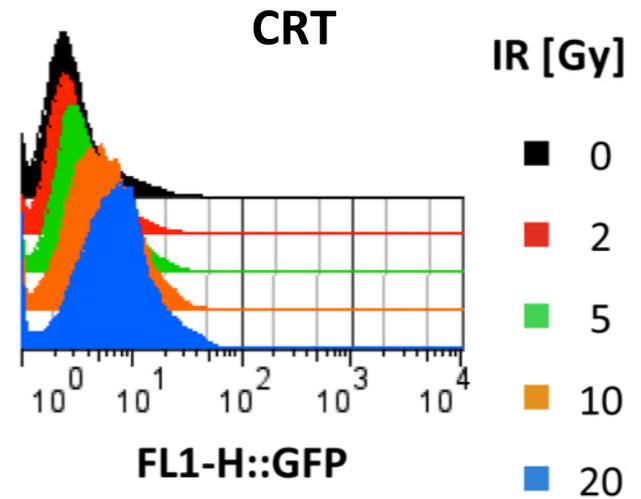
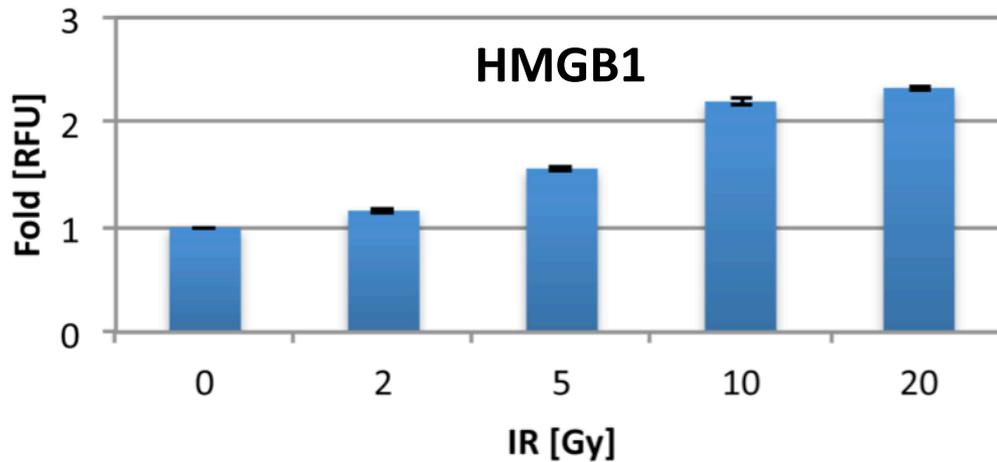
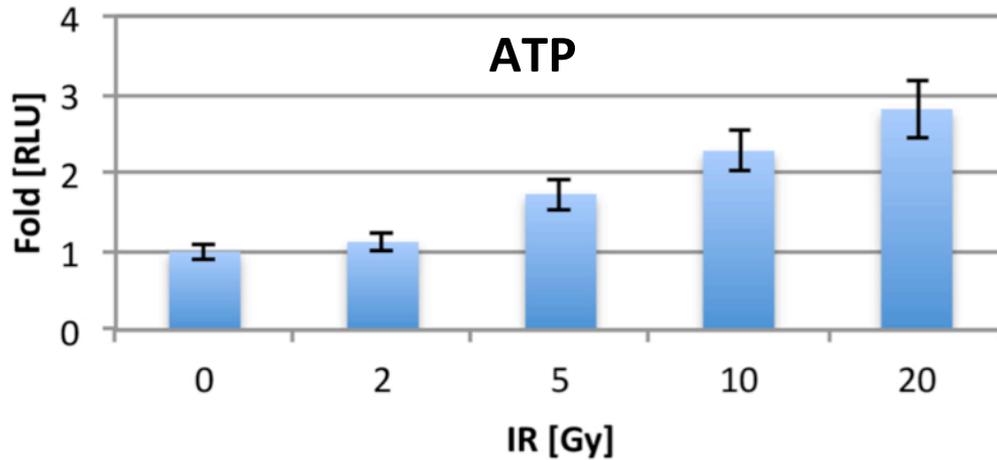


CRT, the “eat me” signal calreticulin translocates to cell surface (Obeid et al., Nat Med 2007, 13:54-61; Cell Death Differ 2007, 14:1848)

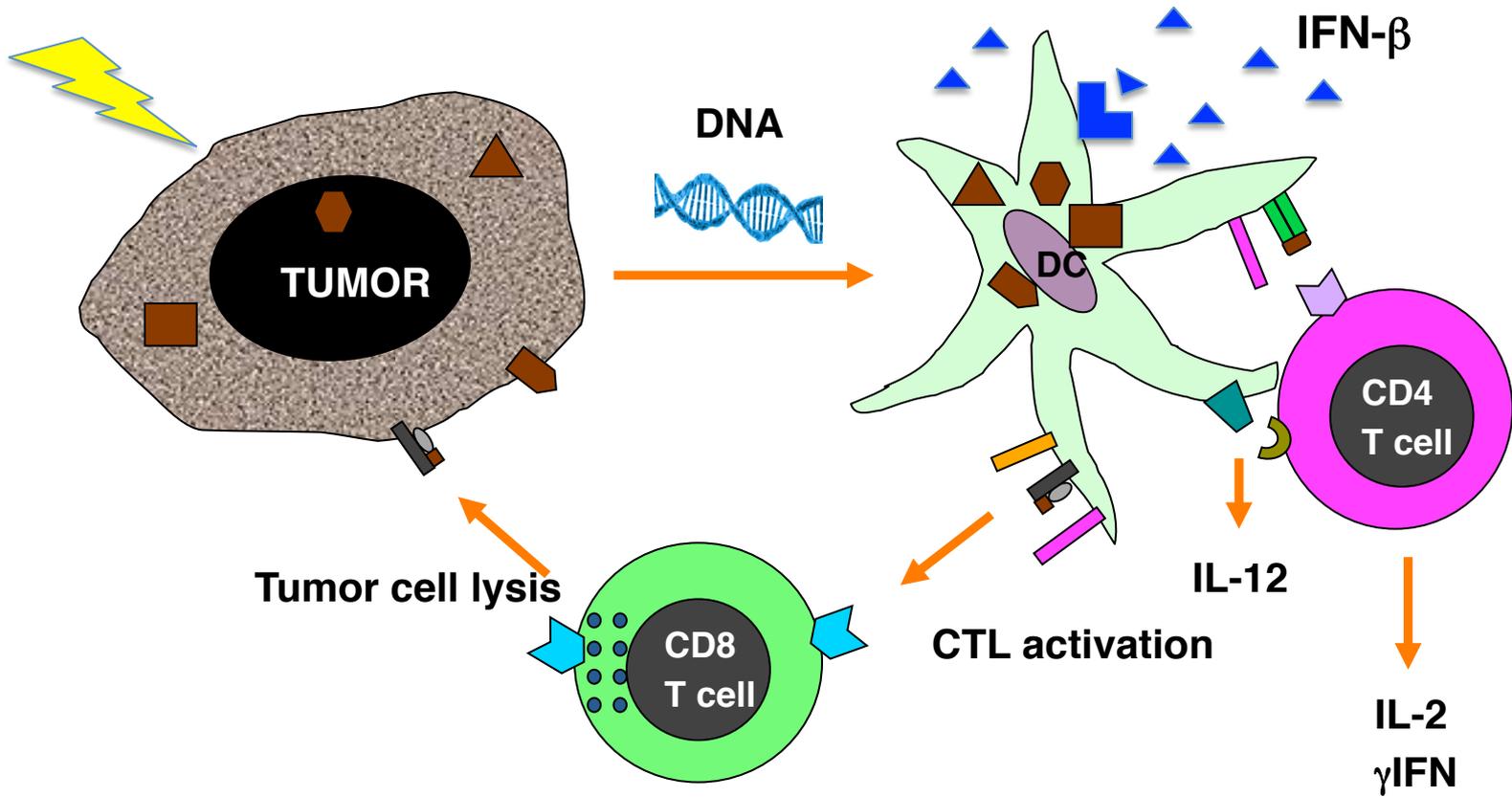
HMGB-1, a damage associated molecular pattern (DAMP) binds to TLR4 to promote cross-presentation of tumor-derived antigens (Apetoh et al., Nat Med 2007, 13:1050)

ATP released by dying cells binds to P2RX7 purinergic receptor leading to inflammasome activation and IL-1β production (Ghiringhelli et al., Nat Med 2009, 15:1170)

Immunogenic cell death (ICD)



Cross-priming of anti-tumor T cells

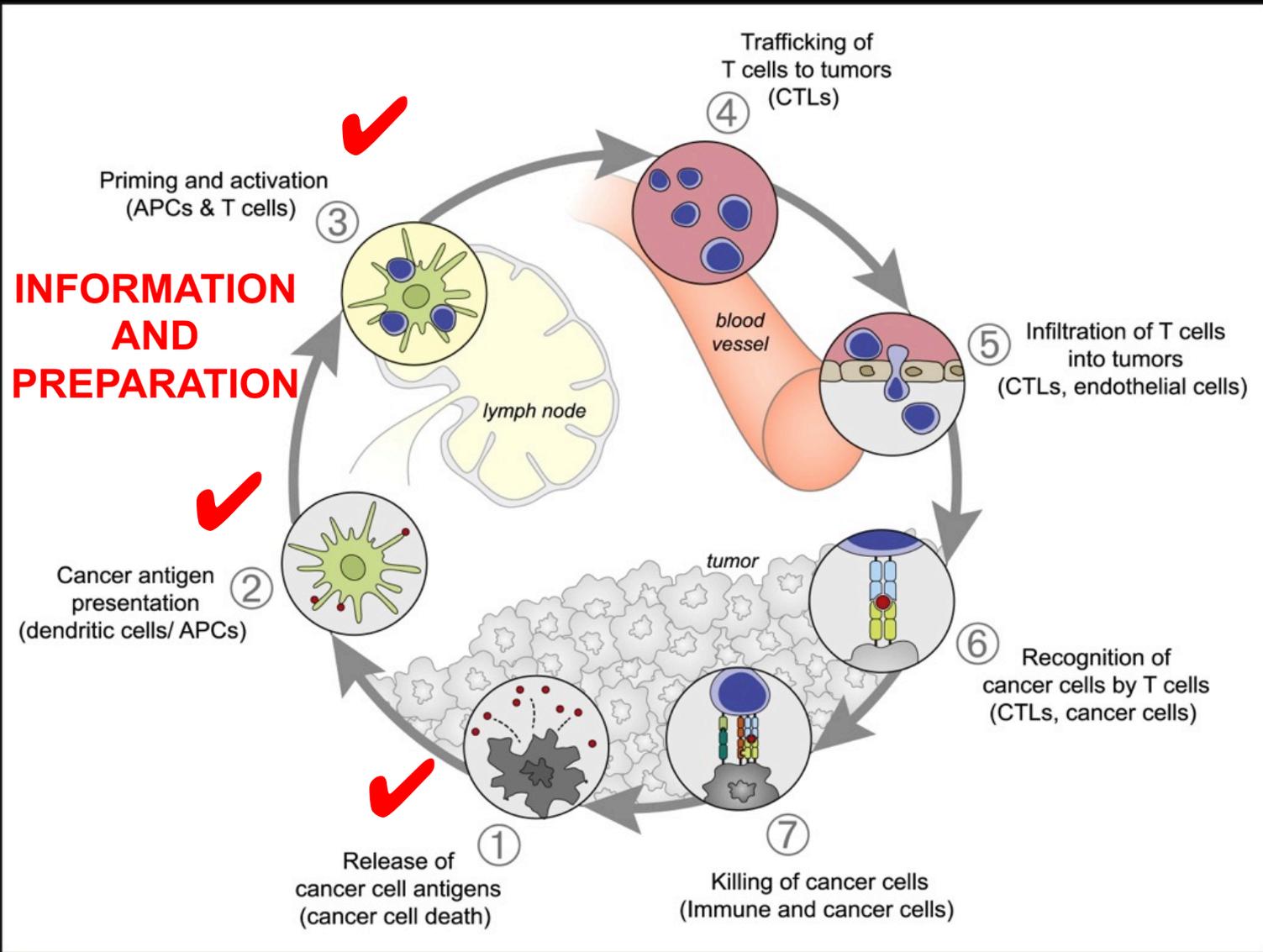


IFN type I is produced by DC infiltrating irradiated tumors (Burnette et al., *Cancer Res* 2011; 71(7); 2488–96)

STING-dependent cytosolic DNA sensing by in irradiated tumors (Deng et al., *Immunity* 2014; 41; 843)

The Cancer-Immunity Cycle

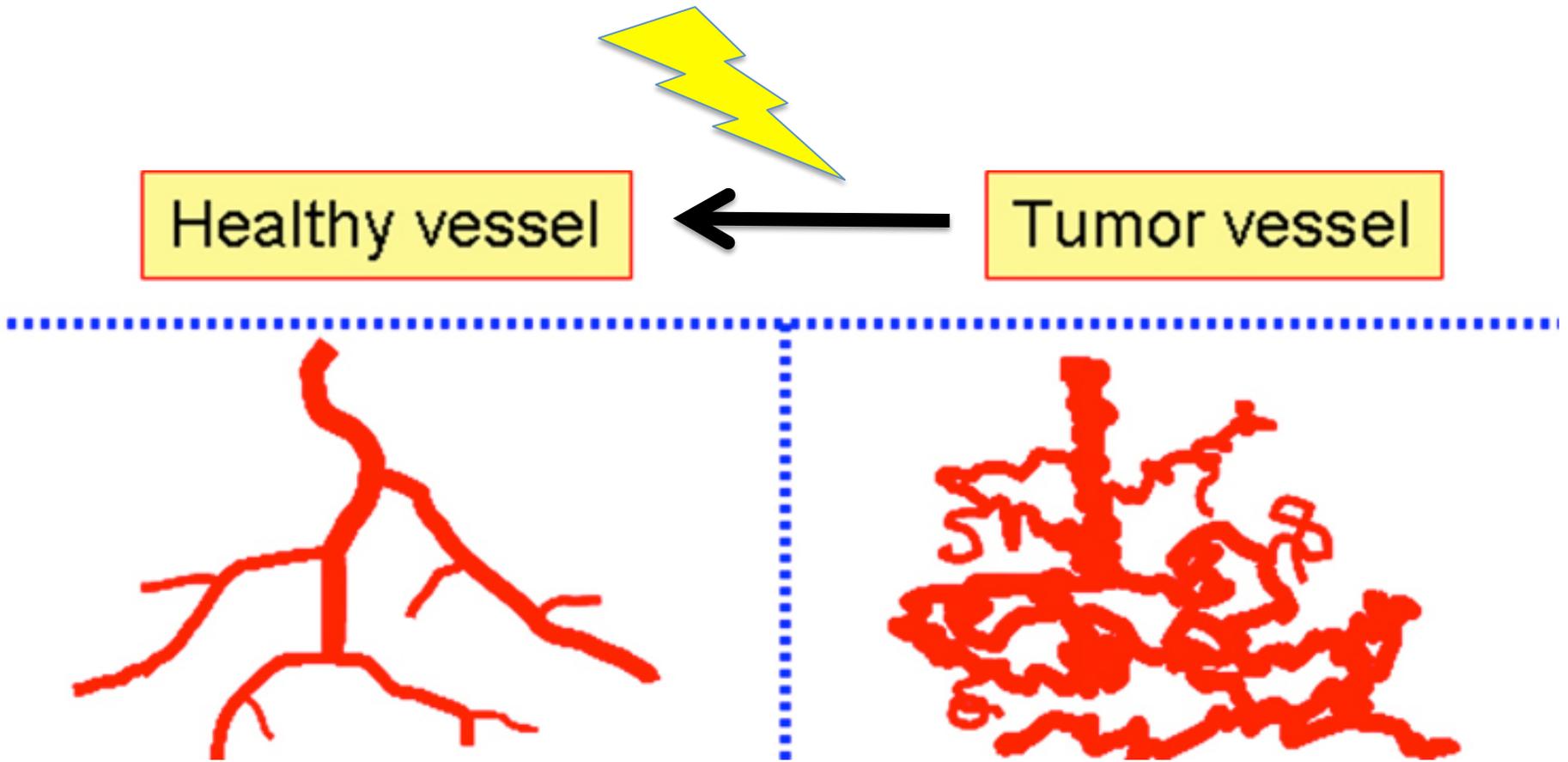
(Chen and Mellman, *Immunity* 2013)



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Vascular barrier: radiation-induced vascular normalization



Increased expression by irradiated tumor and stromal cells of:

Chemokines (e.g, CXCL16, CXCL10, CXCL9)

Cytokines (e.g., IL-1 β , TNF α , IFN type I)

Adhesion molecule (ICAM-1, VCAM)

MHC class I molecules

Stress-induced ligand (NKG2D ligands)

Tumor antigens (e.g., CEA, MUC-1)

Death Receptors (Fas/CD95)

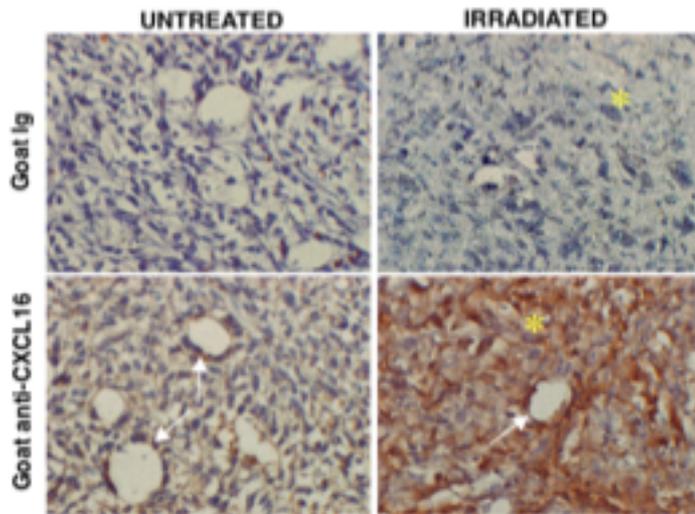


Increased T cell recruitment and infiltration

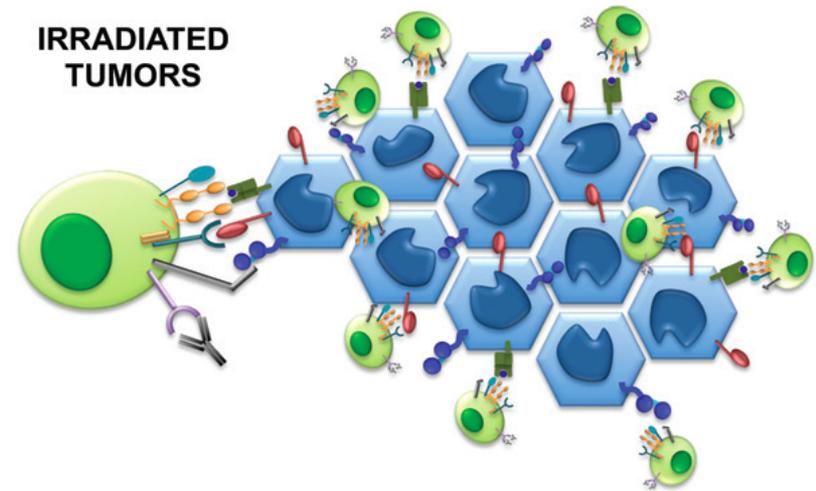
Increased interactions between effector T cells and tumor cells

Increased tumor cell killing

Induction of CXCL16 and NKG2D ligands in the tumor by RT is required for tumor rejection by CD8 T cells



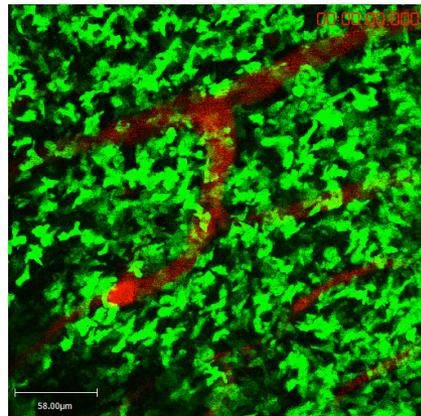
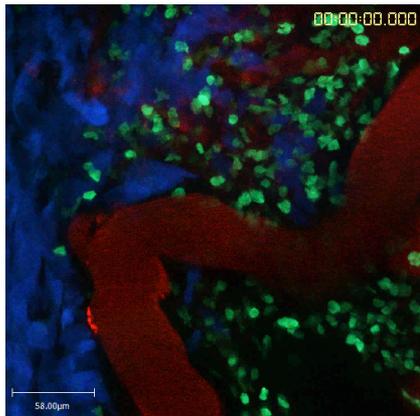
Matsumura *et al.*, *J Immunol* 2008



Demaria *et al.*, *Oncolimmunology* 2:3, e23127 2013

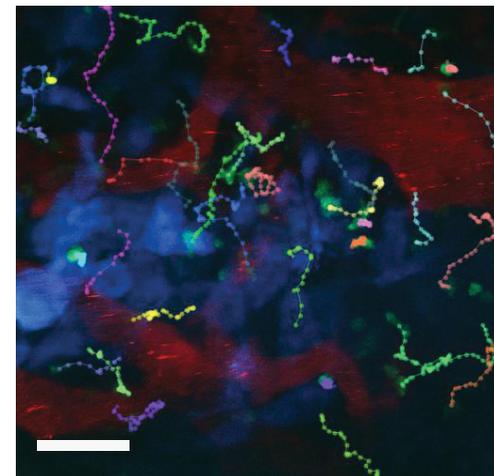
Anti-CTLA-4

RT + Anti-CTLA-4



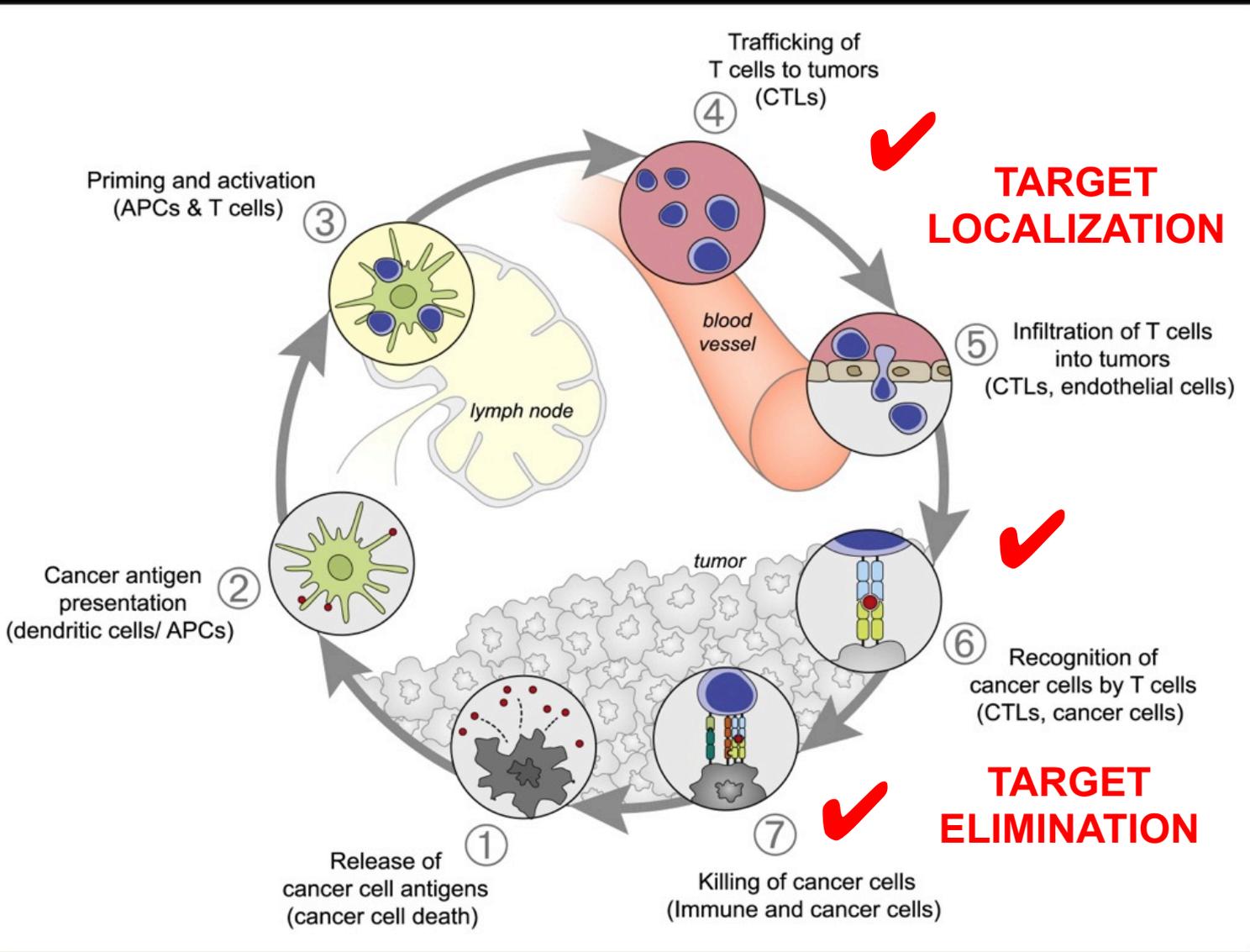
Ruocco *et al.*, *J Clin Invest* 122:3718-30 2012

Blood vessels
T cells
Tumor cells



The Cancer-Immunity Cycle

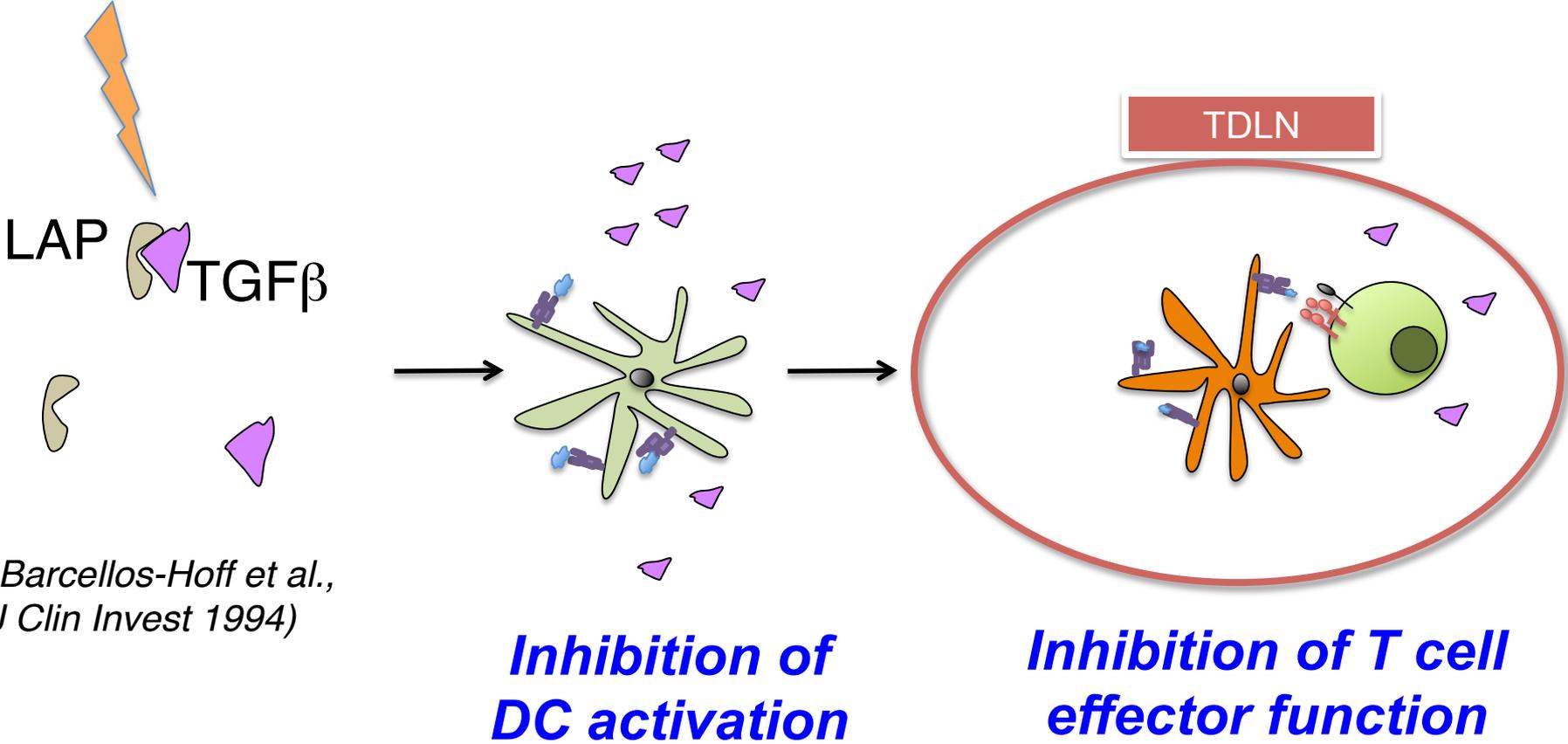
(Chen and Mellman, *Immunity* 2013)



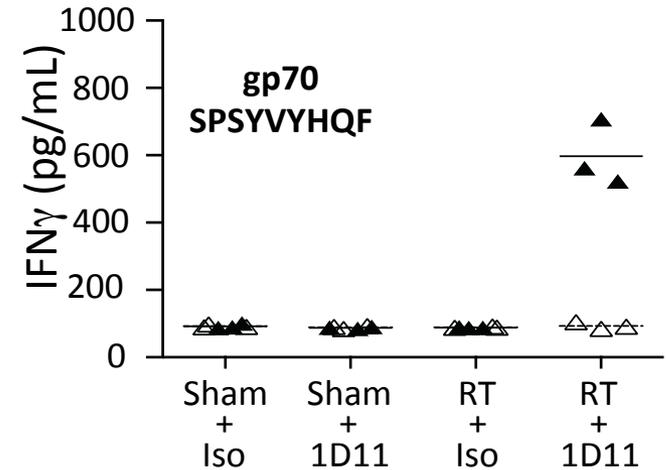
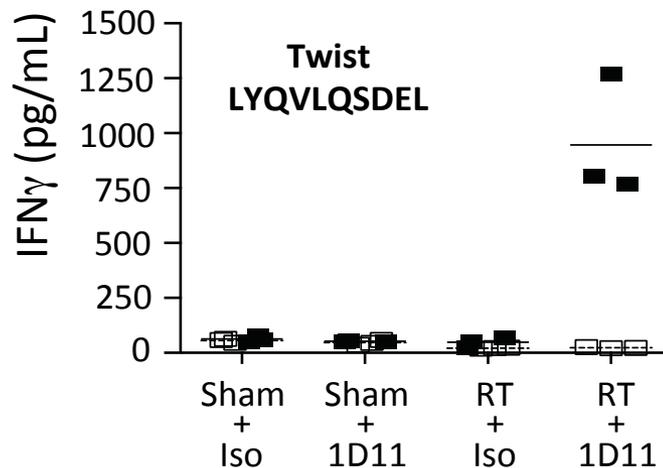
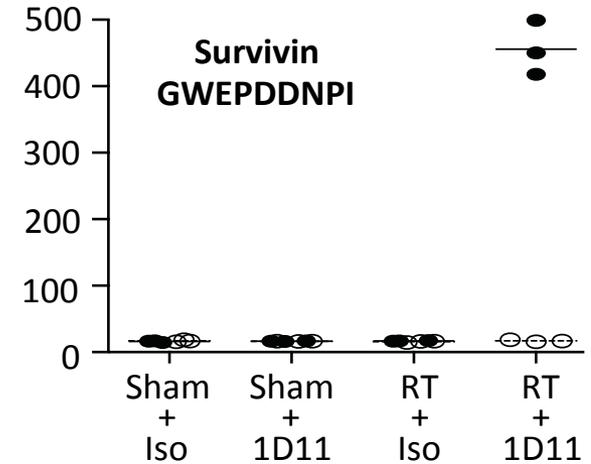
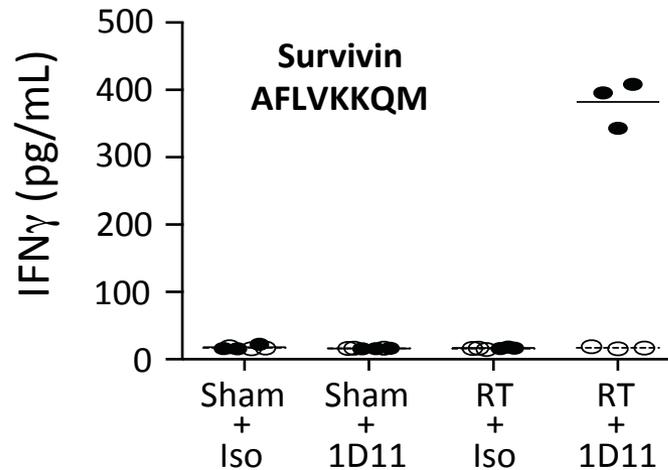
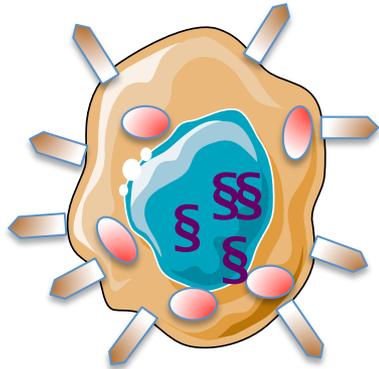
Effects of radiation that can hinder the development of anti-tumor immunity

- Induction of Treg cells (*Schaue et al., Front Oncol 2012*)
- Activation of TGF β (*Barcellos-Hoff et al., J Clin Invest 1994*)
- Up-regulation of PD-L1 (*Deng et al., J Clin Invest 2014*)
- Induction of pro-tumorigenic M2 macrophages (*Tsai et al., IJROBP 2007*)
- Induction/activation of STAT-3, Bcl-XL, VEGF (*Ho et al., Cancer Sci 2010; Kargiotis et al., J Neurooncol 2010*)

RT-induced negative regulators



TGF β neutralization is required for RT-induced tumor-specific T cell priming

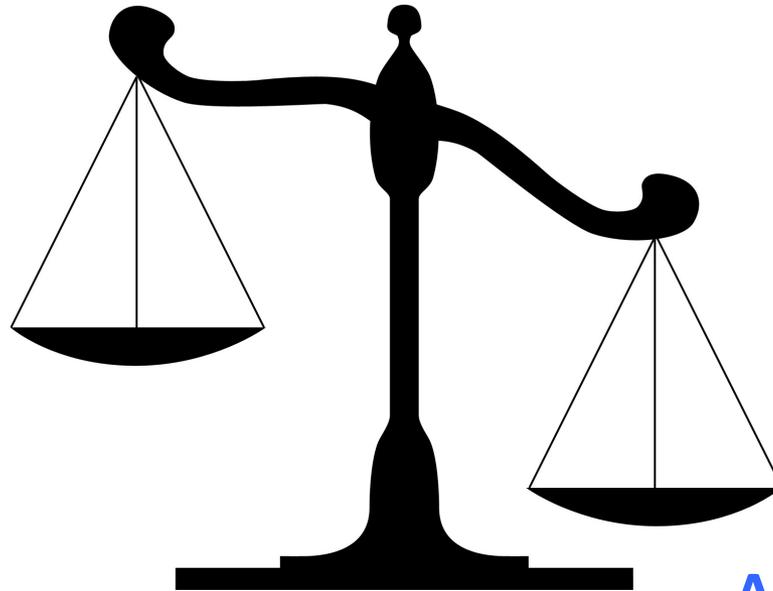


Acute
inflammation

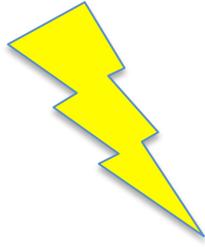


Immunosuppression

**Pro-inflammatory
Tumor rejection**



**Anti-inflammatory
Immunosuppression**



**Pro-inflammatory
Tumor rejection**

**Anti-inflammatory
Immunosuppression**

? Radiation dose and
fractionation

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Pre-clinical combinations of RT and immunotherapy

Increasing APC

Flt3-Ligand

Chakravarty et al, Cancer Res 1999

Demaria et al. IJROBP 2004

Exogenous DC

Nikitina et al., Int J Cancer 2001

Teitz-Tennenbaum et al, Cancer Res 2003

Kim et al., Int J Cancer 2004

Increasing danger signals

TLR9 agonists

Milas et al., Cancer Res 2004

Mason et al., Clin Cancer Res 2005

Zhang et al., PlosOne 2012

TLR7 agonists

Dewan et al., Clin Cancer Res 2012

Dovedi et al., Blood 2013

Pre-clinical combinations of RT and immunotherapy

Improving T cell co-stimulation

Anti-CD137 mAb

Shi et al., Anticancer Res 2006

Newcomb et al., Radiat Res 2010

Anti-OX40 mAb

Gough et al., J. Immunother 2010

Blocking immune checkpoints

Anti-CTLA-4 mAb

Demaria et al., Clin Cancer Res 2005

Dewan et al., Clin Cancer Res 2009

Anti-PD-1 mAb

Verbrugge et al., Cancer Res 2012

Zeng et al., IJROBP 2013

Anti-PDL-1 mAb

Deng et al., J Clin Invest 2014

Dovedi et al., Cancer Res 2014

Pre-clinical combinations of RT and immunotherapy

Priming anti-tumor T cells

Vaccinia and avipox
CEA vaccines

Chakraborty et al., Cancer Res 2004

GM-CSF tumor cell
vaccine (GVAX)

Newcomb et al., Clin Cancer Res 2006

Adoptive T cell transfer

Effector CD8 T cells

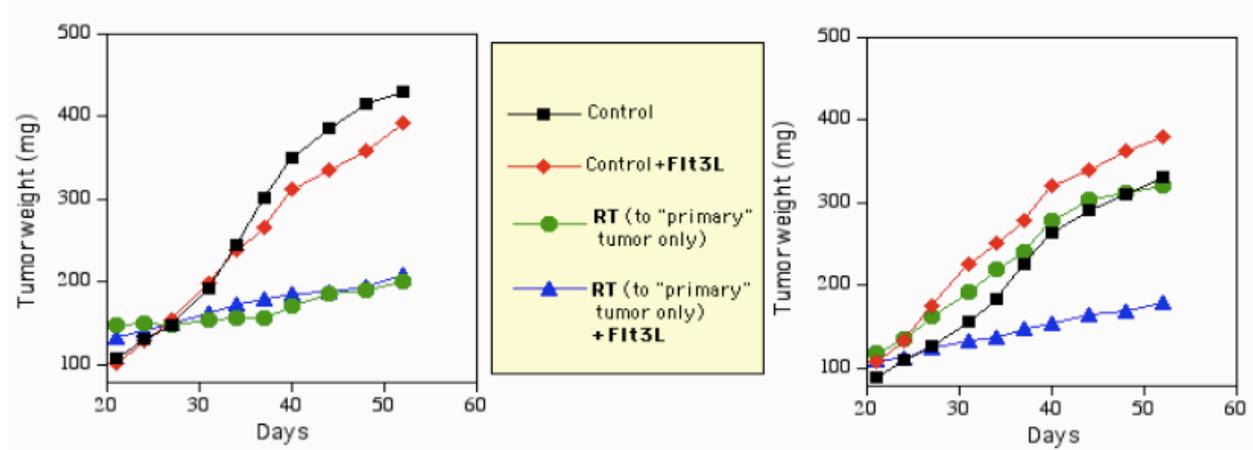
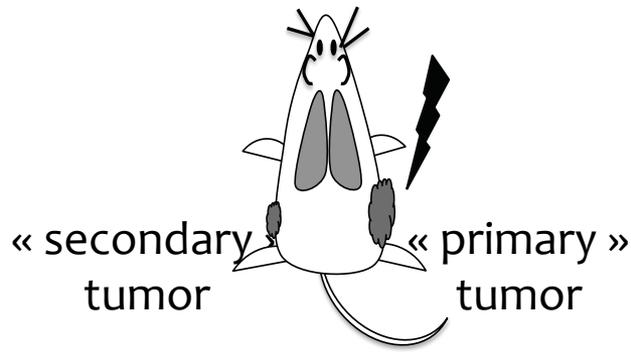
Chakraborty et al., J Immunol 2003

Reits et al., J Exp Med 2006

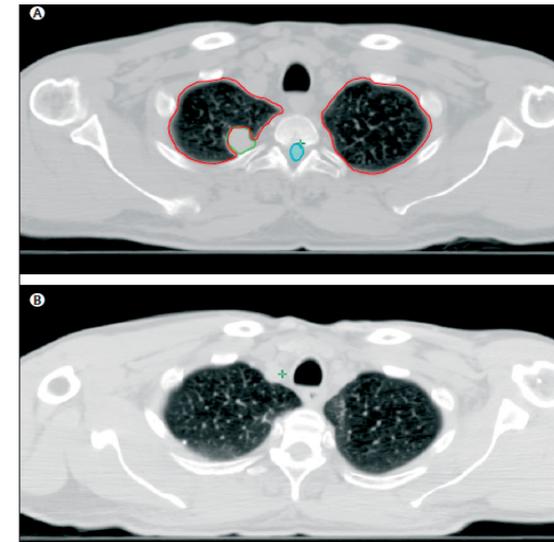
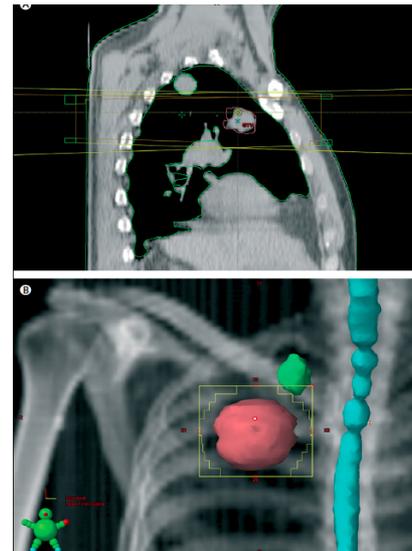
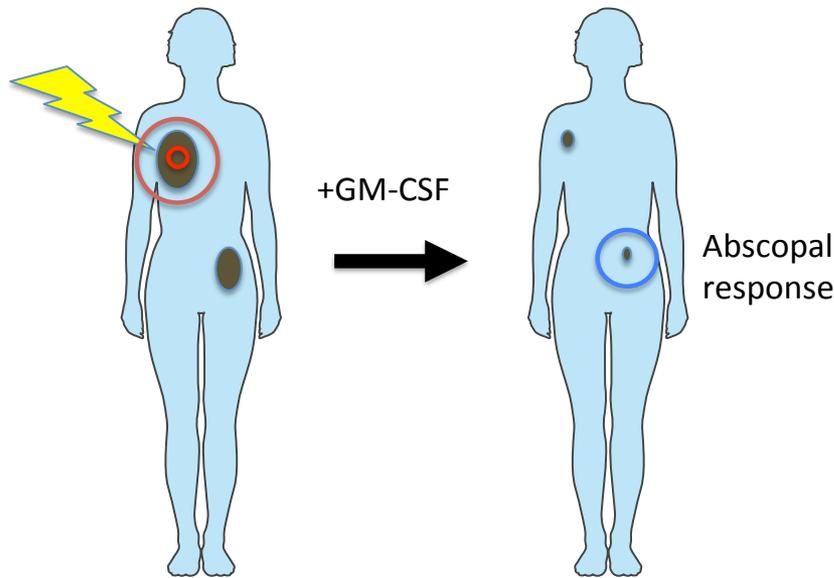
CD8 and CD4 T cells

Klug et al., Cancer Cell 2013

DC growth factors in combination with RT induce abscopal effects in mice and patients



Demaria et al., *Int J Radiation Oncology Biol Phys* 2004



Golden et al., *Lancet Oncology* 2015, *In press*

Clinical Translation

DC administration

Hepatocellular carcinoma: DC i.t. 2 days after one dose radiation: partial response in 2/14 patients (*Chi et al., J Immunother 2005*)

Sarcoma: DC i.t. during multi-fraction neoadjuvant radiation: tumor-specific immune responses in 9/17 patients (*Finkelstein et al., IJROBP 2012*)

TLR9 agonists

Lymphoma: phase I/II study 2 Gy x 2 plus i.t. synthetic CpG. Abscopal responses in 27% with 1 complete response, 3 partial responses, and 8 patients with stable disease (*Brody et al., JCO 2009*)

Second trial performed in Mycosis fungoides (*Kim et al., Blood. 2012*).

TLR7 agonists

Breast cancer with cutaneous metastases: phase I/II study 6 Gy x 5 plus topical imiquimod (NCT01421017).

High dose IL-2

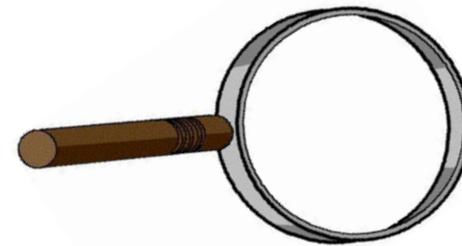
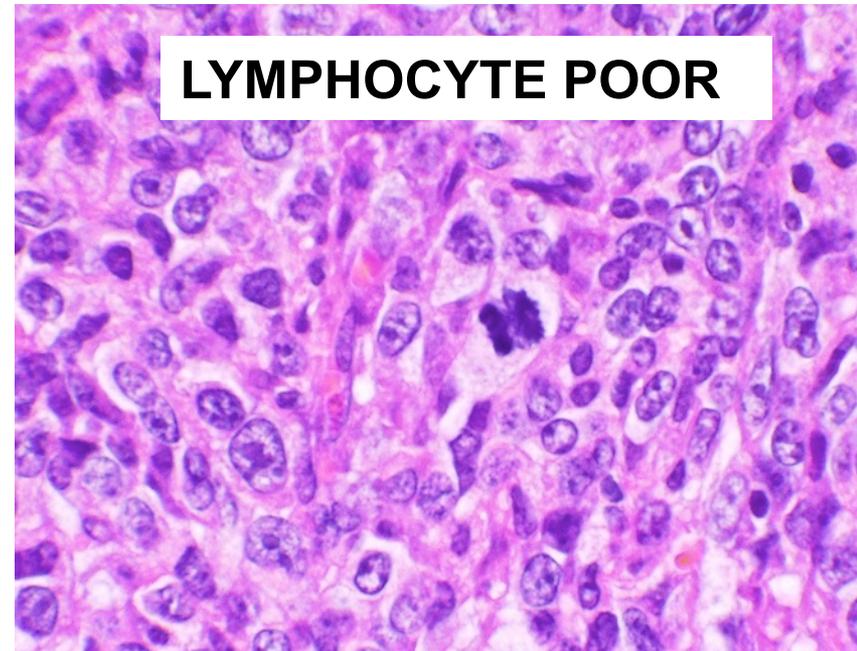
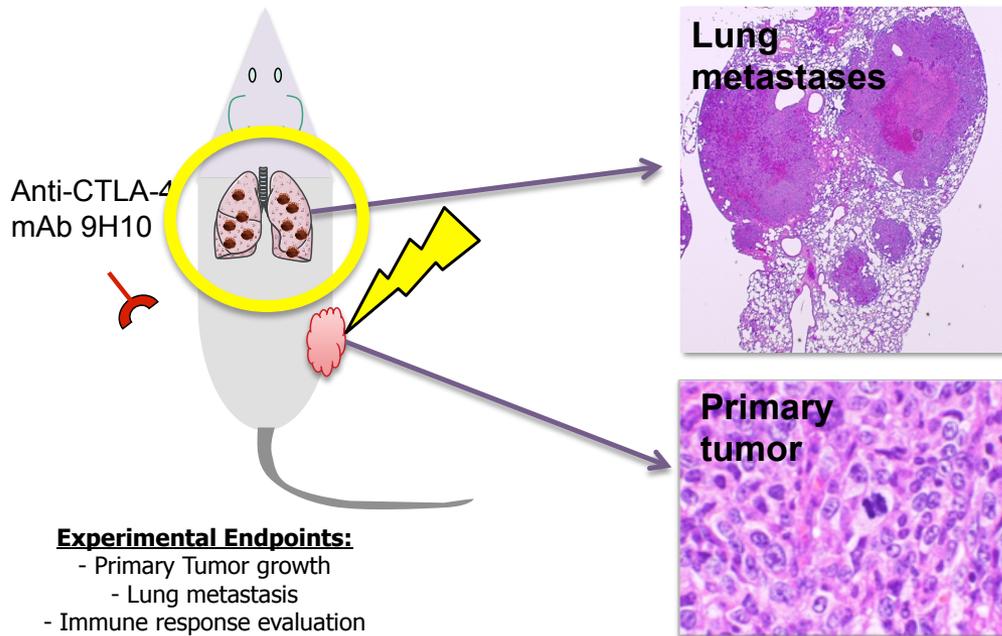
Melanoma and RCC (phase I): SBRT followed by high dose IL-2. Response rate of 66%. Patients who responded showed increase in effector memory T cells (*Seung et al., Sci Transl Med. 2012*).

Phase II randomized study of SBRT (20 Gy) and high-dose IL-2 versus IL-2 alone in patients with metastatic melanoma is ongoing (NCT01416831).

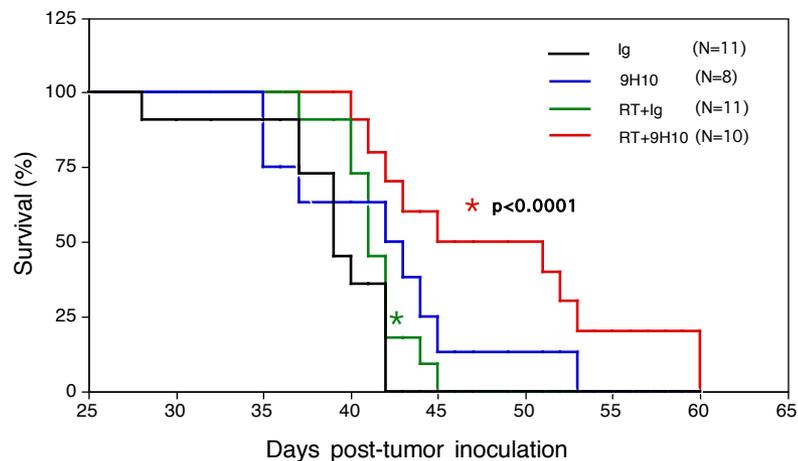
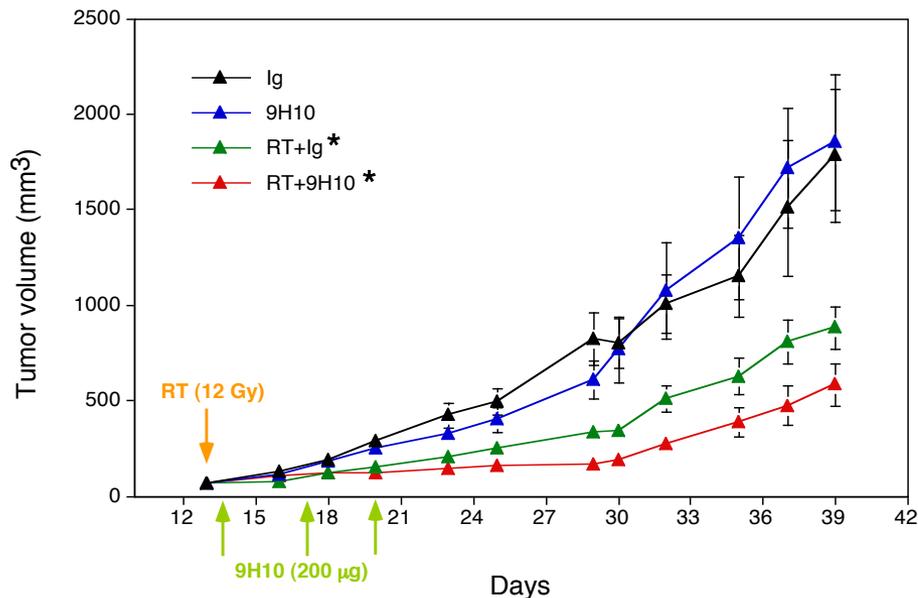
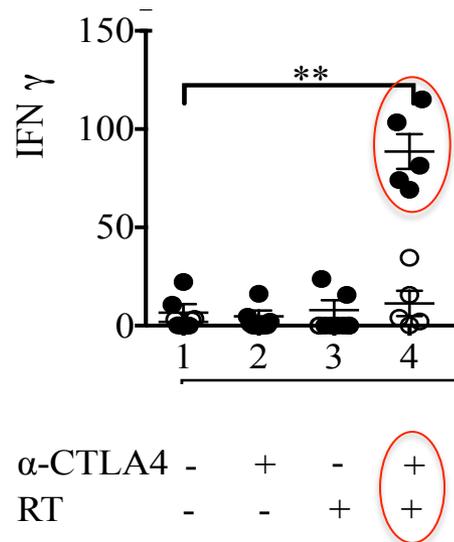
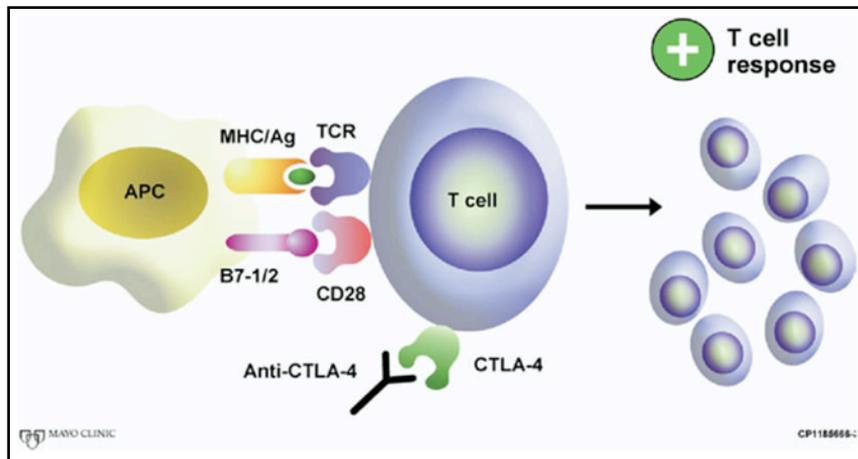
Phase II in metastatic RCC (NCT01896271).

Mouse model of anti-CTLA-4-refractory cancer

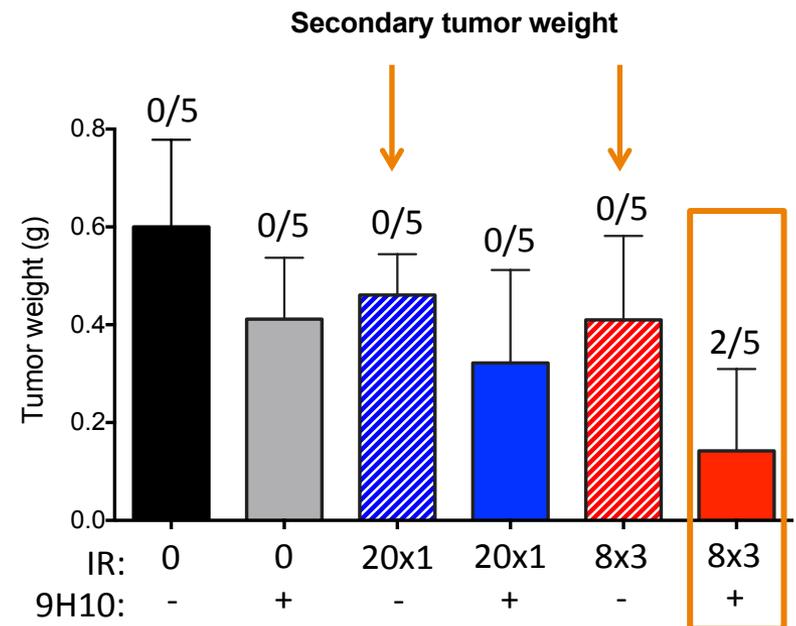
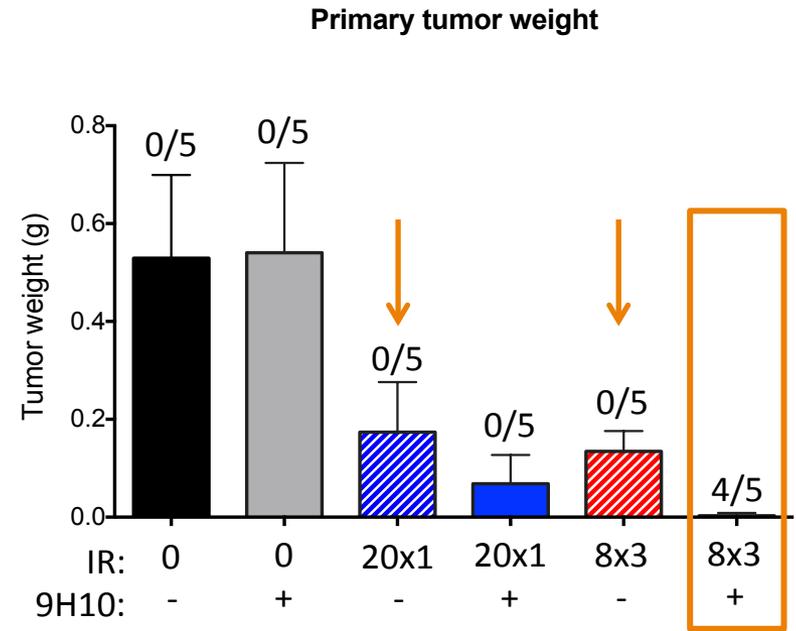
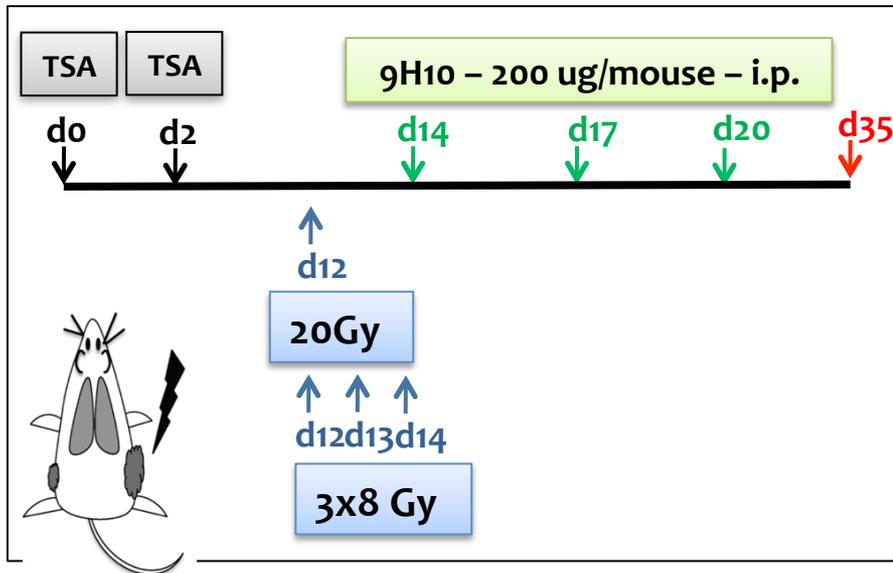
4T1 mouse model of metastatic breast cancer



Radiotherapy synergizes with CTLA-4 blockade



Fractionated but not single dose RT elicits an abscopal response in combination with anti-CTLA-4



Anti-CTLA-4 (Ipilimumab)

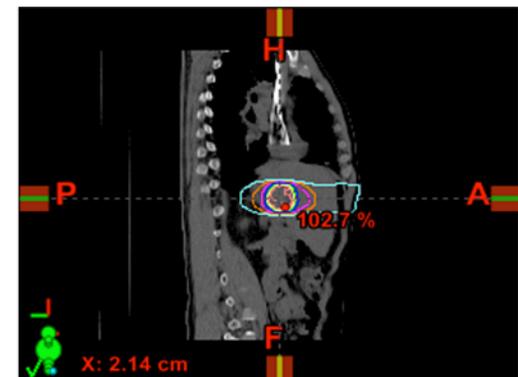
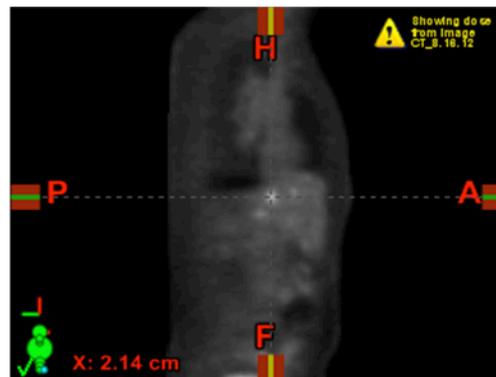
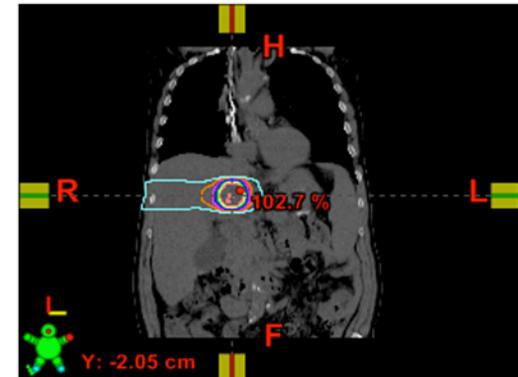
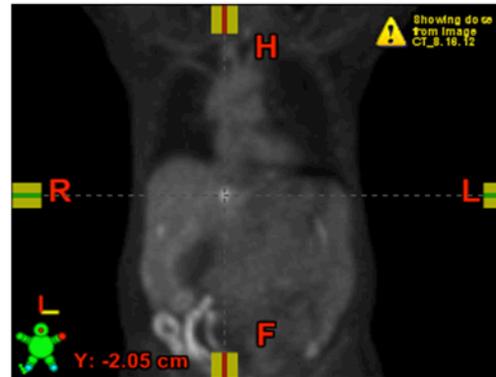
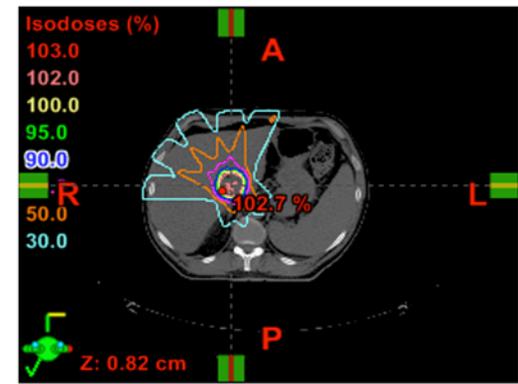
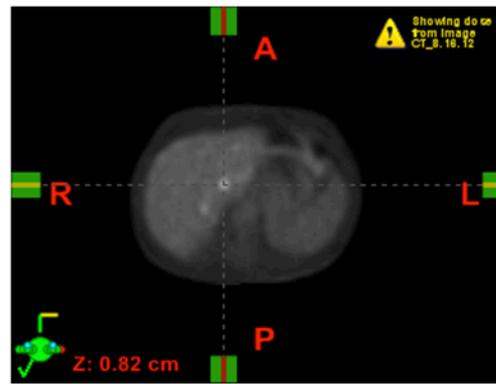
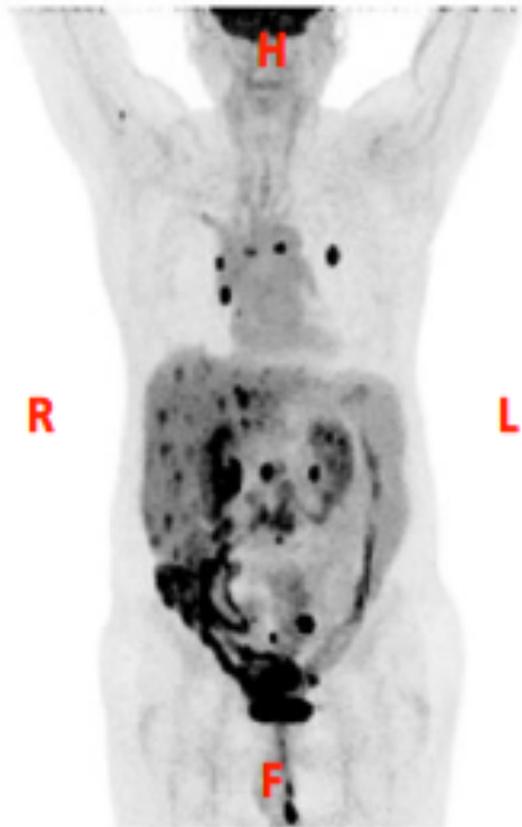
Metastatic melanoma: Case report of a patient with progressive disease on Ipilimumab who showed a dramatic abscopal response after radiotherapy (9.5 Gy x 3) to one paraspinal mass, accompanied by immunological changes (*Postow et al., NEJM 2012*).

Phase III randomized trial in prostate cancer (8Gy x 1 to bone metastasis +/- Ipi) with no significant difference in OS but benefit of Ipi in subgroup with good prognostic features (*Kwon et al., Lancet Oncol 2014*)

Currently >20 trials testing combination of radiotherapy and Ipilimumab are underway.

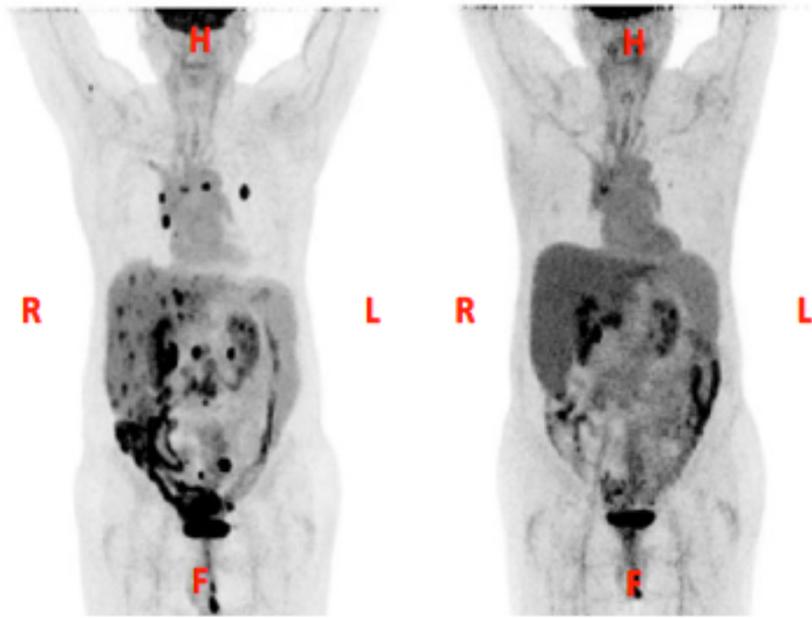
Patient with Metastatic NSCLC

Progressing after 3 lines of chemo and chest RT: Multiple lung, bone and liver metastasis



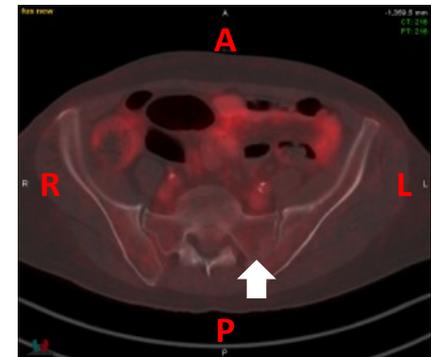
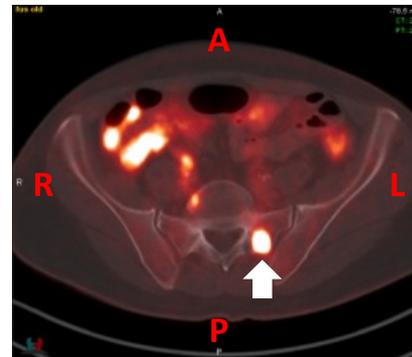
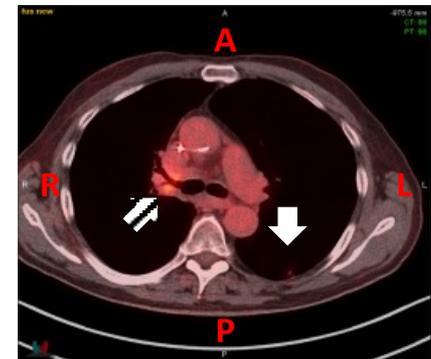
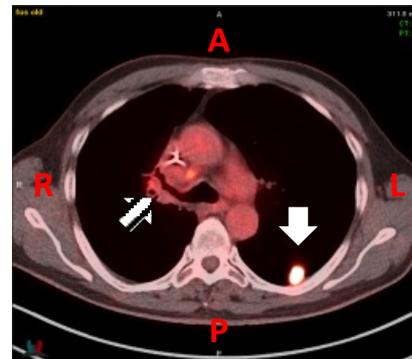
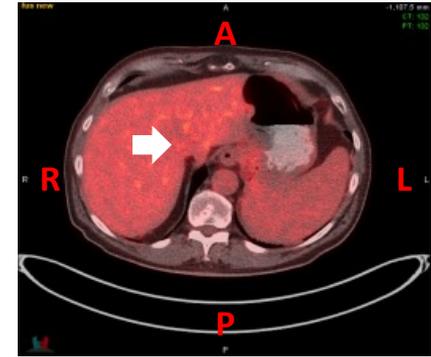
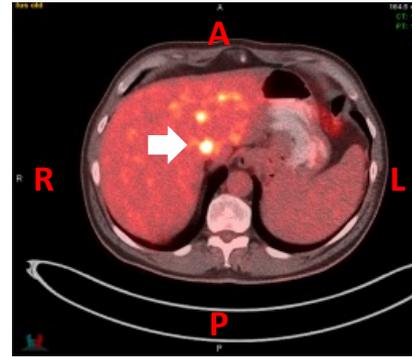
RT to one liver met
6 Gy X 5 (TD 30 GY)
Ipilimumab, 3 mg/Kg, after first RT
q3 weeks, X 4 cycles

Abscopal response in a NSCLC patient treated with local RT and Ipilimumab



August 2012 PET/CT

January 2013 PET/CT



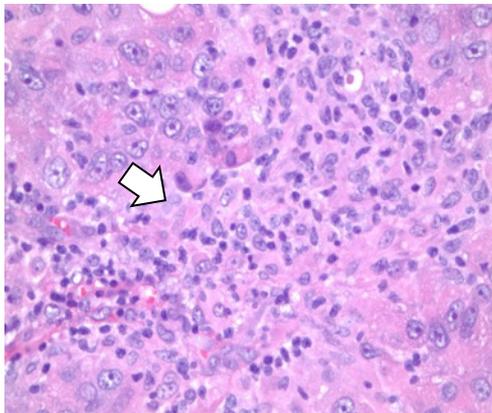
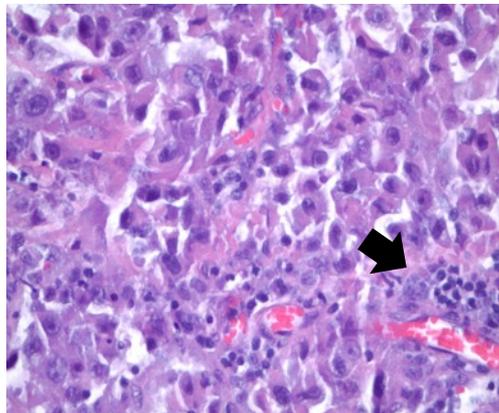
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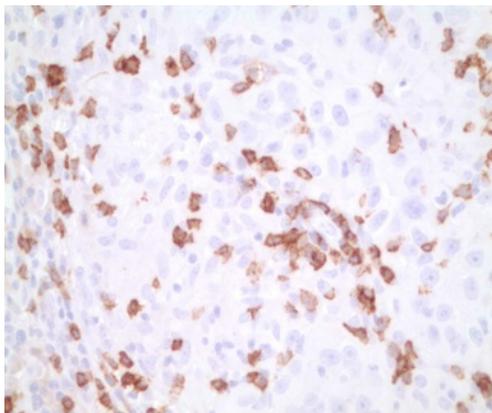
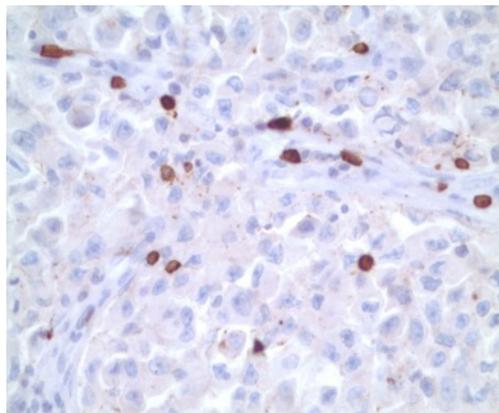
2010

2013

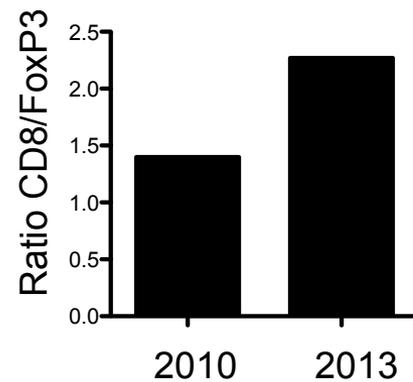
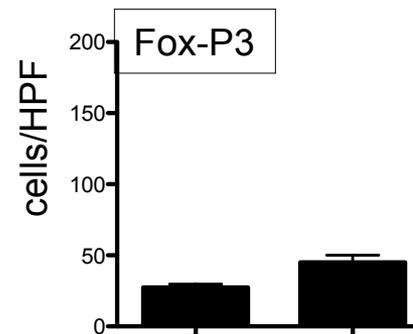
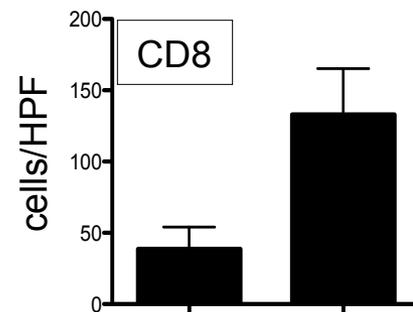
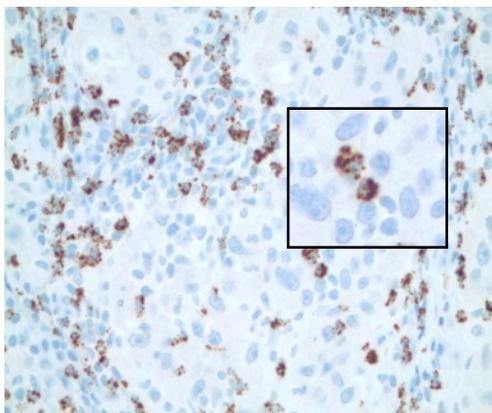
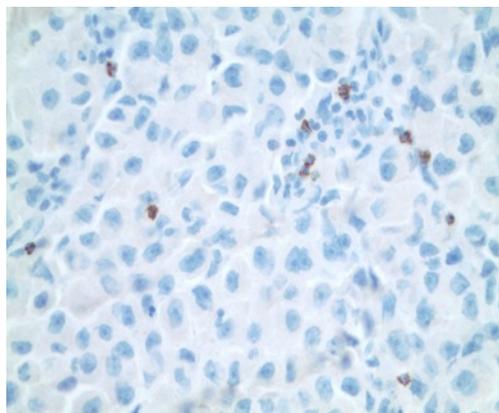
H&E



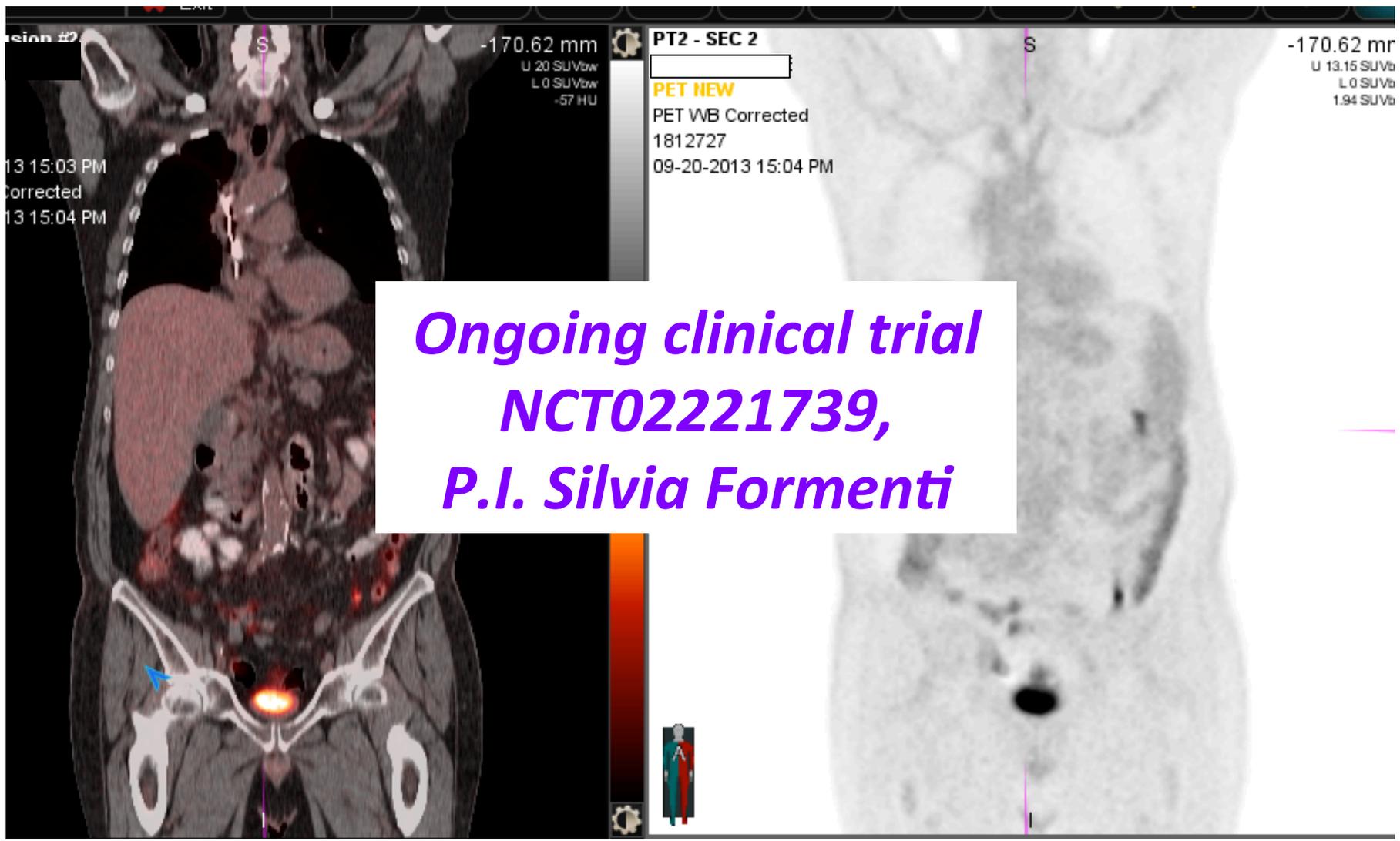
CD8



TIA-1



Clinical and radiological CR at one year: currently NED at >2 years



Lessons and Take Home Messages

- RT has positive effects at all steps of anti-tumor immune response
- Radiotherapy is a suitable partner for combination with multiple immunotherapies
- Several trials testing combinations of RT and immunotherapy are ongoing

Open Questions

- Dose and fractionation matter
- Tumor type/histology
- Sequencing of RT and immunotherapy
- Location of metastasis irradiated (e.g., visceral vs bone)