

# Society for Immunotherapy of Cancer (SITC)

## Immunotherapy for Treatment of Hematologic Malignancies

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## Disclosure



National Institute  
of Allergy and  
Infectious Diseases



# Bone Marrow Transplantation (BMT) Hematopoietic Stem Cell Transplantation (HSCT)

## Applications

*Hematological disorders*

*Hematological malignances*

*Solid tumors*

## Complications

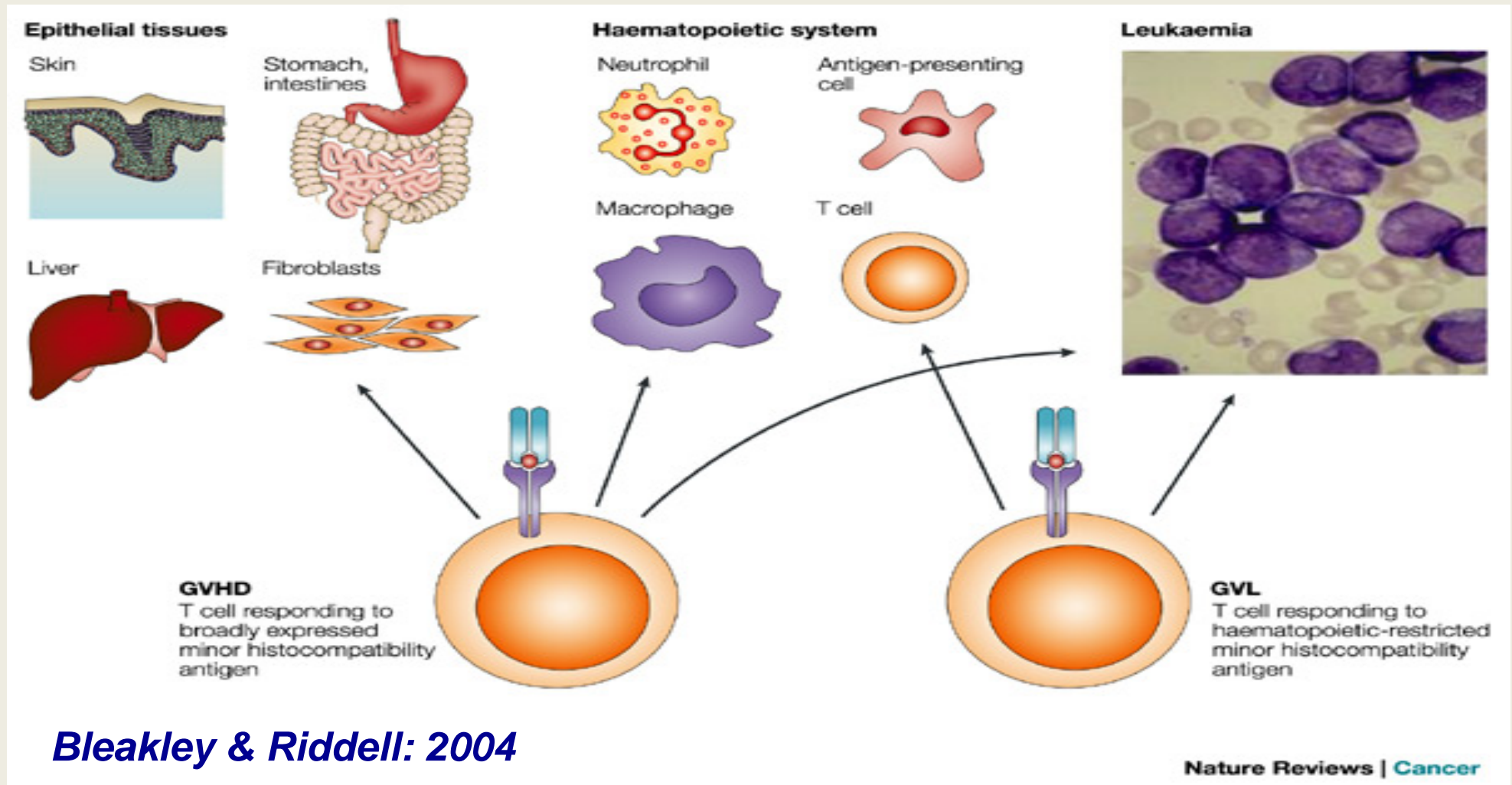
*Graft-versus-host diseases*

*Tumor relapse*

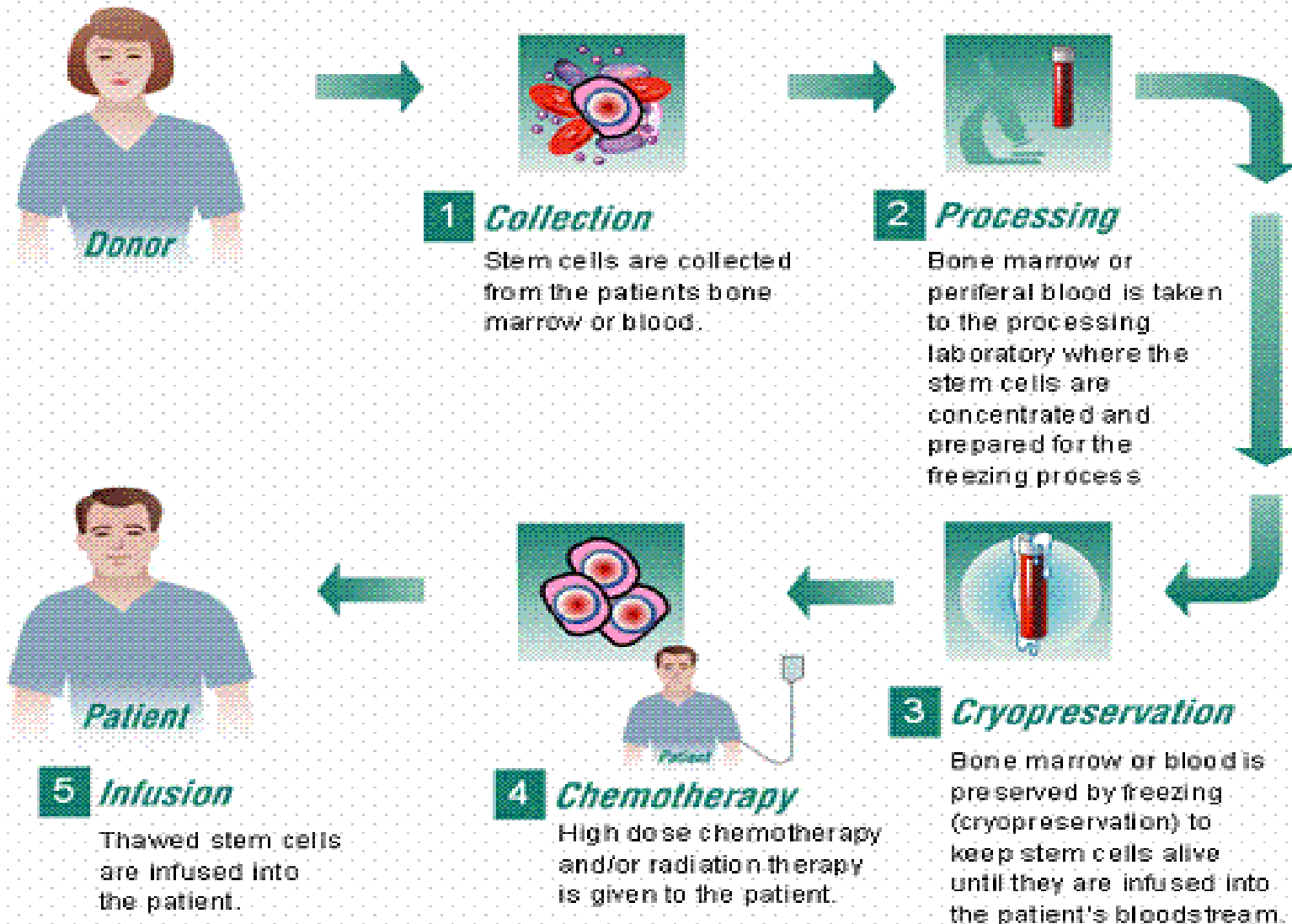
*Infection*

*Graft rejection*

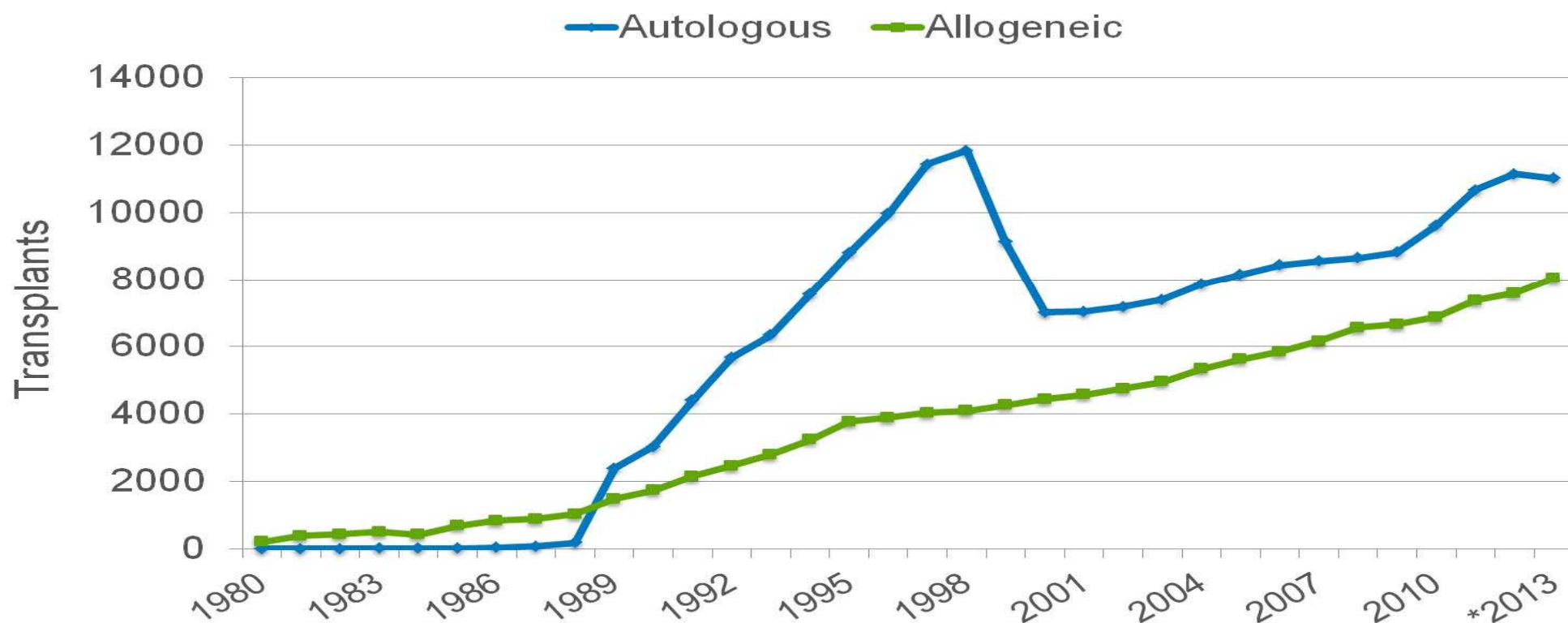
# Separating GVHD and GVL effects



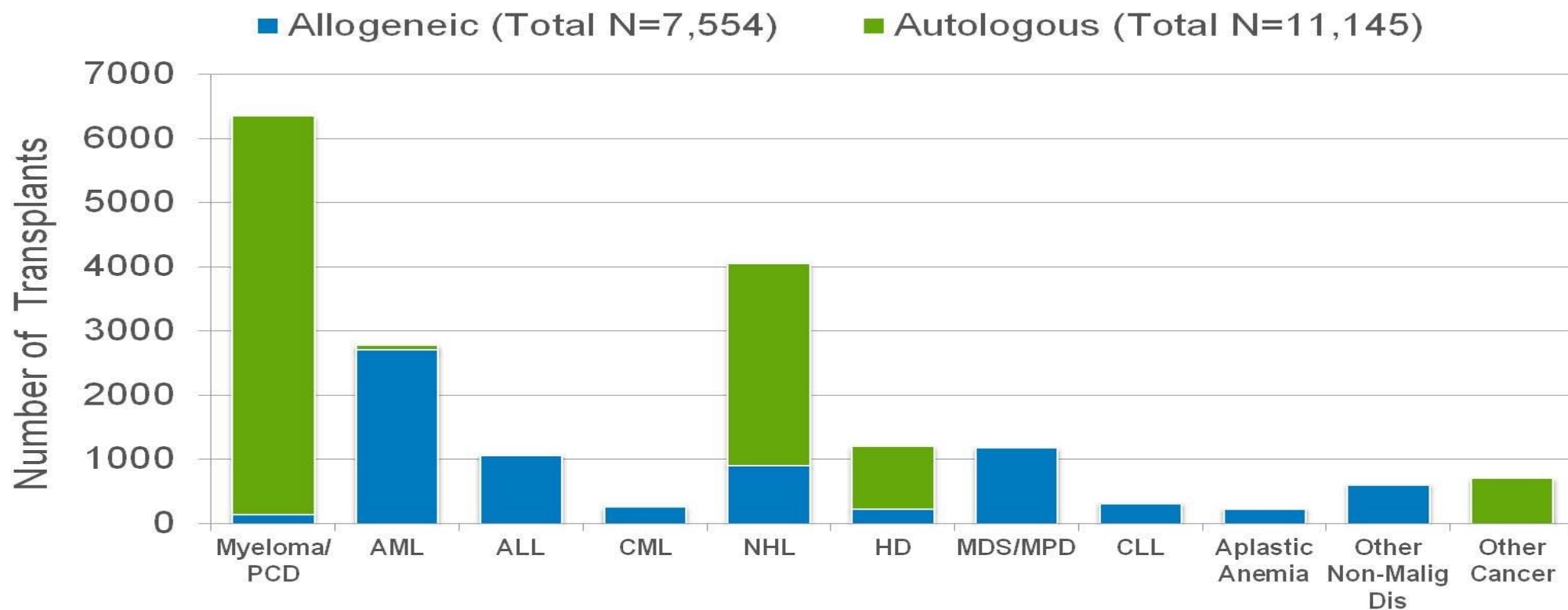
## The Allogeneic Transplant Process



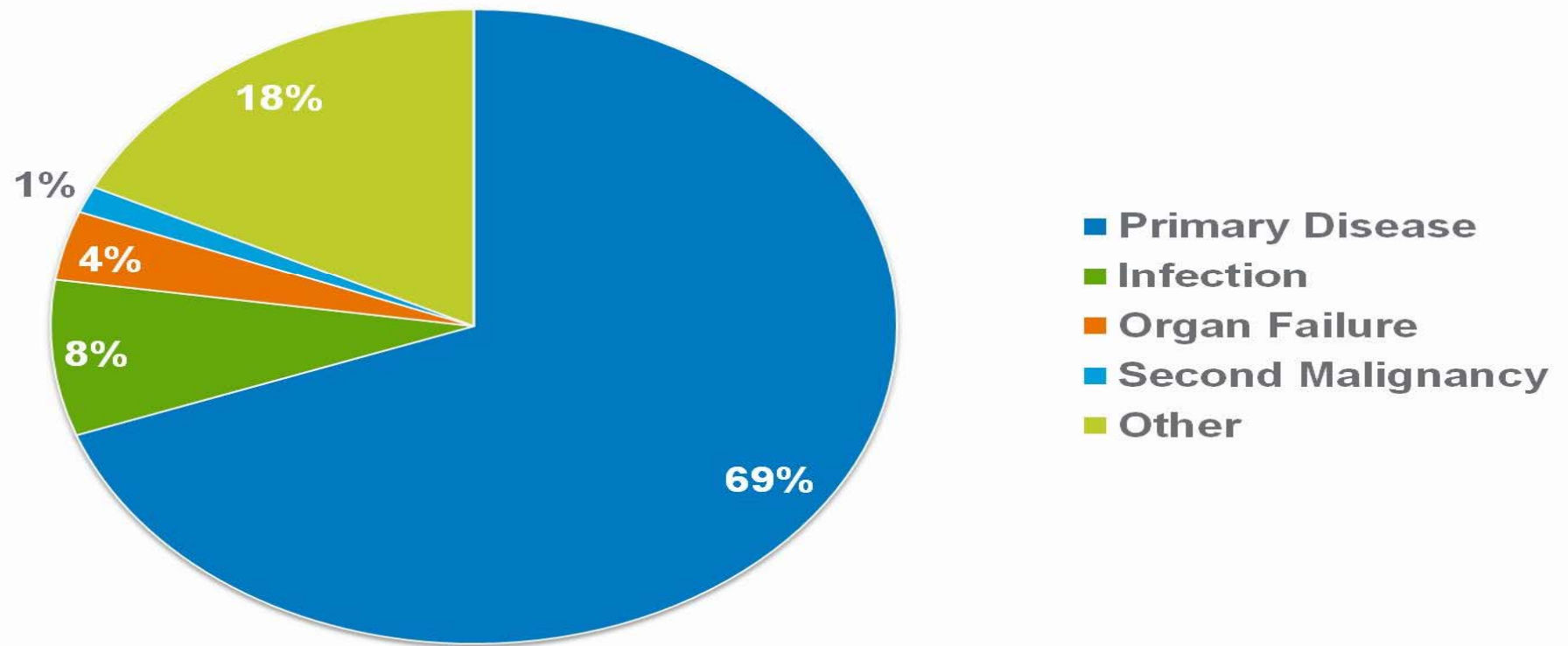
# Annual Number of Transplant Recipients in the US by Transplant Type



# Indications for Hematopoietic Stem Cell Transplants in the US, 2012

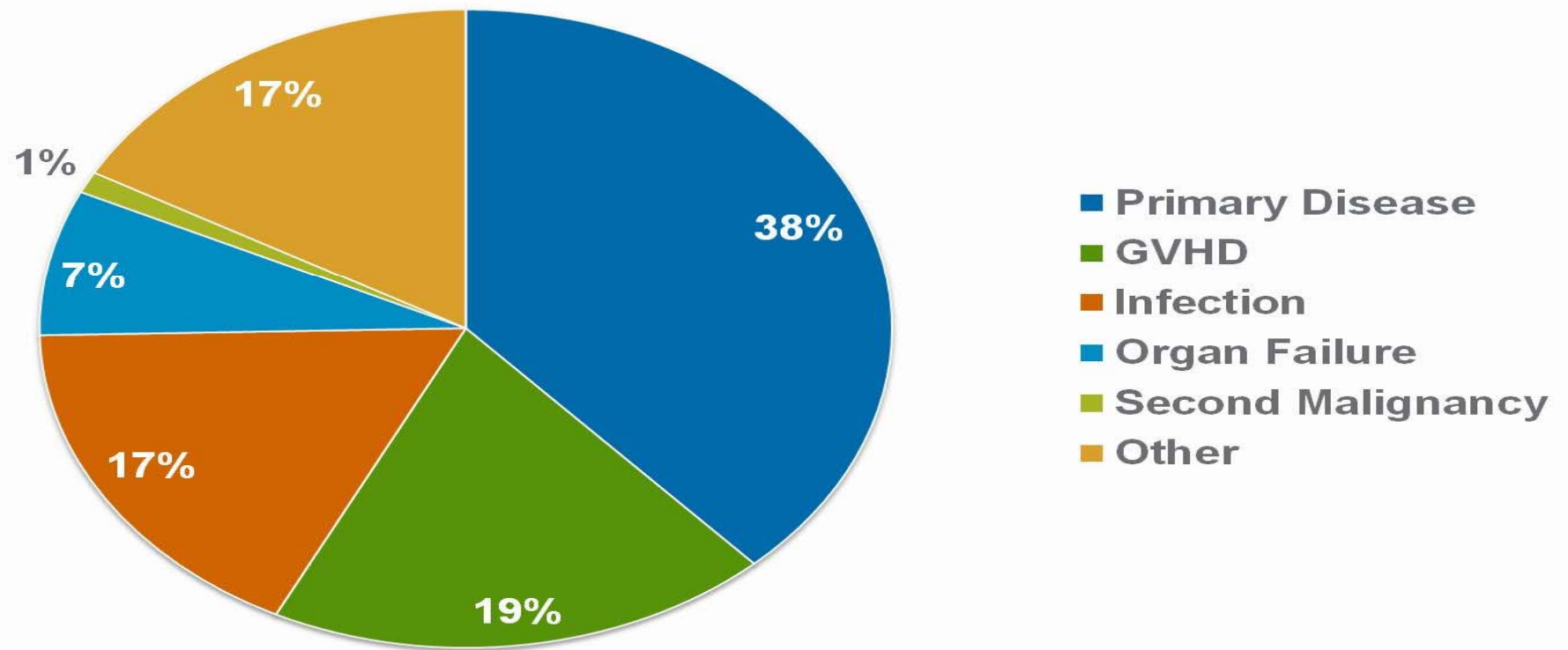


# Causes of Death after Autologous Transplants done in 2010-2011





# Causes of Death after Unrelated Donor Transplants done in 2010-2011

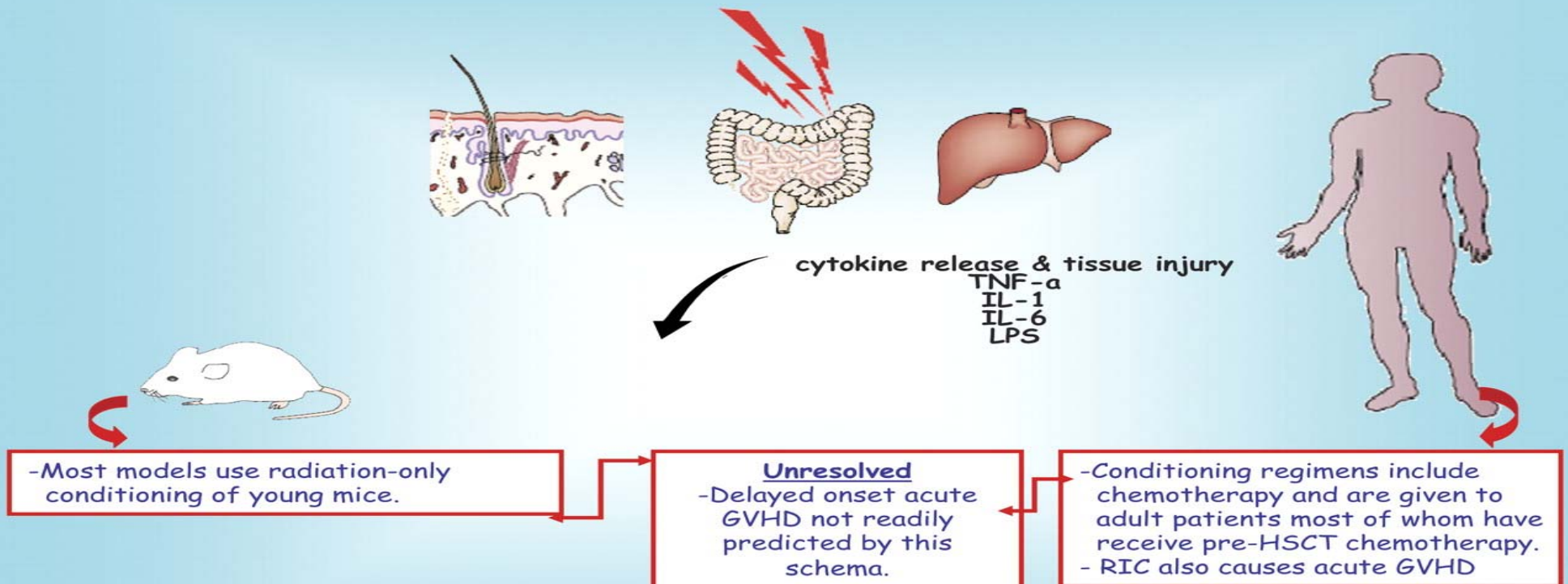


# **Billingham's GVHD Criteria**

- **Genetic disparity between donor and host**
- **Immune deficiency**
- **Immune cells present in donor graft**

# Priming of the immune response

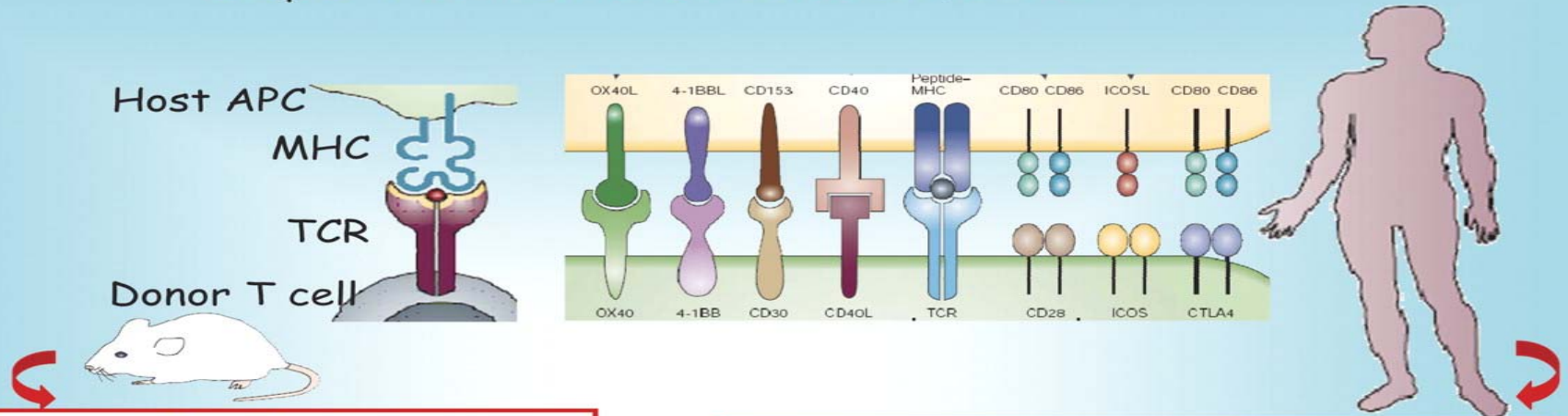
## Step 1: Priming of the immune response



**Socie and Blazar: 2009, BLOOD**

# T-cell activation and costimulation

## Step 2: T-cell activation and co-stimulation



- MiH antigenic disparities that cause acute GVHD have been identified
- Host APCs proven to initiate GVHD
- Critical role of positive and inhibitory co-stimulatory molecules proven

- HLA antigen disparities that cause acute GVHD have been identified. Only a few MiHa antigens that cause GVHD have been identified.
- Host APCs in GVHD not well studied to date
- Role of positive and inhibitory co-stimulatory molecules expression in vivo not well studied

### Issues

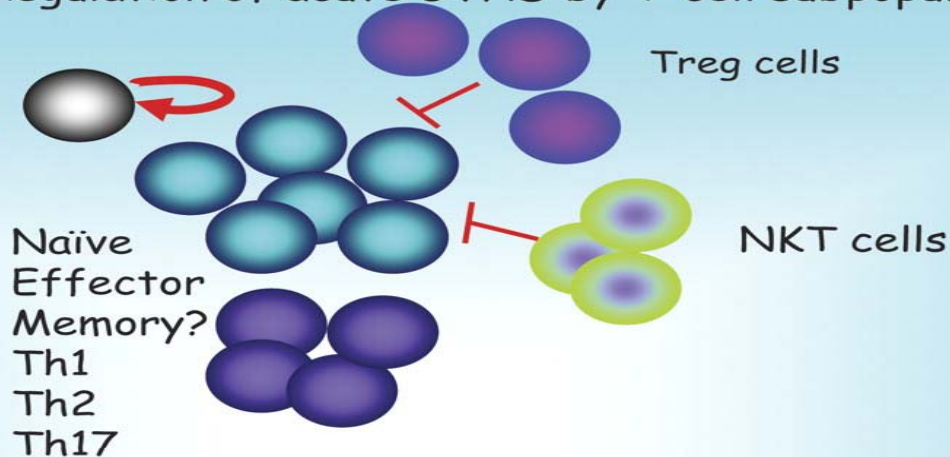
Reagents tested in mice or large animals may have different functional effects due to distinct species-specific expression patterns or targeted epitopes (e.g. cytokine syndrome see with anti-CD28 Ab in humans but not large or small animals)



# Regulation of acute GVHD by T-cell subsets

## Step 3: Regulation of acute GVHD by T-cell subpopulations

Stemness of  
GVHD-inducing  
T-cell?



Naïve  
Effector  
Memory?  
Th1  
Th2  
Th17

NKT cells

- Critical role of naïve T-cells
- CD4+ or CD8+ subsets sufficient for GVHD generation in some models
- Stemness of T-cell inducing GVHD identified
- Th1 vs. Th2 vs Th17 cells known to cause organ specific GVHD effects

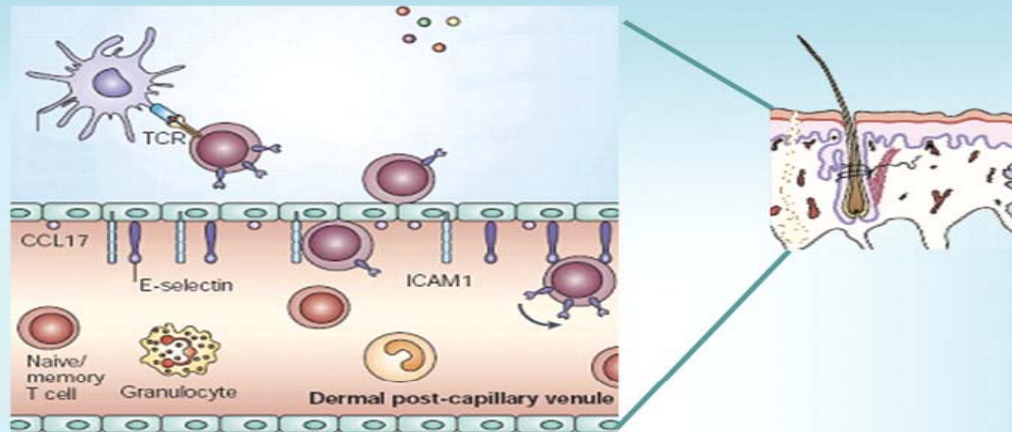
- Naïve vs. memory T-cells?
- Role of Treg, NKT and Th17 cells only partially established

### Issues

- Influence of distinct memory cell repertoire differences between species as a result of specific environmental, pathogen or antigenic exposure that results in different proportions of non-alloreactive vs alloreactive memory T-cells
- Role of differential expression of antigens on activated or suppressor T-cells (e.g. HLA-DR expression CD4+ T cells and CD28 expression on CD8+ suppressor T-cells in humans vs. mice)

# T-cell trafficking

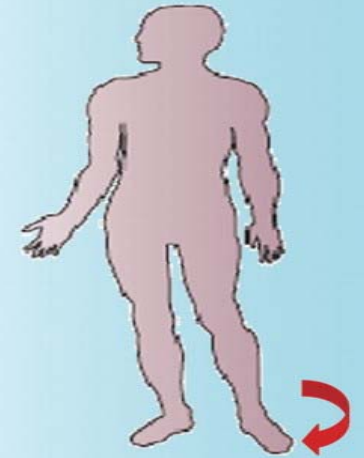
## Step 4: T-cell trafficking



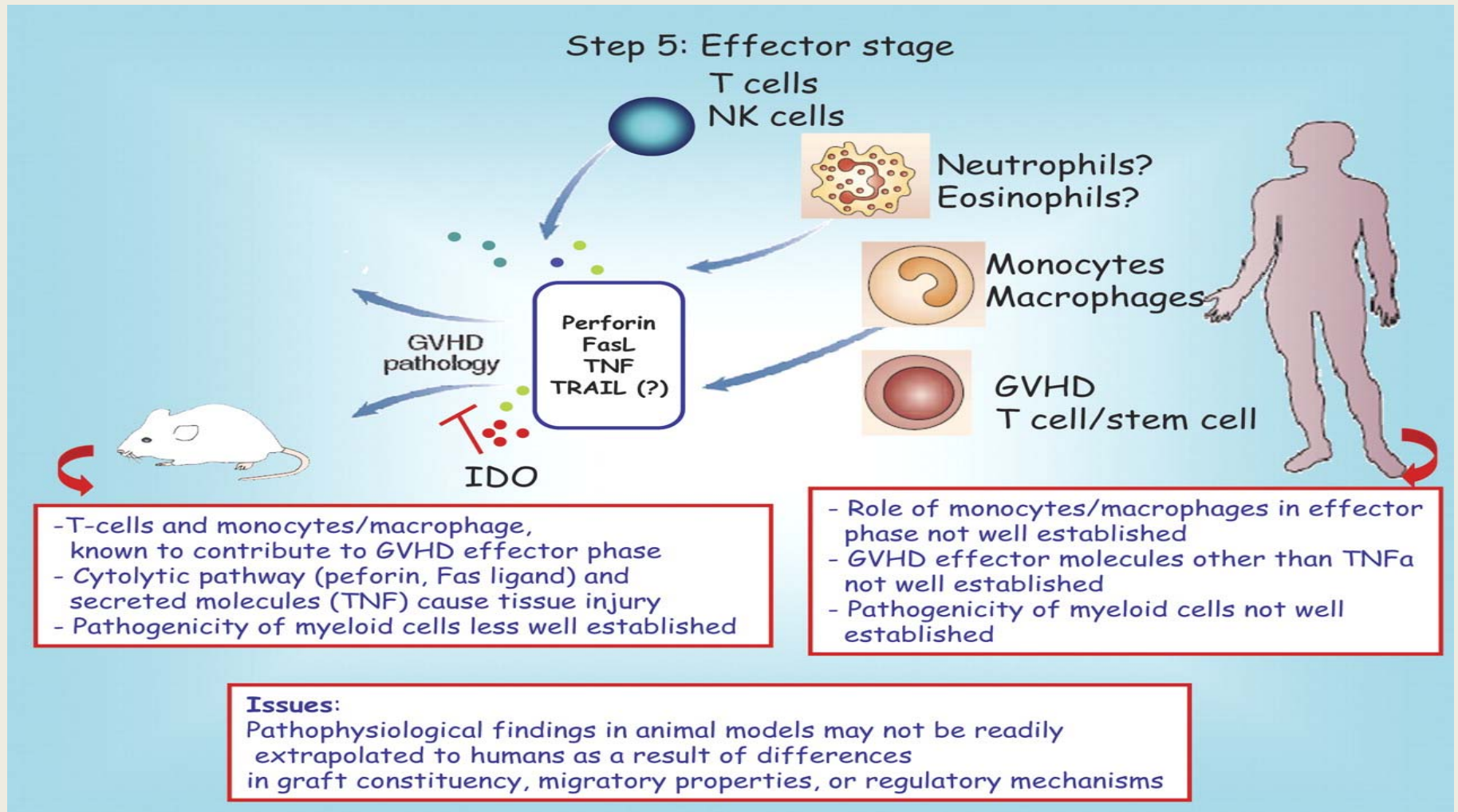
- Secondary lymphoid organs facilitate GVHD initiation.
- Parenchymal organs amplify GVHD
- Homing mechanisms (chemokine, selectin and adhesion molecules) well established

Issue  
Chemokine/receptor redundancy hinder clinical applications

- GVHD initiation and amplification sites not well established
- Homing mechanisms (chemokines, selectin, adhesion molecules) not well established

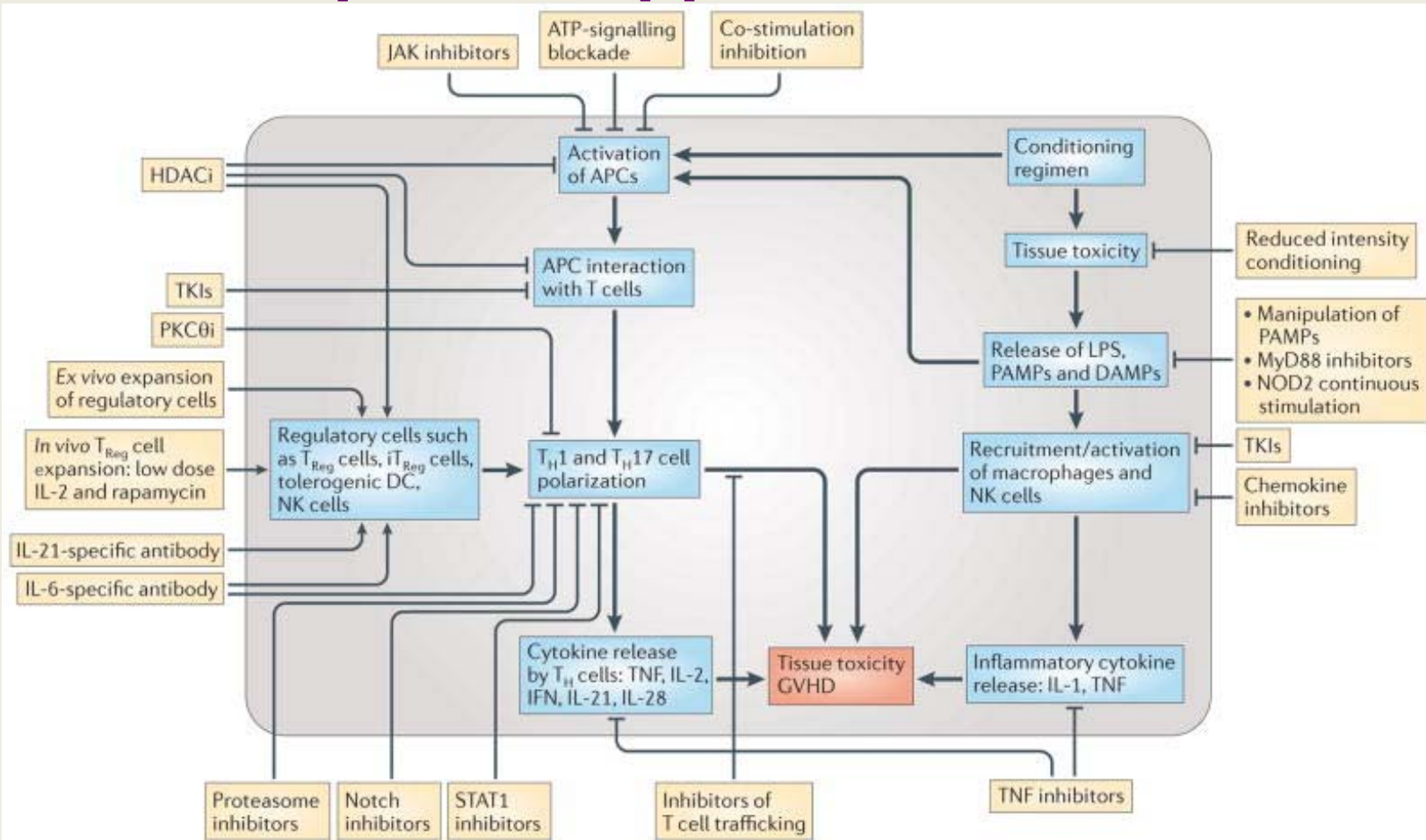


# Effector Phase





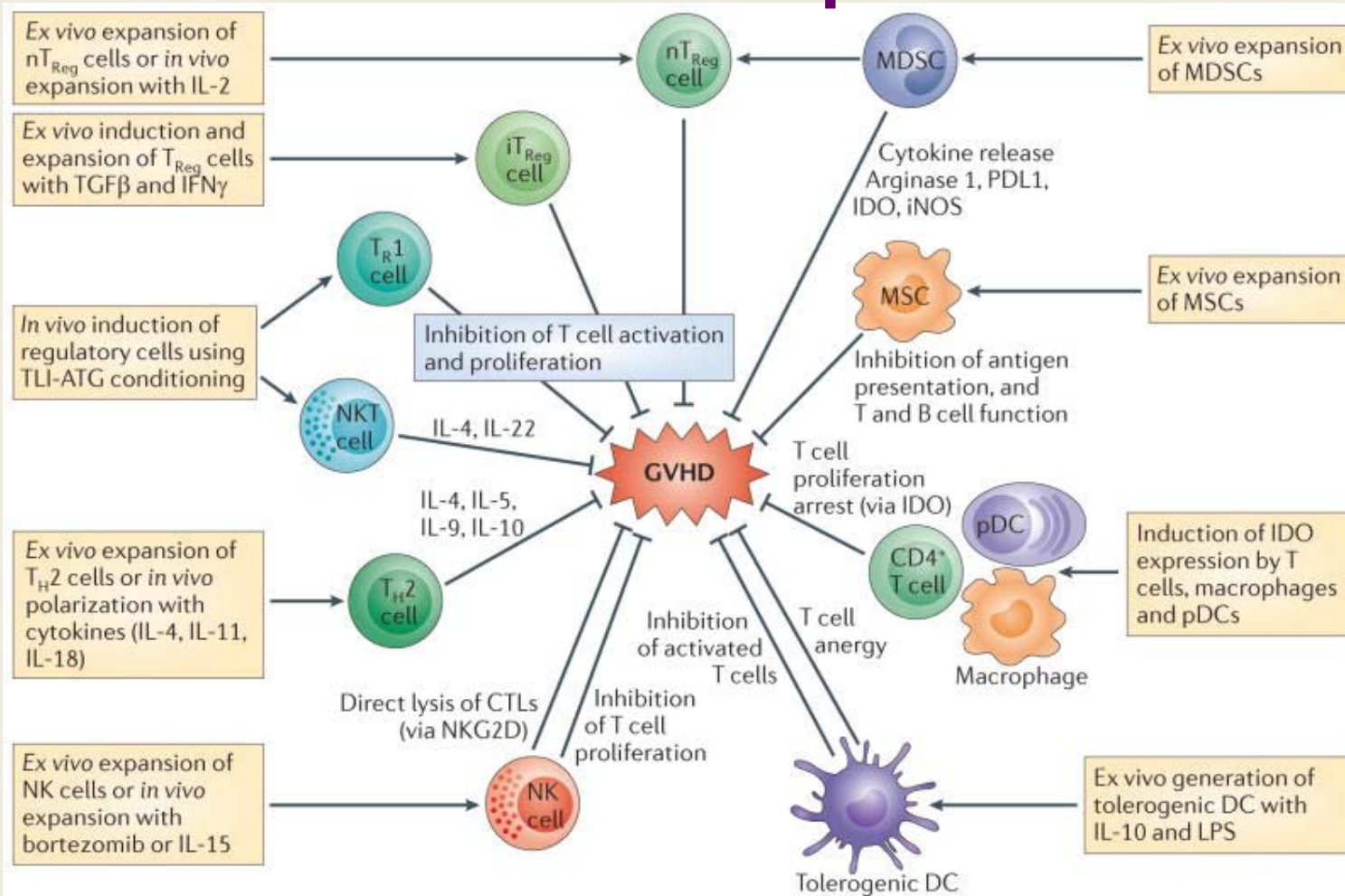
# Therapeutic Approaches for GVHD



Blazar BR et al: 2012, Nat Rev Immunol



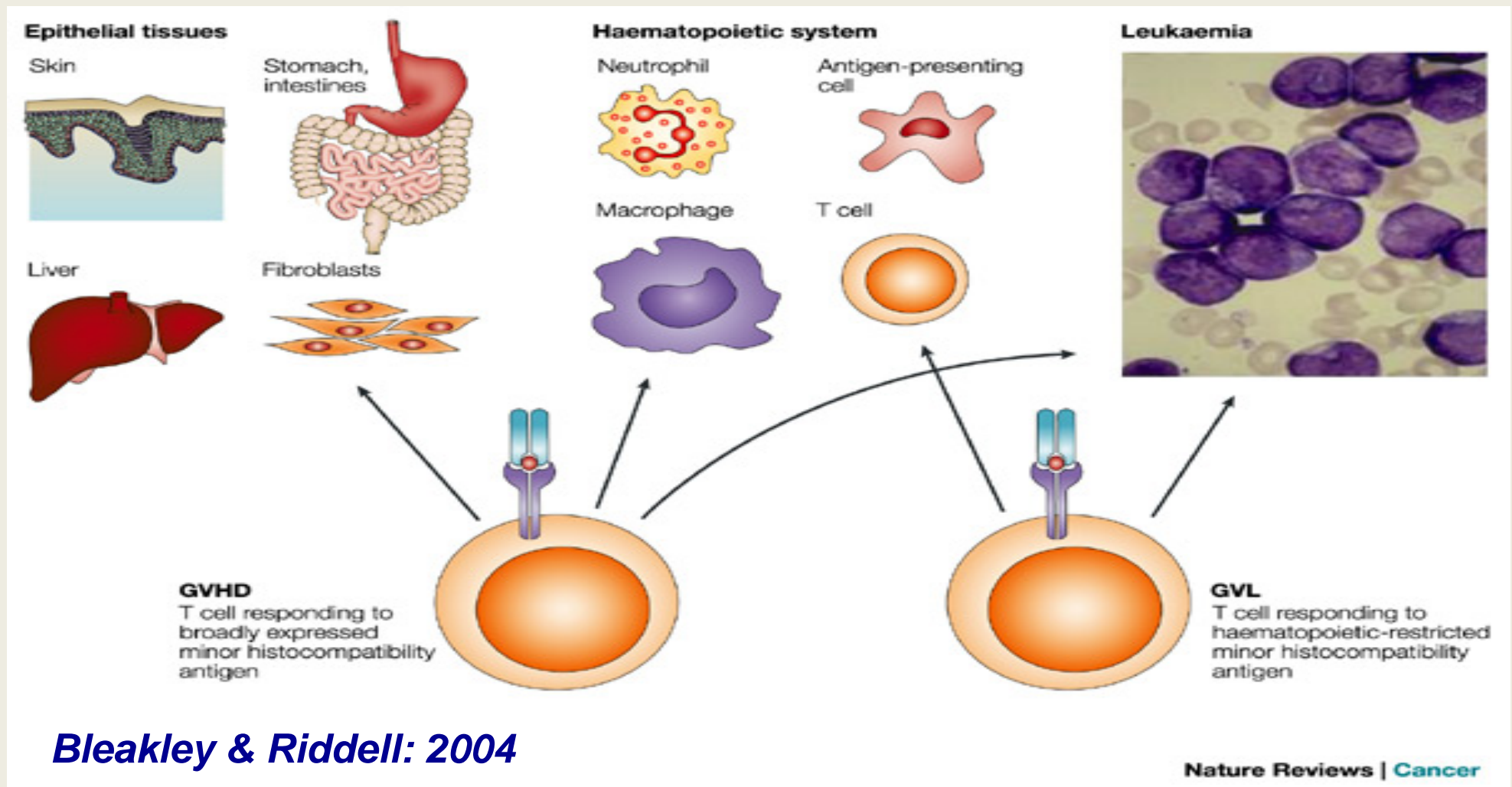
# Cellular Immunotherapies in GVHD



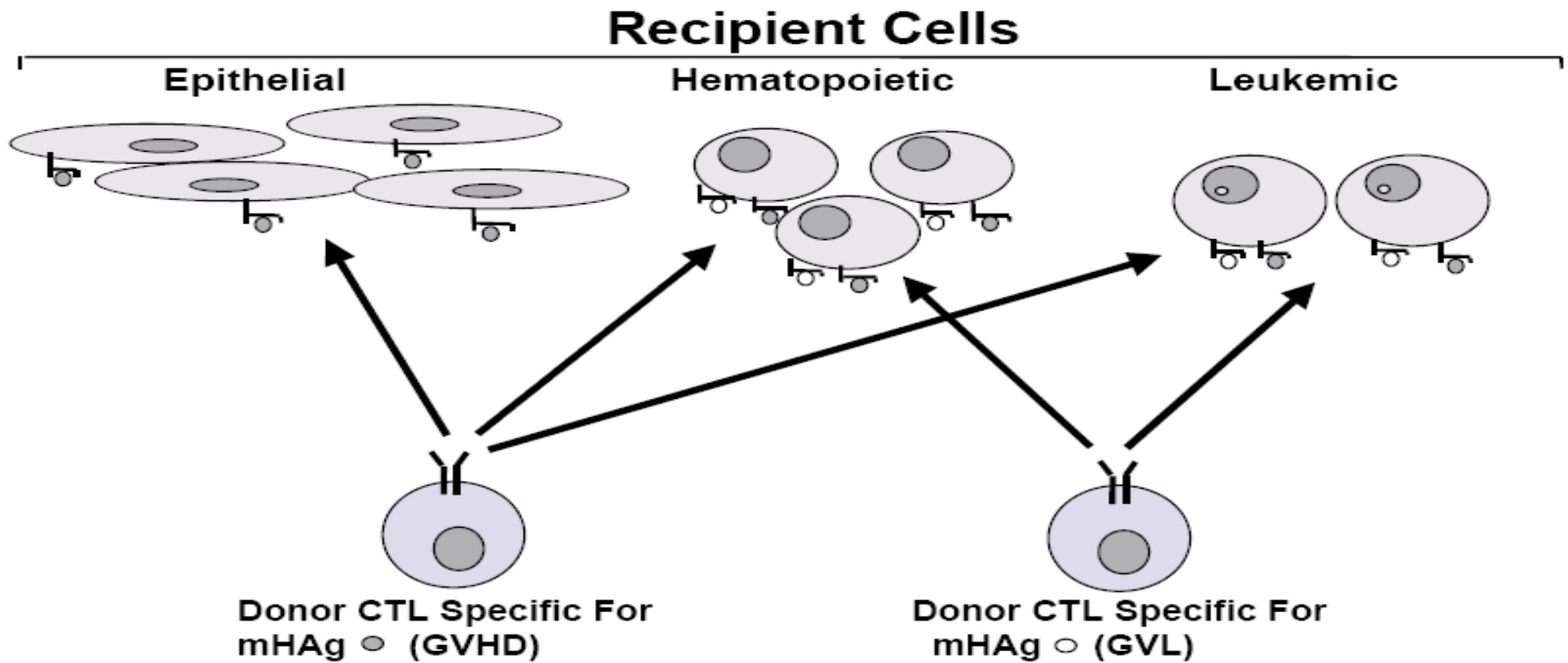
Blazar BR et al: 2012, Nat Rev Immunol

Nature Reviews | Immunology

# Separating GVHD and GVL effects



## Tissue-specific mHAg may separate GVL and GVHD



*Riddell SR et al: Cancer Control, 2002*

# Human mHAgS

Table 1 | **Human minor histocompatibility antigens**

Minor histocompatibility antigen	HLA restriction	Gene/chromosome	Peptide sequence	Tissue distribution	Identification technique	References
HA-1	HLA A201	<i>KIAA0223</i> /19p13	VLHDDLLEA	Haematopoietic	HPLC with mass spectrometry	25
HA-1	HLA B60	<i>KIAA0223</i> /19p13	KECVLHDDL	Haematopoietic	Polymorphic-peptide screening	26
HA-2	HLA A201	<i>MYOG1</i> /7	YIGEVLSV	Haematopoietic	HPLC with mass spectrometry	27,28
HA-3	HLA A1	<i>LBC</i> /15q24-25	VTEPGTAQY	Ubiquitous	HPLC with mass spectrometry	29
HA-8	HLA A201	<i>KIAA0020</i> /9	RTLDKVLEV	Ubiquitous	HPLC with mass spectrometry	30
HB-1	HLA B44	5q32	EEKRGSLHWW	Haematopoietic, especially B-cell leukaemias	cDNA-expression cloning	31,32
UGT2B17	HLA 2902	<i>UGT2B17</i> /4q13	AELLNIPFLY	Ubiquitous	cDNA expression cloning	33
BCL2A1	HLA A24	<i>BCL2A1</i> /15q24.3	DYLQYVKQI	Haematopoietic	Genetic-linkage analysis	34
BCL2A1	HLA B4403	<i>BCL2A1</i> /15q24.3	KEFEDDIINW	Haematopoietic	Genetic-linkage analysis	34
HY B7	HLA B702	<i>SMCY</i>	SPSVDKARAEL	Ubiquitous	HPLC with mass spectrometry	35
HY A2	HLA A201	<i>SMCY</i>	FIDSYICQV	Ubiquitous	HPLC with mass spectrometry	36
HY A1	HLA A101	<i>DFFRY</i>	IVDCLTEMY	Ubiquitous	HPLC with mass spectrometry	37
HY B60	HLA B60	<i>UTY</i>	RESEESVSL	Ubiquitous	cDNA-expression cloning	38
HY B8	HLA B8	<i>UTY</i>	LPHNHTDL	Ubiquitous	cDNA-expression cloning	39
HY DQ5	HLA DQ5	<i>DBY</i>	HIENFSDIDMGE	Ubiquitous	cDNA-expression cloning	40
HY DRB3	HLA DRB3	<i>RPS4Y</i>	VIKVNDTVQI	Not reported	cDNA-expression cloning	41

HLA, human leukocyte antigen; HPLC, high-performance liquid chromatography.

***Bleakley & Riddell: 2004***

# HY-directed responses

**Table 8. Effect of donor/recipient sex on death**

Donor/recipient sex	Adjusted hazard ratio*	95% CI	P
Female/male	1	NA	NA
Male/female	0.82	0.72-0.94	.004
Female/female	0.81	0.72-0.92	.002
Male/male	0.87	0.78-0.97	.01

NA indicates not applicable.

\*Adjusted for patient and donor age, GVHD prophylaxis, disease status, conditioning regimen, and patient/donor CMV serostatus.

***Randolph SB et al: Blood, 2004***

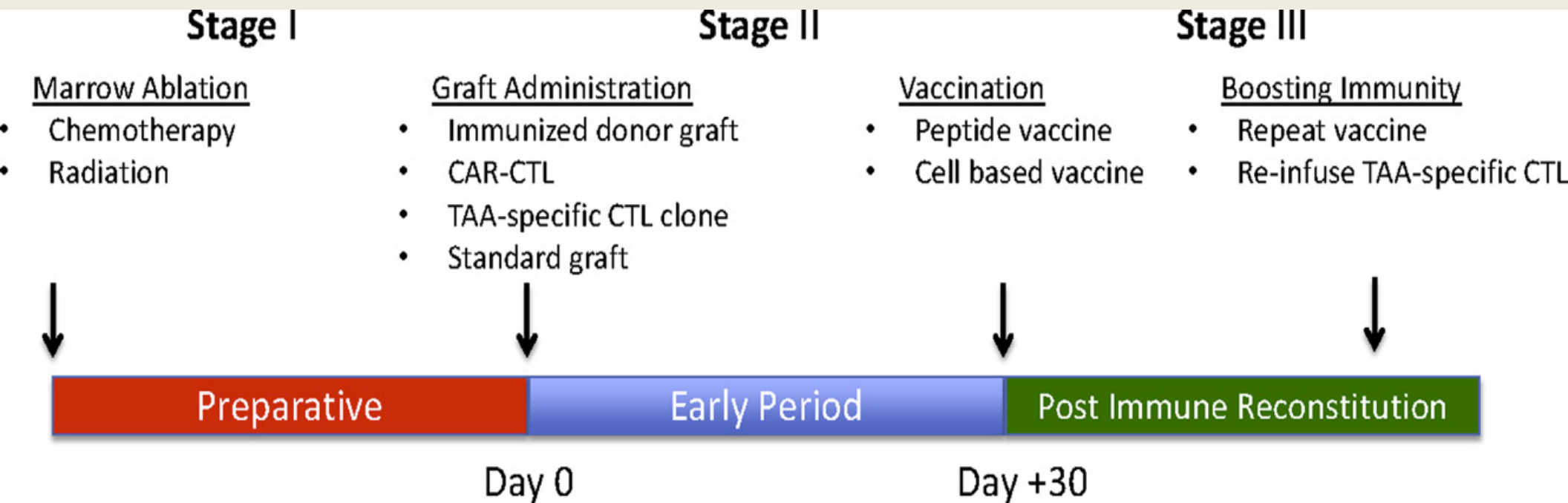


# Leukemia Associated Antigens

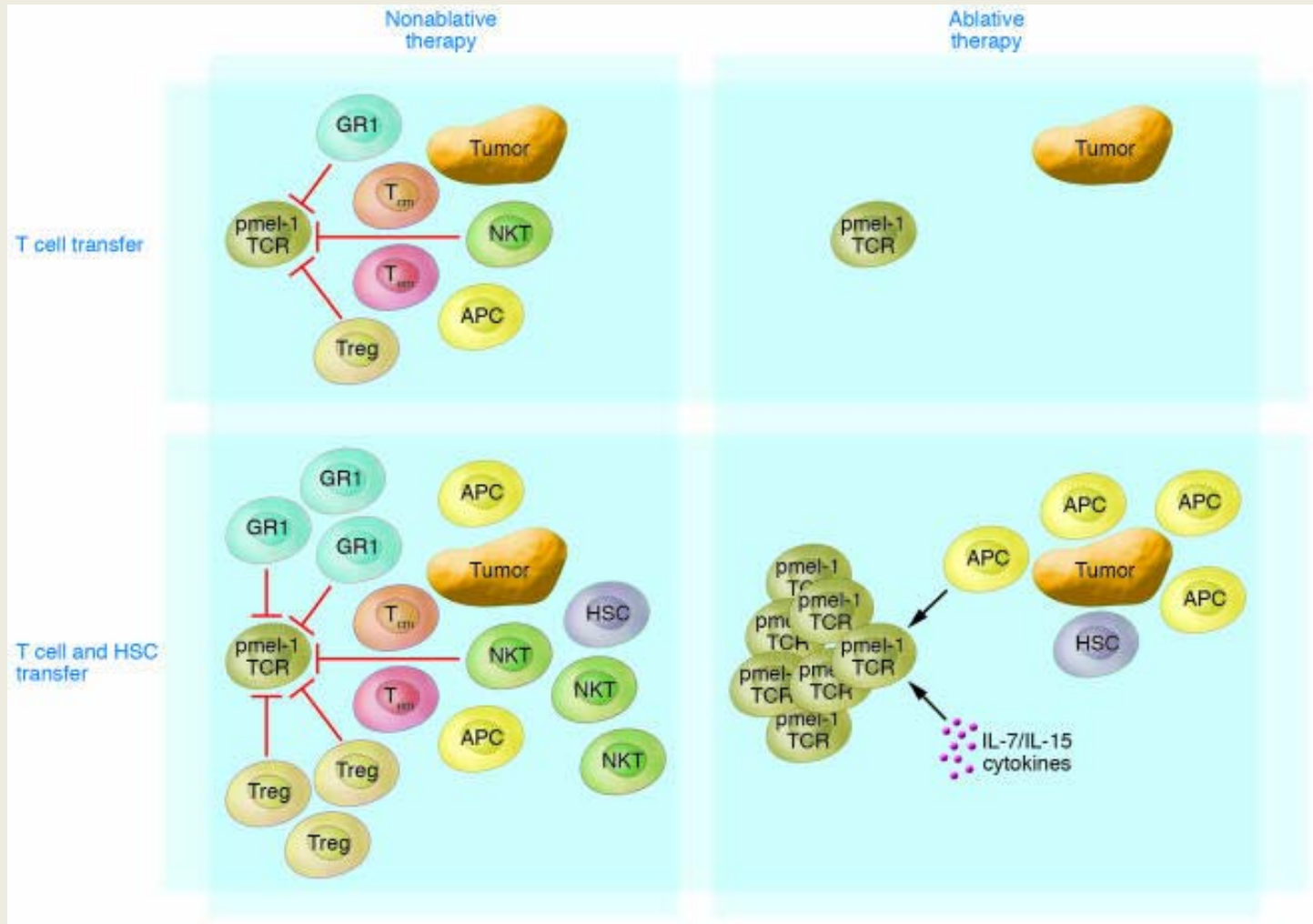
Leukemia antigens				
Class	Example(s)	Pros		Cons
<b>Tumor-associated antigens (TAAs)</b>	WT-1 [25], hTERT [26], PRAME [27], HMMR/Rhamm [28]	- multiple candidates identified- often shared by > 1 malignancy	- present on normal tissue	<b>- self antigens that generate low-avidity T-cells- must be successfully processed and presented by the MHC of the malignant cell</b>
<b>Tumor differentiation antigens</b>	PR1 [29], CG1 [30], CD33 [31]	- more restricted distribution than TAAs	- present on subset of normal cells, which can include hematopoietic stem cells	
<b>Cancer testis (CT) antigens</b>	Cyclin-A1 [32], NY-ESO-1 [33] , MAGE [34]	- frequently restricted to non-essential tissues and tumor	- few identified in leukemia	<b>- must be successfully processed and presented by the MHC</b>
<b>Minor histocompatibility antigens (mHAs)</b>	HA-1 [35], ACC1 [36], T4A [37], LB-LY75-IK [38]	- result in high avidity allo T-cells since epitopes are foreign to donor- some are largely restricted to hematopoietic compartment	- Necessitate rescue with mHA-negative stem cells to restore normal hematopoiesis- need for allogeneic TCRs	
<b>Tumor-specific antigens (neoantigens)</b>	BCR-ABL [39], FLT3-ITD [40], B-cell receptor idiotype [41]	- result in high avidity autologous T-cells- many derive from proteins critical in leukomogenesis	- individual-specific- few identified in leukemia since mutation rate is low	
<b>Oncoviral antigens</b>	HTLV-I Tax protein [42]	- generate very high-avidity T-cells	- only relevant to virus-initiated malignancies	<b>- require CAR for targeting, which can mediate on-target, off-tumor adverse effects</b>
<b>Extracellular antigens</b>	CD19 (see CD19 section), Lewis Y [43], CD22 [44], ROR1 [45]	<b>-MHC-independent-</b> interaction with CAR is high-affinity	- many are present on normal tissues	

*Garber HR et al: Mol & Cell Therapies, 2014:2:25*

# Tumor-associated Antigen-targeting Therapies



# Myeloablation and HSC Enhance Immunotherapy



Anasetti & Mule: *J Clin Invest*, 2007



## **Research in Yu's Lab**

- **Control T-cell differentiation**
- **Modify T-cell costimulation**
- **Target kinases for T- and B-cell activation**
- **Adoptive T-cell therapy: effector and regulatory**
- **Combinational therapy with HSC and T cells for solid tumor, such as melanoma**

## **Take Home Messages**

- **HSCT is a classic cancer immunotherapy through GVL effect**
- **GVHD is a major complication**
- **Separating GVH and GVL response is a major challenge**
- **Anti-tumor effect can be enhanced by targeting mHAg or TAA**
- **Combinational therapy with HSC and T cells increases the efficacy of immunotherapy**

# Questions

**1. Hematopoietic stem cell transplantation (HSCT) is a therapeutic procedure. What is the major application of HSCT?**

- A. Genetic or congenital defects***
- B. Hematological disorders***
- C. Hematological malignances***
- D. Solid tumors***

## Questions

**2. It is a major challenge to suppress GVHD while preserving GVT effect after allogeneic HSCT. Targeting what antigen(s) is promoting GVT effect while limiting GVHD?**

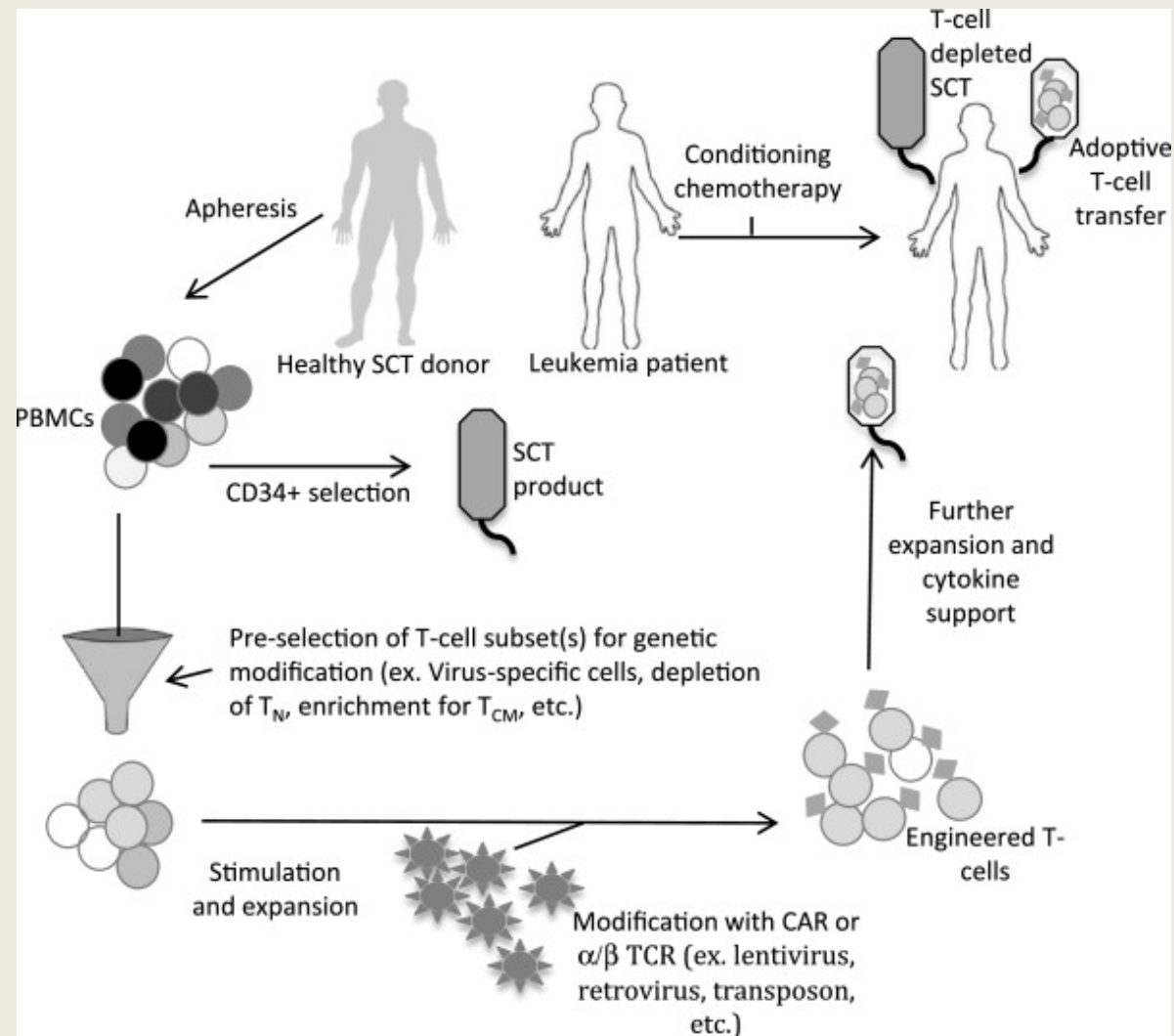
- A. MHC antigens***
- B. Minor histocompatibility antigens (miHA)***
- C. Tumor associated antigens (TAA)***
- D. miHA and TAA***

## Questions

**3. How does HSCT enhance adoptive T-cell immunotherapy against cancer?**

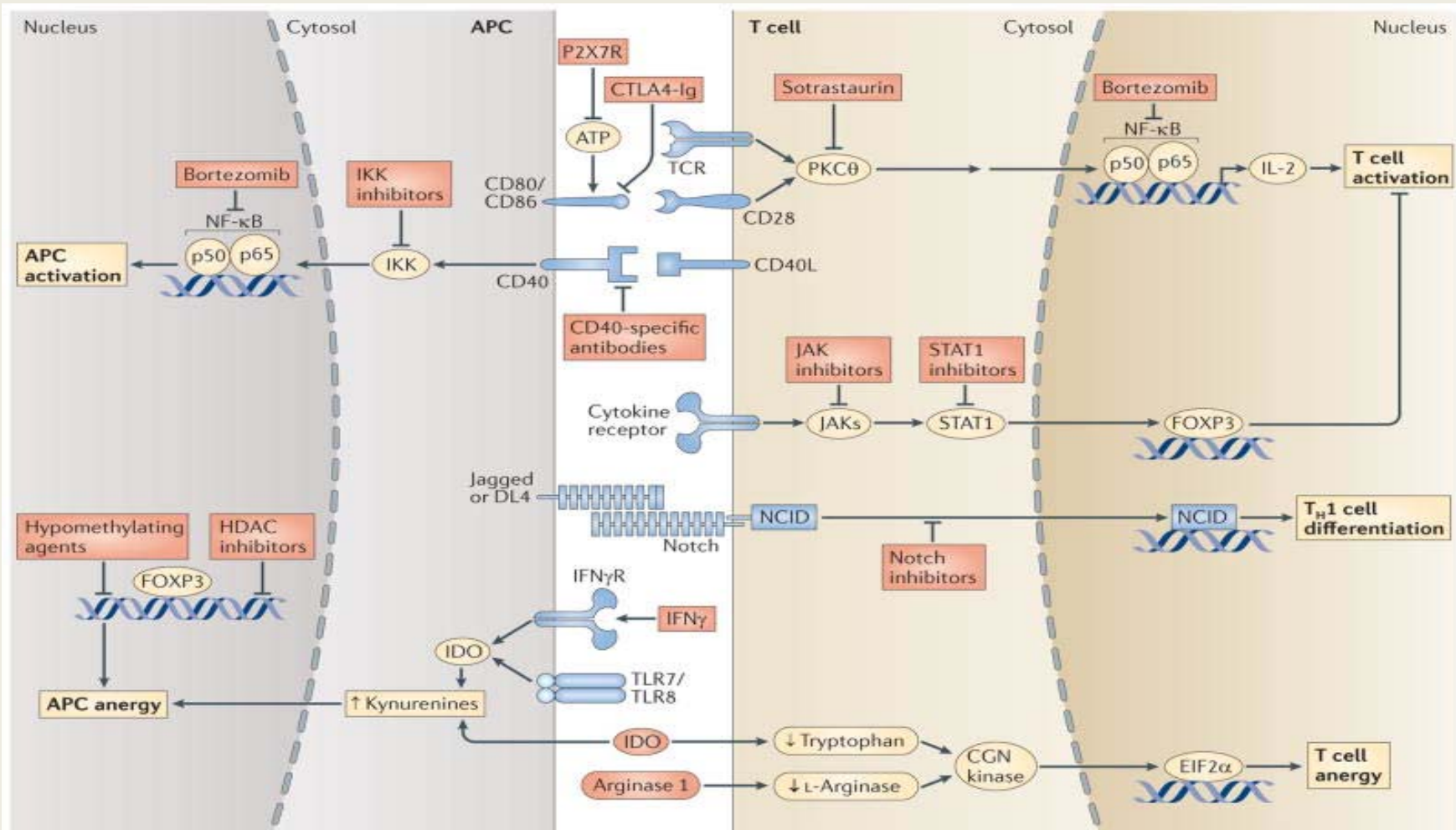
- A. Myeloablative conditioning depletes inhibitory cells, such as Tregs, NKT, and suppressive monocytes***
- B. Enhances cytokine availability due to removing cytokine sink***
- C. HSC produces APCs and T-cell homeostatic cytokines (e.g. IL-7 and IL-15)***
- D. All of above***

# Adoptive Immunotherapy with donor T cells



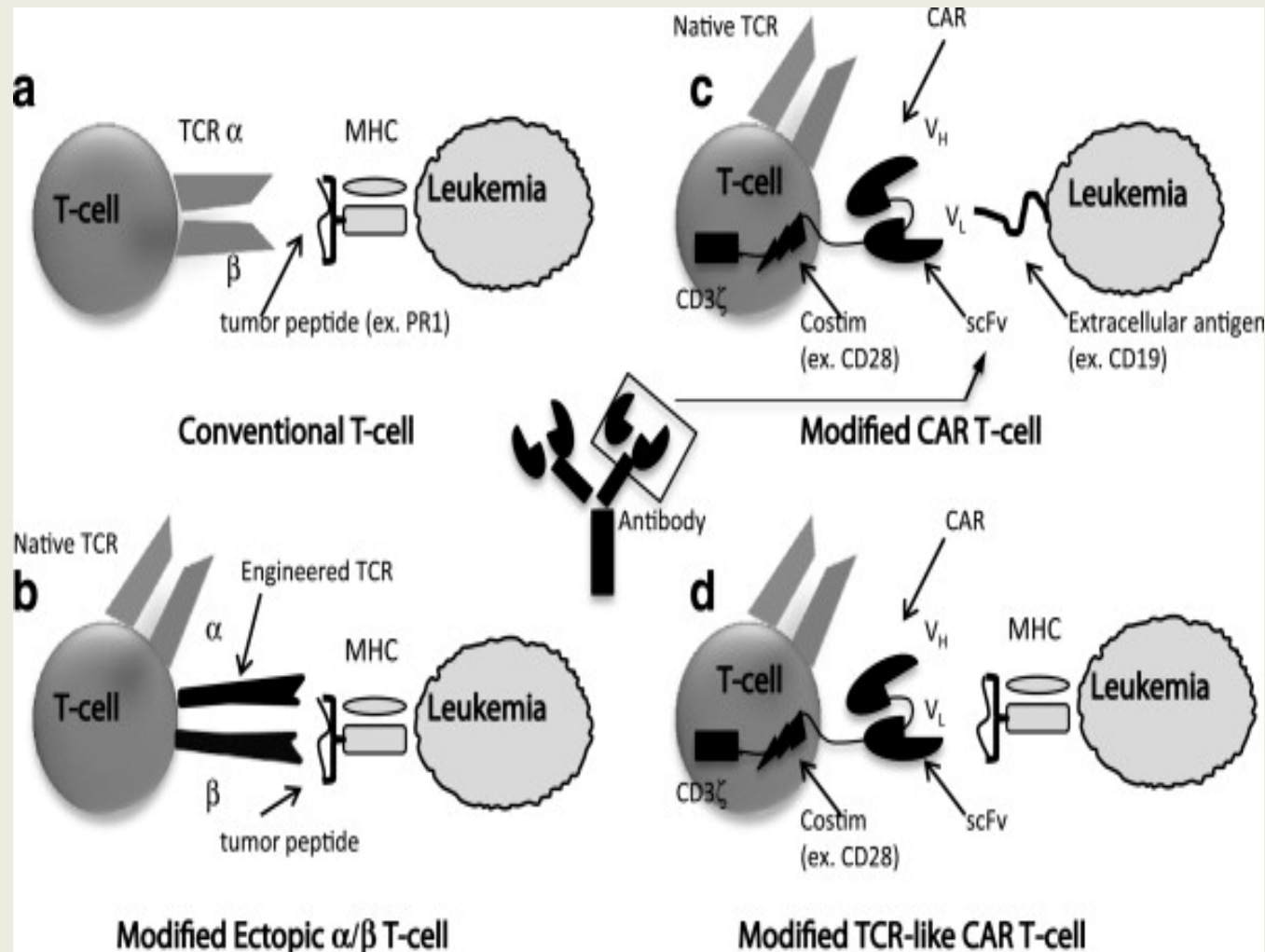
*Garber HR et al:  
Mol & Cell Therapies  
2014;2:25*

# T cell/APC Interaction and Intervention



Blazar BR et al: 2012, Nat Rev Immunol

# T cell Approaches to Target Leukemia Antigens



*Garber HR et al:  
Mol & Cell Therapies  
2014;2:25*



# Stem Cell Sources for Allogeneic Transplants by Year

